

LIST OF NEUROSCIENCE TECHNIQUES TO STUDY BRAIN ACTIVITY IN VIVO

After the LLM Screening, it is necessary to merge **both Step 2** and **Step 3** into a single list, again screening for the techniques that fit the established criteria and also looking for duplicates. All the while creating a properly structured list, and also already tagging if they are Invasive [**I**] or non-invasive [**NI**], techniques which are minimally invasive (for instance, require only a contrast injection or isotope) are tagged as [**I***].

New additions while performing the merge of both Step 2 and 3 and checking on selected papers were added to the list, but are pointed at the end as additions and where did they come from (tagged as “§”).

Note: some items are listed due to their importance but are not counted as they can be seen as advancements or specific approaches within the same technique.

Any feedback is more than welcomed at ricardojunior@unistra.fr

1. ELECTROMAGNETIC

Microscopic Resolutions

Intracellular or Juxtacellular

- 1.1. Patch-Clamp [**I**] (1)
https://doi.org/10.1007/978-1-0716-0818-0_1
<https://doi.org/10.3390/s21041448>
- 1.2. Sharp Electrode Intracellular Recordings [**I**] (2)
[https://doi.org/10.1016/S0165-0270\(03\)00126-2](https://doi.org/10.1016/S0165-0270(03)00126-2)
- 1.3. Juxtacellular Recording and Labeling [**I**] (3)
<https://doi.org/10.1016/j.jneumeth.2006.02.004>
<https://doi.org/10.1038/nprot.2014.161>

Extracellular

- 1.4. Tungsten Microelectrodes [**I**] (4)
<https://doi.org/10.3389/fnins.2021.691788>
- 1.5. Stereotrodes [**I**] (5)
[https://doi.org/10.1016/0165-0270\(83\)90097-3](https://doi.org/10.1016/0165-0270(83)90097-3)
<https://doi.org/10.1016/j.jneumeth.2013.11.013>
- 1.6. Tetrodes [**I**] (6)
[https://doi.org/10.1016/S0165-0270\(02\)00092-4](https://doi.org/10.1016/S0165-0270(02)00092-4)
- 1.7. Multielectrode Arrays [**I**]
 - 1.7.1. Michigan Probes [**I**] (7)
<https://doi.org/10.1109/10.7273>
<https://doi.org/10.1016/j.heares.2008.01.010>

1.7.2. Utah Arrays [I]	(8)
https://doi.org/10.1016/S0013-4694(96)95176-0	
1.7.3. Floating Microelectrode Arrays [I]	(9)
https://doi.org/10.1016/j.jneumeth.2006.09.005	
1.8. Neuropixels [I]	(10)
https://doi.org/10.1016/j.conb.2018.01.009	
1.9. CMOS-Integrated Neural Probes [I]	(11)
https://doi.org/10.1126/sciadv.adf9524	
https://doi.org/10.1016/j.snr.2024.100206	
1.10. Fishbone Nanowire Arrays [I]	(12)
https://doi.org/10.1002/adma.202504171	
1.11. Nitrogen-Vacancy Diamond Sensors [I]	(13)
https://doi.org/10.1073/pnas.1601513113	
https://doi.org/10.1038/s41598-023-39539-y	
# Middle Term Resolutions and Multimodal Approaches	
1.12. Flexible Bioelectronic Neural Interfaces [I]	(14)
https://doi.org/10.1038/s41563-020-0679-7	
1.13. Optotetrodes [I]	(15)
https://doi.org/10.1038/nn.2992	
1.14. Transparent Graphene Microelectrodes Arrays [I]	(16)
https://doi.org/10.1038/s41467-018-04457-5	
1.15. Electrical Impedance Tomography [NI/I]	(17)
https://link.springer.com/article/10.1186/s40779-022-00370-7	
https://doi.org/10.1016/j.neuroimage.2019.05.023	
https://doi.org/10.1006/nimg.2000.0698	
# Macroscopic Resolutions	
1.16. Electroencephalography [NI]	(18)
https://doi.org/10.1038/nrn3241	
https://doi.org/10.1016/j.ijpsycho.2015.05.004	
1.17. Electrooculogram [NI]	(19)
https://doi.org/10.1038/nrn3241	
1.16.1. Micro Electrooculogram [I]	(20)
https://doi.org/10.1038/s41378-023-00597-x	
1.18. Magnetoencephalography [NI]	
https://doi.org/10.1038/nrn3241	
https://doi.org/10.3390/brainsci12060788	
1.17.1 SQUID (the typical one) [NI]	(21)
1.17.2. Optically Pumped Magnetometer MEG (OPM-MEG) [NI]	(22)
https://doi.org/10.1038/s41398-024-03047-y	
1.19. Stentrodes [I]	(23)
https://doi.org/10.1038/nbt.3428	
https://doi.org/10.1016/j.jneumeth.2025.110471	
https://doi.org/10.1126/science.adh3916	
1.20. Stereoelectroencephalography [I]	(24)
https://doi.org/10.1097/wnp.0000000000000249	
https://doi.org/10.1016/j.seizure.2019.01.021	

1.21. Deep Brain Stimulation [I] (25)

<https://doi.org/10.3389/fpsy.2023.1080260>

<https://doi.org/10.1016/j.neurom.2022.01.017>

1.22. Neural Dust [I] (26)

<https://doi.org/10.1016/j.conb.2017.12.010>

<https://doi.org/10.3389/fnins.2021.599549>

2. MOLECULES SENSING

Microdialysis and Push-Pull Perfusion

2.1. Microdialysis [I] (27)

<https://doi.org/10.1021/acsami.2c02740>

<https://doi.org/10.1039/C4AN01974A>

2.2. Push-pull Perfusion [I] (28)

<https://doi.org/10.1021/acsami.2c02740>

<https://pubs.acs.org/doi/10.1021/acs.analchem.8b02468>

Note: both 2.1. and 2.2. can be associated with the following techniques

2.3. + High Performance Liquid Chromatography [I] (29)

<https://doi.org/10.1021/ac4023605>

2.4. + Column Liquid Chromatography [I] (30)

<https://doi.org/10.1021/acsami.2c02740>

2.5. + Capillary Electrophoresis [I] (31)

<https://doi.org/10.1016/j.jneumeth.2004.03.025>

2.6. + Mass Spectrometry [I] (32)

<https://doi.org/10.1016/j.jpba.2005.07.025>

<https://doi.org/10.1208/s12248-017-0114-4>

2.7. + Electrochemical Workstation [I] (33)

<https://doi.org/10.1021/acs.nanolett.2c00289>

Carbon Fiber Microelectrodes

2.8. Voltammetry [I] (34)

<https://doi.org/10.1021/acsami.2c02740>

<https://doi.org/10.1016/j.neubiorev.2005.02.003>

<https://doi.org/10.1523/JNEUROSCI.14-01-00442.1994>

2.8.1. Differential Pulse Voltammetry

2.8.2. Cyclic Voltammetry

2.8.3. Fast-Scan Cyclic Voltammetry

[https://doi.org/10.1016/0306-4522\(88\)90255-2](https://doi.org/10.1016/0306-4522(88)90255-2)

<https://doi.org/10.1039/C9AN01925A>

2.9. Amperometry [I] (35)

<https://doi.org/10.1021/acsami.2c02740>

2.9.1. High-Speed Chronoamperometry

<https://doi.org/10.3390/cells11152454>

2.10. Liquid-Liquid Interface Microsensors [I] (36)

<https://doi.org/10.1021/acssensors.1c00978>

<https://doi.org/10.1021/ac504151e>

Indirect Sensing

2.11. Enzyme-Based sensors [I] (37)

<https://doi.org/10.1111/j.1471-4159.2006.03673.x>

<https://doi.org/10.1021/ac403250w>

2.12. Aptamer-based sensors [I] (38)

<https://doi.org/10.1021/acsptsci.4c00579>

2.13. Molecularly Imprinted Polymers Sensors [I] (39)

<https://doi.org/10.1016/j.snb.2011.07.052>

<https://doi.org/10.1016/j.trac.2018.08.002>

Other sensing techniques

2.14. Electron Paramagnetic Resonance (EPR) Oximetry [I] (40)

<https://doi.org/10.1007/s12013-017-0798-1>

<https://doi.org/10.4103/2045-9912.202911>

2.15. Hydrogen Clearance CBF Measurement [I] (41)

<https://doi.org/10.1038/jcbfm.1986.83>

3. POSITRON EMISSION TOMOGRAPHY

3.1. Total-Body PET Imaging [I*] (42)

<https://doi.org/10.1259/bjr.20220357>

<https://doi.org/10.2967/jnumed.119.231845>

<https://doi.org/10.1007/s00259-022-06057-4>

3.2. Metabolic Tracing with Short-Lived Isotopes (Dynamic PET) [I*] (43)

<https://doi.org/10.1002/acn3.51546R>

<https://doi.org/10.1007/s00259-023-06542-4>

3.3. μ -opioid Receptor PET ([^{11}C]carfentanil) [I*] (44)

<https://doi.org/10.1007/s00259-024-06746-2>

3.4. Dopamine D2/D3 Receptor Occupancy PET [I*] (45)

<https://doi.org/10.1038/s42003-022-03434-5>

<https://doi.org/10.1177/0271678X231210128>

3.5. Cerebral Metabolic Rate of Oxygen (CMRO₂) PET [I*] (46)

<https://doi.org/10.1016/j.neuroimage.2020.117136>

<https://doi.org/10.2967/jnumed.120.260521>

3.6. GABA-A Receptor PET ([^{18}F]flumazenil) [I*] (47)

<https://doi.org/10.3390/ph16030417>

3.7. Serotonin Transporter PET ([^{11}C]DASB) [I*] (48)

<https://doi.org/10.3390/ijms26010252>

3.8. Oxygen-15 Water PET ([^{15}O]H₂O PET) [I*] (49)

<https://doi.org/10.1053/j.semnuclmed.2023.08.002>

<https://doi.org/10.1016/j.neuroimage.2013.11.044>

4. NON-FLUORESCENCE OPTICAL TECHNIQUES

Infrared

4.1. Thermal Infrared Imaging / Thermoencephaloscopy [NI] (50)

<https://doi.org/10.1038/srep17471>

[https://doi.org/10.1016/S0301-0082\(98\)00038-0](https://doi.org/10.1016/S0301-0082(98)00038-0)

<https://doi.org/10.1111/psyp.12243>

4.2. Functional Near-Infrared Spectroscopy [NI] (51)

<https://doi.org/10.1111/nyas.13948>

4.2.1. Time-Domain Near-Infrared Spectroscopy (TD-NIRS) [NI] (52)

<https://doi.org/10.1038/s41598-024-68555-9>

<https://doi.org/10.1117/1.JBO.27.7.074710> *

<https://doi.org/10.1117/1.NPh.10.1.013504> *

4.2.2. Functional Diffuse Optical Tomography (fDOT) [NI] (53)

<https://doi.org/10.3390/s25072040> *

<https://doi.org/10.1038/s41598-025-85858-7>

<https://doi.org/10.1117/1.NPh.6.3.035007>

4.3. Diffuse Correlation Spectroscopy [NI] (54)

<https://doi.org/10.1016/j.neuroimage.2013.06.017>

<https://doi.org/10.1007/s12028-018-0573-1>

Intrinsic Signals

4.4. Optical Intrinsic Signal [I] (55)

<https://doi.org/10.1117/1.NPh.10.2.020601>

<https://doi.org/10.1016/j.xpro.2021.100779> *

<https://doi.org/10.1038/324361a0>

<https://doi.org/10.1146/annurev.ne.08.030185.001403>

4.4.1. Hyperspectral Imaging of Intrinsic Signals (56)

<https://doi.org/10.1117/1.NPh.2.4.045003>

https://doi.org/10.1007/978-3-319-91287-5_3

4.5. Laser Speckle Contrast Imaging [I] (57)

<https://doi.org/10.1117/1.NPh.10.2.020601>

<https://doi.org/10.1097/00004647-200103000-00002>

4.6. Spatial Frequency Domain Imaging [I] (58)

<https://doi.org/10.1117/1.NPh.10.2.020601>

<https://doi.org/10.1117/1.3088140>

<https://doi.org/10.1117/1.NPh.8.2.025001>

4.6.1. Structured Illumination Diffuse Optical Tomography [NI] (59)

<https://doi.org/10.1117/1.NPh.4.2.021102>

4.7. Laser Doppler Flowmetry [I] (60)

<https://doi.org/10.1111/micc.12884>

<https://doi.org/10.1002/lpor.202401016> *

<https://doi.org/10.1007/s12028-017-0472-x>

4.8. Optical Coherence Tomography [I/NI] (61)

<https://doi.org/10.1126/science.1957169>

<https://doi.org/10.1016/j.jneumeth.2008.11.026>

4.8.1. Speckle-Modulated Optical Coherence Tomography (OCT) [I] (62)

<https://doi.org/10.1038/ncomms15845>

<https://doi.org/10.1063/5.0278271>

<https://doi.org/10.1093/cercor/bhac388>

4.8.2. Doppler Optical Coherence Tomography (D-OCT) [I] (63)

<https://doi.org/10.1364/BOE.5.003217>

<https://doi.org/10.1021/acsphotonics.4c00856>

4.8.3. Visible Light Optical Coherence Tomography (vis-OCT) [I] (64)

<https://doi.org/10.1364/BOE.6.003941>
<https://doi.org/10.1364/BOE.6.001429>
<https://doi.org/10.1117/1.JBO.22.12.121707>

5. ULTRASOUND AND PHOTOACOUSTICS

Ultrasound (Purely)

5.1. Transcranial Doppler [NI] (65)

<https://doi.org/10.33549/physiolres.935413>
<https://doi.org/10.1186/s41984-021-00114-0>
<https://doi.org/10.1159/000103113>

5.2. Functional Ultrasound (fUS) Imaging [NI/I] (66)

<https://doi.org/10.1016/j.cobme.2021.100286>
<https://doi.org/10.1146/annurev-neuro-111020-100706>
<https://doi.org/10.1016/j.neuroscience.2021.03.005>
<https://doi.org/10.1038/nmeth.1641>

5.2.1. Microbubble-Enhanced fUS (Contrast-Enhanced fUS) [I] (67)

<https://doi.org/10.1016/j.neuroimage.2015.09.037>
<https://doi.org/10.1016/j.cobme.2021.100286>

5.2.2. Functional Ultrasound Microscopy (68)

<https://doi.org/10.1038/s41592-022-01549-5>

Photoacoustic

5.3. Photoacoustic Imaging (PAI) [I/NI] (69)

<https://doi.org/10.1016/j.pacs.2019.05.001>
<https://doi.org/10.1063/1.2195024>
<https://doi.org/10.3109/13813450312331337649>

5.3.1. Photoacoustic Computed Tomography (PACT) [I/NI] (70)

<https://doi.org/10.1038/nbt839>
<https://doi.org/10.1002/jbio.201700024>
<https://doi.org/10.1364/BOE.423707>

5.3.2. Functional Photoacoustic Microscopy (fPAM) [I] (71)

<https://doi.org/10.1038/s41377-022-00836-2>
<https://doi.org/10.1016/j.neuroimage.2021.118260>

5.4 Voltage-Sensitive Photoacoustic Imaging [I] (72)

<https://doi.org/10.1002/lpor.202400165>

6. MAGNETIC RESONANCE IMAGING

Hemodynamic

6.1. Blood Oxygen Dependent Levels (BOLD) [NI] (73)

<https://doi.org/10.1073/pnas.87.24.9868>
<https://doi.org/10.1038/jcbfm.2012.23>
<https://doi.org/10.1016/j.tins.2024.12.010>
<https://doi.org/10.1002/nbm.70076>
<https://doi.org/10.1523/JNEUROSCI.23-10-03963.2003>

6.1.1. Functional Connectivity MRI (fcMRI) [NI]

- <https://doi.org/10.1016/j.neuroimage.2023.120249>
<https://doi.org/10.1038/s42003-023-05629-w>
 6.1.2. Resting-State fMRI (rs-fMRI) [NI]
<https://doi.org/10.1002/jmri.28894>
- 6.2. Arterial Spin Labeling [NI] (74)
<https://doi.org/10.1002/mrm.1910230106> (first)
<https://doi.org/10.1002/mrm.29381> *
<https://doi.org/10.1073/pnas.89.1.212>
 6.2.1. Continuous Arterial Spin Labeling (CASL) [NI]
<https://doi.org/10.3389/fradi.2022.929533>
 6.2.2. Pulsed Arterial Spin Labeling (PASL) [NI]
<https://doi.org/10.3389/fradi.2022.929533>
- 6.3. Vascular-Space-Occupancy (VASO) [NI] (75)
<https://doi.org/10.1002/mrm.10519>
<https://doi.org/10.1016/j.neuroimage.2021.118868>
 6.3.1. VASO and Perfusion (VAPER) [NI] (76)
<https://doi.org/10.1016/j.neuroimage.2019.116358>
https://doi.org/10.1162/imag_a_00140
- 6.4. Laminar fMRI [NI]
<https://doi.org/10.1016/j.neuroimage.2017.02.063>
<https://doi.org/10.1093/psyrad/kkx027>
<https://doi.org/10.1016/j.cobme.2021.100288>
<https://doi.org/10.1016/j.neuroimage.2017.07.004>
- 6.5. Exogenous Labeling [I]
<https://doi.org/10.3348/kjr.2014.15.5.554>
 6.5.1. Dynamic Susceptibility Contrast (DSC)-MRI [NI/I] (77)
<https://doi.org/10.1038/jcbfm.2010.4>
<https://doi.org/10.1007/s10334-009-0190-2>
<https://doi.org/10.1038/s41598-024-58086-8> [NI]
 6.5.2. Dynamic Contrast-Enhanced (DCE)-MR [I] (78)
<http://www.ajnr.org/content/27/7/1467>
<https://doi.org/10.2174/1874440001105010090>
<https://doi.org/10.7554/eLife.89611.4>
- 6.6. Susceptibility-Weighted Imaging (SWI) [NI] (79)
<https://doi.org/10.1016/j.neuroimage.2012.01.020>
<https://doi.org/10.1016/j.neuroimage.2007.11.046>
- 6.7. Intravoxel Incoherent Motion (IVIM) MRI [NI] (80)
<https://doi.org/10.1371/journal.pone.0117706>
<https://doi.org/10.1002/nbm.3780>
<https://doi.org/10.1016/j.neuroimage.2017.12.062>
- 6.8. Phase-Contrast MRI [NI] (81)
<https://doi.org/10.1148/rg.2020190039>
<https://doi.org/10.1186/s40809-016-0019-0>
 6.8.1. Displacement Spectrum (DiSpect) Imaging (82)
<https://doi.org/10.1002/mrm.28882>
- # Non Hemodynamic**
- 6.9. Functional Magnetic Resonance Spectroscopy [NI] (83)

<https://doi.org/10.1016/j.neuroimage.2023.120194>

<https://doi.org/10.1177/0271678X221076570>

<https://doi.org/10.1002/nbm.4314>

6.9.1. ^{31}P MRS [NI]

<https://doi.org/10.1002/nbm.70043>

6.9.2. ^{13}C MRS [I]

<https://doi.org/10.1007/s11064-022-03538-8>

<https://doi.org/10.1038/s41598-019-38981-1>

6.10. Functional Quantitative Susceptibility Mapping (fQSM) [NI] (84)

<https://doi.org/10.1016/j.neuroimage.2021.117924>

6.11. ^{23}Na Sodium MRI [NI] (85)

<https://doi.org/10.1016/j.neuroimage.2018.09.071>

https://archive.ismrm.org/2025/2503_bFbEoFQ1W.html

6.12. Hyperpolarized ^{13}C Metabolic MRI [I] (86)

<https://doi.org/10.1073/pnas.1613345114>

<https://doi.org/10.1016/j.neuroimage.2022.119284>

6.13. Chemical Exchange Saturation Transfer (CEST) fMRI [NI] (87)

<https://doi.org/10.1038/s41598-019-40986-9>

<https://doi.org/10.1002/jmri.27850>

6.14. Magnetic Resonance Elastography (MRE) [NI] (88)

<https://doi.org/10.3389/fbioe.2021.666456>

<https://archive.ismrm.org/2014/0871.html>

6.15. Apparent Diffusion Coefficient fMRI [NI] (89)

<https://doi.org/10.1038/s42003-025-07889-0>

<https://doi.org/10.1038/s41467-025-60357-5>

<https://doi.org/10.3389/fnhum.2013.00817>

6.16. Molecular fMRI [I] (90)

<https://doi.org/10.1016/j.jneumeth.2021.109372>

7. LABELED OPTICAL TECHNIQUES

Indicators and Sensors

<https://doi.org/10.1146/annurev-anchem-061522-044819>

7.1. Calcium Indicators [I]

<https://doi.org/10.1111/j.1476-5381.2010.00988.x>

<https://doi.org/10.1016/j.cell.2021.12.007>

7.1.1. Calcium Indicator Dyes (Ratiometric / Nonratiometric) [I] (91)

<https://doi.org/10.1073/pnas.1507110112>

[https://doi.org/10.1016/S0143-4160\(98\)90085-9](https://doi.org/10.1016/S0143-4160(98)90085-9)

7.1.2. Genetically Encoded Calcium Indicators (GECIs) [I] (92)

<https://doi.org/10.1016/j.neures.2020.05.013> (GCaMP variants)

7.2. Voltage Indicators [I]

<https://doi.org/10.3390/colorants3040025> (General)

7.2.1. Voltage-Sensitive Dye Imaging (VSDI) (Ratiometric / Nonratiometric) [I] (93)

<https://doi.org/10.1016/j.ccllet.2020.12.060>

<https://doi.org/10.1002/lpor.202400165>

7.2.2. Genetically Encoded Voltage Indicators (GEVIs) [I] (94)

https://doi.org/10.1007/978-981-15-8763-4_12

7.3. Other Ion Indicators [I] (95)

<https://doi.org/10.1111/j.1476-5381.2010.00988.x>

<https://doi.org/10.1016/j.cell.2021.12.007>

7.3.1. Genetically Encoded Chloride Indicators (Cl-Sensor) [I]

<https://doi.org/10.1038/s41467-023-37433-9>

7.3.2. Genetically Encoded Potassium Indicators (GEPs) [I]

<https://doi.org/10.1038/s41598-024-62993-1>

7.3.3. pH-Sensitive Fluorescent Proteins [I]

<https://doi.org/10.1523/JNEUROSCI.0670-07.2007>

[https://doi.org/10.1016/S0896-6273\(04\)00144-8](https://doi.org/10.1016/S0896-6273(04)00144-8)

7.4. Second-Messenger Sensors [I] (96)

7.4.1. Genetically Encoded cAMP/PKA/Second-Messenger Sensors (e.g., Pink Flamindo, G-Flamp) [I]

<https://doi.org/10.1038/s41467-022-32994-7>

<https://doi.org/10.1016/j.celrep.2017.12.022>

7.5. Neurotransmitter/Neuropeptide Sensors [I] (97)

7.5.1. Genetically Encoded Dopamine Indicators (GRAB-DA, dLight1) [I]

<https://doi.org/10.1038/s41592-023-02100-w>

7.5.2. Genetically Encoded Glutamate Indicators (iGluSnFR) [I]

<https://doi.org/10.1038/s41592-023-01863-6>

<https://doi.org/10.1111/jnc.15608> (reviews)

7.5.3. Genetically Encoded Acetylcholine Indicators (GACH) [I]

<https://doi.org/10.1038/s41586-023-06492-9>

7.5.4. Genetically Encoded Serotonin Sensors (GRAB-5HT) [I]

<https://doi.org/10.1038/s41593-021-00823-7>

<https://doi.org/10.1038/s41592-024-02188-8>

7.5.5. Genetically Encoded Norepinephrine Sensors (GRAB-NE) [I]

<https://doi.org/10.1016/j.neuron.2019.02.037>

<https://doi.org/10.1016/j.neuron.2024.03.001>

7.5.6. Genetically Encoded GABA Indicators (iGABASnFR) [I]

<https://doi.org/10.1111/jnc.15608>

<https://doi.org/10.1016/j.ebiom.2021.103272>

<https://doi.org/10.1038/s41592-019-0471-2>

7.5.7. Genetically Encoded Histamine Sensors (GRAB-HA) [I]

<https://doi.org/10.1016/j.neuron.2023.02.024>

7.5.8. Genetically Encoded Adenosine Sensors (GRAB-Ado) [I]

<https://doi.org/10.1038/s41586-022-05407-4>

<https://doi.org/10.1038/s41467-025-59530-7>

7.5.9. Fluorescent False Neurotransmitters (FFNs) [I]

<https://doi.org/10.1021/acscchemneuro.1c00580>

<https://doi.org/10.1038/s41467-018-05075-x>

7.5.10. Synaptophysin-pHluorin (SypHy) [I]

<https://doi.org/10.1523/JNEUROSCI.0670-07.2007>

[https://doi.org/10.1016/S0896-6273\(04\)00144-8](https://doi.org/10.1016/S0896-6273(04)00144-8)

7.6. Metabolic and Other Sensors [I] (98)

7.6.1. Genetically Encoded ATP Indicators (iATPSnFR) [I]

<https://doi.org/10.1016/j.neuron.2021.11.027>

<https://doi.org/10.1038/s41586-021-03497-0>

7.6.2. Genetically Encoded Lactate Sensors [I]

<https://doi.org/10.1038/s41467-025-64484-x>

7.6.3. Genetically Encoded Nitric Oxide Sensors (geNOps) [I]

<https://doi.org/10.1016/j.celrep.2023.113514>

Microscopy Modalities

7.7. One-Photon Imaging (Miniscope / Widefield) [I] (99)

<https://doi.org/10.1016/j.conb.2011.12.002>

<https://doi.org/10.1016/j.cell.2022.02.017>

7.8. Two-Photon Microscopy [I] (100)

<https://doi.org/10.1016/j.conb.2011.12.002>

<https://doi.org/10.1016/j.cell.2022.02.017>

7.9. Three-Photon Microscopy [I] (101)

<https://doi.org/10.1038/s41583-025-00937-y>

<https://doi.org/10.1038/s42003-025-08079-8>

7.10. Fiber Photometry [I] (102)

<https://doi.org/10.1080/15476278.2025.2489667>

7.11. Mesoscopic Calcium Imaging (Widefield, not Miniscope) [I] (103)

<https://doi.org/10.3390/biology11111601>

<https://doi.org/10.3389/fnins.2023.1210199>

<https://doi.org/10.1371/journal.pone.0185759>

7.12. Light-Sheet Fluorescence Microscopy (LSFM) [I] (104)

<https://doi.org/10.1038/nmeth.2434>

<https://doi.org/10.1016/j.jneumeth.2018.07.011>

<https://doi.org/10.1016/j.conb.2018.03.007>

<https://doi.org/10.1146/annurev-neuro-070918-050357>

7.13. Swept Confocally-Aligned Planar Excitation (SCAPE) Microscopy [I] (105)

<https://doi.org/10.1038/nphoton.2014.323>

<https://doi.org/10.1038/s41587-020-0628-7>

7.14. Adaptive Optics for In Vivo Neural Imaging [I] (106)

<https://doi.org/10.3389/fnins.2022.880859>

<https://doi.org/10.3389/fnins.2023.1188614>

7.15. Oblique Plane Microscopy (OPM) [I] (107)

<https://doi.org/10.1038/s41467-023-43741-x>

7.16. Multifocal/Multibeam Two-Photon Microscopy [I] (108)

<https://doi.org/10.1364/BOE.9.003678>

<https://doi.org/10.3389/fncel.2019.00039>

<https://doi.org/10.1364/BOE.514826>

7.17. Line-Scanning Temporal Focusing Microscopy (mostTF) [I] (109)

<https://doi.org/10.1038/s41598-024-57208-6>

<https://doi.org/10.1364/BOE.9.005654>

7.18. Light Field Microscopy [I] (110)

<https://doi.org/10.1016/j.jneumeth.2021.109083>

7.19. Fluorescence Lifetime Imaging Microscopy (FLIM) [I] (111)

<https://doi.org/10.1364/OPTICA.426870>

<https://doi.org/10.1038/s41598-020-77737-0>

7.20. Total Internal Reflection Fluorescence (TIRF) Microscopy [I] (112)

<https://doi.org/10.1021/acsnano.3c04489>

- 7.21. Random Access Microscopy [I] (113)
<https://doi.org/10.1038/nmeth.4033>
<https://iopscience.iop.org/article/10.1088/2515-7647/ad2e0d>
- 7.22. Multispectral Optoacoustic Tomography (MSOT) [I] (114)
<https://doi.org/10.1016/j.pacs.2021.100285>
- 7.23. Bioluminescent Voltage Imaging (e.g., LOTUS-V) [I] (115)
<https://doi.org/10.1038/s41598-019-43897-x>
<https://doi.org/10.1117/1.nph.11.2.024203>
- 7.24. Digital Holographic Microscopy [I] (116)
<https://doi.org/10.1038/s41467-023-36889-z>
<https://doi.org/10.1016/j.conb.2018.03.006>

8. INTERFERENCE TECHNIQUES

<https://doi.org/10.3389/fnbeh.2021.820017>

Electrical Stimulation

- 8.1. Cortical Stimulation [I] (117)
<https://doi.org/10.1016/j.yebeh.2009.03.001> (1870)
- 8.2. Microstimulation [I] (118)
<https://doi.org/10.1038/s41551-024-01299-z>
<https://doi.org/10.1002/adhm.202100119>
- 8.2.1 Supracortical Microstimulation [I]
<https://doi.org/10.1146/annurev-bioeng-103023-072855>
- 8.3. Temporal Interference Simulation [NI] (119)
<https://doi.org/10.3389/fnhum.2023.1266753>
<https://doi.org/10.1016/j.cell.2017.05.024>
<https://doi.org/10.1109/EMBC46164.2021.9629968>
- 8.4. Transcranial Electrical Stimulation [NI]
<https://doi.org/10.3390/biomedicines10102333>
<https://doi.org/10.1371/journal.pbio.3001973>
- 8.4.1. Transcranial Direct Current Stimulation (tDCS) [NI] (120)
<https://doi.org/10.3389/fnhum.2025.1640565>
- 8.4.2. Transcranial Alternating Current Stimulation (tACS) [NI] (121)
<https://doi.org/10.3389/fnhum.2025.1640565>
- 8.4.3. Transcranial Random Noise Stimulation (RNS) [NI] (122)
<https://doi.org/10.1038/s41598-019-51553-7>
<https://doi.org/10.3389/fncel.2017.00162>
- 8.5. Transcranial Magnetic Stimulation [NI] (123)
<https://doi.org/10.1007/s12264-021-00781-x>
<https://doi.org/10.1146/annurev-psych-081120-013144>
- 8.5.1. Deep Transcranial Magnetic Stimulation (dTMS) [NI] (124)
<https://doi.org/10.5498/wjp.v13.i9.607>
<https://doi.org/10.1016/j.jneumeth.2021.109261>
<https://doi.org/10.1016/j.jneumeth.2020.108709>
<https://doi.org/10.1523/ENEURO.0163-17.2018>
- 8.6. Peripheral Nerve Stimulation [I]
- 8.6.1. Galvanic Vestibular Stimulation (GVS) [I] (125)

<https://doi.org/10.1152/jn.00035.2019>
<https://doi.org/10.3389/fneur.2012.00117>
[https://doi.org/10.1016/0006-8993\(82\)90990-8](https://doi.org/10.1016/0006-8993(82)90990-8) (1982)
<https://doi.org/10.1016/j.brainresbull.2004.07.008>
 8.6.2. Trigeminal Nerve Stimulation (TNS) [I] (126)
<https://doi.org/10.1186/s42234-023-00128-z>
<https://doi.org/10.3171/jns.2002.97.5.1179>
<https://doi.org/10.1007/s00221-018-5338-8>
 8.6.3. Vagus Nerve Stimulation (VNS) [I] (127)
<https://doi.org/10.1093/cercor/bhab158>
<https://doi.org/10.3390/brainsci12091137>

Optogenetics and Optogenetics-Like

<https://doi.org/10.1002/advs.202413817>
 8.7. Optogenetics [I] (128)
<https://doi.org/10.1017/S0033583523000033>
<https://doi.org/10.3389/fncel.2021.778900>
<https://doi.org/10.1007/978-981-15-8763-4>
 8.7.1. Channelrhodopsin-2 (ChR2)
 8.7.2. Halorhodopsin (NpHR)
 8.7.3. Archaeorhodopsin (Arch)
 8.7.4. Anion-conducting channelrhodopsins (ACRs)
 8.7.5. Photoswitchable Voltage-Gated Ion Channels
 8.7.6. Photoswitchable Ligand-Gated Ion Channels
 8.7.7. Optical Switch Protein Conjugates
 8.7.8. Holographic Optogenetic Stimulation [I] (129)
<https://doi.org/10.1038/s41593-021-00902-9>
<https://doi.org/10.1523/JNEUROSCI.1785-18.2018>
 8.7.9. X-ray Optogenetics [NI/I] (130)
<https://doi.org/10.1038/s41467-021-24717-1>
 8.8. Chemogenetics [I] (131)
<https://doi.org/10.1177/10738584221134587>
<https://doi.org/10.1002/glia.24390>
<https://doi.org/10.1523/JNEUROSCI.0625-23.2023>
<https://doi.org/10.1038/s41593-020-0661-3>
 8.9. Mechano/Sonogenetics [NI] (132)
<https://doi.org/10.1016/j.brs.2022.09.002>
<https://doi.org/10.1002/anie.202317112>
 8.10. Magnetogenetics [NI] (133)
<https://doi.org/10.15252/emboj.201797177>
<https://doi.org/10.1186/s12951-024-02616-z>
<https://doi.org/10.1038/s41565-024-01694-2>
<https://doi.org/10.1038/s41467-021-25837-4>
<https://doi.org/10.7554/eLife.27069>
<https://doi.org/10.1038/s41563-022-01281-7>
 8.11. Thermogenetics [NI/I] (134)
<https://doi.org/10.1002/advs.202413817>
 8.12. Upconversion Nanoparticle-Based Neural Imaging [I] (135)

<https://doi.org/10.1126/science.aag1144>
<https://doi.org/10.1016/j.biomaterials.2017.07.017>

Other Interference Techniques

- 8.12. Cortical Cooling [I] (136)
<https://doi.org/10.3389/fnsys.2011.00053>
[https://doi.org/10.1016/0006-8993\(77\)90734-X](https://doi.org/10.1016/0006-8993(77)90734-X)
- 8.13. Lidocaine Inactivation [I] (137)
[https://doi.org/10.1016/S0165-0270\(97\)02229-2](https://doi.org/10.1016/S0165-0270(97)02229-2)
- 8.14. Muscimol Inactivation [I] (138)
<https://doi.org/10.1016/j.jneumeth.2008.01.033>
- 8.15. Transcranial Focused Ultrasound [NI] (139)
<https://doi.org/10.3389/fnhum.2021.749162>
<https://doi.org/10.1371/journal.pone.0288654>
<https://doi.org/10.1007/s13534-024-00369-0>
- 8.15.1. Focused Ultrasound Blood–Brain Barrier Opening (FUS-BBBO) [NI] (140)
<https://doi.org/10.1002%2Fmms.101>
<https://doi.org/10.1148/radiol.14140245>
<https://doi.org/10.1093/brain/awab460>
<https://doi.org/10.1038/s41598-018-25904-9>
- 8.16. Optoacoustic Neuromodulation [I] (141)
<https://doi.org/10.1038/s41467-020-14706-1>
- 8.17. Infrared Neural Stimulation [NI/I] (142)
<https://doi.org/10.1073/pnas.2015685118>
<https://doi.org/10.1038/s41598-021-89163-x>
<https://doi.org/10.1016/j.brs.2023.01.1678>
- 8.18. Transcranial Photobiomodulation (tPBM) [NI] (143)
<https://doi.org/10.1038/s41598-019-42693-x>
<https://doi.org/10.1126/sciadv.abq3211>
<https://doi.org/10.1364/BOE.402047>
- 8.19 Photothermal Neuromodulation (non-genetic) [I] (144)
<https://doi.org/10.1002/admi.202400873>
<https://doi.org/10.1021/acsnano.4c01037>
- 8.20. Iontronic microfluidic probes / microfluidic interconnection [I] (145)
<https://doi.org/10.1038/s41378-021-00295-6>
<https://doi.org/10.1002/sml.202410906>
- 8.21. Neuromodulation via Magnetic Nanodiscs [I] (146)
<https://doi.org/10.1038/s41565-024-01798-9>
<https://doi.org/10.3389/fnhum.2025.1489940>
- 8.22. Terahertz Neural Control [NI] (147)
<https://doi.org/10.7554/eLife.97444.3>
DOI: [10.4103/NRR.NRR-D-23-00872](https://doi.org/10.4103/NRR.NRR-D-23-00872)

9. Miscellaneous Techniques

(or they are a mix/interface of the above or not encompassed by the above classification)

- 9.1. Thermal Diffusion Flowmetry [I] (148)
<https://doi.org/10.1089/neu.2018.6309>
- 9.2. Magnetic Particle Imaging (MPI) [I] (149)

<https://doi.org/10.18416%2FIJMPI.2020.2009009>

<https://doi.org/10.1016/j.neuroimage.2018.05.004>

9.3. Near-Infrared Fluorescence Imaging of Hemodynamics [I] (150)

<https://doi.org/10.1371/journal.pone.0048383>

9.4. Second Harmonic Generation (SHG) Microscopy [I] (151)

<https://doi.org/10.1073/pnas.1004748107> (*pivotal, not brain*)

<https://doi.org/10.1021/acsphotonics.9b01749> (*brain dynamics*)

ADDITIONS:

1. Diffuse Correlation Spectroscopy

from <https://doi.org/10.1111/micc.12884>

2. Laminar Functional Magnetic Resonance Imaging

from <https://doi.org/10.1093/psyrad/kkae027>

3. Phase-Contrast Based Approaches MRI

from <https://doi.org/10.1002/mrm.28882>

4. Thermal Diffusion Flowmetry

<https://doi.org/10.1089/neu.2018.6309>

from <https://doi.org/10.1111/micc.12884>

EXCLUDED after recheck:

1. Deep Label-Free Microscopy (DLFM)

<https://doi.org/10.1038/nm.3495> (after rechecking its time frames)

2. Microvascular Volumetric Pulsatility Mapping

<https://doi.org/10.1038/s44161-025-00722-1>

<https://doi.org/10.3389/fnagi.2025.1486775>

<https://doi.org/10.1177/0271678X20980652>

(the literature does not show functional studies to the moment)

3. Magnetic Resonance Fingerprinting (MRF)

<https://doi.org/10.1002/jmri.29812> (It is not a technique per se, but more a “pipeline” optimization method. EXTREMELY interesting and useful, but here excluded)
