

## *LIST OF NEUROSCIENCE TECHNIQUES TO STUDY BRAIN ACTIVITY IN VIVO*

After the LLM Screening, it is necessary to merge **both Step 2 and Step 3** into a single list, again screening for the techniques that fit the established criteria and also looking for duplicates. All the while creating a properly structured list, and also already tagging if they are Invasive [I] or non-invasive [NI], techniques which are minimally invasive (for instance, require only a contrast injection or isotope) are tagged as [I\*].

New additions while performing the merge of both Step 2 and 3 and checking on selected papers were added to the list, but are pointed at the end as additions and where did they come from (tagged as “\$”).

Note: some items are listed due to their importance but are not counted as they can be seen as advancements or specific approaches within the same technique.

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### **1. ELECTROMAGNETIC**

#### **# Microscopic Resolutions**

##### ***Intracellular or Juxtacellular***

1.1. Patch-Clamp [I] (1)

[https://doi.org/10.1007/978-1-0716-0818-0\\_1](https://doi.org/10.1007/978-1-0716-0818-0_1)

<https://doi.org/10.3390/s21041448>

1.2. Sharp Electrode Intracellular Recordings [I] (2)

[https://doi.org/10.1016/S0165-0270\(03\)00126-2](https://doi.org/10.1016/S0165-0270(03)00126-2)

1.3. Juxtacellular Recording and Labeling [I] (3)

<https://doi.org/10.1016/j.jneumeth.2006.02.004>

<https://doi.org/10.1038/nprot.2014.161>

##### ***Extracellular***

1.4. Tungsten Microelectrodes [I] (4)

<https://doi.org/10.3389/fnins.2021.691788>

1.5. Stereotrodes [I] (5)

[https://doi.org/10.1016/0165-0270\(83\)90097-3](https://doi.org/10.1016/0165-0270(83)90097-3)

<https://doi.org/10.1016/j.jneumeth.2013.11.013>

1.6. Tetrodes [I] (6)

[https://doi.org/10.1016/S0165-0270\(02\)00092-4](https://doi.org/10.1016/S0165-0270(02)00092-4)

1.7. Multielectrode Arrays [I]

1.7.1. Michigan Probes [I] (7)

<https://doi.org/10.1109/10.7273>

<https://doi.org/10.1016/j.heares.2008.01.010>

- 1.7.2. Utah Arrays [I] (8)  
[https://doi.org/10.1016/S0013-4694\(96\)95176-0](https://doi.org/10.1016/S0013-4694(96)95176-0)
- 1.7.3. Floating Microelectrode Arrays [I] (9)  
<https://doi.org/10.1016/j.jneumeth.2006.09.005>
- 1.8. Neuropixels [I] (10)  
<https://doi.org/10.1016/j.conb.2018.01.009>
- 1.9. CMOS-Integrated Neural Probes [I] (11)  
<https://doi.org/10.1126/sciadv.adf9524>  
<https://doi.org/10.1016/j.snr.2024.100206>
- 1.10. Fishbone Nanowire Arrays [I] (12)  
<https://doi.org/10.1002/adma.202504171>
- 1.11. Nitrogen-Vacancy Diamond Sensors [I] (13)  
<https://doi.org/10.1073/pnas.1601513113>  
<https://doi.org/10.1038/s41598-023-39539-y>

### # Middle Term Resolutions and Multimodal Approaches

- 1.12. Flexible Bioelectronic Neural Interfaces [I] (14)  
<https://doi.org/10.1038/s41563-020-0679-7>
- 1.13. Optotetrodes [I] (15)  
<https://doi.org/10.1038/nn.2992>
- 1.14. Transparent Graphene Microelectrodes Arrays [I] (16)  
<https://doi.org/10.1038/s41467-018-04457-5>
- 1.15. Electrical Impedance Tomography [NI/I] (17)  
<https://link.springer.com/article/10.1186/s40779-022-00370-7>  
<https://doi.org/10.1016/j.neuroimage.2019.05.023>  
<https://doi.org/10.1006/nimg.2000.0698>

### # Macroscopic Resolutions

- 1.16. Electroencephalography [NI] (18)  
<https://doi.org/10.1038/nrn3241>  
<https://doi.org/10.1016/j.ijpsycho.2015.05.004>
- 1.17. Electrocorticogram [NI] (19)  
<https://doi.org/10.1038/nrn3241>
- 1.16.1. Micro Electrocorticogram [I] (20)  
<https://doi.org/10.1038/s41378-023-00597-x>
- 1.18. Magnetoencephalography [NI]  
<https://doi.org/10.1038/nrn3241>  
<https://doi.org/10.3390/brainsci12060788>
- 1.17.1 SQUID (the typical one) [NI] (21)  
    1.17.2. Optically Pumped Magnetometer MEG (OPM-MEG) [NI] (22)  
<https://doi.org/10.1038/s41398-024-03047-y>
- 1.19. Stentrodes [I] (23)  
<https://doi.org/10.1038/nbt.3428>  
<https://doi.org/10.1016/j.jneumeth.2025.110471>  
<https://doi.org/10.1126/science.adh3916>
- 1.20. Stereoelectroencephalography [I] (24)  
<https://doi.org/10.1097/wnp.0000000000000249>  
<https://doi.org/10.1016/j.seizure.2019.01.021>

- 1.21. Deep Brain Stimulation [I] (25)  
<https://doi.org/10.3389/fpsyg.2023.1080260>  
<https://doi.org/10.1016/j.neurom.2022.01.017>
- 1.22. Neural Dust [I] (26)  
<https://doi.org/10.1016/j.conb.2017.12.010>  
<https://doi.org/10.3389/fnins.2021.599549>

## 2. MOLECULES SENSING

### # Microdialysis and Push-Pull Perfusion

- 2.1. Microdialysis [I] (27)  
<https://doi.org/10.1021/acsami.2c02740>  
<https://doi.org/10.1039/C4AN01974A>
- 2.2. Push-pull Perfusion [I] (28)  
<https://doi.org/10.1021/acsami.2c02740>  
<https://pubs.acs.org/doi/10.1021/acs.analchem.8b02468>
- Note:** both 2.1. and 2.2. can be associated with the following techniques
- 2.3. + High Performance Liquid Chromatography [I] (29)  
<https://doi.org/10.1021/ac4023605>
- 2.4. + Column Liquid Chromatography [I] (30)  
<https://doi.org/10.1021/acsami.2c02740>
- 2.5. + Capillary Electrophoresis [I] (31)  
<https://doi.org/10.1016/j.jneumeth.2004.03.025>
- 2.6. + Mass Spectrometry [I] (32)  
<https://doi.org/10.1016/j.jpba.2005.07.025>  
<https://doi.org/10.1208/s12248-017-0114-4>
- 2.7. + Electrochemical Workstation [I] (33)  
<https://doi.org/10.1021/acs.nanolett.2c00289>

### # Carbon Fiber Microelectrodes

- 2.8. Voltammetry [I] (34)  
<https://doi.org/10.1021/acsami.2c02740>  
<https://doi.org/10.1016/j.neubiorev.2005.02.003>  
<https://doi.org/10.1523/JNEUROSCI.14-01-00442.1994>
- 2.8.1. Differential Pulse Voltammetry
- 2.8.2. Cyclic Voltammetry
- 2.8.3. Fast-Scan Cyclic Voltammetry  
[https://doi.org/10.1016/0306-4522\(88\)90255-2](https://doi.org/10.1016/0306-4522(88)90255-2)  
<https://doi.org/10.1039/C9AN01925A>
- 2.9. Amperometry [I] (35)  
<https://doi.org/10.1021/acsami.2c02740>
- 2.9.1. High-Speed Chronoamperometry  
<https://doi.org/10.3390/cells11152454>
- 2.10. Liquid-Liquid Interface Microsensors [I] (36)  
<https://doi.org/10.1021/acssensors.1c00978>  
<https://doi.org/10.1021/ac504151e>

### # Indirect Sensing

- 2.11. Enzyme-Based sensors [I] (37)  
<https://doi.org/10.1111/j.1471-4159.2006.03673.x>  
<https://doi.org/10.1021/ac403250w>
- 2.12. Aptamer-based sensors [I] (38)  
<https://doi.org/10.1021/acspctsci.4c00579>
- 2.13. Molecularly Imprinted Polymers Sensors [I] (39)  
<https://doi.org/10.1016/j.snb.2011.07.052>  
<https://doi.org/10.1016/j.trac.2018.08.002>

#### # Other sensing techniques

- 2.14. Electron Paramagnetic Resonance (EPR) Oximetry [I] (40)  
<https://doi.org/10.1007/s12013-017-0798-1>  
<https://doi.org/10.4103/2045-9912.202911>
- 2.15. Hydrogen Clearance CBF Measurement [I] (41)  
<https://doi.org/10.1038/jcbfm.1986.83>

### 3. POSITRON EMISSION TOMOGRAPHY

- 3.1. Total-Body PET Imaging [I\*] (42)  
<https://doi.org/10.1259/bjr.20220357>  
<https://doi.org/10.2967/jnumed.119.231845>  
<https://doi.org/10.1007/s00259-022-06057-4>
- 3.2. Metabolic Tracing with Short-Lived Isotopes (Dynamic PET) [I\*] (43)  
<https://doi.org/10.1002/acn3.51546R>  
<https://doi.org/10.1007/s00259-023-06542-4>
- 3.3.  $\mu$ -opioid Receptor PET ( $[^{11}\text{C}]$ carfentanil) [I\*] (44)  
<https://doi.org/10.1007/s00259-024-06746-2>
- 3.4. Dopamine D2/D3 Receptor Occupancy PET [I\*] (45)  
<https://doi.org/10.1038/s42003-022-03434-5>  
<https://doi.org/10.1177/0271678X231210128>
- 3.5. Cerebral Metabolic Rate of Oxygen (CMRO<sub>2</sub>) PET [I\*] (46)  
<https://doi.org/10.1016/j.neuroimage.2020.117136>  
<https://doi.org/10.2967/jnumed.120.260521>
- 3.6. GABA-A Receptor PET ( $[^{18}\text{F}]$ flumazenil) [I\*] (47)  
<https://doi.org/10.3390/ph16030417>
- 3.7. Serotonin Transporter PET ( $[^{11}\text{C}]$ DASB) [I\*] (48)  
<https://doi.org/10.3390/ijms26010252>
- 3.8. Oxygen-15 Water PET ( $[^{15}\text{O}]$ H<sub>2</sub>O PET) [I\*] (49)  
<https://doi.org/10.1053/j.semnuclmed.2023.08.002>  
<https://doi.org/10.1016/j.neuroimage.2013.11.044>

### 4. NON-FLUORESCENCE OPTICAL TECHNIQUES

#### # Infrared

- 4.1. Thermal Infrared Imaging / Thermoencephaloscopy [NI] (50)  
<https://doi.org/10.1038/srep17471>  
[https://doi.org/10.1016/S0301-0082\(98\)00038-0](https://doi.org/10.1016/S0301-0082(98)00038-0)  
<https://doi.org/10.1111/psyp.12243>
- 4.2. Functional Near-Infrared Spectroscopy [NI] (51)

- <https://doi.org/10.1111/nyas.13948>
- 4.2.1. Time-Domain Near-Infrared Spectroscopy (TD-NIRS) [NI] (52)
- <https://doi.org/10.1038/s41598-024-68555-9>
- <https://doi.org/10.1117/1.JBO.27.7.074710> \*
- <https://doi.org/10.1117/1.NPh.10.1.013504> \*
- 4.2.2. Functional Diffuse Optical Tomography (fDOT) [NI] (53)
- <https://doi.org/10.3390/s25072040> \*
- <https://doi.org/10.1038/s41598-025-85858-7>
- <https://doi.org/10.1117/1.NPh.6.3.035007>
- 4.3. Diffuse Correlation Spectroscopy [NI] (54)
- <https://doi.org/10.1016/j.neuroimage.2013.06.017>
- <https://doi.org/10.1007/s12028-018-0573-1>

## # Intrinsic Signals

- 4.4. Optical Intrinsic Signal [I] (55)
- <https://doi.org/10.1117/1.NPh.10.2.020601>
- <https://doi.org/10.1016/j.xpro.2021.100779> \*
- <https://doi.org/10.1038/324361a0>
- <https://doi.org/10.1146/annurev.ne.08.030185.001403>
- 4.4.1. Hyperspectral Imaging of Intrinsic Signals (56)
- <https://doi.org/10.1117/1.NPh.2.4.045003>
- [https://doi.org/10.1007/978-3-319-91287-5\\_3](https://doi.org/10.1007/978-3-319-91287-5_3)
- 4.5. Laser Speckle Contrast Imaging [I] (57)
- <https://doi.org/10.1117/1.NPh.10.2.020601>
- <https://doi.org/10.1097/00004647-200103000-00002>
- 4.6. Spatial Frequency Domain Imaging [I] (58)
- <https://doi.org/10.1117/1.NPh.10.2.020601>
- <https://doi.org/10.1117/1.3088140>
- <https://doi.org/10.1117/1.NPh.8.2.025001>
- 4.6.1. Structured Illumination Diffuse Optical Tomography [NI] (59)
- <https://doi.org/10.1117/1.NPh.4.2.021102>
- 4.7. Laser Doppler Flowmetry [I] (60)
- <https://doi.org/10.1111/micc.12884>
- <https://doi.org/10.1002/lpor.202401016> \*
- <https://doi.org/10.1007/s12028-017-0472-x>
- 4.8. Optical Coherence Tomography [I/NI] (61)
- <https://doi.org/10.1126/science.1957169>
- <https://doi.org/10.1016/j.jneumeth.2008.11.026>
- 4.8.1. Speckle-Modulated Optical Coherence Tomography (OCT) [I] (62)
- <https://doi.org/10.1038/ncomms15845>
- <https://doi.org/10.1063/5.0278271>
- <https://doi.org/10.1093/cercor/bhac388>
- 4.8.2. Doppler Optical Coherence Tomography (D-OCT) [I] (63)
- <https://doi.org/10.1364/BOE.5.003217>
- <https://doi.org/10.1021/acsphotonics.4c00856>
- 4.8.3. Visible Light Optical Coherence Tomography (vis-OCT) [I] (64)

<https://doi.org/10.1364/BOE.6.003941>  
<https://doi.org/10.1364/BOE.6.001429>  
<https://doi.org/10.1117/1.JBO.22.12.121707>

## 5. ULTRASOUND AND PHOTOACOUSTICS

### # Ultrasound (Purely)

5.1. Transcranial Doppler [NI] (65)

<https://doi.org/10.33549/physiolres.935413>

<https://doi.org/10.1186/s41984-021-00114-0>

<https://doi.org/10.1159/000103113>

5.2. Functional Ultrasound (fUS) Imaging [NI/I] (66)

<https://doi.org/10.1016/j.cobme.2021.100286>

<https://doi.org/10.1146/annurev-neuro-111020-100706>

<https://doi.org/10.1016/j.neuroscience.2021.03.005>

<https://doi.org/10.1038/nmeth.1641>

5.2.1. Microbubble-Enhanced fUS (Contrast-Enhanced fUS) [I] (67)

<https://doi.org/10.1016/j.neuroimage.2015.09.037>

<https://doi.org/10.1016/j.cobme.2021.100286>

5.2.2. Functional Ultrasound Microscopy (68)

<https://doi.org/10.1038/s41592-022-01549-5>

### # Photoacoustic

5.3. Photoacoustic Imaging (PAI) [I/NI] (69)

<https://doi.org/10.1016/j.pacs.2019.05.001>

<https://doi.org/10.1063/1.2195024>

<https://doi.org/10.3109/13813450312331337649>

5.3.1. Photoacoustic Computed Tomography (PACT) [I/NI] (70)

<https://doi.org/10.1038/nbt839>

<https://doi.org/10.1002/jbio.201700024>

<https://doi.org/10.1364/BOE.423707>

5.3.2. Functional Photoacoustic Microscopy (fPAM) [I] (71)

<https://doi.org/10.1038/s41377-022-00836-2>

<https://doi.org/10.1016/j.neuroimage.2021.118260>

5.4 Voltage-Sensitive Photoacoustic Imaging [I] (72)

<https://doi.org/10.1002/lpor.202400165>

## 6. MAGNETIC RESONANCE IMAGING

### # Hemodynamic

6.1. Blood Oxygen Dependent Levels (BOLD) [NI] (73)

<https://doi.org/10.1073/pnas.87.24.9868>

<https://doi.org/10.1038/jcbfm.2012.23>

<https://doi.org/10.1016/j.tins.2024.12.010>

<https://doi.org/10.1002/nbm.70076>

<https://doi.org/10.1523/JNEUROSCI.23-10-03963.2003>

6.1.1. Functional Connectivity MRI (fcMRI) [NI]

<a href="https://doi.org/10.1016/j.neuroimage.2023.120249">https://doi.org/10.1016/j.neuroimage.2023.120249</a>	
<a href="https://doi.org/10.1038/s42003-023-05629-w">https://doi.org/10.1038/s42003-023-05629-w</a>	
6.1.2. Resting-State fMRI (rs-fMRI) [NI]	
<a href="https://doi.org/10.1002/jmri.28894">https://doi.org/10.1002/jmri.28894</a>	
6.2. Arterial Spin Labeling [NI]	(74)
<a href="https://doi.org/10.1002/mrm.1910230106">https://doi.org/10.1002/mrm.1910230106</a> (first)	
<a href="https://doi.org/10.1002/mrm.29381">https://doi.org/10.1002/mrm.29381</a> *	
<a href="https://doi.org/10.1073/pnas.89.1.212">https://doi.org/10.1073/pnas.89.1.212</a>	
6.2.1. Continuous Arterial Spin Labeling (CASL) [NI]	
<a href="https://doi.org/10.3389/fradi.2022.929533">https://doi.org/10.3389/fradi.2022.929533</a>	
6.2.2. Pulsed Arterial Spin Labeling (PASL) [NI]	
<a href="https://doi.org/10.3389/fradi.2022.929533">https://doi.org/10.3389/fradi.2022.929533</a>	
6.3. Vascular-Space-Occupancy (VASO) [NI]	(75)
<a href="https://doi.org/10.1002/mrm.10519">https://doi.org/10.1002/mrm.10519</a>	
<a href="https://doi.org/10.1016/j.neuroimage.2021.118868">https://doi.org/10.1016/j.neuroimage.2021.118868</a>	
6.3.1. VASO and Perfusion (VAPER) [NI]	(76)
<a href="https://doi.org/10.1016/j.neuroimage.2019.116358">https://doi.org/10.1016/j.neuroimage.2019.116358</a>	
<a href="https://doi.org/10.1162/imag_a_00140">https://doi.org/10.1162/imag_a_00140</a>	
6.4. Laminar fMRI [NI]	
<a href="https://doi.org/10.1016/j.neuroimage.2017.02.063">https://doi.org/10.1016/j.neuroimage.2017.02.063</a>	
<a href="https://doi.org/10.1093/psyrad/kkae027">https://doi.org/10.1093/psyrad/kkae027</a>	
<a href="https://doi.org/10.1016/j.cobme.2021.100288">https://doi.org/10.1016/j.cobme.2021.100288</a>	
<a href="https://doi.org/10.1016/j.neuroimage.2017.07.004">https://doi.org/10.1016/j.neuroimage.2017.07.004</a>	
6.5. Exogenous Labeling [I]	
<a href="https://doi.org/10.3348/kjr.2014.15.5.554">https://doi.org/10.3348/kjr.2014.15.5.554</a>	
6.5.1. Dynamic Susceptibility Contrast (DSC)-MRI [NI/I]	(77)
<a href="https://doi.org/10.1038/jcbfm.2010.4">https://doi.org/10.1038/jcbfm.2010.4</a>	
<a href="https://doi.org/10.1007/s10334-009-0190-2">https://doi.org/10.1007/s10334-009-0190-2</a>	
<a href="https://doi.org/10.1038/s41598-024-58086-8">https://doi.org/10.1038/s41598-024-58086-8</a> [NI]	
6.5.2. Dynamic Contrast-Enhanced (DCE)-MR [I]	(78)
<a href="http://www.ajnr.org/content/27/7/1467">http://www.ajnr.org/content/27/7/1467</a>	
<a href="https://doi.org/10.2174/1874440001105010090">https://doi.org/10.2174/1874440001105010090</a>	
<a href="https://doi.org/10.7554/eLife.89611.4">https://doi.org/10.7554/eLife.89611.4</a>	
6.6. Susceptibility-Weighted Imaging (SWI) [NI]	(79)
<a href="https://doi.org/10.1016/j.neuroimage.2012.01.020">https://doi.org/10.1016/j.neuroimage.2012.01.020</a>	
<a href="https://doi.org/10.1016/j.neuroimage.2007.11.046">https://doi.org/10.1016/j.neuroimage.2007.11.046</a>	
6.7. Intravoxel Incoherent Motion (IVIM) MRI [NI]	(80)
<a href="https://doi.org/10.1371/journal.pone.0117706">https://doi.org/10.1371/journal.pone.0117706</a>	
<a href="https://doi.org/10.1002/nbm.3780">https://doi.org/10.1002/nbm.3780</a>	
<a href="https://doi.org/10.1016/j.neuroimage.2017.12.062">https://doi.org/10.1016/j.neuroimage.2017.12.062</a>	
6.8. Phase-Contrast MRI [NI]	(81)
<a href="https://doi.org/10.1148/rg.2020190039">https://doi.org/10.1148/rg.2020190039</a>	
<a href="https://doi.org/10.1186/s40809-016-0019-0">https://doi.org/10.1186/s40809-016-0019-0</a>	
6.8.1. Displacement Spectrum (DiSpect) Imaging	(82)
<a href="https://doi.org/10.1002/mrm.28882">https://doi.org/10.1002/mrm.28882</a>	
# Non Hemodynamic	
6.9. Functional Magnetic Resonance Spectroscopy [NI]	(83)

<a href="https://doi.org/10.1016/j.neuroimage.2023.120194">https://doi.org/10.1016/j.neuroimage.2023.120194</a>	
<a href="https://doi.org/10.1177/0271678X221076570">https://doi.org/10.1177/0271678X221076570</a>	
<a href="https://doi.org/10.1002/nbm.4314">https://doi.org/10.1002/nbm.4314</a>	
6.9.1. $^{31}\text{P}$ MRS [NI]	
<a href="https://doi.org/10.1002/nbm.70043">https://doi.org/10.1002/nbm.70043</a>	
6.9.2. $^{13}\text{C}$ MRS [I]	
<a href="https://doi.org/10.1007/s11064-022-03538-8">https://doi.org/10.1007/s11064-022-03538-8</a>	
<a href="https://doi.org/10.1038/s41598-019-38981-1">https://doi.org/10.1038/s41598-019-38981-1</a>	
6.10. Functional Quantitative Susceptibility Mapping (fQSM) [NI]	(84)
<a href="https://doi.org/10.1016/j.neuroimage.2021.117924">https://doi.org/10.1016/j.neuroimage.2021.117924</a>	
6.11. $^{23}\text{Na}$ Sodium MRI [NI]	(85)
<a href="https://doi.org/10.1016/j.neuroimage.2018.09.071">https://doi.org/10.1016/j.neuroimage.2018.09.071</a>	
<a href="https://archive.ismrm.org/2025/2503_bFbEoFQ1W.html">https://archive.ismrm.org/2025/2503_bFbEoFQ1W.html</a>	
6.12. Hyperpolarized $^{13}\text{C}$ Metabolic MRI [I]	(86)
<a href="https://doi.org/10.1073/pnas.1613345114">https://doi.org/10.1073/pnas.1613345114</a>	
<a href="https://doi.org/10.1016/j.neuroimage.2022.119284">https://doi.org/10.1016/j.neuroimage.2022.119284</a>	
6.13. Chemical Exchange Saturation Transfer (CEST) fMRI [NI]	(87)
<a href="https://doi.org/10.1038/s41598-019-40986-9">https://doi.org/10.1038/s41598-019-40986-9</a>	
<a href="https://doi.org/10.1002/jmri.27850">https://doi.org/10.1002/jmri.27850</a>	
6.14. Magnetic Resonance Elastography (MRE) [NI]	(88)
<a href="https://doi.org/10.3389/fbioe.2021.666456">https://doi.org/10.3389/fbioe.2021.666456</a>	
<a href="https://archive.ismrm.org/2014/0871.html">https://archive.ismrm.org/2014/0871.html</a>	
6.15. Apparent Diffusion Coefficient fMRI [NI]	(89)
<a href="https://doi.org/10.1038/s42003-025-07889-0">https://doi.org/10.1038/s42003-025-07889-0</a>	
<a href="https://doi.org/10.1038/s41467-025-60357-5">https://doi.org/10.1038/s41467-025-60357-5</a>	
<a href="https://doi.org/10.3389/fnhum.2013.00817">https://doi.org/10.3389/fnhum.2013.00817</a>	
6.16. Molecular fMRI [I]	(90)
<a href="https://doi.org/10.1016/j.jneumeth.2021.109372">https://doi.org/10.1016/j.jneumeth.2021.109372</a>	

## 7. LABELED OPTICAL TECHNIQUES

### # Indicators and Sensors

<https://doi.org/10.1146/annurev-anchem-061522-044819>

#### 7.1. Calcium Indicators [I]

<https://doi.org/10.1111/j.1476-5381.2010.00988.x>

<https://doi.org/10.1016/j.cell.2021.12.007>

7.1.1. Calcium Indicator Dyes (Ratiometric / Nonratiometric) [I] (91)

<https://doi.org/10.1073/pnas.1507110112>

[https://doi.org/10.1016/S0143-4160\(98\)90085-9](https://doi.org/10.1016/S0143-4160(98)90085-9)

7.1.2. Genetically Encoded Calcium Indicators (GECIs) [I] (92)

<https://doi.org/10.1016/j.neures.2020.05.013> (GCaMP variants)

#### 7.2. Voltage Indicators [I]

<https://doi.org/10.3390/colorants3040025> (General)

7.2.1. Voltage-Sensitive Dye Imaging (VSDI) (Ratiometric / Nonratiometric) [I] (93)

<https://doi.org/10.1016/j.cclet.2020.12.060>

<https://doi.org/10.1002/lpor.202400165>

7.2.2. Genetically Encoded Voltage Indicators (GEVIs) [I] (94)

[https://doi.org/10.1007/978-981-15-8763-4\\_12](https://doi.org/10.1007/978-981-15-8763-4_12)

7.3. Other Ion Indicators [I]	(95)
<a href="https://doi.org/10.1111/j.1476-5381.2010.00988.x">https://doi.org/10.1111/j.1476-5381.2010.00988.x</a>	
<a href="https://doi.org/10.1016/j.cell.2021.12.007">https://doi.org/10.1016/j.cell.2021.12.007</a>	
7.3.1. Genetically Encoded Chloride Indicators (Cl-Sensor) [I]	
<a href="https://doi.org/10.1038/s41467-023-37433-9">https://doi.org/10.1038/s41467-023-37433-9</a>	
7.3.2. Genetically Encoded Potassium Indicators (GEPIs) [I]	
<a href="https://doi.org/10.1038/s41598-024-62993-1">https://doi.org/10.1038/s41598-024-62993-1</a>	
7.3.3. pH-Sensitive Fluorescent Proteins [I]	
<a href="https://doi.org/10.1523/JNEUROSCI.0670-07.2007">https://doi.org/10.1523/JNEUROSCI.0670-07.2007</a>	
<a href="https://doi.org/10.1016/S0896-6273(04)00144-8">https://doi.org/10.1016/S0896-6273(04)00144-8</a>	
7.4. Second-Messenger Sensors [I]	(96)
7.4.1. Genetically Encoded cAMP/PKA/Second-Messenger Sensors (e.g., Pink Flamindo, G-Flamp) [I]	
<a href="https://doi.org/10.1038/s41467-022-32994-7">https://doi.org/10.1038/s41467-022-32994-7</a>	
<a href="https://doi.org/10.1016/j.celrep.2017.12.022">https://doi.org/10.1016/j.celrep.2017.12.022</a>	
7.5. Neurotransmitter/Neuropeptide Sensors [I]	(97)
7.5.1. Genetically Encoded Dopamine Indicators (GRAB-DA, dLight1) [I]	
<a href="https://doi.org/10.1038/s41592-023-02100-w">https://doi.org/10.1038/s41592-023-02100-w</a>	
7.5.2. Genetically Encoded Glutamate Indicators (iGluSnFR) [I]	
<a href="https://doi.org/10.1038/s41592-023-01863-6">https://doi.org/10.1038/s41592-023-01863-6</a>	
<a href="https://doi.org/10.1111/jnc.15608">https://doi.org/10.1111/jnc.15608</a> (reviews)	
7.5.3. Genetically Encoded Acetylcholine Indicators (GACh) [I]	
<a href="https://doi.org/10.1038/s41586-023-06492-9">https://doi.org/10.1038/s41586-023-06492-9</a>	
7.5.4. Genetically Encoded Serotonin Sensors (GRAB-5HT) [I]	
<a href="https://doi.org/10.1038/s41593-021-00823-7">https://doi.org/10.1038/s41593-021-00823-7</a>	
<a href="https://doi.org/10.1038/s41592-024-02188-8">https://doi.org/10.1038/s41592-024-02188-8</a>	
7.5.5. Genetically Encoded Norepinephrine Sensors (GRAB-NE) [I]	
<a href="https://doi.org/10.1016/j.neuron.2019.02.037">https://doi.org/10.1016/j.neuron.2019.02.037</a>	
<a href="https://doi.org/10.1016/j.neuron.2024.03.001">https://doi.org/10.1016/j.neuron.2024.03.001</a>	
7.5.6. Genetically Encoded GABA Indicators (iGABA-SnFR) [I]	
<a href="https://doi.org/10.1111/jnc.15608">https://doi.org/10.1111/jnc.15608</a>	
<a href="https://doi.org/10.1016/j.ebiom.2021.103272">https://doi.org/10.1016/j.ebiom.2021.103272</a>	
<a href="https://doi.org/10.1038/s41592-019-0471-2">https://doi.org/10.1038/s41592-019-0471-2</a>	
7.5.7. Genetically Encoded Histamine Sensors (GRAB-HA) [I]	
<a href="https://doi.org/10.1016/j.neuron.2023.02.024">https://doi.org/10.1016/j.neuron.2023.02.024</a>	
7.5.8. Genetically Encoded Adenosine Sensors (GRAB-Ado) [I]	
<a href="https://doi.org/10.1038/s41586-022-05407-4">https://doi.org/10.1038/s41586-022-05407-4</a>	
<a href="https://doi.org/10.1038/s41467-025-59530-7">https://doi.org/10.1038/s41467-025-59530-7</a>	
7.5.9. Fluorescent False Neurotransmitters (FFNs) [I]	
<a href="https://doi.org/10.1021/acschemneuro.1c00580">https://doi.org/10.1021/acschemneuro.1c00580</a>	
<a href="https://doi.org/10.1038/s41467-018-05075-x">https://doi.org/10.1038/s41467-018-05075-x</a>	
7.5.10. Synaptophysin-pHluorin (SypHy) [I]	
<a href="https://doi.org/10.1523/JNEUROSCI.0670-07.2007">https://doi.org/10.1523/JNEUROSCI.0670-07.2007</a>	
<a href="https://doi.org/10.1016/S0896-6273(04)00144-8">https://doi.org/10.1016/S0896-6273(04)00144-8</a>	
7.6. Metabolic and Other Sensors [I]	(98)
7.6.1. Genetically Encoded ATP Indicators (iATPSnFR) [I]	
<a href="https://doi.org/10.1016/j.neuron.2021.11.027">https://doi.org/10.1016/j.neuron.2021.11.027</a>	
<a href="https://doi.org/10.1038/s41586-021-03497-0">https://doi.org/10.1038/s41586-021-03497-0</a>	

### 7.6.2. Genetically Encoded Lactate Sensors [I]

<https://doi.org/10.1038/s41467-025-64484-x>

### 7.6.3. Genetically Encoded Nitric Oxide Sensors (geNOps) [I]

<https://doi.org/10.1016/j.celrep.2023.113514>

## # Microscopy Modalities

- 7.7. One-Photon Imaging (Miniscope / Widefield) [I] (99)  
<https://doi.org/10.1016/j.conb.2011.12.002>  
<https://doi.org/10.1016/j.cell.2022.02.017>
- 7.8. Two-Photon Microscopy [I] (100)  
<https://doi.org/10.1016/j.conb.2011.12.002>  
<https://doi.org/10.1016/j.cell.2022.02.017>
- 7.9. Three-Photon Microscopy [I] (101)  
<https://doi.org/10.1038/s41583-025-00937-y>  
<https://doi.org/10.1038/s42003-025-08079-8>
- 7.10. Fiber Photometry [I] (102)  
<https://doi.org/10.1080/15476278.2025.2489667>
- 7.11. Mesoscopic Calcium Imaging (Widefield, not Miniscope) [I] (103)  
<https://doi.org/10.3390/biology11111601>  
<https://doi.org/10.3389/fnins.2023.1210199>  
<https://doi.org/10.1371/journal.pone.0185759>
- 7.12. Light-Sheet Fluorescence Microscopy (LSFM) [I] (104)  
<https://doi.org/10.1038/nmeth.2434>  
<https://doi.org/10.1016/j.jneumeth.2018.07.011>  
<https://doi.org/10.1016/j.conb.2018.03.007>  
<https://doi.org/10.1146/annurev-neuro-070918-050357>
- 7.13. Swept Confocally-Aligned Planar Excitation (SCAPE) Microscopy [I] (105)  
<https://doi.org/10.1038/nphoton.2014.323>  
<https://doi.org/10.1038/s41587-020-0628-7>
- 7.14. Adaptive Optics for In Vivo Neural Imaging [I] (106)  
<https://doi.org/10.3389/fnins.2022.880859>  
<https://doi.org/10.3389/fnins.2023.1188614>
- 7.15. Oblique Plane Microscopy (OPM) [I] (107)  
<https://doi.org/10.1038/s41467-023-43741-x>
- 7.16. Multifocal/Multibeam Two-Photon Microscopy [I] (108)  
<https://doi.org/10.1364/BOE.9.003678>  
<https://doi.org/10.3389/fncel.2019.00039>  
<https://doi.org/10.1364/BOE.514826>
- 7.17. Line-Scanning Temporal Focusing Microscopy (mosTF) [I] (109)  
<https://doi.org/10.1038/s41598-024-57208-6>  
<https://doi.org/10.1364/BOE.9.005654>
- 7.18. Light Field Microscopy [I] (110)  
<https://doi.org/10.1016/j.jneumeth.2021.109083>
- 7.19. Fluorescence Lifetime Imaging Microscopy (FLIM) [I] (111)  
<https://doi.org/10.1364/OPTICA.426870>  
<https://doi.org/10.1038/s41598-020-77737-0>
- 7.20. Total Internal Reflection Fluorescence (TIRF) Microscopy [I] (112)  
<https://doi.org/10.1021/acsnano.3c04489>

- 7.21. Random Access Microscopy [I] (113)  
<https://doi.org/10.1038/nmeth.4033>  
<https://iopscience.iop.org/article/10.1088/2515-7647/ad2e0d>
- 7.22. Multispectral Optoacoustic Tomography (MSOT) [I] (114)  
<https://doi.org/10.1016/j.pacs.2021.100285>
- 7.23. Bioluminescent Voltage Imaging (e.g., LOTUS-V) [I] (115)  
<https://doi.org/10.1038/s41598-019-43897-x>  
<https://doi.org/10.1117/1.nph.11.2.024203>
- 7.24 Digital Holographic Microscopy [I] (116)  
<https://doi.org/10.1038/s41467-023-36889-z>  
<https://doi.org/10.1016/j.conb.2018.03.006>

## 8. INTERFERENCE TECHNIQUES

<https://doi.org/10.3389/fnbeh.2021.820017>

### # Electrical Stimulation

- 8.1. Cortical Stimulation [I] (117)  
<https://doi.org/10.1016/j.yebeh.2009.03.001> (1870)
- 8.2. Microstimulation [I] (118)  
<https://doi.org/10.1038/s41551-024-01299-z>  
<https://doi.org/10.1002/adhm.202100119>
  - 8.2.1 Supracortical Microstimulation [I]  
<https://doi.org/10.1146/annurev-bioeng-103023-072855>
- 8.3. Temporal Interference Simulation [NI] (119)  
<https://doi.org/10.3389/fnhum.2023.1266753>  
<https://doi.org/10.1016/j.cell.2017.05.024>  
<https://doi.org/10.1109/EMBC46164.2021.9629968>
- 8.4. Transcranial Electrical Stimulation [NI]  
<https://doi.org/10.3390/biomedicines10102333>  
<https://doi.org/10.1371/journal.pbio.3001973>
  - 8.4.1. Transcranial Direct Current Stimulation (tDCS) [NI] (120)  
<https://doi.org/10.3389/fnhum.2025.1640565>
  - 8.4.2. Transcranial Alternating Current Stimulation (tACS) [NI] (121)  
<https://doi.org/10.3389/fnhum.2025.1640565>
  - 8.4.3. Transcranial Random Noise Stimulation (RNS) [NI] (122)  
<https://doi.org/10.1038/s41598-019-51553-7>  
<https://doi.org/10.3389/fncel.2017.00162>
- 8.5. Transcranial Magnetic Stimulation [NI] (123)  
<https://doi.org/10.1007/s12264-021-00781-x>  
<https://doi.org/10.1146/annurev-psych-081120-013144>
  - 8.5.1. Deep Transcranial Magnetic Stimulation (dTMS) [NI] (124)  
<https://doi.org/10.5498/wjp.v13.i9.607>  
<https://doi.org/10.1016/j.jneumeth.2021.109261>  
<https://doi.org/10.1016/j.jneumeth.2020.108709>  
<https://doi.org/10.1523/ENEURO.0163-17.2018>
- 8.6. Peripheral Nerve Stimulation [I]  
8.6.1. Galvanic Vestibular Stimulation (GVS) [I] (125)

- <https://doi.org/10.1152/jn.00035.2019>  
<https://doi.org/10.3389/fneur.2012.00117>  
[https://doi.org/10.1016/0006-8993\(82\)90990-8](https://doi.org/10.1016/0006-8993(82)90990-8) (1982)  
<https://doi.org/10.1016/j.brainresbull.2004.07.008>
- 8.6.2. Trigeminal Nerve Stimulation (TNS) [I] (126)
- <https://doi.org/10.1186/s42234-023-00128-z>  
<https://doi.org/10.3171/jns.2002.97.5.1179>  
<https://doi.org/10.1007/s00221-018-5338-8>
- 8.6.3. Vagus Nerve Stimulation (VNS) [I] (127)
- <https://doi.org/10.1093/cercor/bhab158>  
<https://doi.org/10.3390/brainsci12091137>
- ### # Optogenetics and Optogenetics-Like
- <https://doi.org/10.1002/advs.202413817>
- 8.7. Optogenetics [I] (128)
- <https://doi.org/10.1017/S0033583523000033>  
<https://doi.org/10.3389/fncel.2021.778900>  
<https://doi.org/10.1007/978-981-15-8763-4>
- 8.7.1. Channelrhodopsin-2 (ChR2)  
 8.7.2. Halorhodopsin (NpHR)  
 8.7.3. Archaerhodopsin (Arch)  
 8.7.4. Anion-conducting channelrhodopsins (ACRs)  
 8.7.5. Photoswitchable Voltage-Gated Ion Channels  
 8.7.6. Photoswitchable Ligand-Gated Ion Channels  
 8.7.7. Optical Switch Protein Conjugates  
 8.7.8. Holographic Optogenetic Stimulation [I] (129)
- <https://doi.org/10.1038/s41593-021-00902-9>  
<https://doi.org/10.1523/JNEUROSCI.1785-18.2018>
- 8.7.9. X-ray Optogenetics [NI/I] (130)
- <https://doi.org/10.1038/s41467-021-24717-1>
- 8.8. Chemogenetics [I] (131)
- <https://doi.org/10.1177/10738584221134587>  
<https://doi.org/10.1002/glia.24390>  
<https://doi.org/10.1523/JNEUROSCI.0625-23.2023>  
<https://doi.org/10.1038/s41593-020-0661-3>
- 8.9. Mechano/Sonogenetics [NI] (132)
- <https://doi.org/10.1016/j.brs.2022.09.002>  
<https://doi.org/10.1002/anie.202317112>
- 8.10. Magnetogenetics [NI] (133)
- <https://doi.org/10.15252/embj.201797177>  
<https://doi.org/10.1186/s12951-024-02616-z>  
<https://doi.org/10.1038/s41565-024-01694-2>  
<https://doi.org/10.1038/s41467-021-25837-4>  
<https://doi.org/10.7554/eLife.27069>  
<https://doi.org/10.1038/s41563-022-01281-7>
- 8.11. Thermogenetics [NI/I] (134)
- <https://doi.org/10.1002/advs.202413817>
- 8.12. Upconversion Nanoparticle-Based Neural Imaging [I] (135)

<https://doi.org/10.1126/science.aag1144>  
<https://doi.org/10.1016/j.biomaterials.2017.07.017>

## # Other Interference Techniques

- 8.12. Cortical Cooling [I] (136)  
<https://doi.org/10.3389/fnsys.2011.00053>  
[https://doi.org/10.1016/0006-8993\(77\)90734-X](https://doi.org/10.1016/0006-8993(77)90734-X)
- 8.13. Lidocaine Inactivation [I] (137)  
[https://doi.org/10.1016/S0165-0270\(97\)02229-2](https://doi.org/10.1016/S0165-0270(97)02229-2)
- 8.14. Muscimol Inactivation [I] (138)  
<https://doi.org/10.1016/j.jneumeth.2008.01.033>
- 8.15. Transcranial Focused Ultrasound [NI] (139)  
<https://doi.org/10.3389/fnhum.2021.749162>  
<https://doi.org/10.1371/journal.pone.0288654>  
<https://doi.org/10.1007/s13534-024-00369-0>
- 8.15.1. Focused Ultrasound Blood–Brain Barrier Opening (FUS-BBBO) [NI] (140)  
<https://doi.org/10.1002%2Fmuds.101>  
<https://doi.org/10.1148/radiol.14140245>  
<https://doi.org/10.1093/brain/awab460>  
<https://doi.org/10.1038/s41598-018-25904-9>
- 8.16. Optoacoustic Neuromodulation [I] (141)  
<https://doi.org/10.1038/s41467-020-14706-1>
- 8.17. Infrared Neural Stimulation [NI/I] (142)  
<https://doi.org/10.1073/pnas.2015685118>  
<https://doi.org/10.1038/s41598-021-89163-x>  
<https://doi.org/10.1016/j.brs.2023.01.1678>
- 8.18. Transcranial Photobiomodulation (tPBM) [NI] (143)  
<https://doi.org/10.1038/s41598-019-42693-x>  
<https://doi.org/10.1126/sciadv.abq3211>  
<https://doi.org/10.1364/BOE.402047>
- 8.19 Photothermal Neuromodulation (non-genetic) [I] (144)  
<https://doi.org/10.1002/admi.202400873>  
<https://doi.org/10.1021/acsnano.4c01037>
- 8.20. Iontronic microfluidic probes / microfluidic interconnection [I] (145)  
<https://doi.org/10.1038/s41378-021-00295-6>  
<https://doi.org/10.1002/smll.202410906>
- 8.21. Neuromodulation via Magnetic Nanodiscs [I] (146)  
<https://doi.org/10.1038/s41565-024-01798-9>  
<https://doi.org/10.3389/fnhum.2025.1489940>
- 8.22. Terahertz Neural Control [NI] (147)  
<https://doi.org/10.7554/eLife.97444.3>
- DOI: [10.4103/NRR.NRR-D-23-00872](https://doi.org/10.4103/NRR.NRR-D-23-00872)

## 9. Miscellaneous Techniques

(or they are a mix/interface of the above or not encompassed by the above classification)

- 9.1. Thermal Diffusion Flowmetry [I] (148)  
<https://doi.org/10.1089/neu.2018.6309>
- 9.2. Magnetic Particle Imaging (MPI) [I] (149)

<https://doi.org/10.18416%2FIJMPI.2020.2009009>

<https://doi.org/10.1016/j.neuroimage.2018.05.004>

9.3. Near-Infrared Fluorescence Imaging of Hemodynamics [I] (150)

<https://doi.org/10.1371/journal.pone.0048383>

9.4. Second Harmonic Generation (SHG) Microscopy [I] (151)

<https://doi.org/10.1073/pnas.1004748107> (*pivotal, not brain*)

<https://doi.org/10.1021/acspophotonics.9b01749> (brain dynamics)

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## **ADDITIONS:**

1. Diffuse Correlation Spectroscopy

from <https://doi.org/10.1111/micc.12884>

2. Laminar Functional Magnetic Resonance Imaging

from <https://doi.org/10.1093/psyrad/kkae027>

3. Phase-Contrast Based Approaches MRI

from <https://doi.org/10.1002/mrm.28882>

4. Thermal Diffusion Flowmetry

<https://doi.org/10.1089/neu.2018.6309>

from <https://doi.org/10.1111/micc.12884>

## **EXCLUDED after recheck:**

1. Deep Label-Free Microscopy (DLFM)

<https://doi.org/10.1038/nm.3495> (after rechecking its time frames)

2. Microvascular Volumetric Pulsatility Mapping

<https://doi.org/10.1038/s44161-025-00722-1>

<https://doi.org/10.3389/fnagi.2025.1486775>

<https://doi.org/10.1177/0271678X20980652>

(the literature does not show functional studies to the moment)

3. Magnetic Resonance Fingerprinting (MRF)

<https://doi.org/10.1002/jmri.29812> (It is not a technique per se, but more a “pipeline” optimization method. EXTREMELY interesting and useful, but here excluded)

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