

## Mapping of Raw files to their short names Mapping source: file (user-defined) (automatic shortening of names was not sufficiently short – see 'best effort')

original	short name	
Toni_20140521_GM_QC_01	file 1	521_GM_QC_01
Toni_20140521_GM_QC_02	file 2	521_GM_QC_02
Toni_20140522_GM_QC_01	file 3	522_GM_QC_01
Toni_20140531_FB_QC_02	file 4	531_FB_QC_02
Toni_20140608_FB_qc_01	file 5	608_FB_qc_01

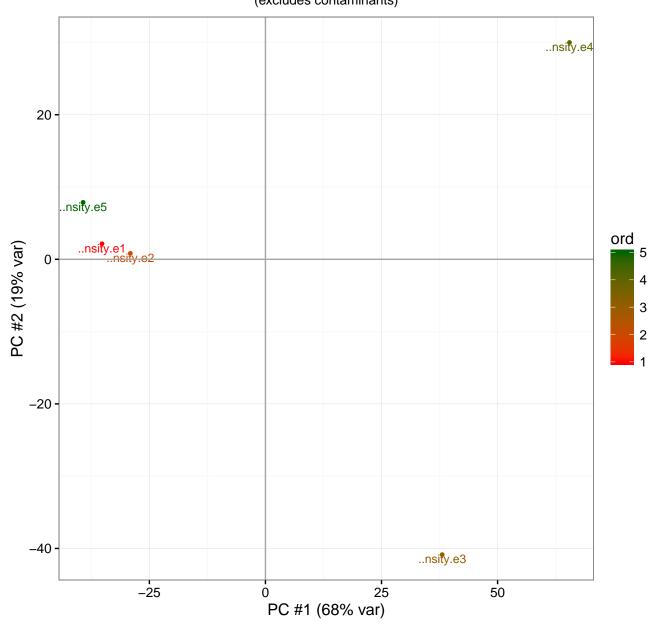
### PAR: parameters

parameter	value	parameter	value
Advanced ratios	False	MS/MS deisotoping (ITMS)	False
Alignment time window [min]	100	MS/MS deisotoping (TOF)	False
Cut peaks	True	MS/MS deisotoping (Unknown)	False
Decoy mode	revert	MS/MS recalibration	False
Discard unmodified counterpa	True	MS/MS tol. (FTMS)	20 ppm
Find dependent peptides	False	MS/MS tol. (ITMS)	0.5 Da
First pass AIF correlation	0.8	MS/MS tol. (TOF)	0.1 Da
Fixed modifications	Carbamidomethyl (C)	MS/MS tol. (Unknown)	0.5 Da
iBAQ	False	Peptides used for protein qu	Razor
iBAQ log fit	False	Protein FDR	0.01
Include contaminants	True	PSM FDR	0.01
Labeled amino acid filtering	True	Re-quantify	True
Match between runs	True	RT shift	False
Matching time window [min]	1	Site FDR	0.01
Min. delta score for modifie	17	Site quantification	Use least modified peptide
Min. delta score for unmodif	0	Site tables	Oxidation (M)Sites.txt
Min. peptide Length	7	Special AAs	KR
Min. peptides	1	Top MS/MS peaks per 100 Da	12
Min. ratio count	2	Top MS/MS peaks per 100 Da	8
Min. razor peptides	1	Top MS/MS peaks per 100 Da	10
Min. score for modified pept	40	Top MS/MS peaks per 100 Da	10
Min. score for unmodified pe	0	Use delta score False	
Min. unique peptides	0	Use Normalized Ratios For Oc	True
Modifications included in pr	Acetyl (Protein N-term) Oxidation (M)	Use only unmodified peptides	True
MS/MS deisotoping (FTMS)	True	Version	1.4.1.2

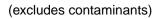
uniprot\_human\_canonical\_and\_isoforms\_20130513.fasta

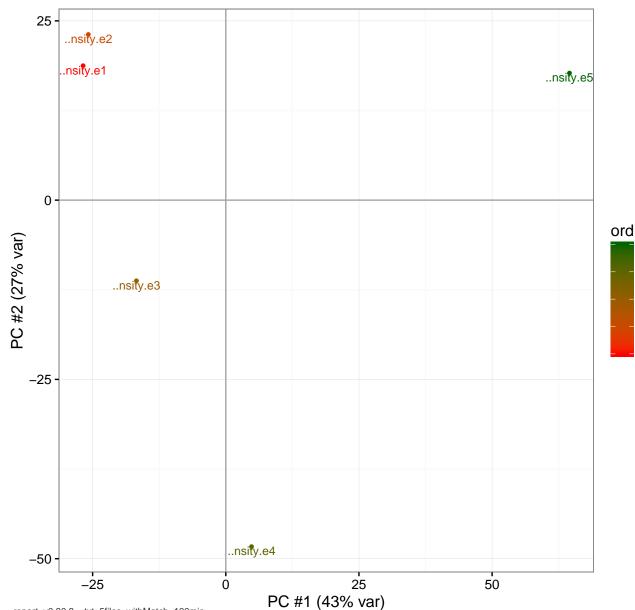
PG: PCA of 'raw intensity'





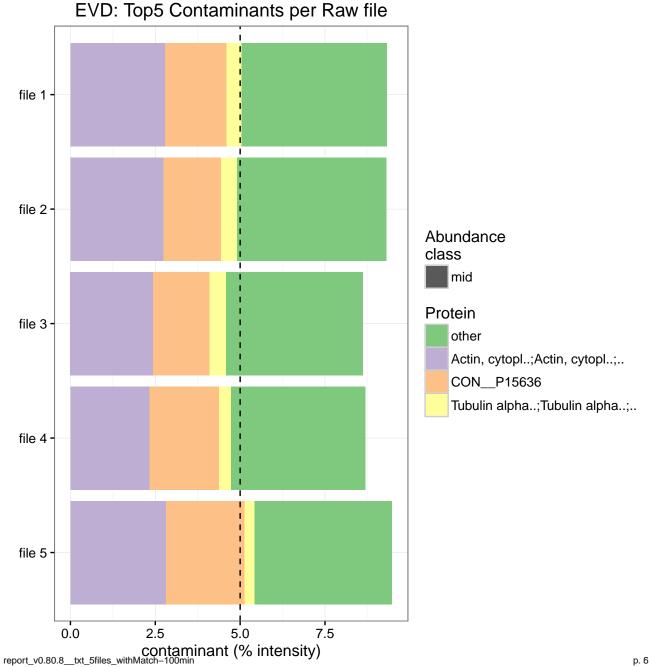
# PG: PCA of 'Ifq intensity'





5

3

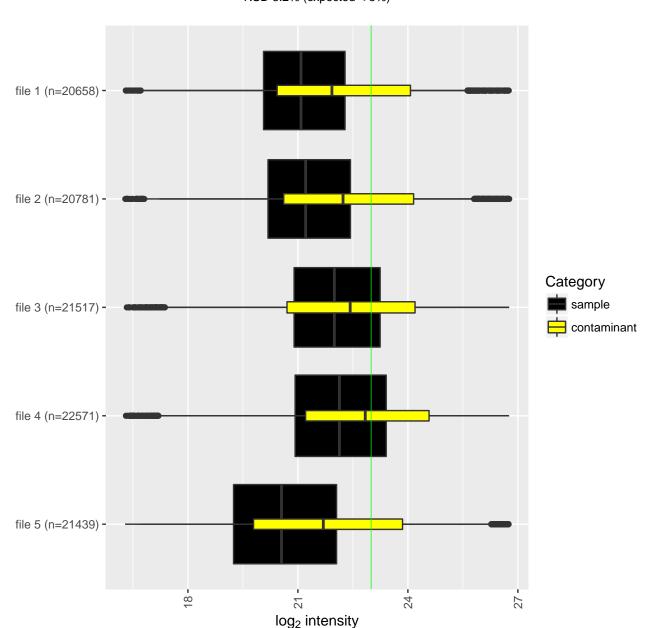


#### **EVD: Contaminants**

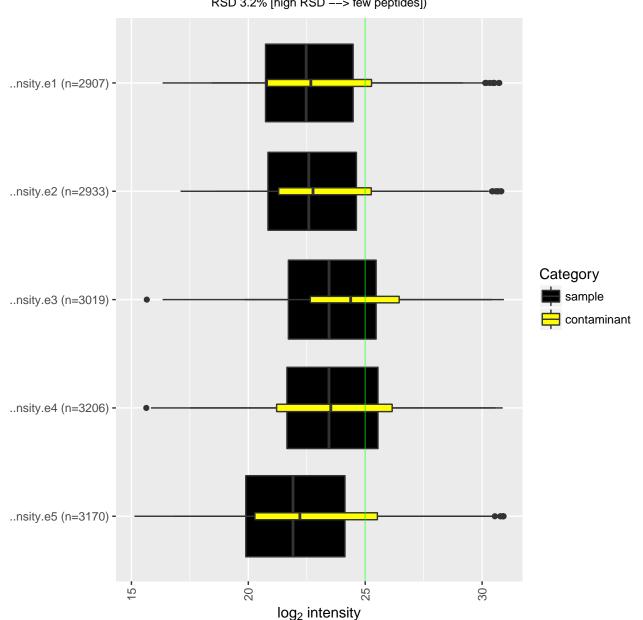
Contaminant 'MYCOPLASMA' was not found in any sample.

Did you use the correct database?

#### EVD: peptide intensity distribution RSD 3.2% (expected < 5%)

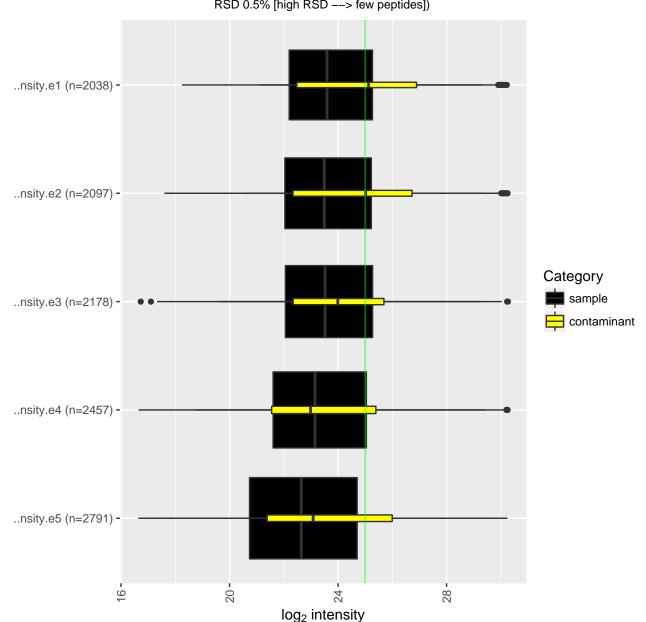


PG: intensity distribution
RSD 3% (w/o zero int.; expected < 5%)
RSD 3.2% [high RSD ---> few peptides])

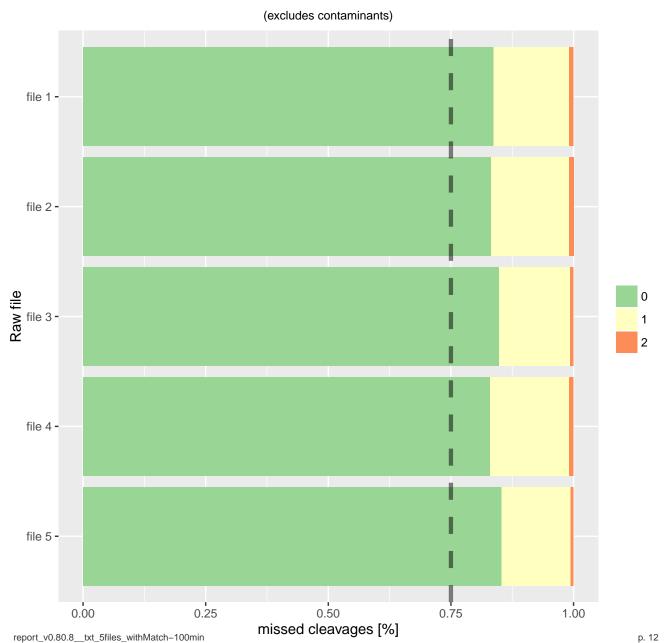


# PG: LFQ intensity distribution RSD 1.7% (w/o zero int.; expected < 5%)

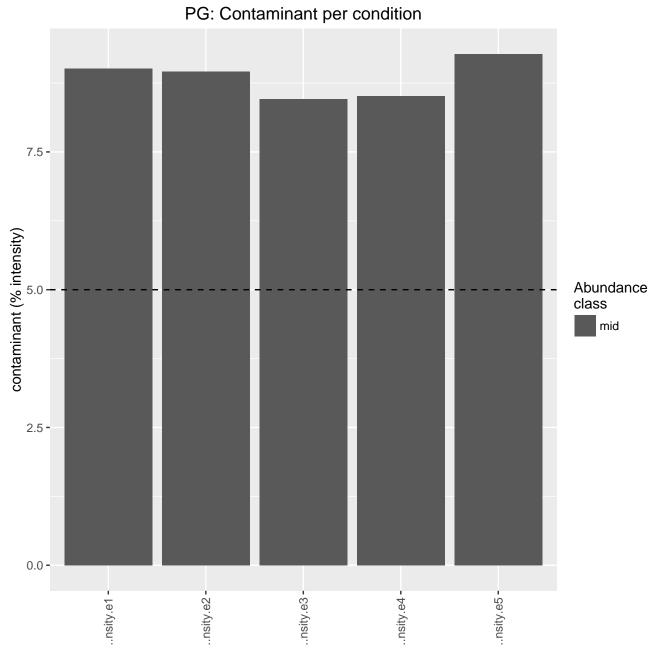
RSD 0.5% [high RSD --> few peptides])

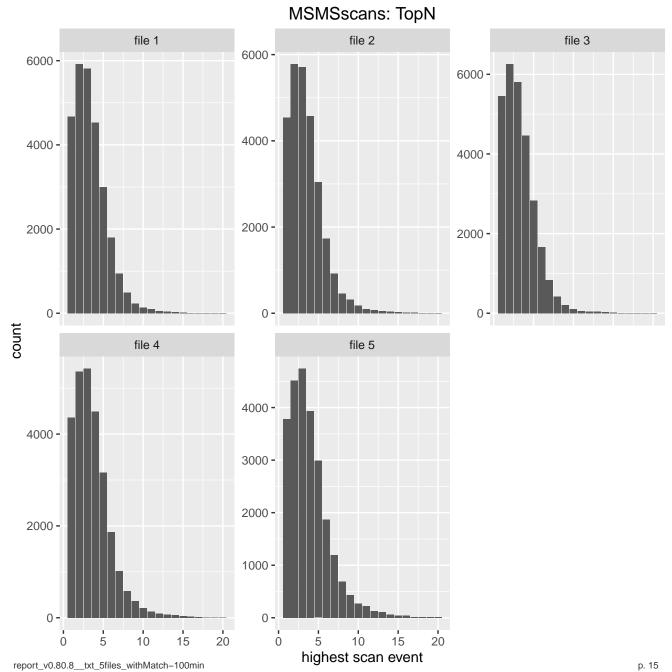


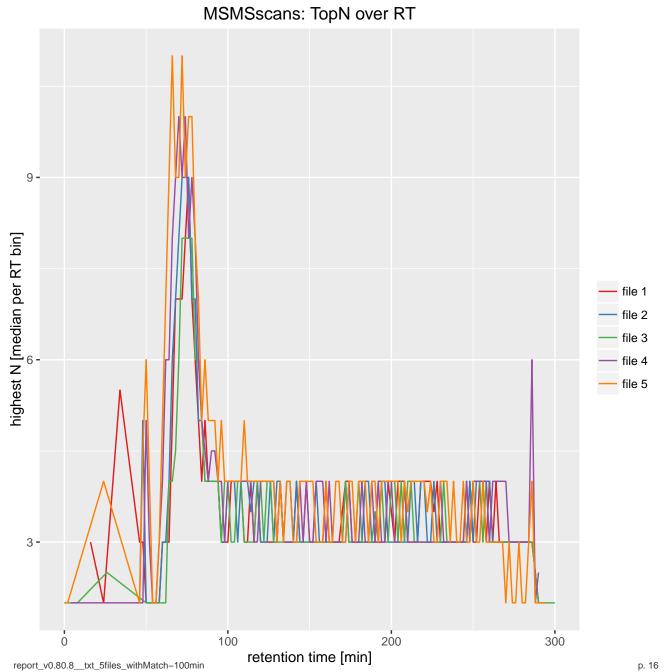
# MSMS: Missed cleavages per Raw file

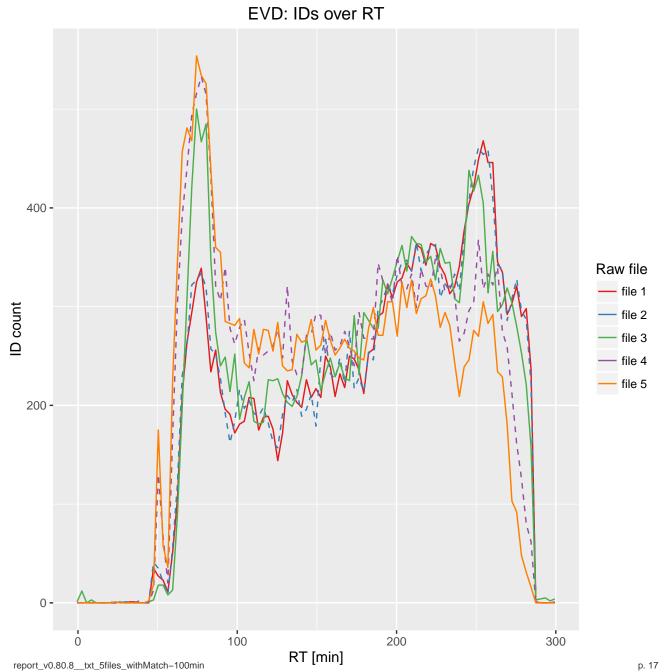


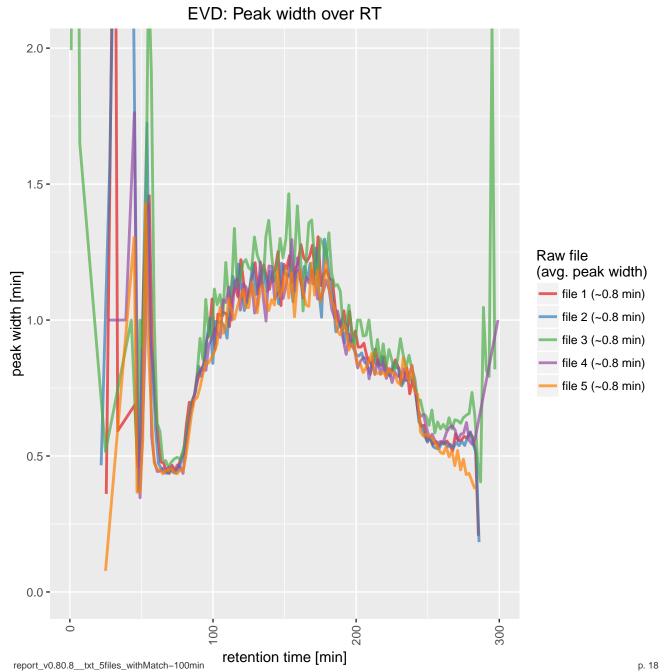
EVD: charge distribution file 1 file 2 charge 2 Raw file file 3 3 4 5 6 7 file 4 file 5 0.50 0.25 0.75 1.00 0.00 fraction [%] p. 13 report\_v0.80.8\_\_txt\_5files\_withMatch-100min





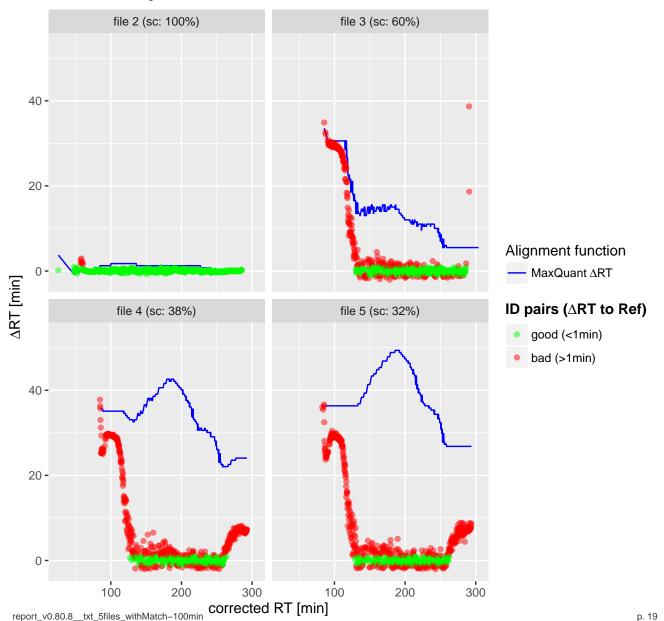




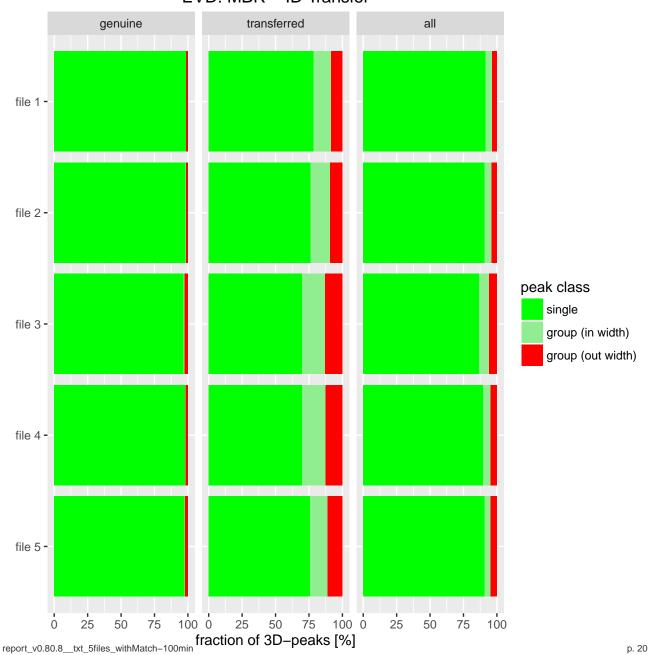


# EVD: MBR – alignment

alignment reference: Toni\_20140521\_GM\_QC\_01

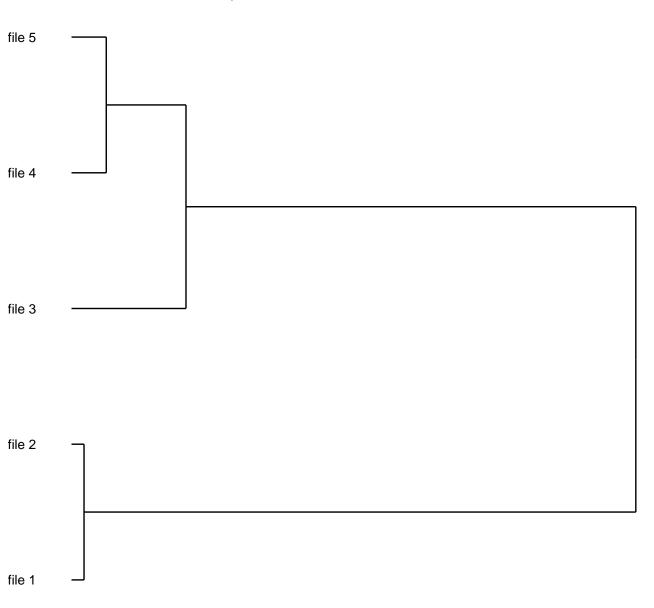


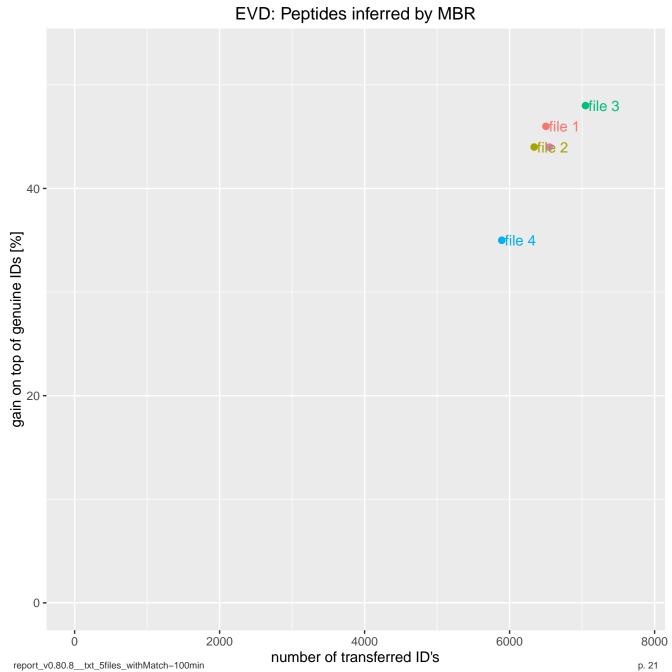
#### EVD: MBR - ID Transfer

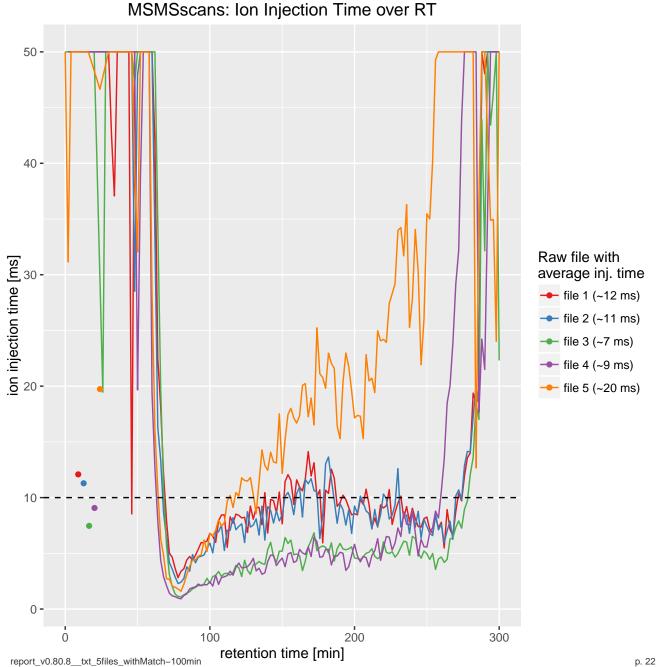


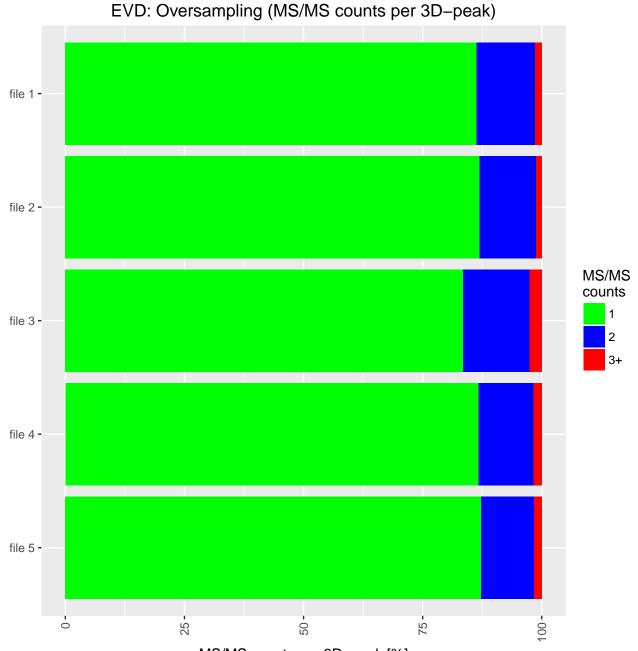
# [experimental] EVD: Clustering Tree of Raw files

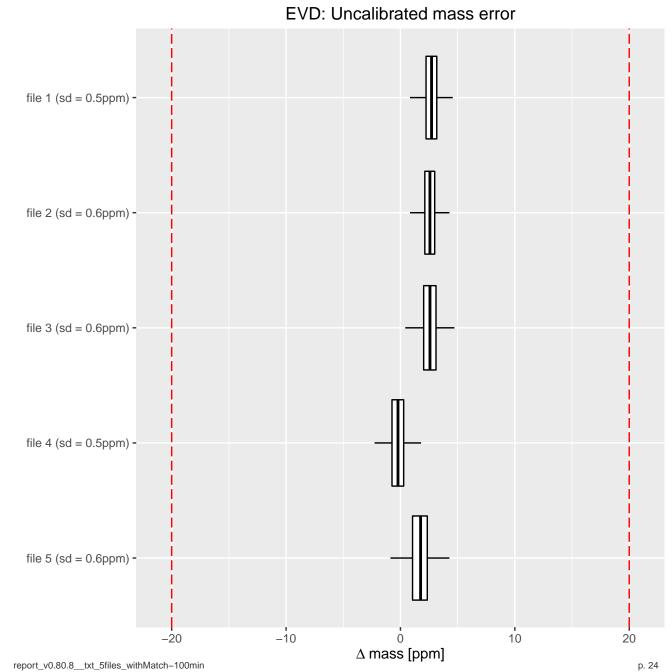
by Correlation of Corrected Retention Times

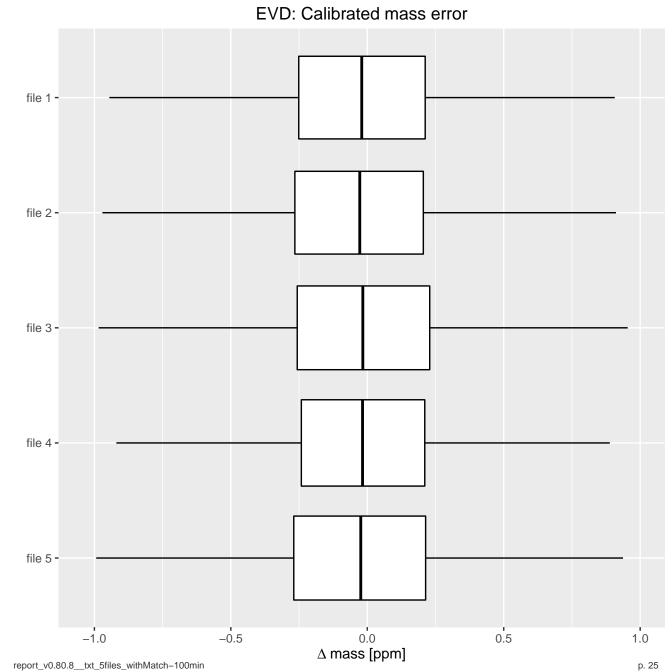




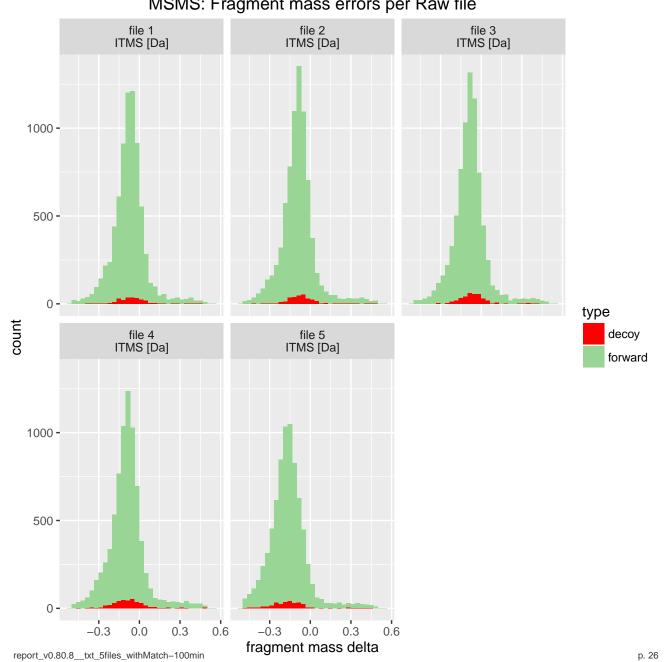


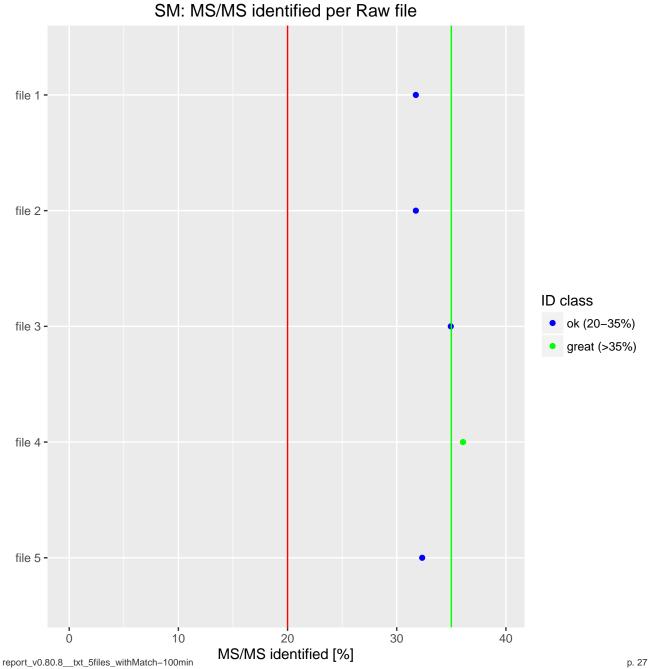




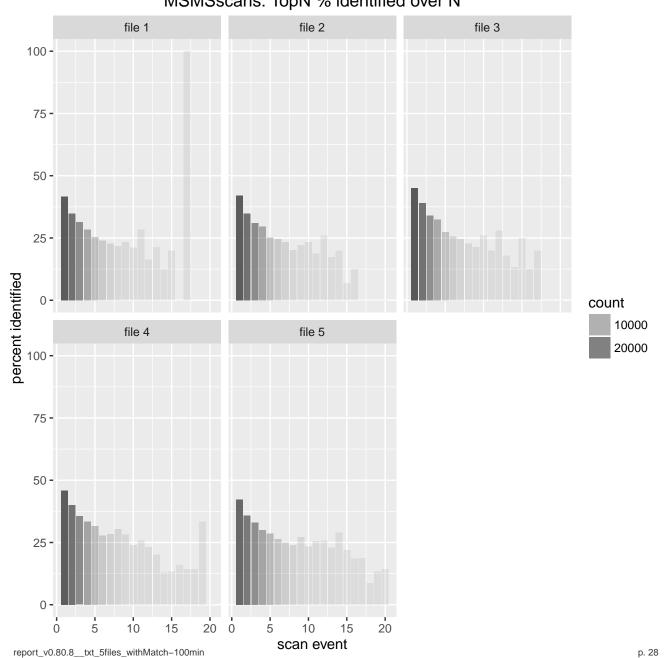


MSMS: Fragment mass errors per Raw file



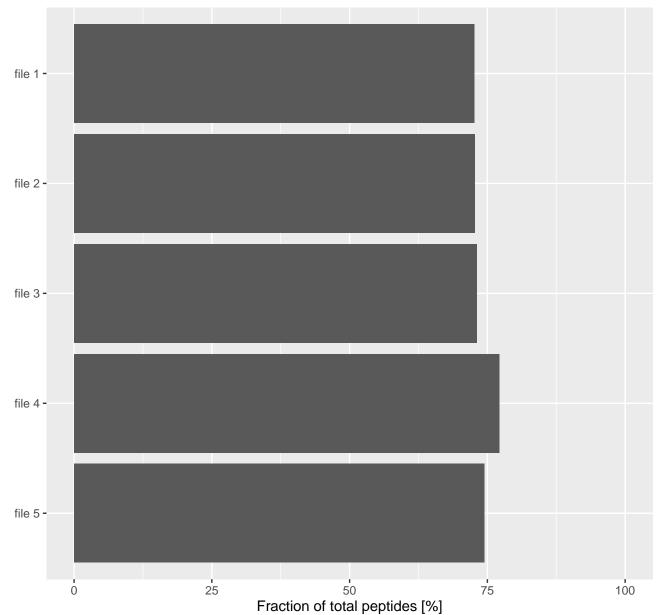


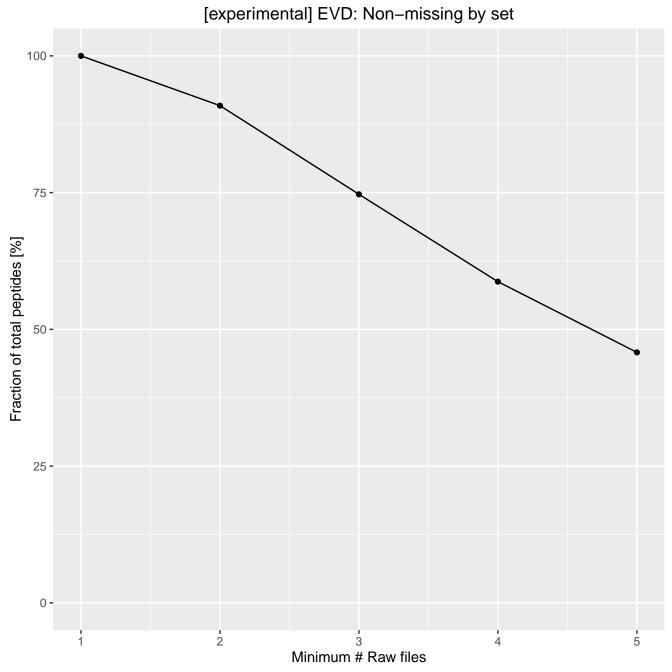
MSMSscans: TopN % identified over N

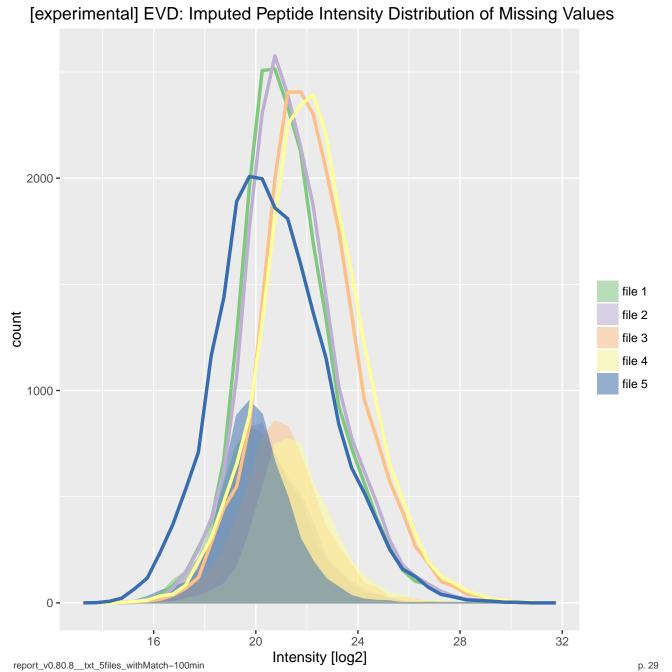


# [experimental] EVD: Non-Missing Peptides

compared to all peptides seen in experiment

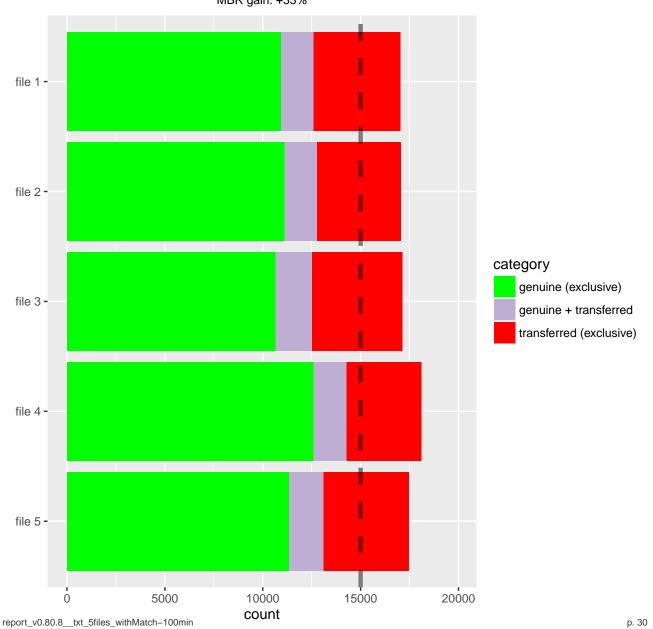






#### EVD: Peptide ID count

MBR gain: +33%



### EVD: ProteinGroups count

