

## *LIST OF NEUROSCIENCE TECHNIQUES TO STUDY BRAIN ACTIVITY IN VIVO*

SEED 1 + SEED 2 cleaned and organized, excluding repetitions or same techniques with different terms

### **SEED 1\_Guide to Research Techniques in Neuroscience**

@book{carter\_guide\_2022, address = {San Diego}, edition = {3rd ed}, title = {Guide to {Research} {Techniques} in {Neuroscience}}, isbn = {978-0-12-818646-6 978-0-323-91561-8}, language = {en}, publisher = {Elsevier Science \& Technology}, author = {Carter, Matt and Essner, Rachel and Goldstein, Nitsan and Iyer, Manasi}, year = {2022}}

### **SEED 2\_Neuromethods**

@Series{Neuromethods,title = {Neuromethods}, publisher = {Springer}, issn = {0893-2336}, issn-electronic = {1940-6045}, url = {https://link.springer.com/series/7657}, note = {A comprehensive series of manuals that offer step-by-step protocols in neuroscience and neurobiology research. Founded in 1986.}}

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**[I] = Invasive [NI] = Non-Invasive**

### **Magnetic Resonance & Computed Tomography (CT) Imaging Techniques**

Functional Magnetic Resonance Imaging *[NI]*

1. Blood Oxygen Dependente Levels (BOLD) *[NI]*
2. quantitative MRI (qMRI) for Manganese (Mn) and Iron (Fe) *[NI]*
3. Dynamic Susceptibility Contrast (DSC)-MRI *[NI]*
4. Arterial Spin Labeling (ASL) *[NI]*
5. Nuclear Magnetic Resonance Spectroscopy *[NI]*  
    <sup>13</sup>C Magnetic Resonance Spectroscopy
6. Positron Emission Tomography (PET) *[NI]*  
        Assay of Enzyme Activity, Neuroreceptors/Neurotransmitters, Messenger Pathways  
        Quantification of Cerebral Blood Flow
7. Single-Photon Emission Computerized Tomography *[NI]*
8. Cerebral Blood Flow with Stable Xenon CT *[NI]*

### **Ultrasound**

9. Transcranial Doppler *[NI]*

### **EEG & MEG**

10. Electroencephalography *[NI]*  
    High-density EEG
11. Magnetoencephalography *[NI]*

### **INVASIVE ELECTRICAL RECORDINGS**

12. Deep Brain Stimulation *[I]*
13. Single Electrodes *[I]*
14. Multielectrode Array *[I]*

## **CELLULAR RECORDINGS / PATCH-CLAMP**

- 15. Patch-Clamp Techniques [I]
  - Voltage-Clamp Fluorometry [I]
  - Patch-Clamp Fluorometry [I]

## **MICROSCOPY / MICROENDOSCOPY**

- 16. One-Photon Microscopy [I]
  - GRIN LENSES
  - Miniscopes
  - mini-mScope Mesoscale Calcium (Ca++)
- 17. Two-Photon Microscopy [I]
- 18. Fiber Photometry [I]
- 19. High-Density Multichannel Fiber Photometry [I]
- 20. Neuro-FITM [I]

## **NEUROSENSING / ELECTROCHEMICAL**

- 21. Microdialysis [I]
- 22. Column Liquid Chromatography [I]
- 23. Push–Pull Superfusion Technique (PPST) [I]
  - Carbon Electrode Surface Chemistry [I]
- 24. Voltammetry [I]
  - Fast-Scan Cyclic (FSC)
- 25. Amperometry [I]
  - High-Speed Chronoamperometry

## **FLUORESCENT ACTIVITY INDICATORS**

### **DYES AND ENCODING**

- 26. Voltage-sensitive dye imaging (VSDI) (Ratiometric / Nonratiometric) [I]
- 27. Calcium Indicator Dyes (Ratiometric / Nonratiometric) [I]
  - Fluorescent Calcium Indicators Dyes
  - acetoxymethyl (AM) ester-based Multi Cell Bolus Loading (MCBL)
- 28. Genetically Encoded Voltage Indicators (GEVIs) [I]
  - Genetically Encoded Ca++ Indicators (GECIs)
  - Sensitive Bioluminescence Reporter
- 29. pH-sensitive Fluorescent Proteins [I]

### **FLUORESCENCE READOUT TECHNIQUES**

- 30. Fluorescence/Föster Resonance Energy Transfer (FRET) [I]
- 31. Bimolecular Fluorescence Complementation (BiFC) [I]
- 32. Fluorescence Recovery After Photobleaching (FRAP) [I]
- 33. Photoactivation/Photoconversion [I]
- 34. Phosphorescence Lifetime (PLIM) [I]
- 35. Multiphoton FRET-FLIM [I]

## **OPTICAL APPROACHES**

- 36. Diffuse Optical Imaging [NI]

37. Near Infrared Spectroscopy [NI]

### # INTERFERENCE

#### ELECTRICAL/MAGNETIC

- 38. Transcranial Magnetic Stimulation [NI]
- 39. Transcranial Direct Current Stimulation (tDCS) [NI]
- 40. Microstimulation [/]
- 41. Electrolytic lesions [/]

#### EXCLUSIVELY GENETIC

- 42. Transgenes from other species [/]
- 43. Designer receptors exclusively activated by designer drugs (DREADDs: hM3Dq, hM4Di) [/]
- 44. RNA Interference (RNAi) [/]
- 45. Morpholinos [/]
- 46. Dominant Negatives [/]
- 47. Single-Cell Electroporation (SCE) (Is an Interference Technique) [/]

#### OPTOGENETICS AND GENETICALLY MEDIATED DERIVATIVES

- 48. Optogenetics [/]
  - Channelrhodopsin-2 (ChR2)
  - Halorhodopsin (NpHR)
  - Archaerhodopsin (Arch)
  - Anion-conducting channelrhodopsins (ACRs)
  - Photoswitchable Voltage-Gated Ion Channels
  - Photoswitchable Ligand-Gated Ion Channels
  - Optical Switch Protein Conjugates
- 49. Magnetothermal Genetic Stimulation [/]

#### ULTRASOUND

- 50. Ultrasonic Neuromodulation [NI]
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#### Excluded after further evaluation

Technique / Reasoning / Notes

1. Autoradiography / <https://doi.org/10.1007/s00441-014-2093-4> / Not In Vivo
2. Measurement of Pial Vessel Hemodynamics / <https://doi.org/10.1080/01616412.1987.11739803> / Restrict used and specifics studies related just to response to other hemodynamic parameters, and soon afterwards substituted by Ultrasound techniques

3. Thermal Changes / [https://doi.org/10.1007/978-1-61779-897-9\\_12](https://doi.org/10.1007/978-1-61779-897-9_12) / Whole-body, no Brain specificity
4. Organized and Nested several within **FLUORESCENT ACTIVITY INDICATORS, DYES and ENCODING** to end with duplicity and ambiguity, and then their READOUTS techniques.
5. Also Organized and Nested Optogenetics Tools, to not overexpress slightly different techniques