

Data importation

Time point objects

Gentotypic layout

1. endpoint

2. S\_timeseries

Raw data

1. Detection of outliers for single observations

2. Correction for spatial trends

3. Outlier detection for series of observations

4. With the cleaned data, re-do the spatial correction

# SPPU Data Analysis Timepoints

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2024-06-09

Set the right working directory.

```
rm(list = ls())

setwd("C:/Users/elise/Documents/Mémoire/Main/Data/Templates/SPPU")
platform <- "SPPU"
```

## Data importation

Reimport the data sets extracted from the Data Preparation and Data Analysis R Markdown.

```
list.files()
```

```
## [1] "endpoint.txt"      "plant_info.txt"    "S_timeseries.txt"  "T_timeseries.txt"
## [5] "timeseries.txt"
```

```

plant_info <- read.table("plant_info.txt", header = TRUE, sep = "\t")
endpoint <- read.table("endpoint.txt", header = TRUE, sep = "\t")
S_timeseries <- read.table("S_timeseries.txt", header = TRUE, sep = "\t")

# plant_info
plant_info <- lapply(plant_info, factor)

# S_timeseries
matching_cols <- intersect(names(S_timeseries), names(plant_info))
S_timeseries[, matching_cols] <- lapply(S_timeseries[, matching_cols], factor)
S_timeseries$Timestamp <- as.POSIXct(S_timeseries$Timestamp, format = "%Y-%m-%d %H:%M:%S")
S_timeseries$Date <- date(S_timeseries$Date)

platform <- "SPPU"

# S_timeseries
df_S_timeseries <- S_timeseries[, colSums(is.na(S_timeseries)) < nrow(S_timeseries)]
genotype_index <- which(colnames(df_S_timeseries) == "Genotype")
variables_S <- colnames(df_S_timeseries[, c(5:(genotype_index - 1))]) # We remove the three first columns that are "Unit.ID", "Time" and "Date"

print(paste(platform, ": The variables for S_timeseries are", paste(variables_S, collapse = ", ", sep = " ")))

```

```
## [1] "SPPU : The variables for S_timeseries are S_Height_cm, S_Height_pixel, S_Area_cm_squared, S_Area_pixel, S_Perimeter_cm, S_Perimeter_pixel, S_Compactness, S_Width_cm, S_Width_pixel"
```

```
S_timeseries$Plant_type <- substr(S_timeseries$Genotype, nchar(as.character(S_timeseries$Genotype)), nchar(as.character(S_timeseries$Genotype)))
```

Get the cleaned endpoint data

```

# We add a Plant_type variable that is H or L, with T being L
S_timeseries$Plant_type <- substr(S_timeseries$Genotype, nchar(as.character(S_timeseries$Genotype)), nchar(as.character(S_timeseries$Genotype)))

S_timeseries$Plant_type <- ifelse(S_timeseries$Plant_type %in% c("T", "L"), "Line",
                                ifelse(S_timeseries$Plant_type == "H", "Hybrid", S_timeseries$Plant_type))

```

## Time point objects

Generation of the timePoints objects using the function "createTimePoints".

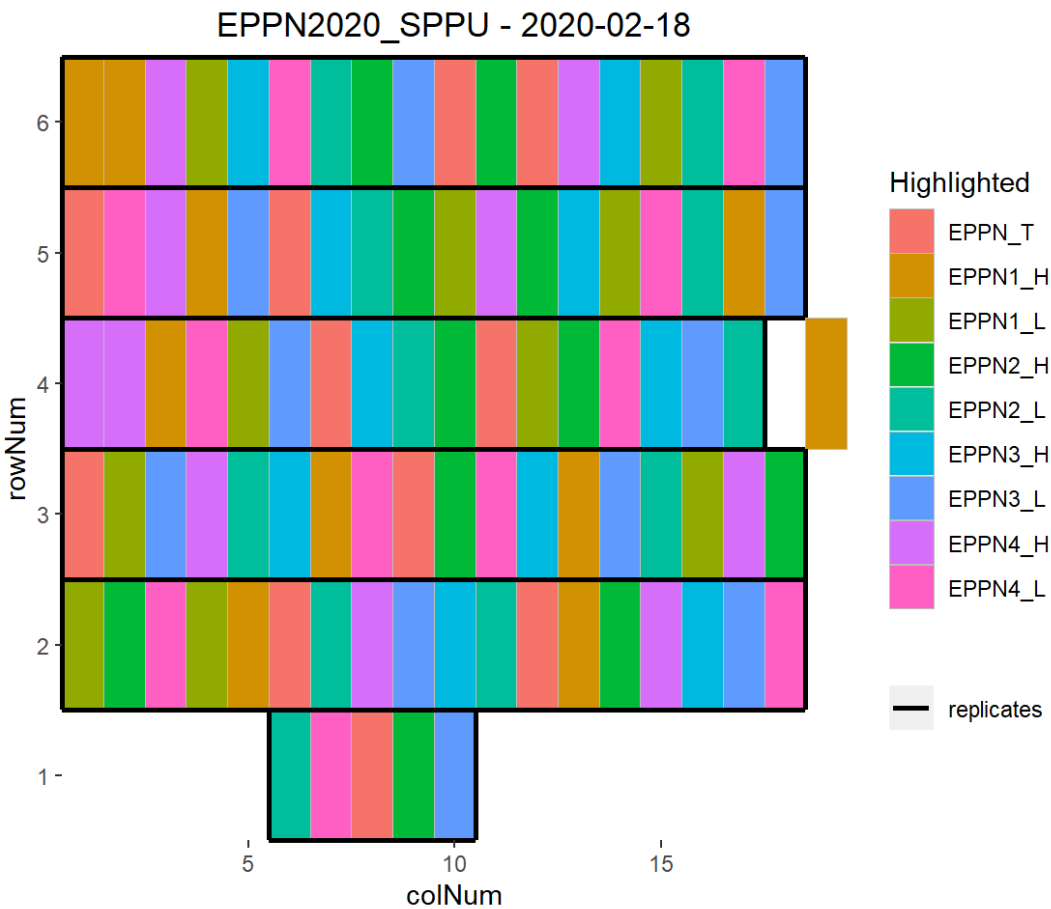
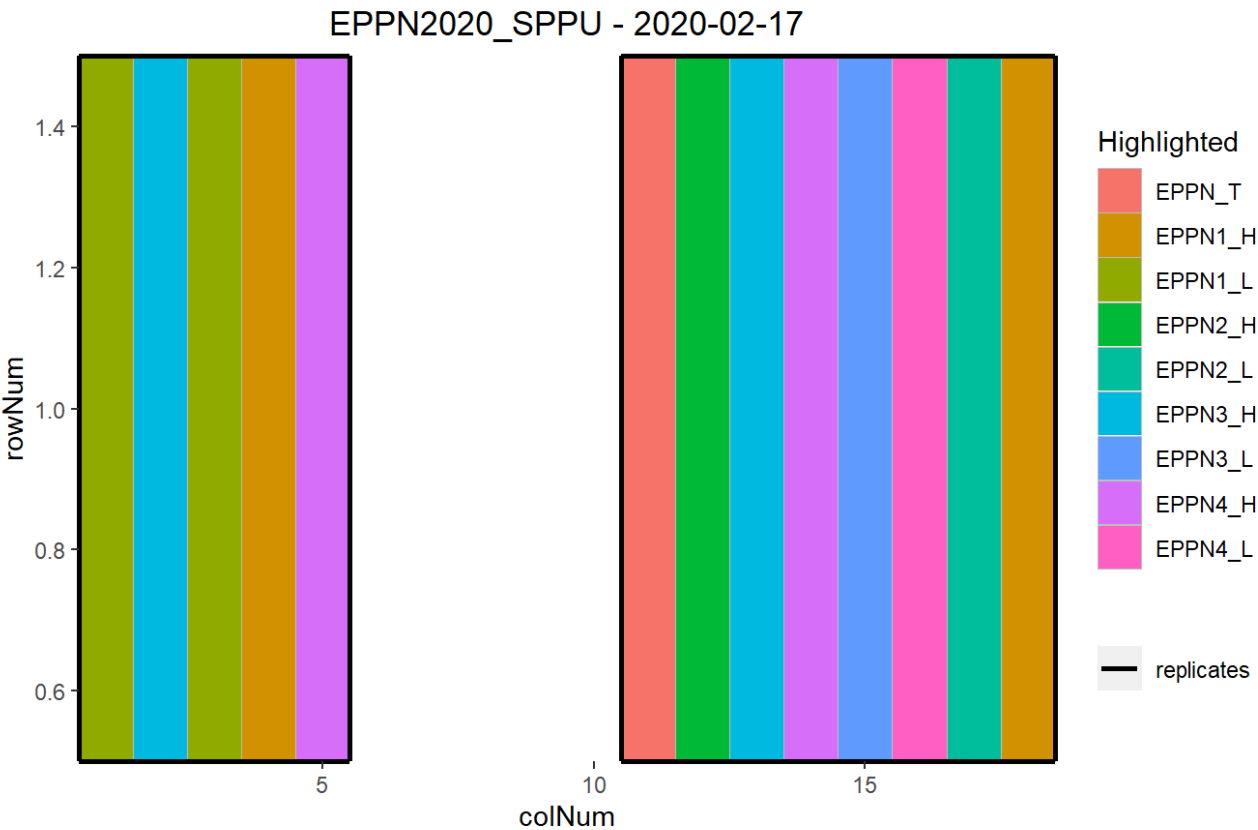
```
timePoint_S <- createTimePoints(dat = S_timeseries,  
                                experimentName = "EPPN2020_SPPU",  
                                genotype = "Genotype",  
                                timePoint = "Date",  
                                plotId = "Unit.ID",  
                                rowNum = "Row",  
                                colNum = "Column",  
                                repId = "Replication",  
                                addCheck = TRUE,  
                                checkGenotypes = "EPPN_T")
```

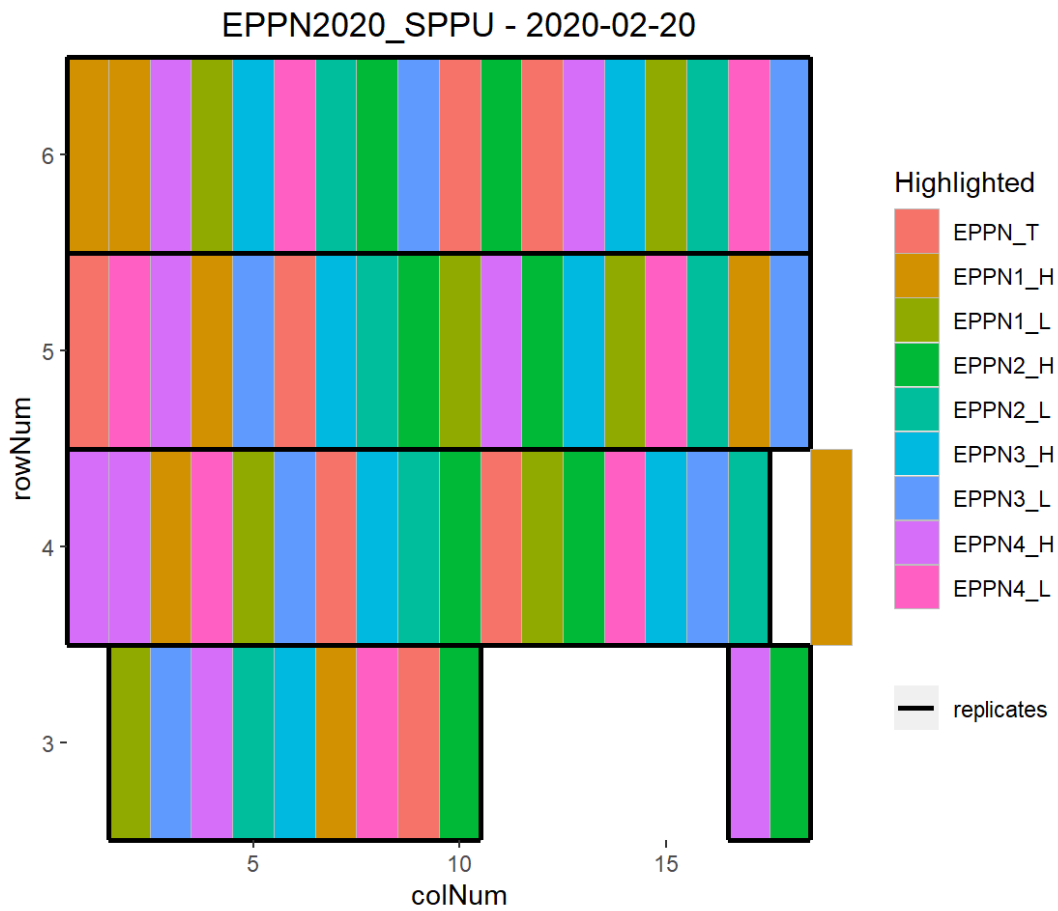
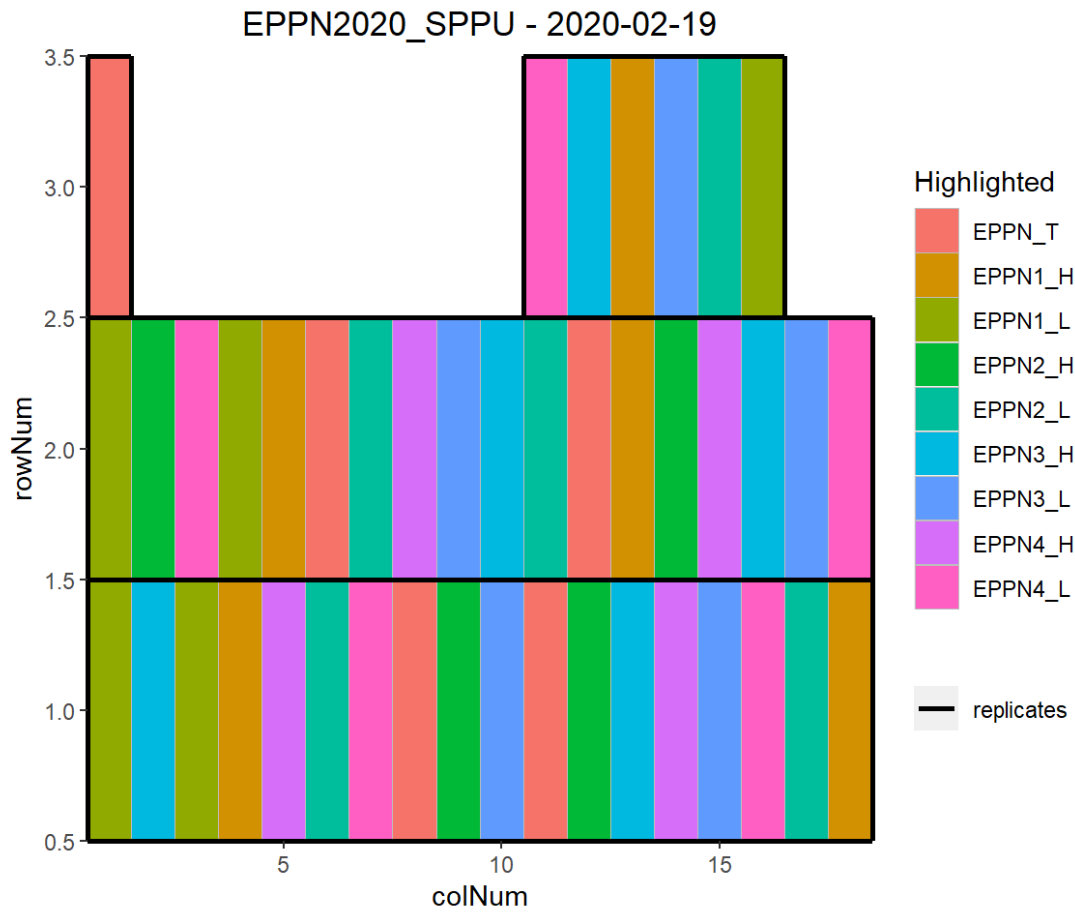
```
## Warning: More than 5 plotIds have observations for less than 50% of the time points.  
The first 5 are printed, to see them all run  
## attr(..., 'plotLimObs') on the output  
## 102, 103, 104, 105, 106
```

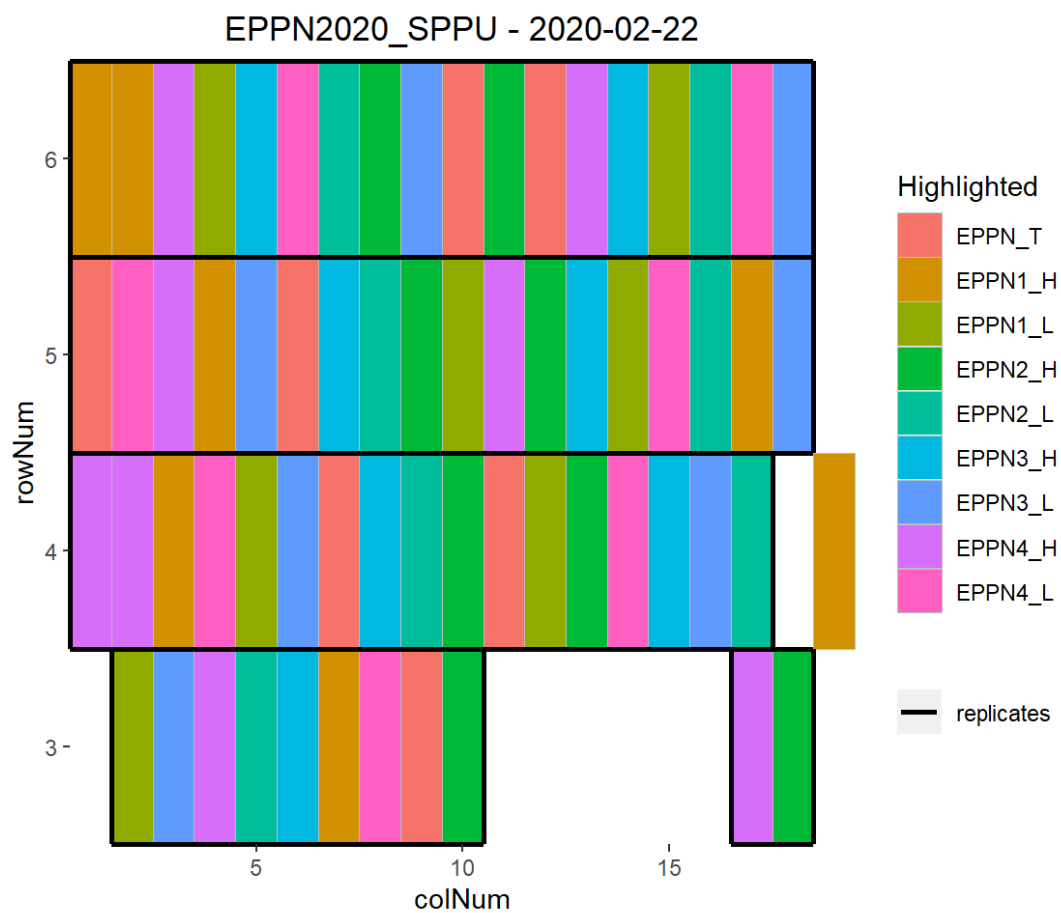
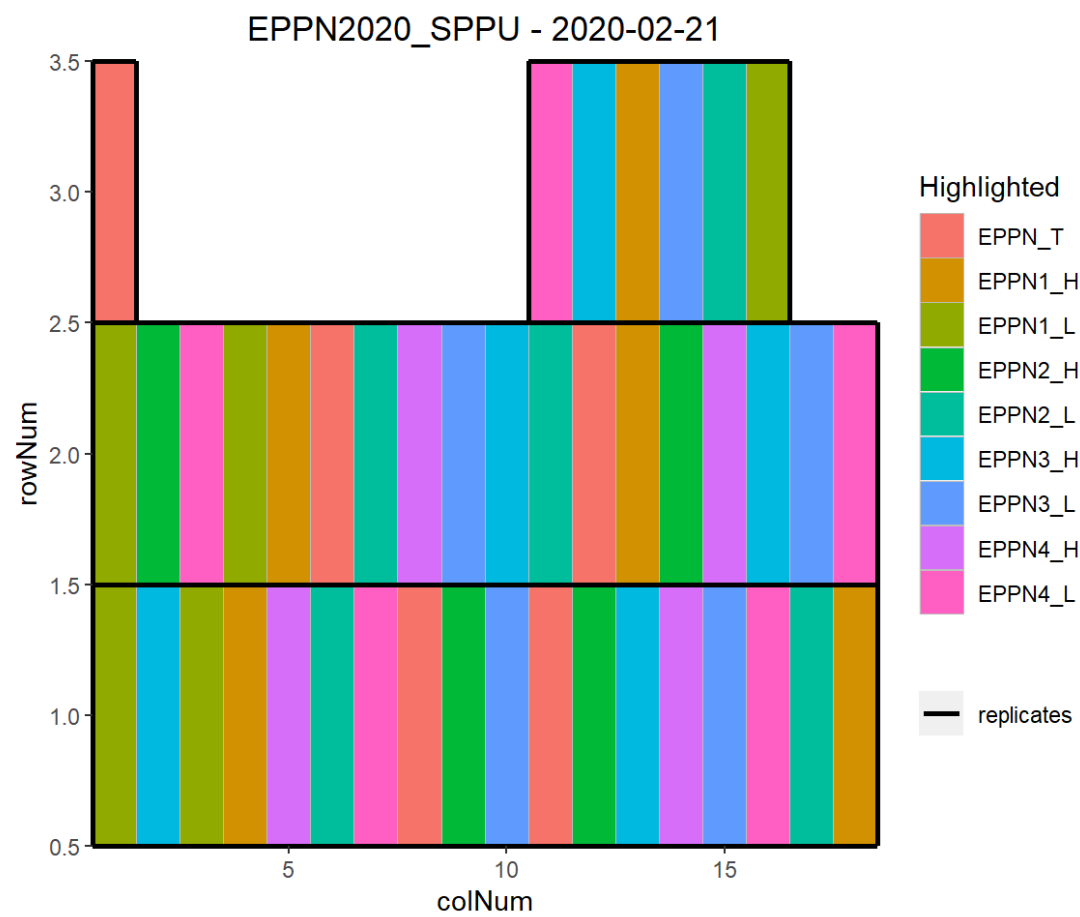
## Gentotypic layout

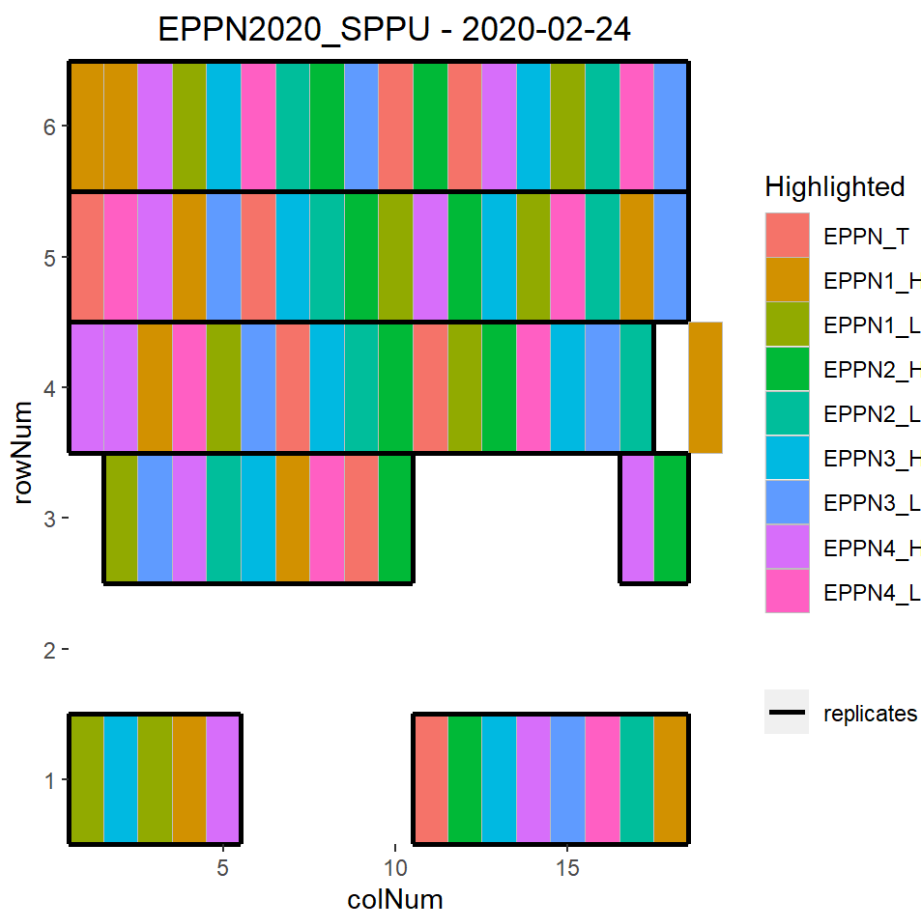
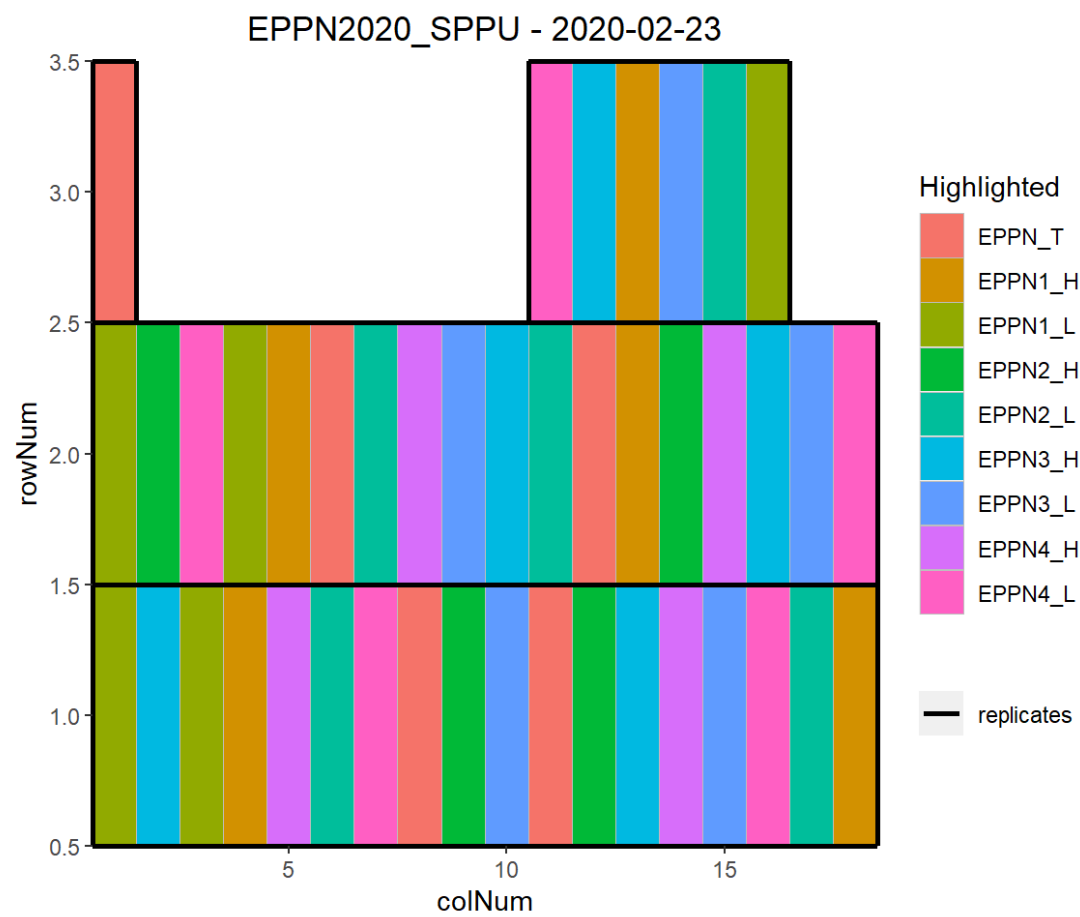
Check the layout of the platforms' genotypes.

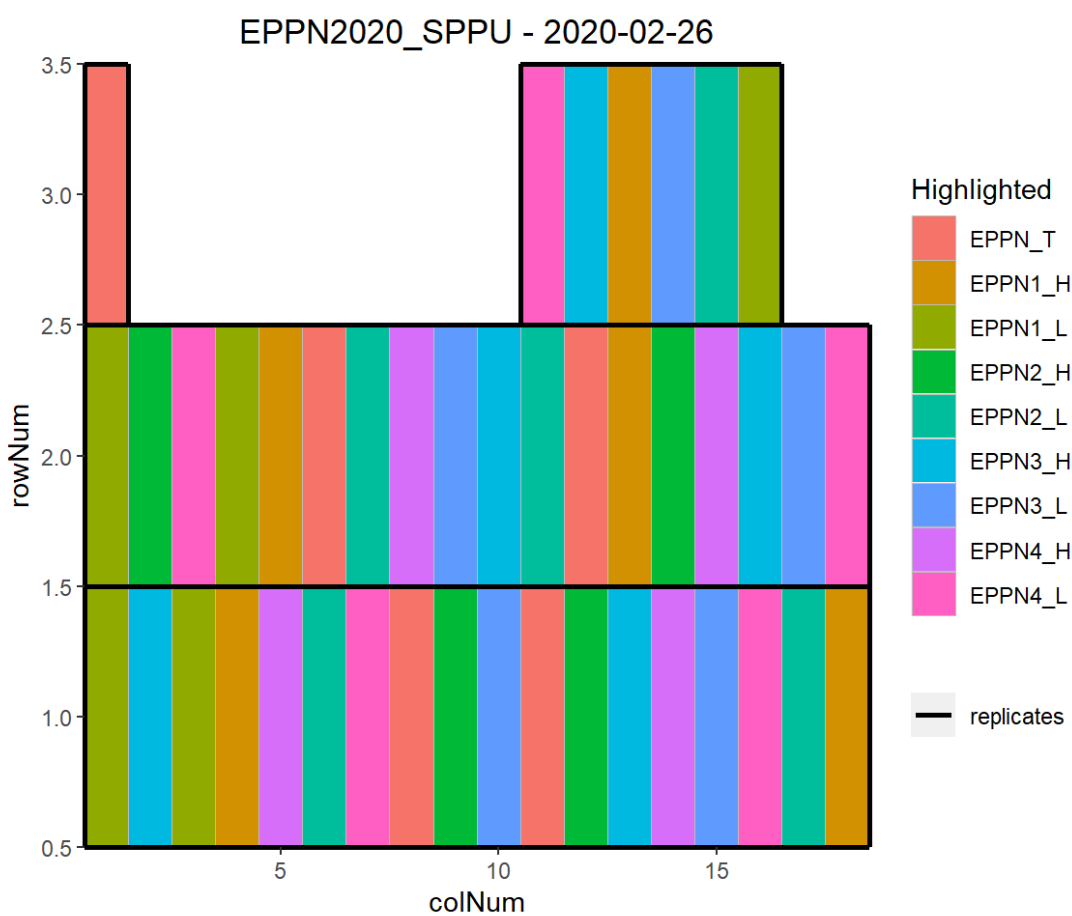
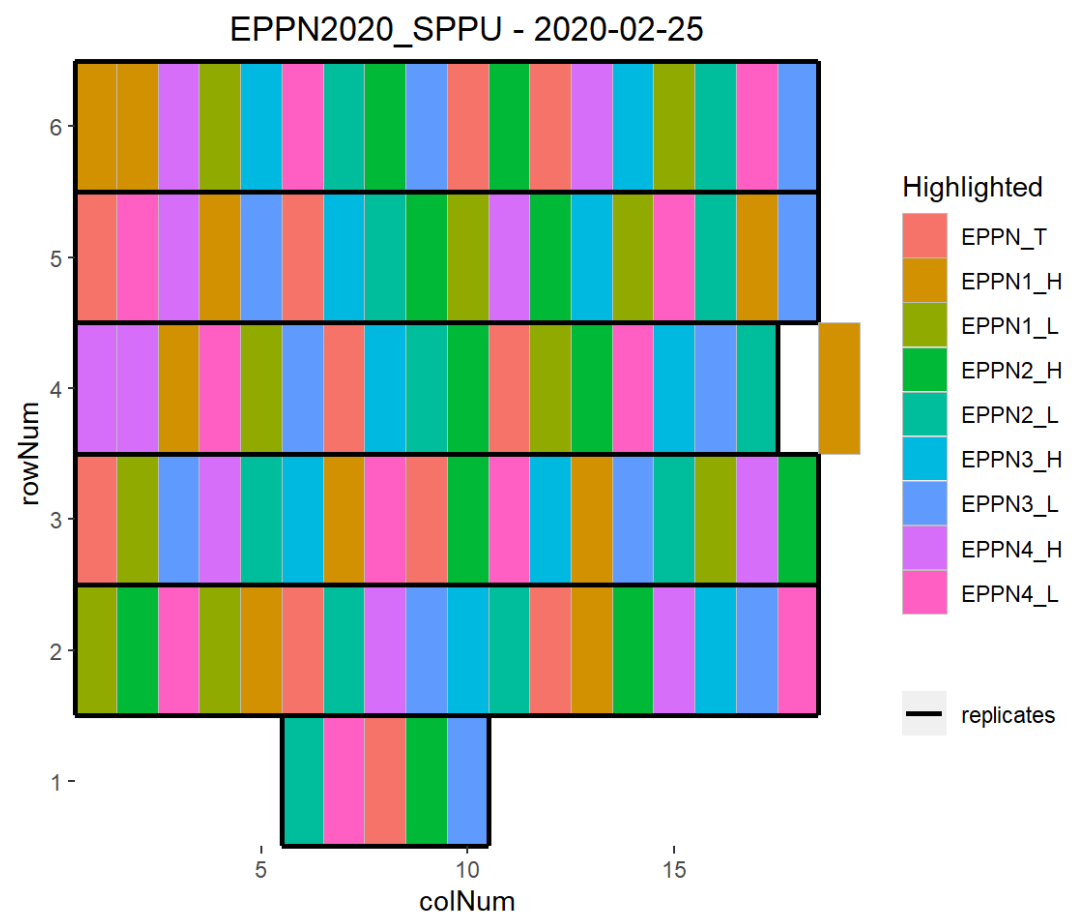
```
genotypes_list <- as.character(unique(S_timeseries$Genotype))  
  
plot(timePoint_S,  
      plotType = "layout",  
      highlight = genotypes_list,  
      showGeno = FALSE)
```



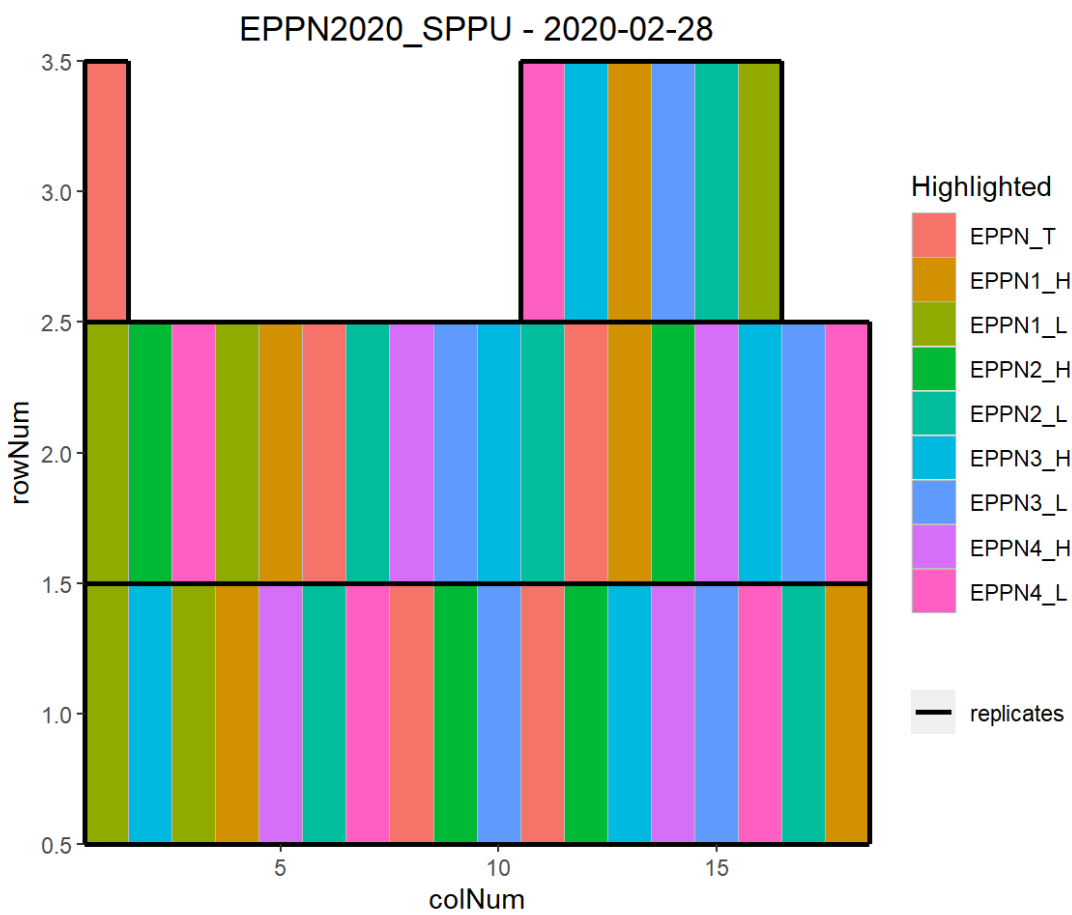
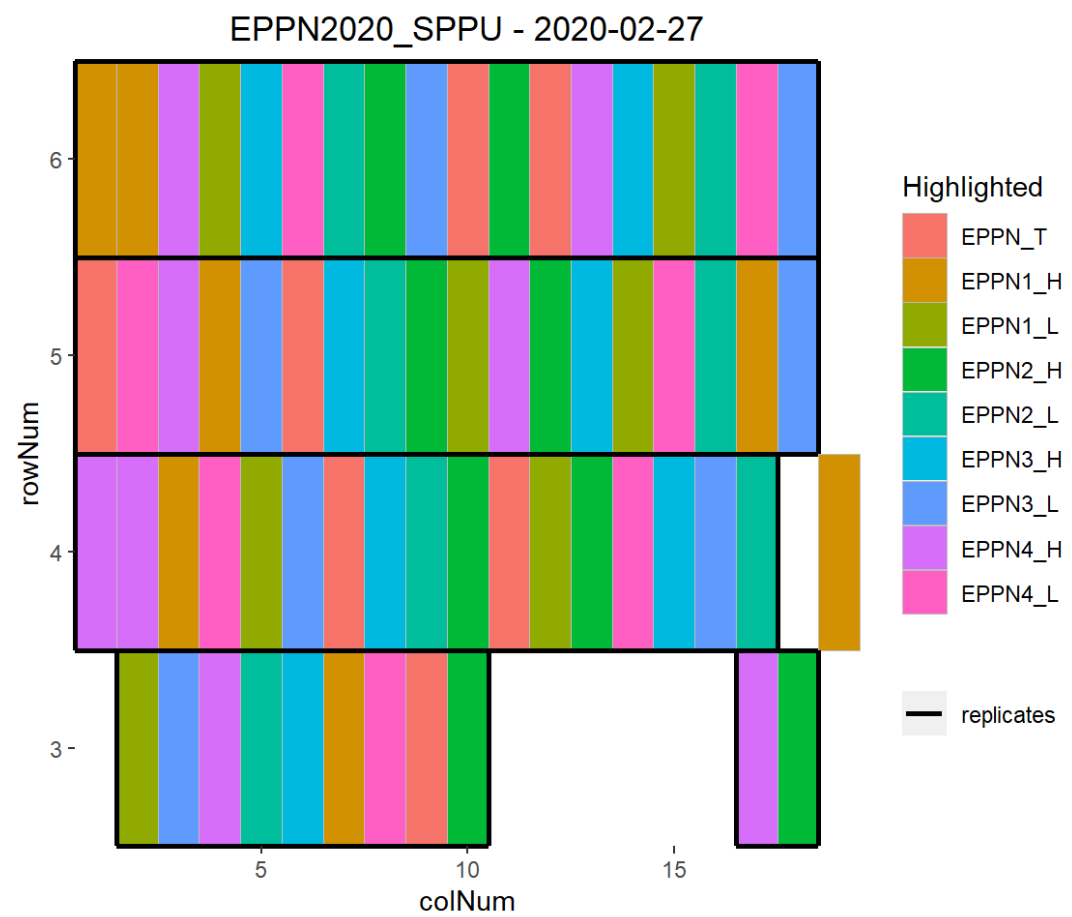


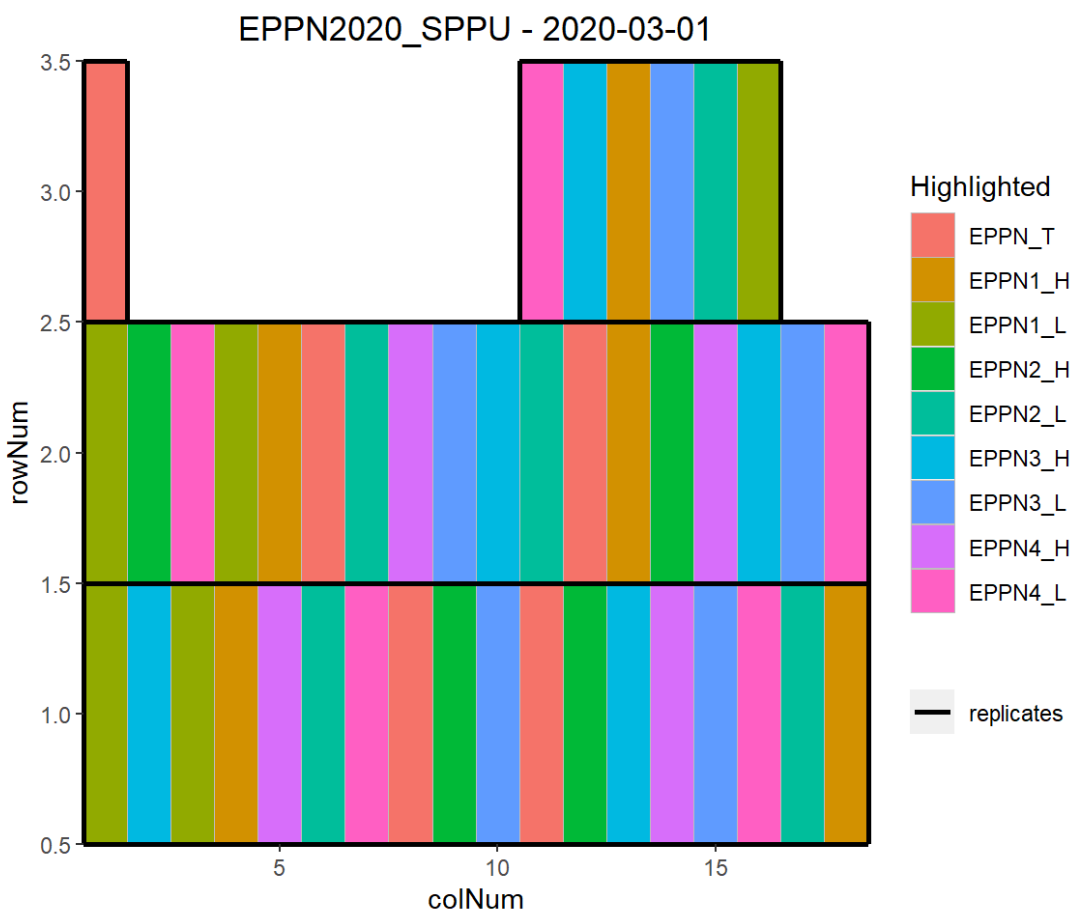
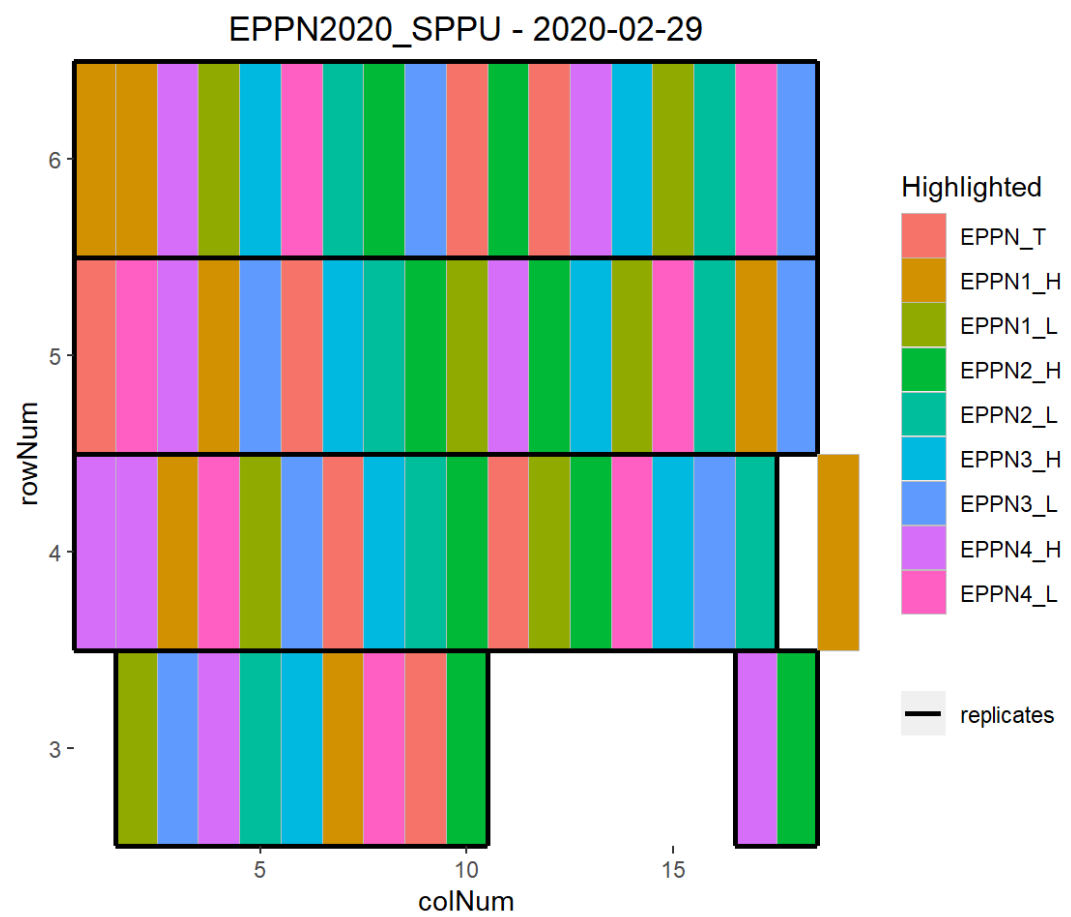


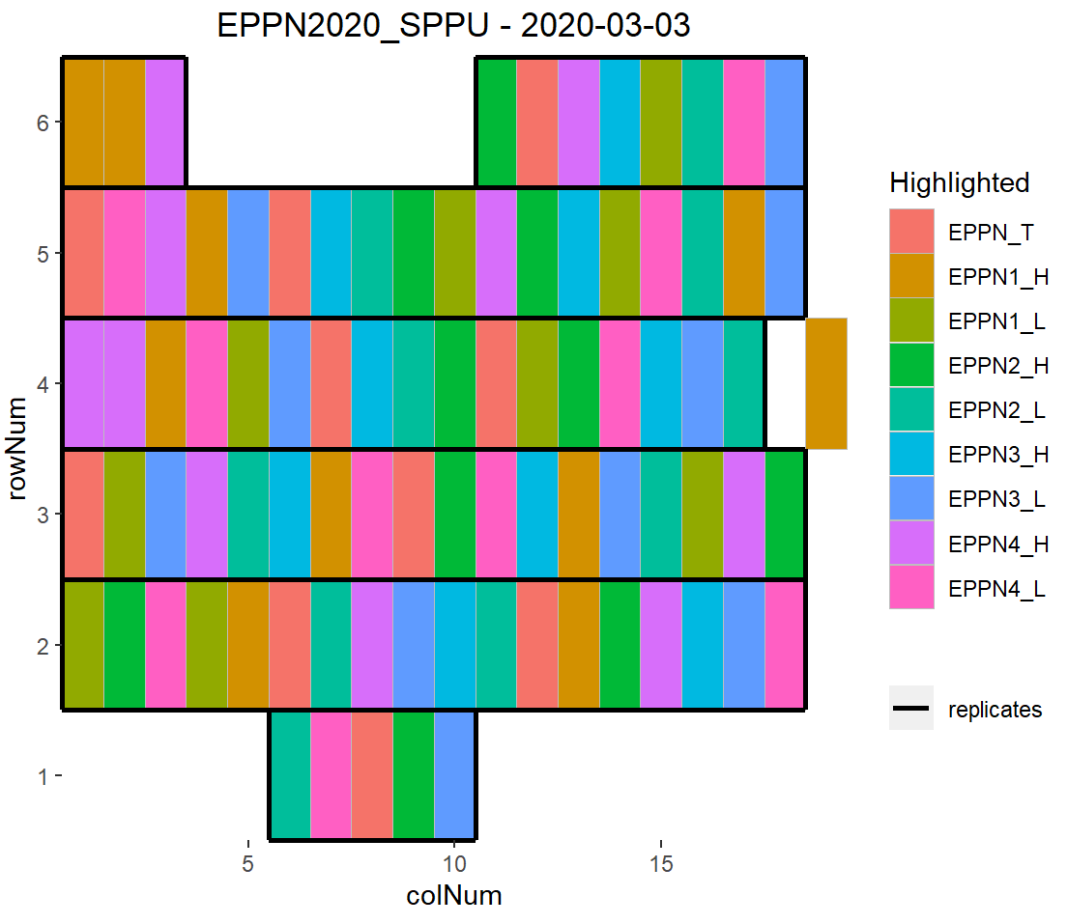
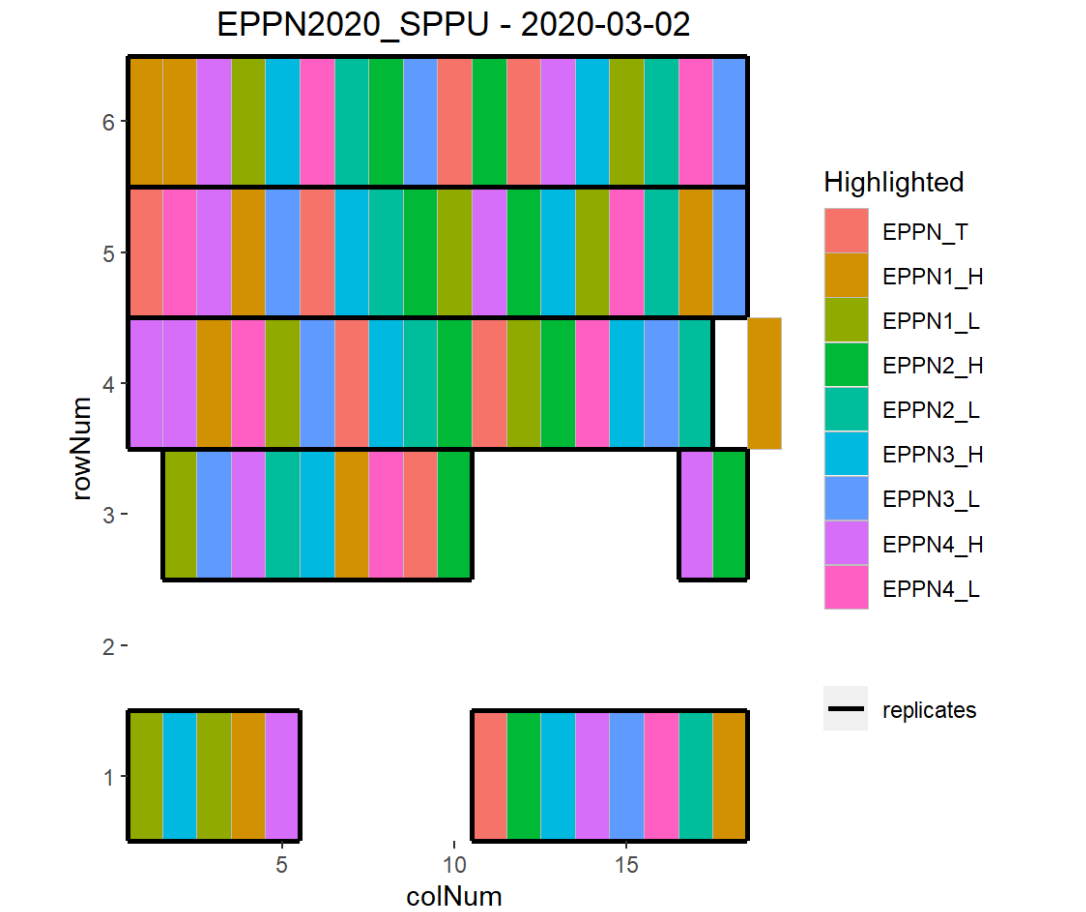


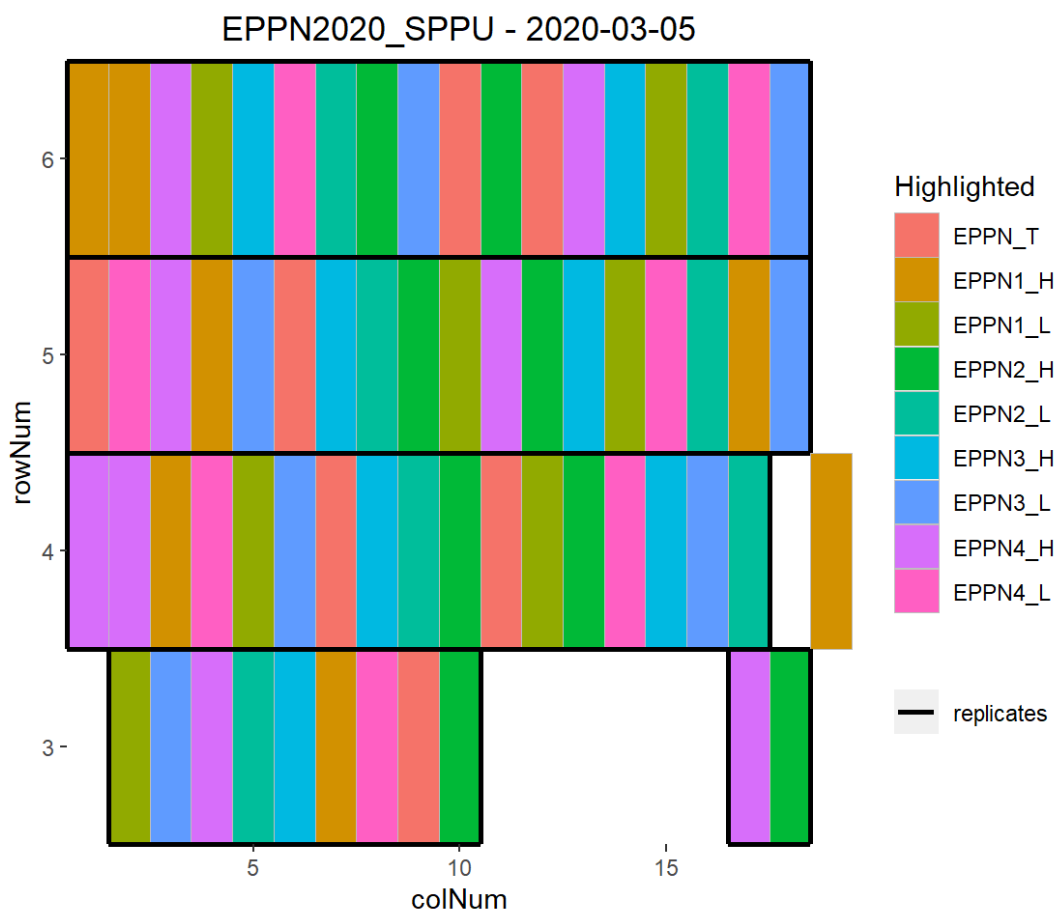
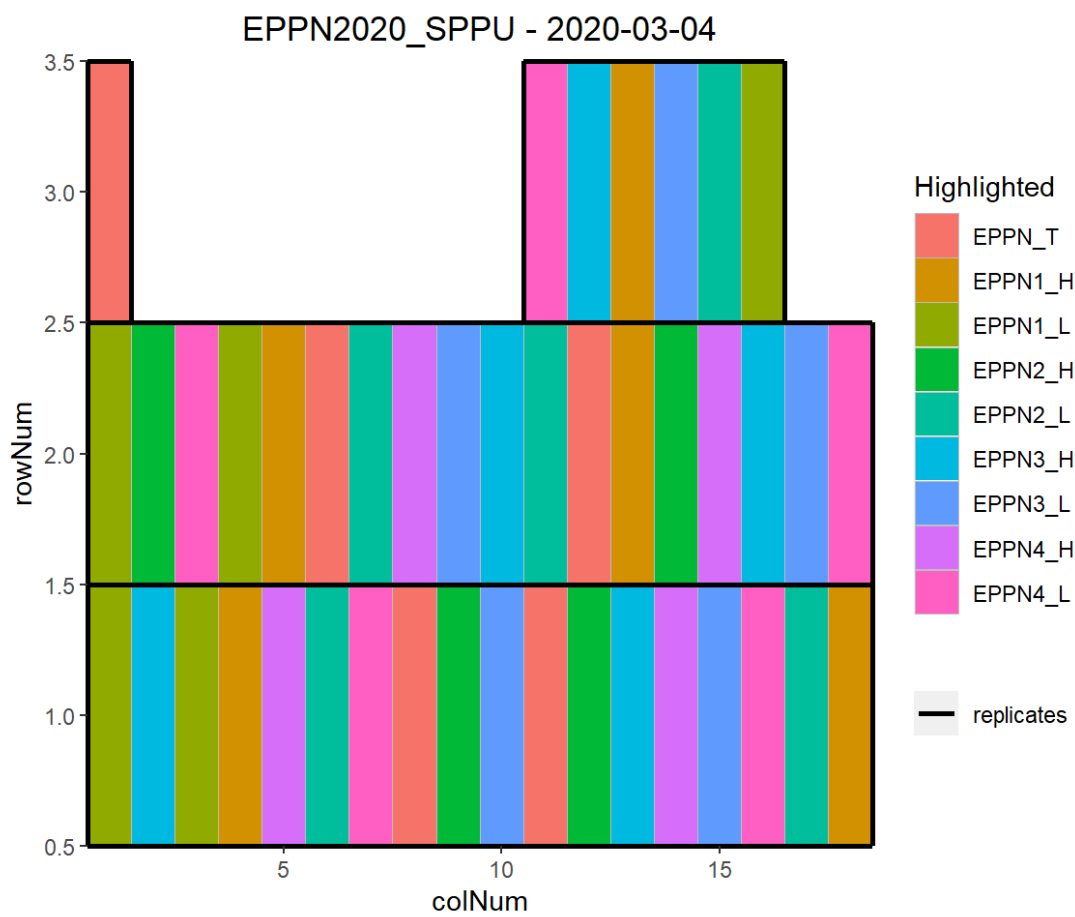


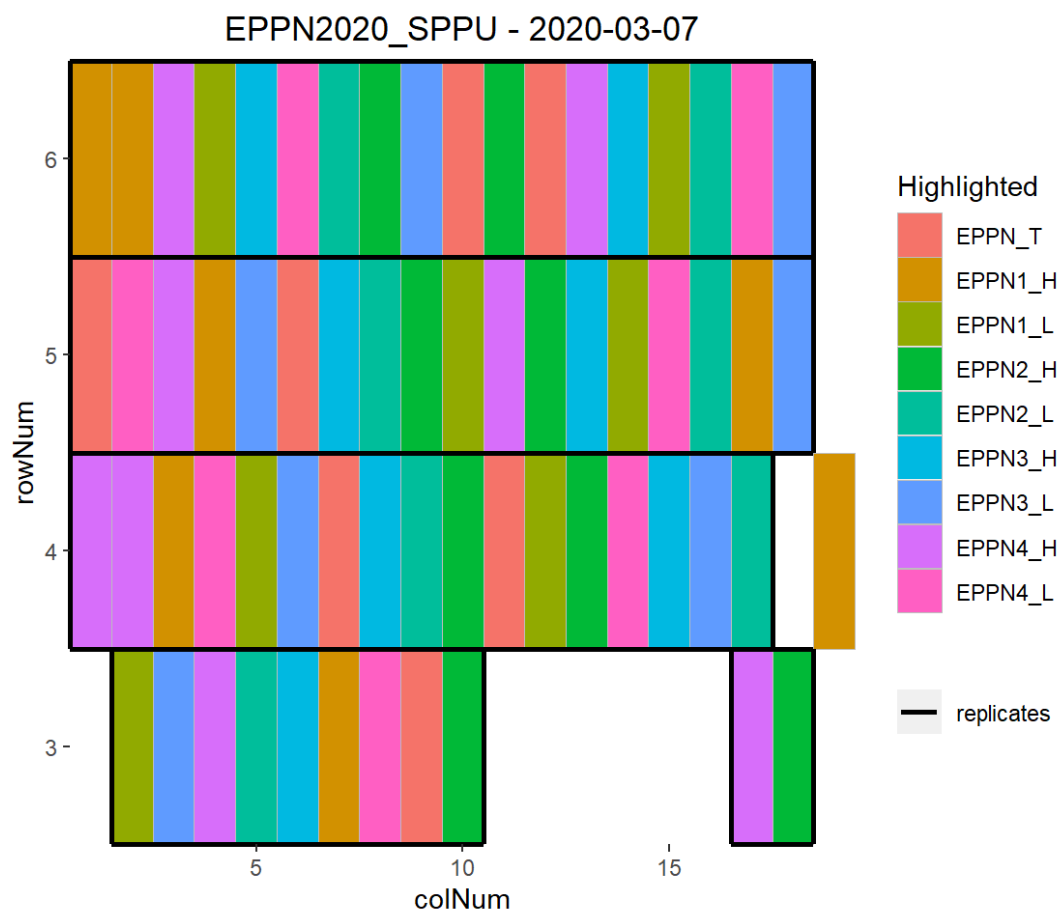
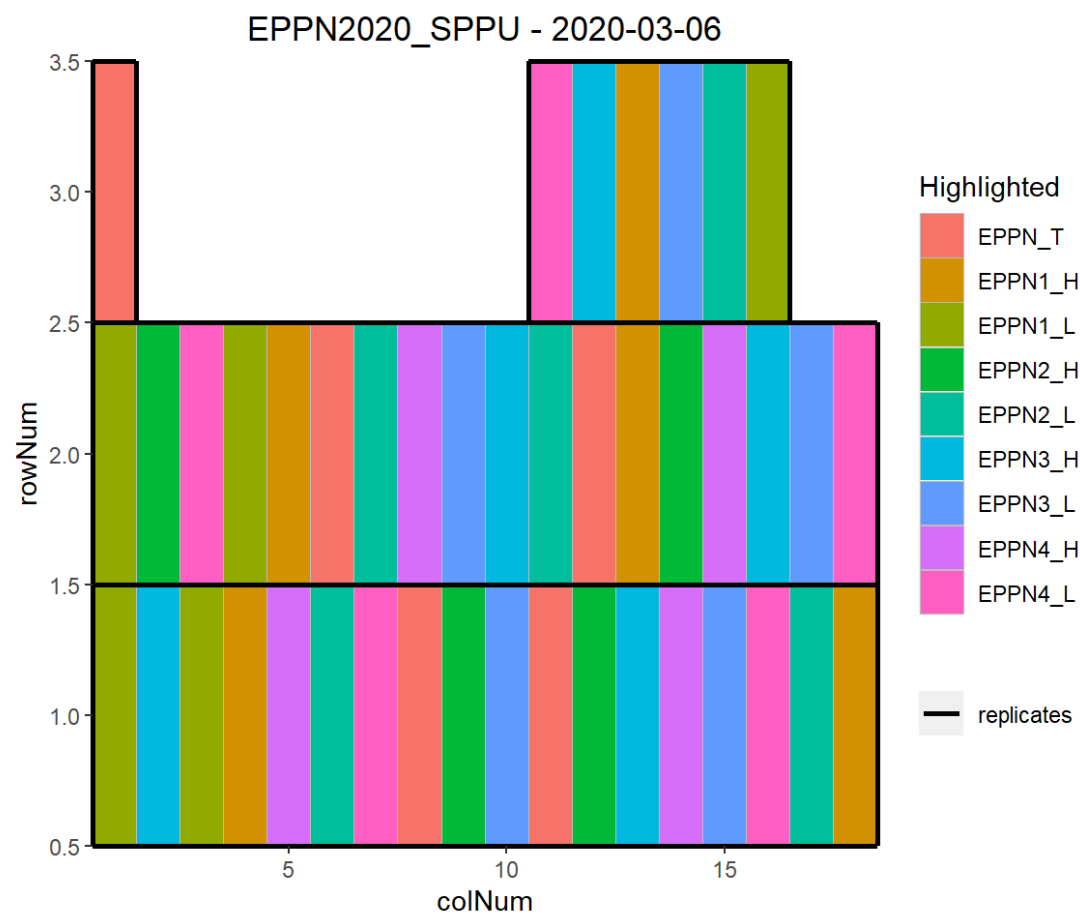


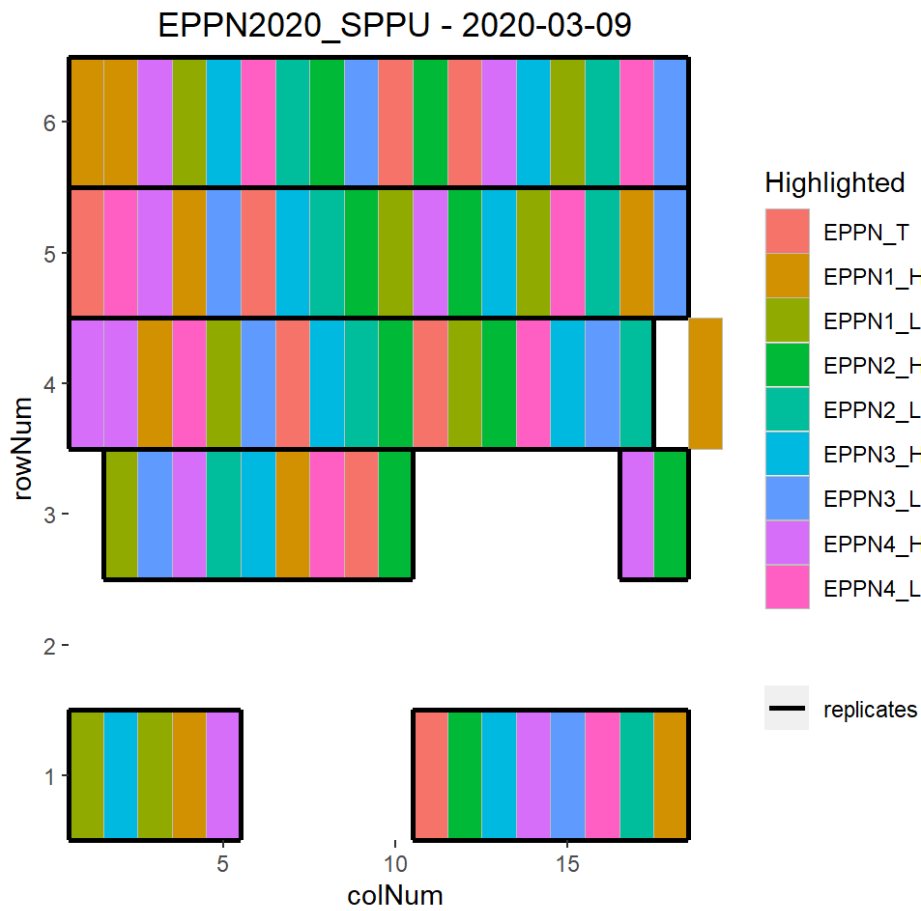
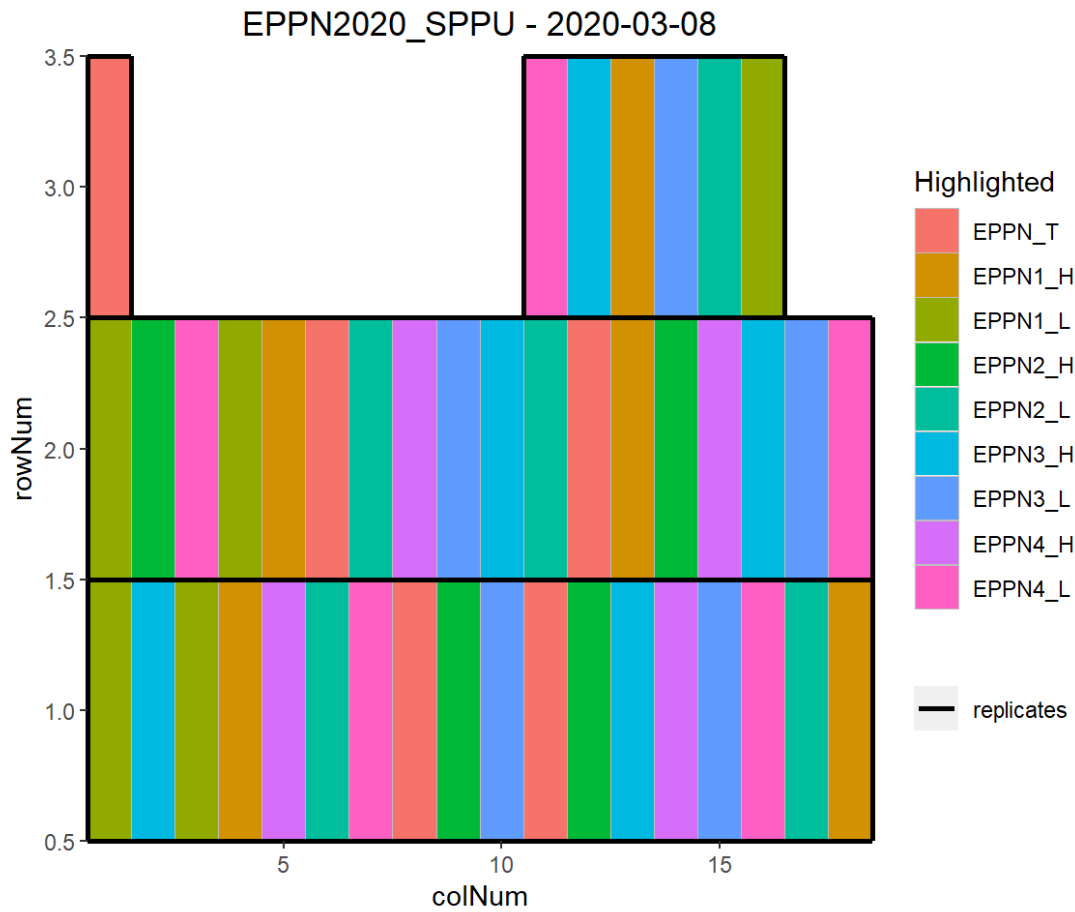


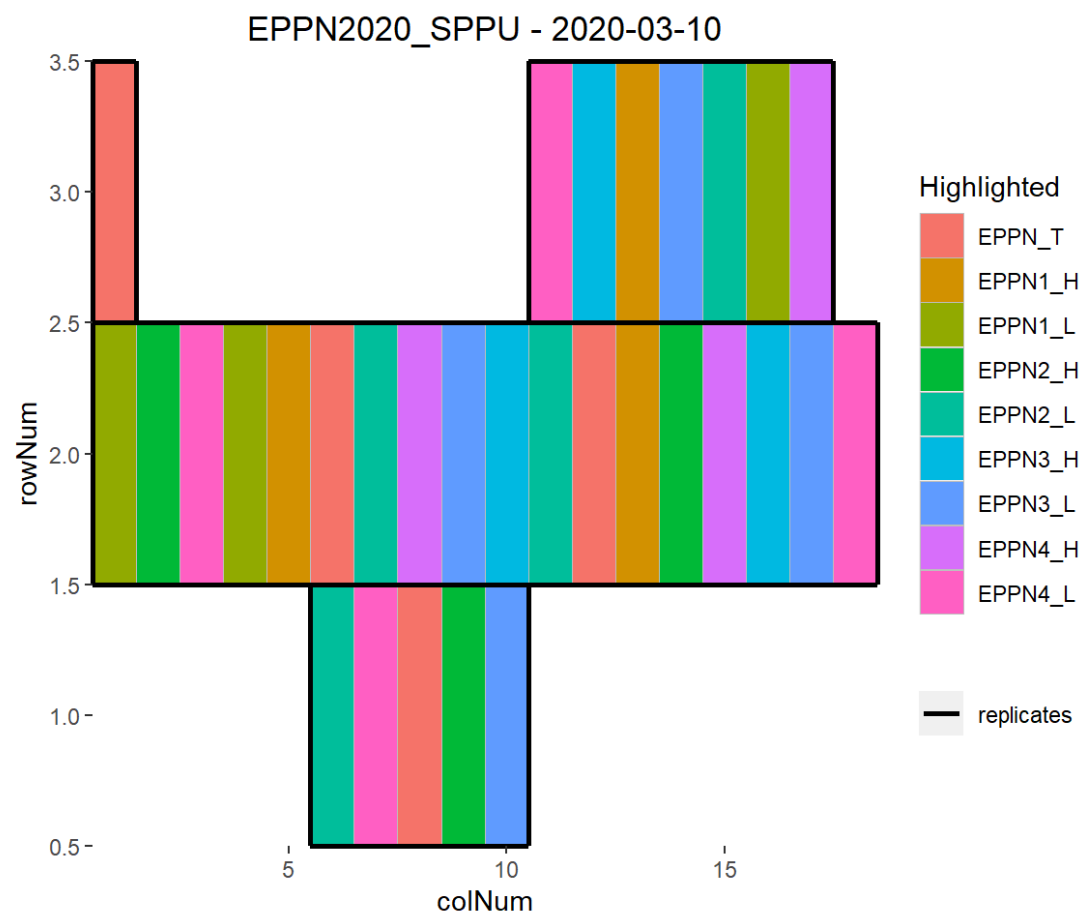












# 1. endpoint

No data

# 2. S\_timeseries

## Raw data

View timePoint object

```
summary(timePoint_S)
```

```
## timePoint_S contains data for experiment EPPN2020_SPPU.  
##  
## It contains 23 time points.  
## First time point: 2020-02-17  
## Last time point: 2020-03-10  
##  
## The following genotypes are defined as check genotypes: EPPN_T.
```

```
getTimePoints(timePoint_S)
```

```
##      timeNumber  timePoint
## 1           1 2020-02-17
## 2           2 2020-02-18
## 3           3 2020-02-19
## 4           4 2020-02-20
## 5           5 2020-02-21
## 6           6 2020-02-22
## 7           7 2020-02-23
## 8           8 2020-02-24
## 9           9 2020-02-25
## 10          10 2020-02-26
## 11          11 2020-02-27
## 12          12 2020-02-28
## 13          13 2020-02-29
## 14          14 2020-03-01
## 15          15 2020-03-02
## 16          16 2020-03-03
## 17          17 2020-03-04
## 18          18 2020-03-05
## 19          19 2020-03-06
## 20          20 2020-03-07
## 21          21 2020-03-08
## 22          22 2020-03-09
## 23          23 2020-03-10
```

```
num_timepoints <- getTimePoints(timePoint_S)
```

## Count the number of observations per trait and time point

We focus on the Height [cm] and Leaf area, because these are the two most common among the platforms.

Height is computed for 6 platforms out of 9 and area for 4 out of 9.

```
var_voulues <- c(variables_S[1], variables_S[3])
traits <- var_voulues

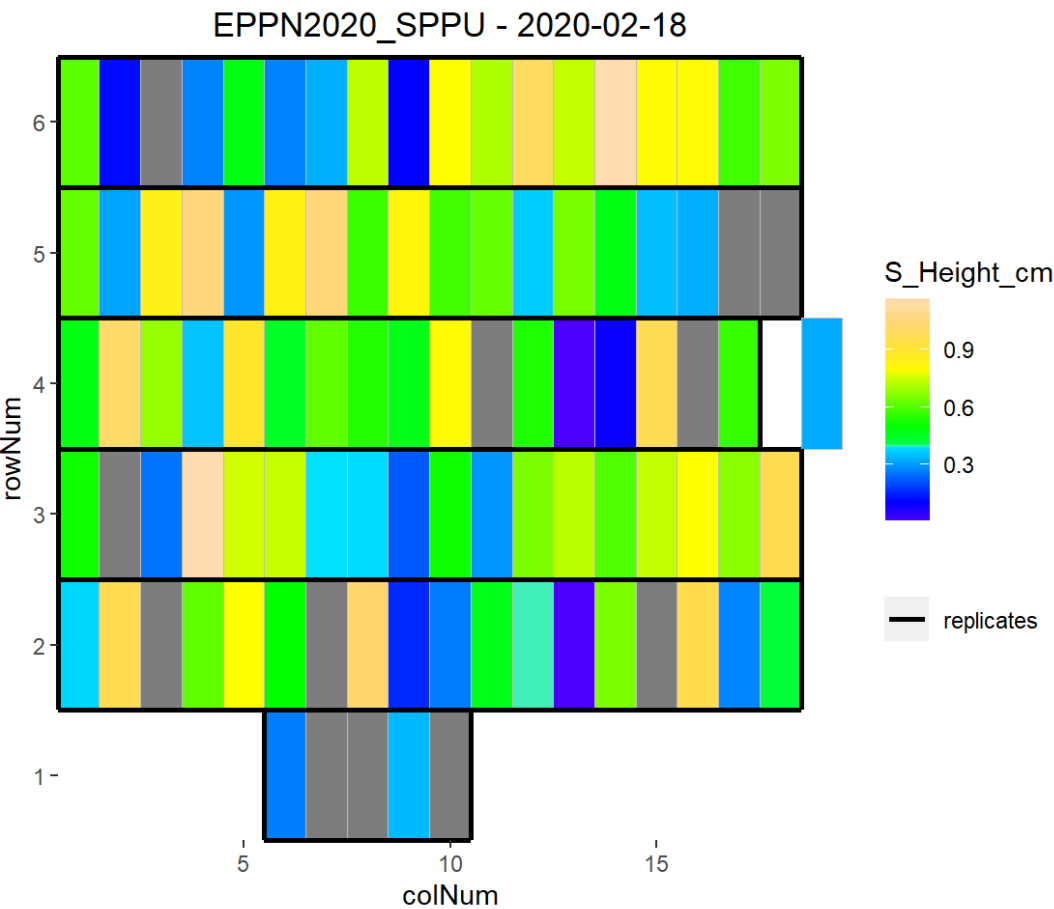
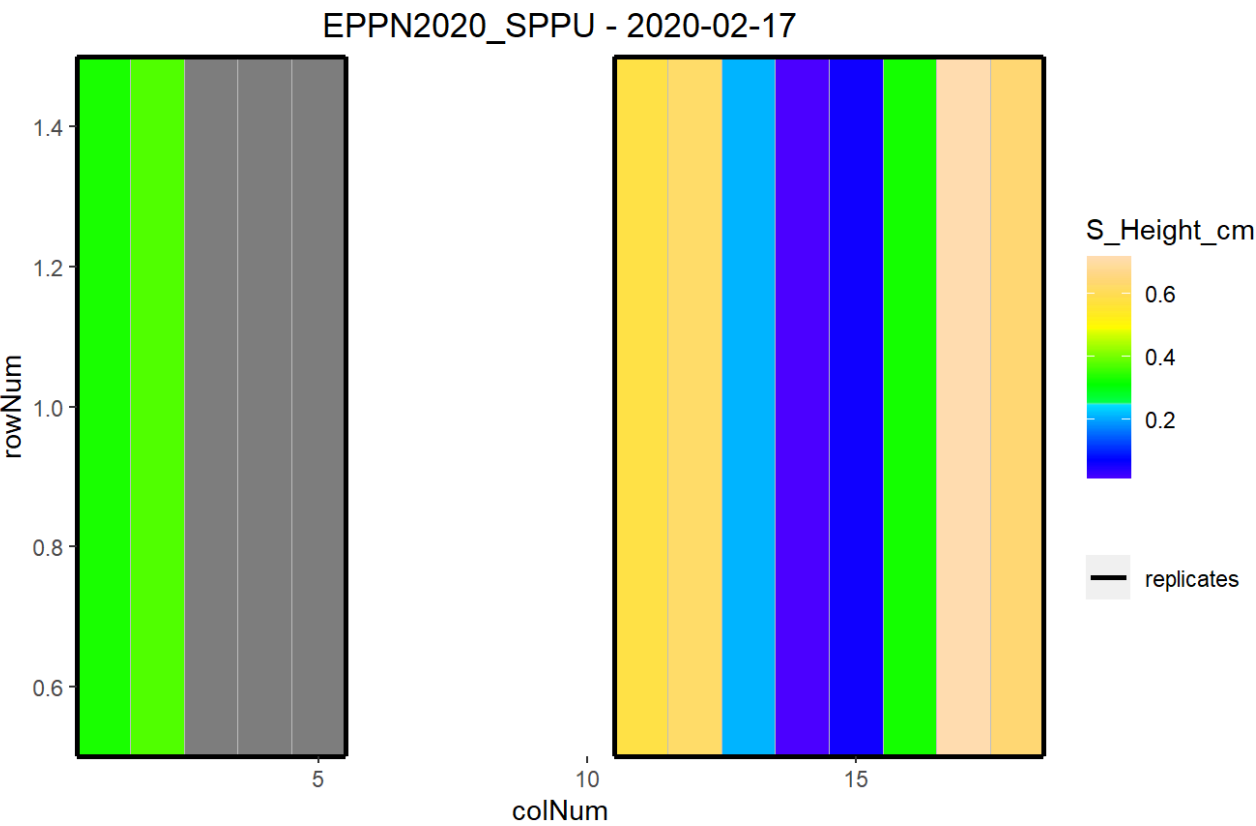
for (trait_name in traits) {
  print(paste("How many observations for", trait_name))
  valid_count <- countValid(timePoint_S, trait_name)
  print(valid_count)
}
```

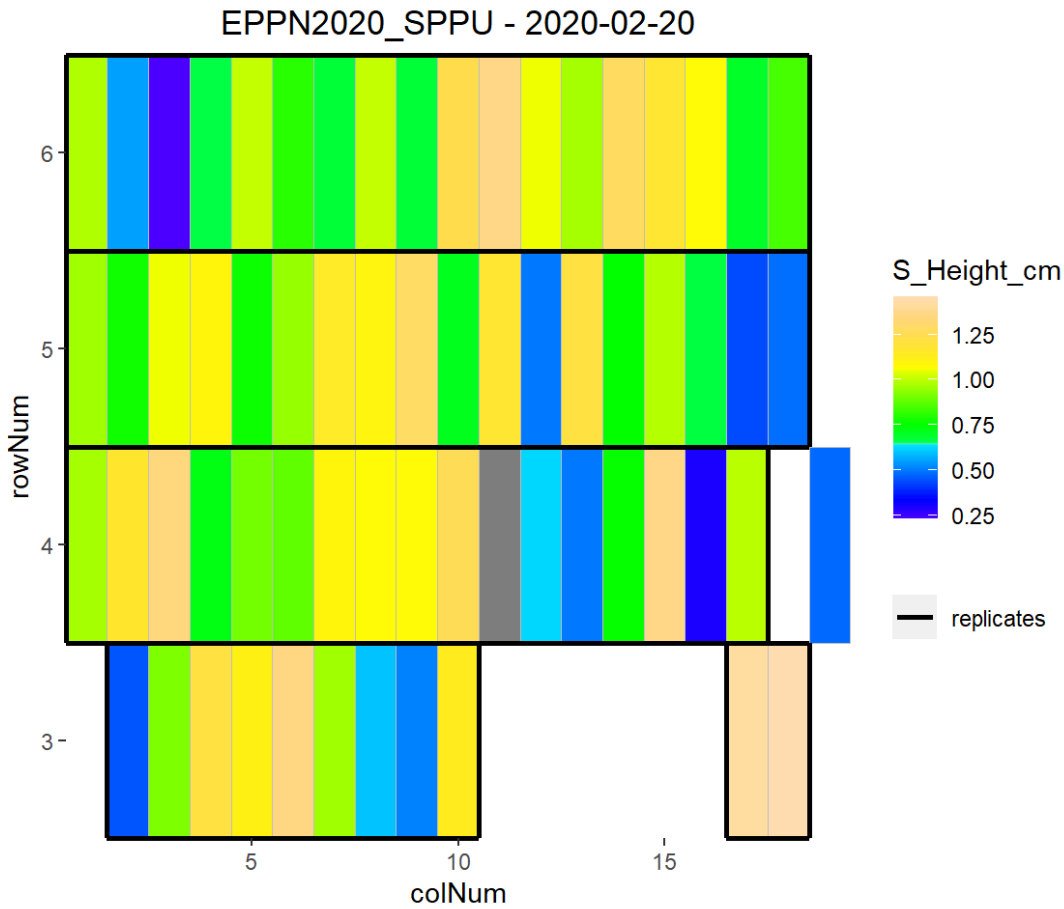
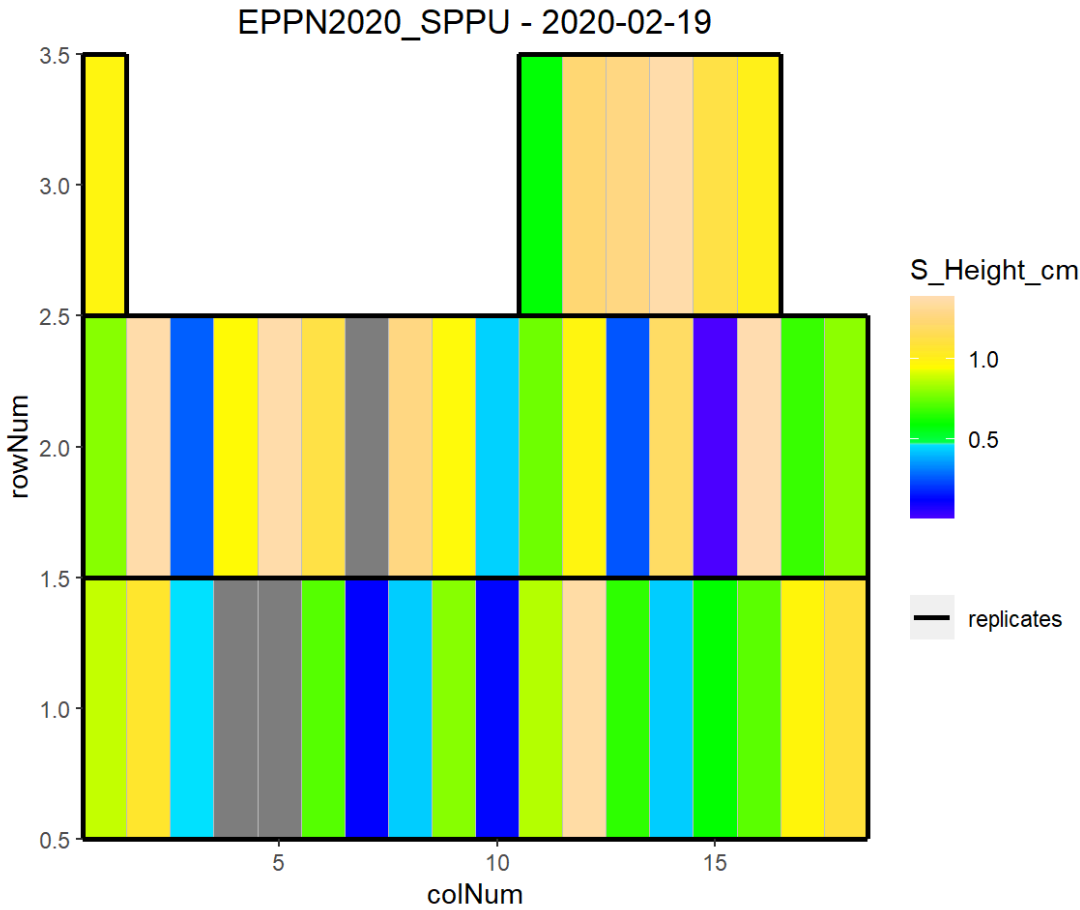


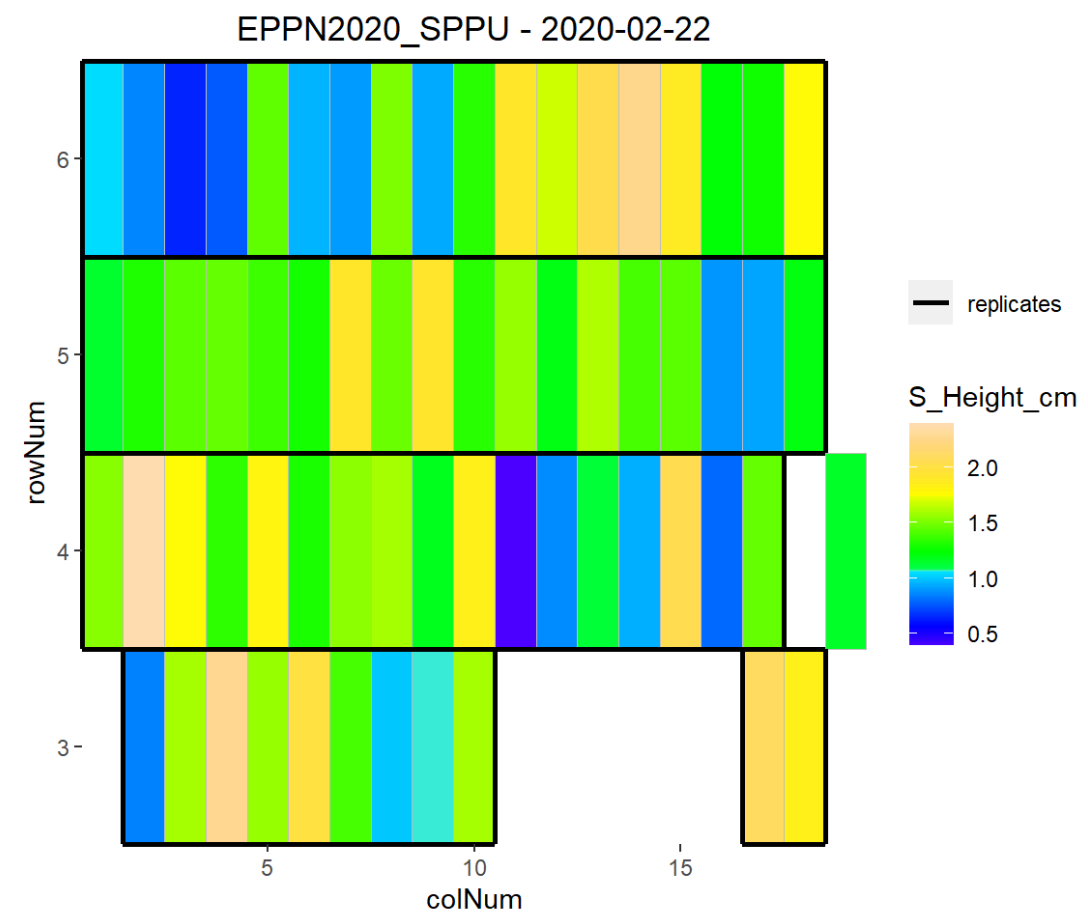
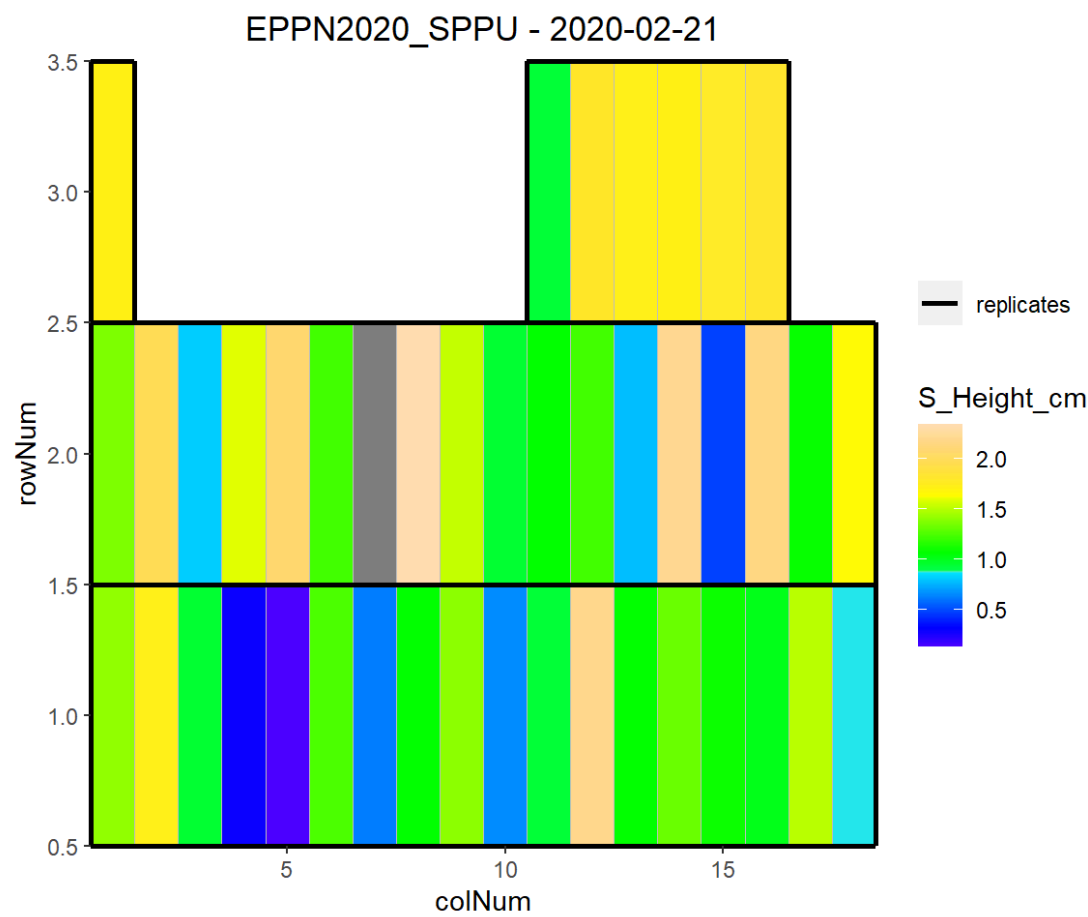
```
## [1] "How many observations for S_Height_cm"
## 2020-02-17 2020-02-18 2020-02-19 2020-02-20 2020-02-21 2020-02-22 2020-02-23
##          10          83          40          64          42          65          42
## 2020-02-24 2020-02-25 2020-02-26 2020-02-27 2020-02-28 2020-02-29 2020-03-01
##          78          94          42          65          42          65          42
## 2020-03-02 2020-03-03 2020-03-04 2020-03-05 2020-03-06 2020-03-07 2020-03-08
##          78          87          42          65          42          65          42
## 2020-03-09 2020-03-10
##          78          30
## [1] "How many observations for S_Area_cmsquared"
## 2020-02-17 2020-02-18 2020-02-19 2020-02-20 2020-02-21 2020-02-22 2020-02-23
##          10          83          40          64          42          65          42
## 2020-02-24 2020-02-25 2020-02-26 2020-02-27 2020-02-28 2020-02-29 2020-03-01
##          78          94          42          65          42          65          42
## 2020-03-02 2020-03-03 2020-03-04 2020-03-05 2020-03-06 2020-03-07 2020-03-08
##          78          87          42          65          42          65          42
## 2020-03-09 2020-03-10
##          78          30
```

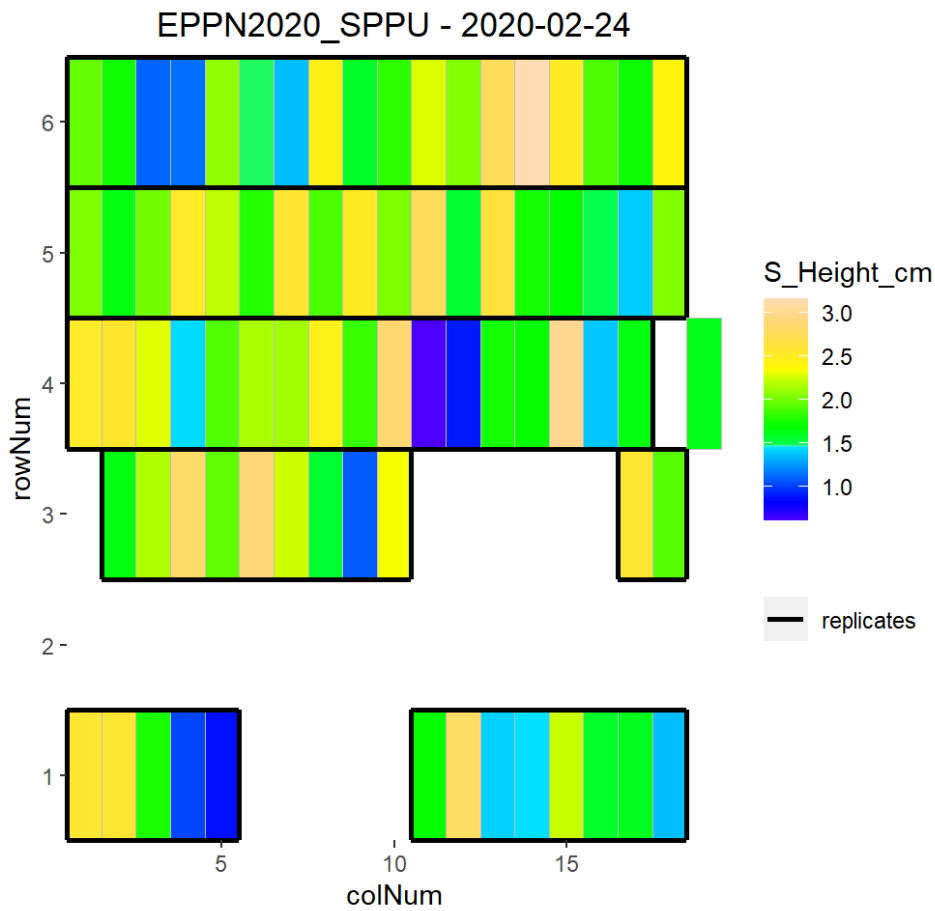
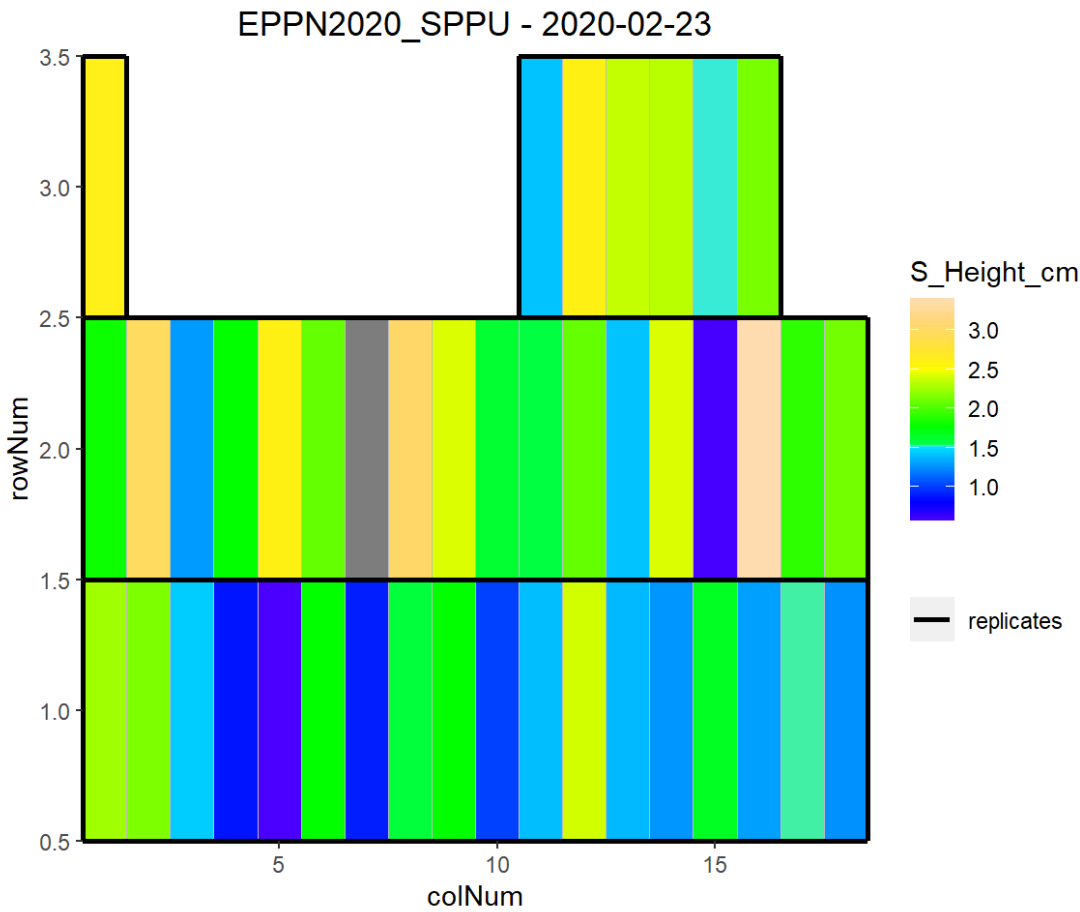
Check the heatmap of the raw data per time point

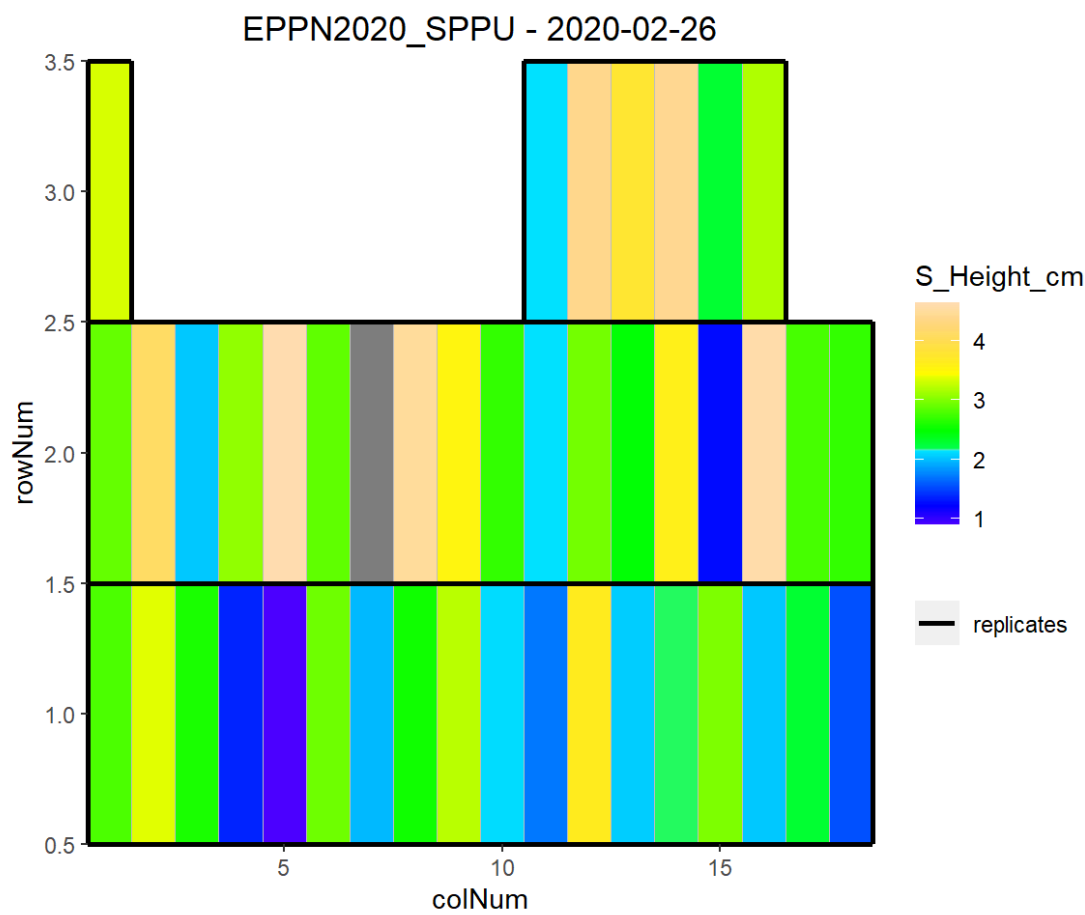
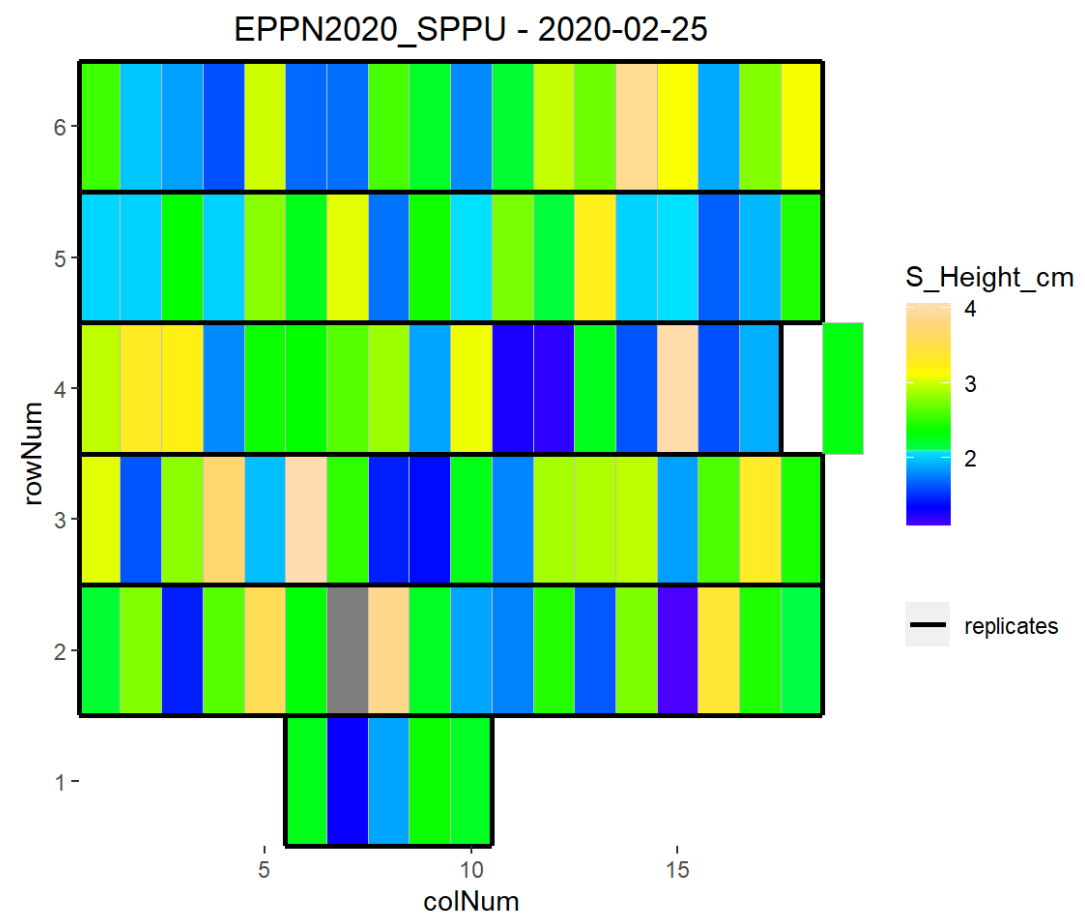
```
for (trait_name in traits) {
  for (tp in 1:length(num_timepoints$timeNumber)) {
    plot(timePoint_S,
         plotType = "layout",
         timePoints = tp,
         traits = trait_name)
  }
}
```

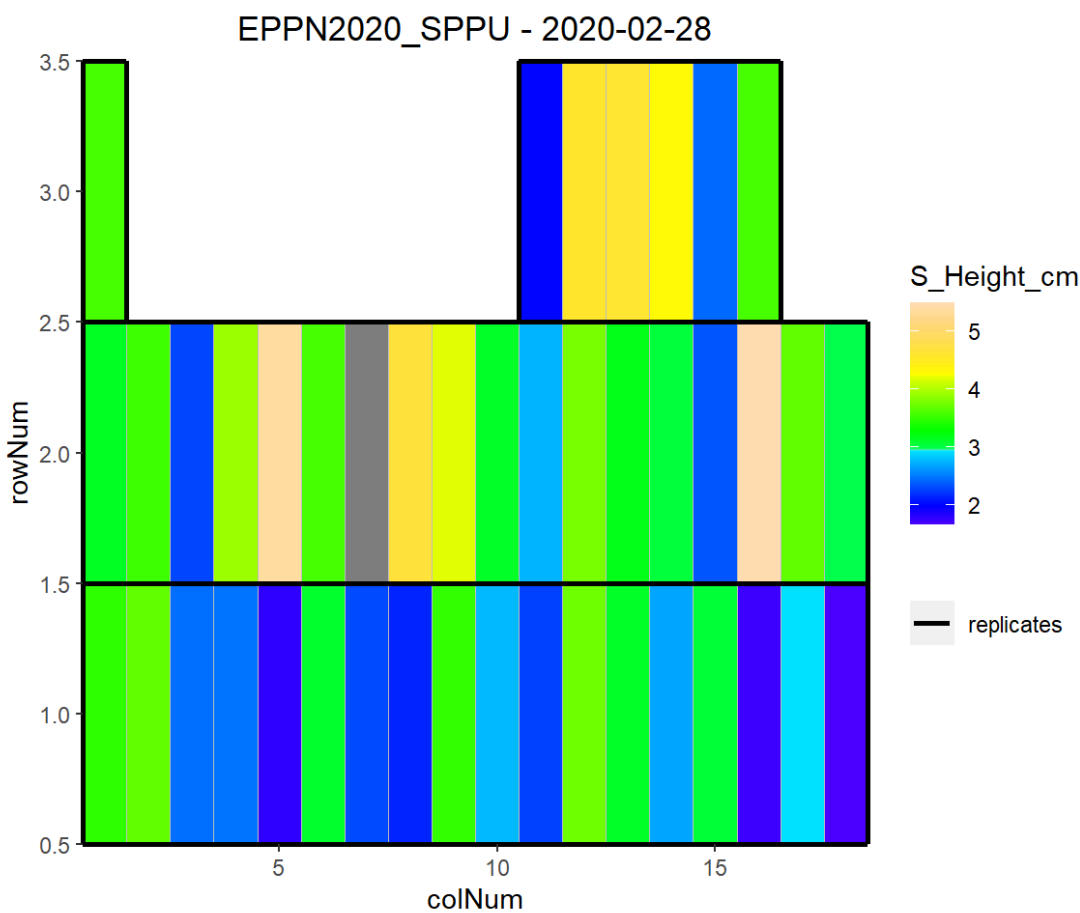
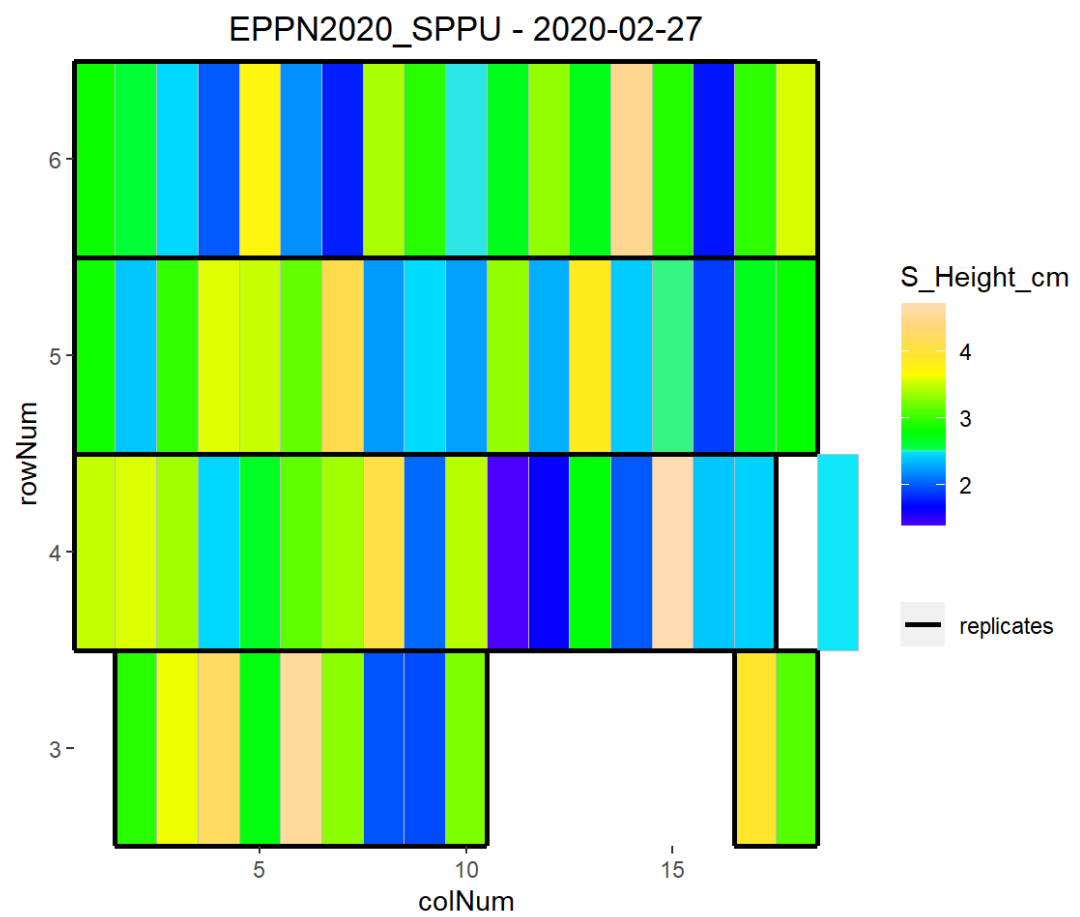


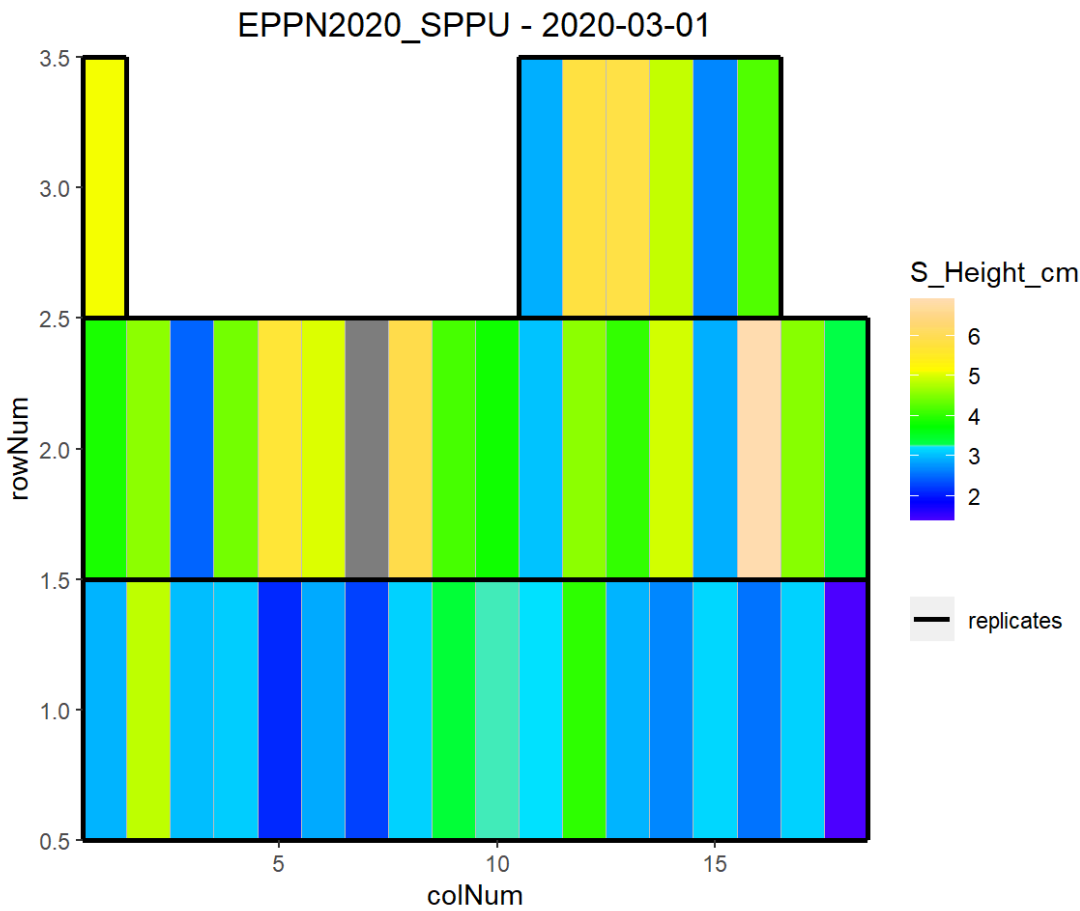
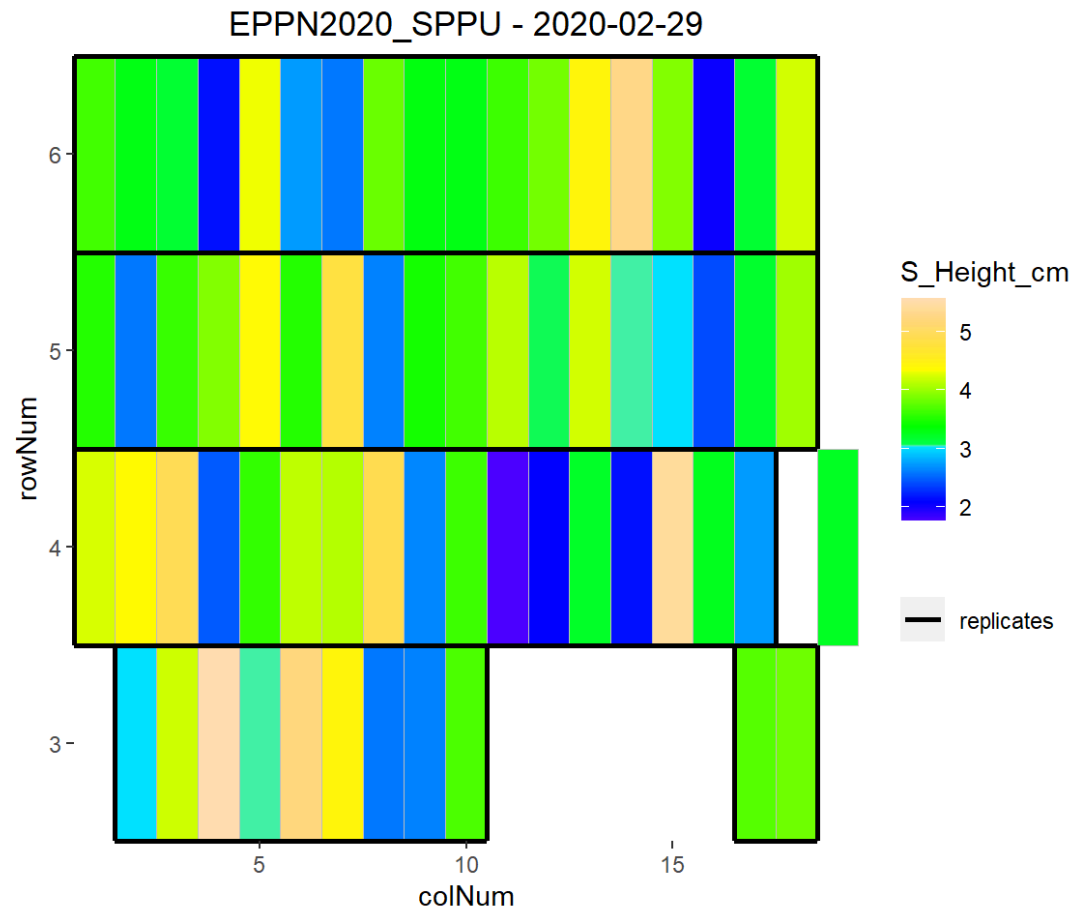




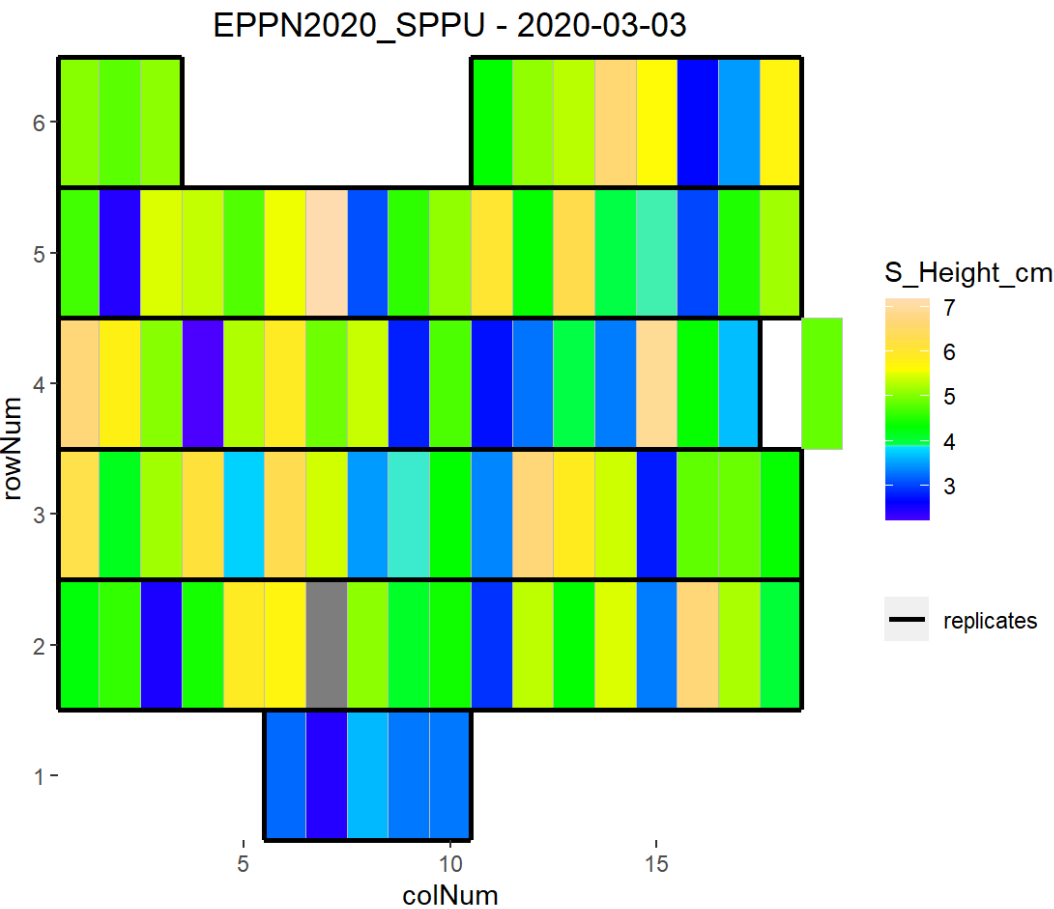
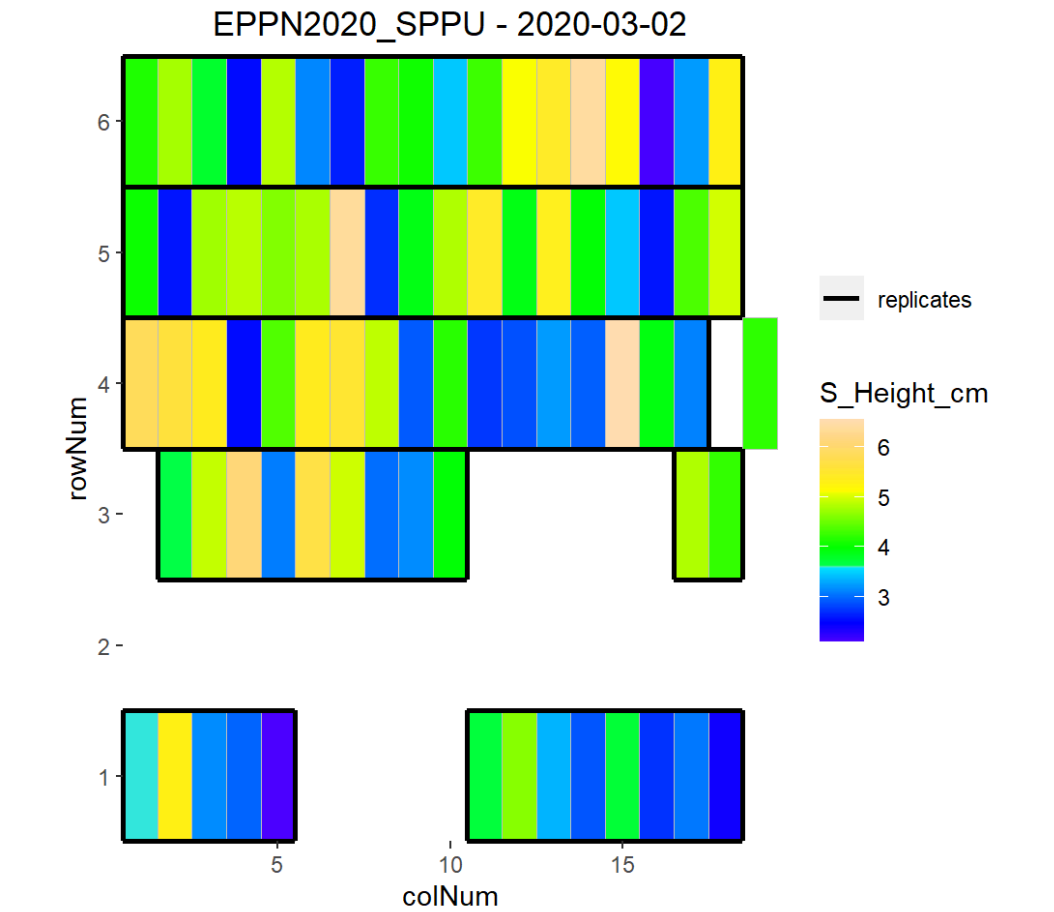


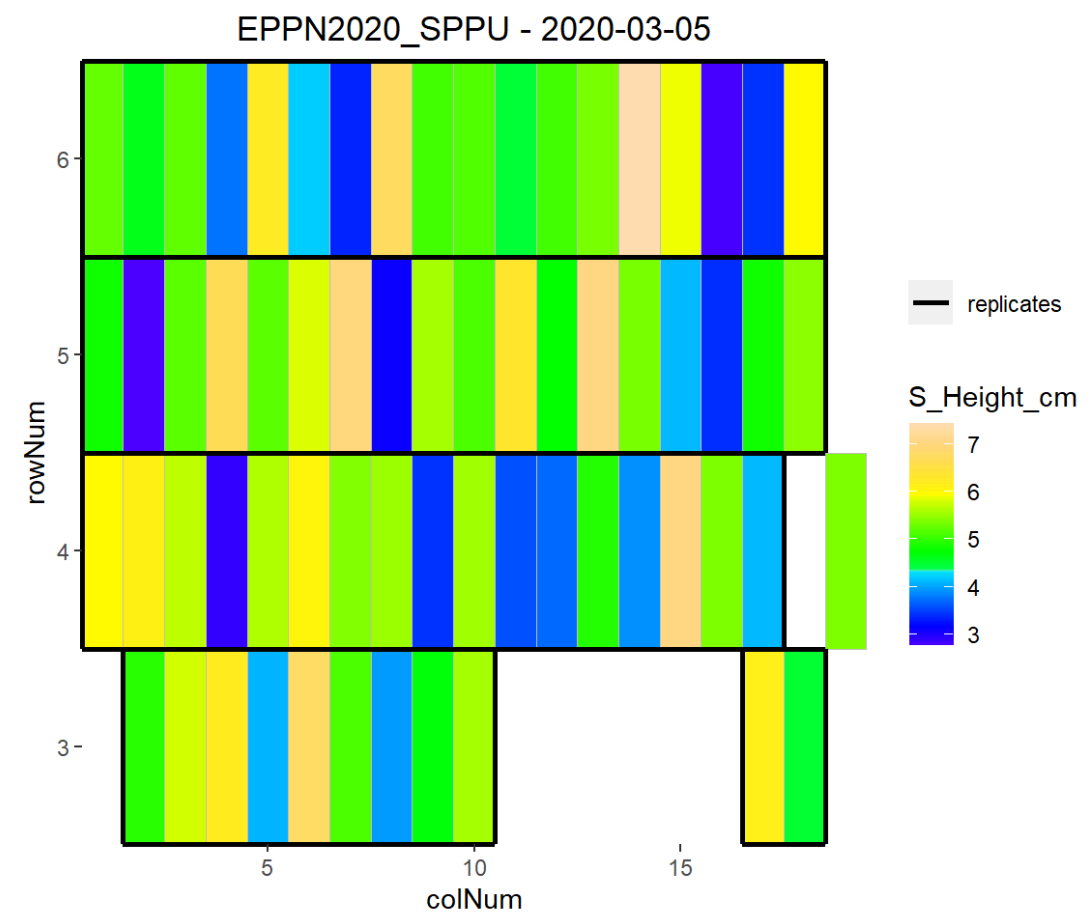
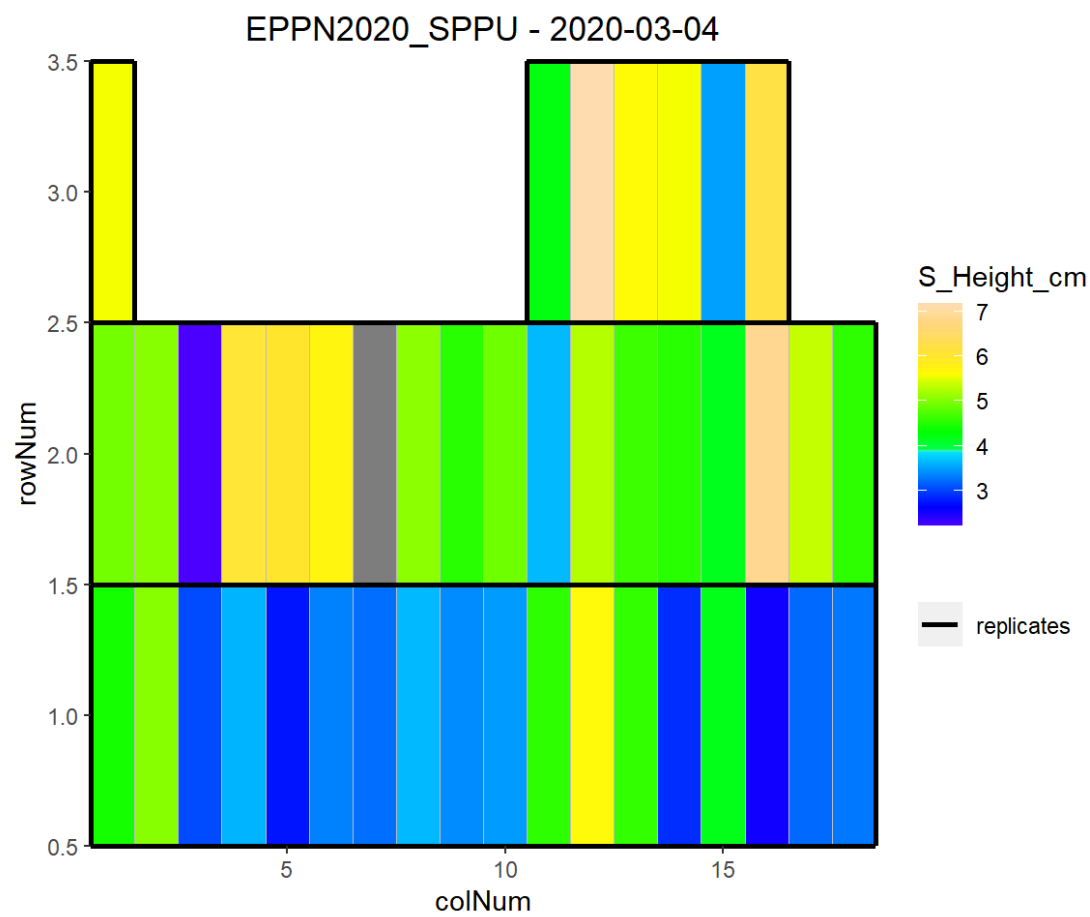


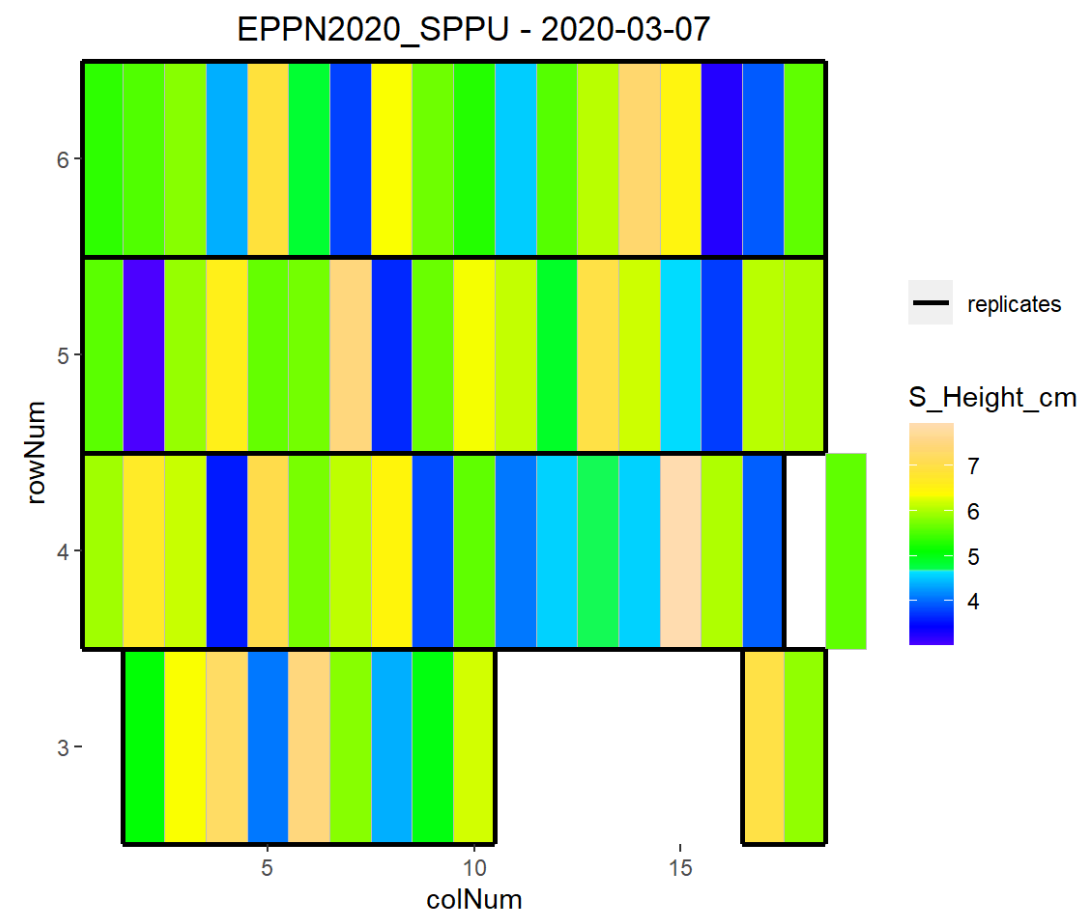
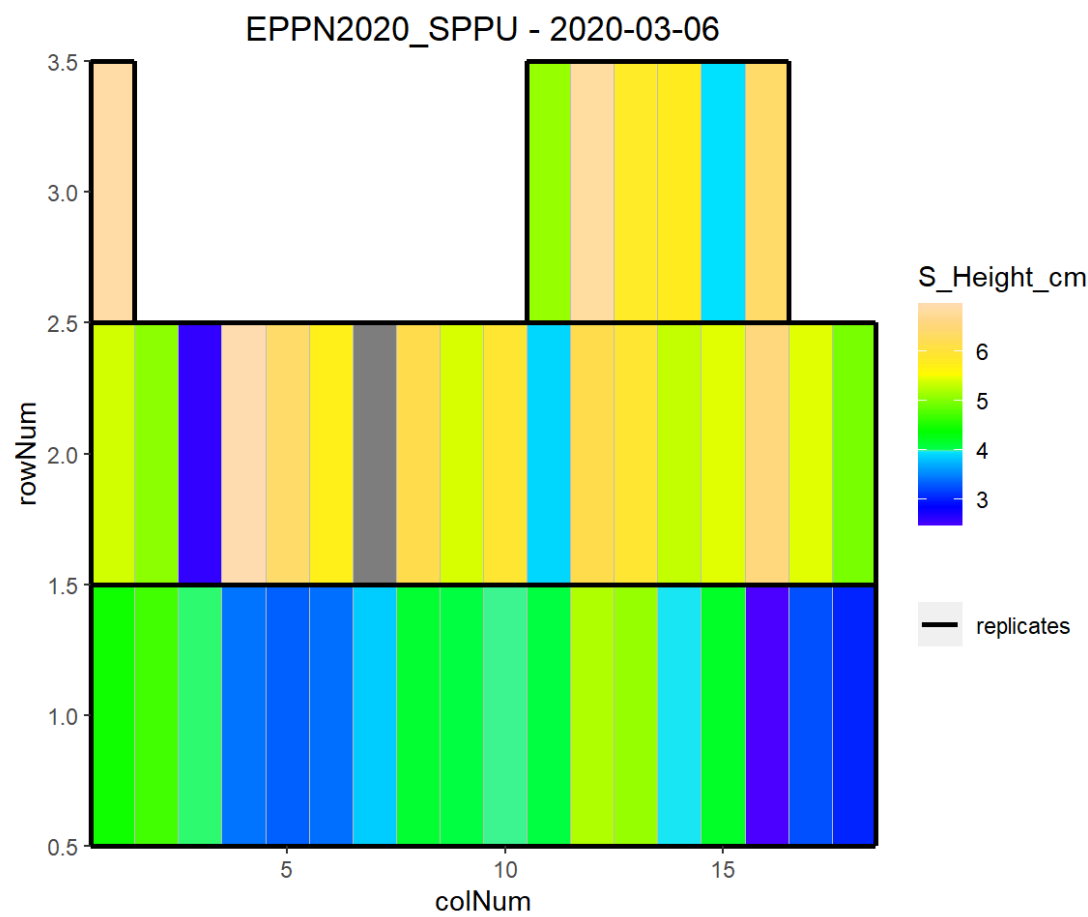


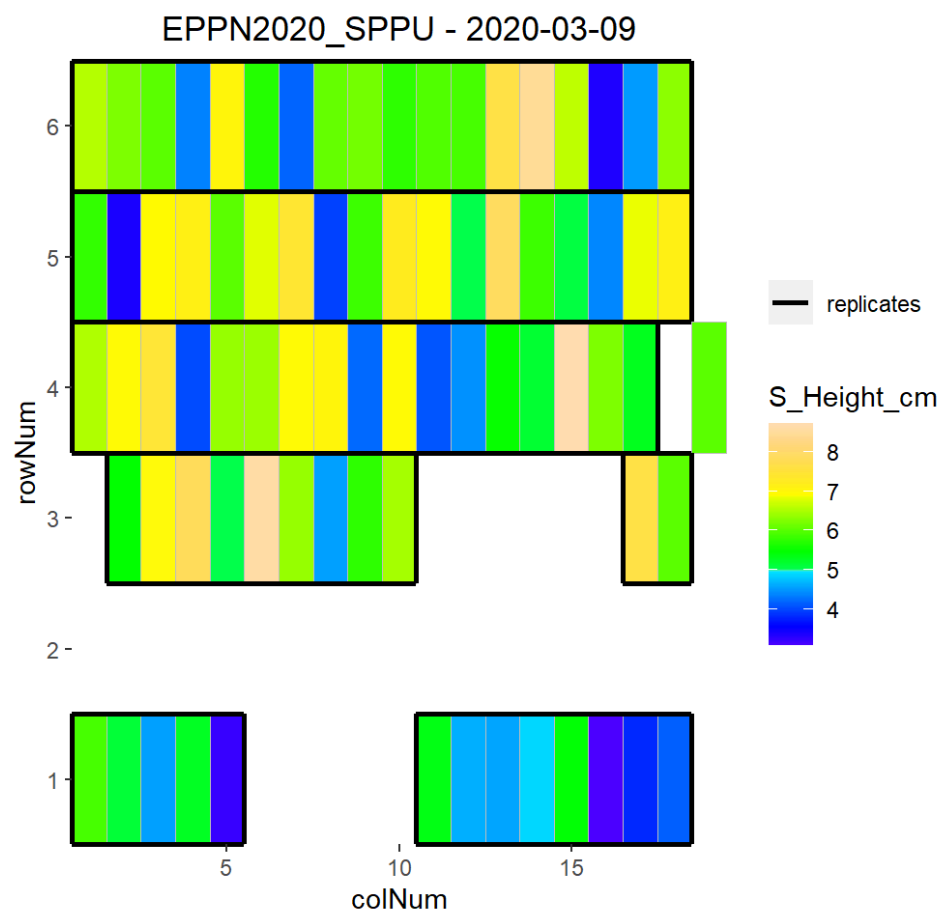
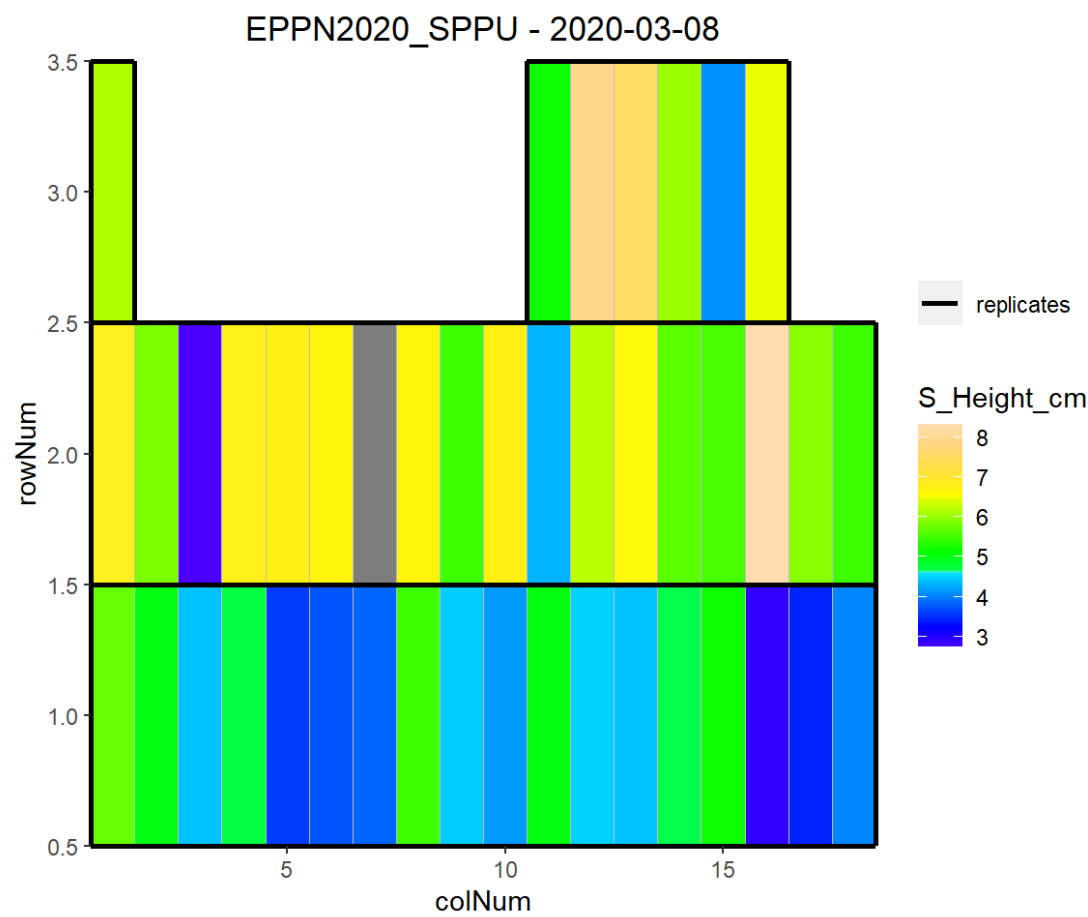


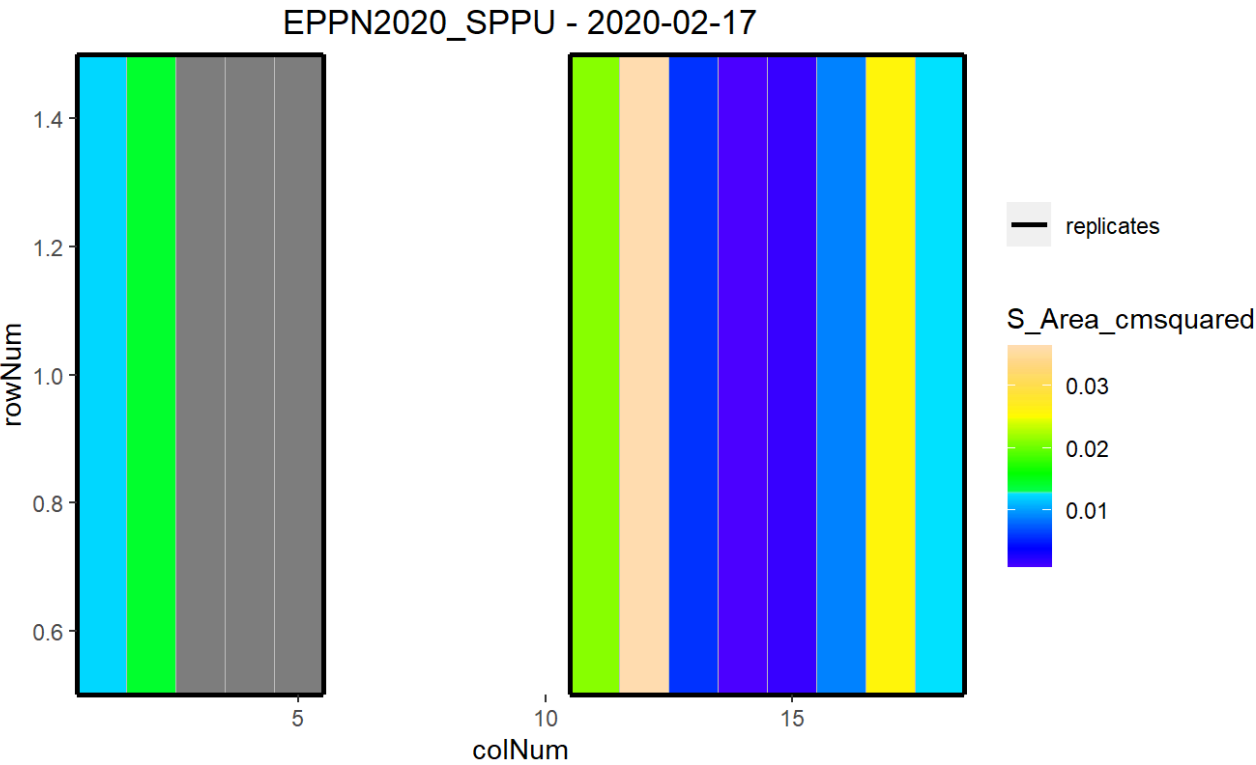
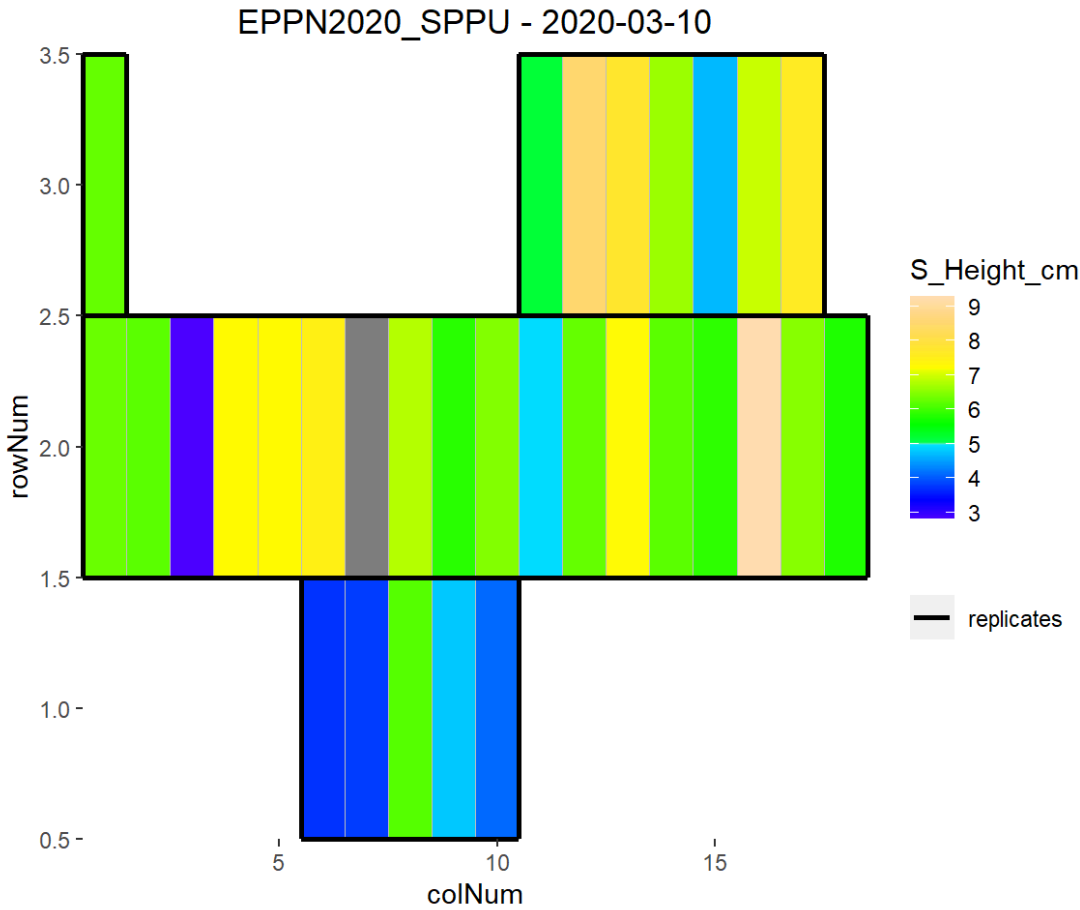


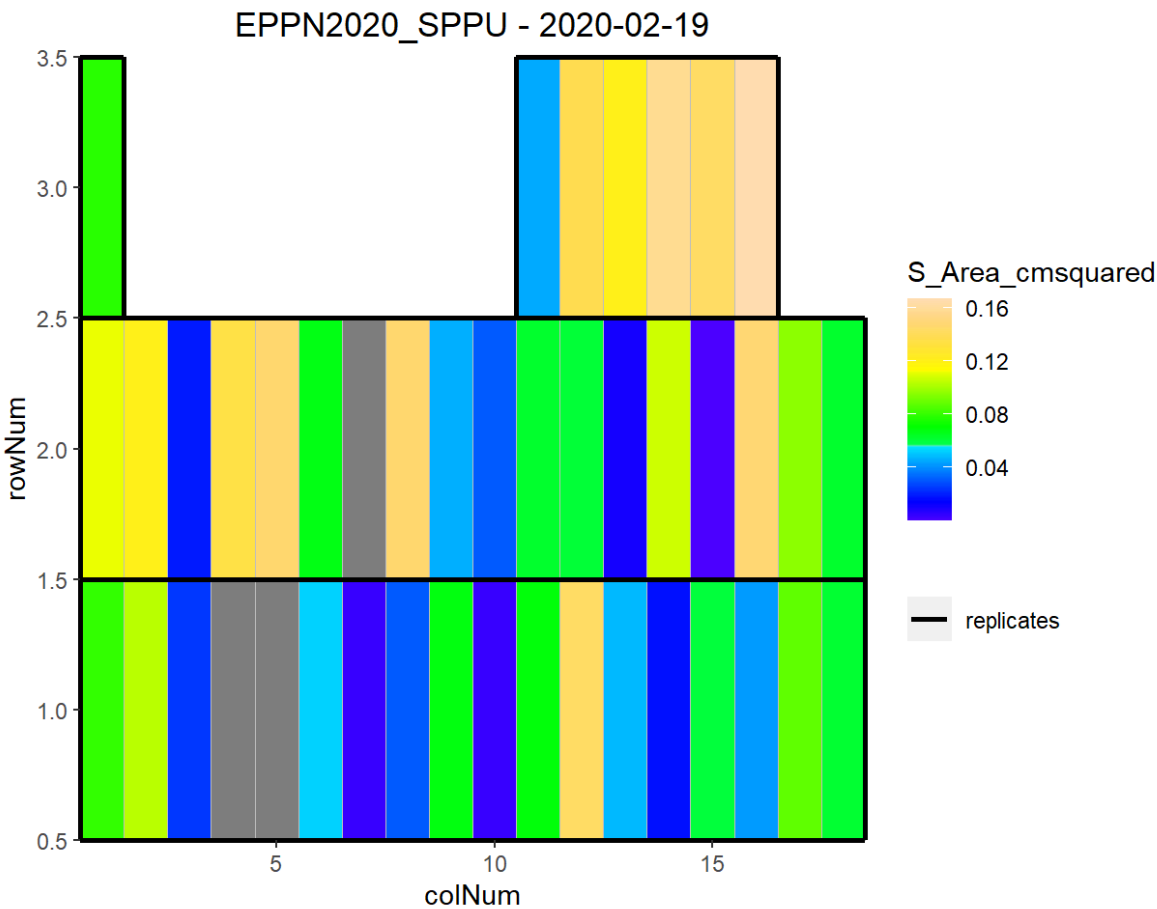
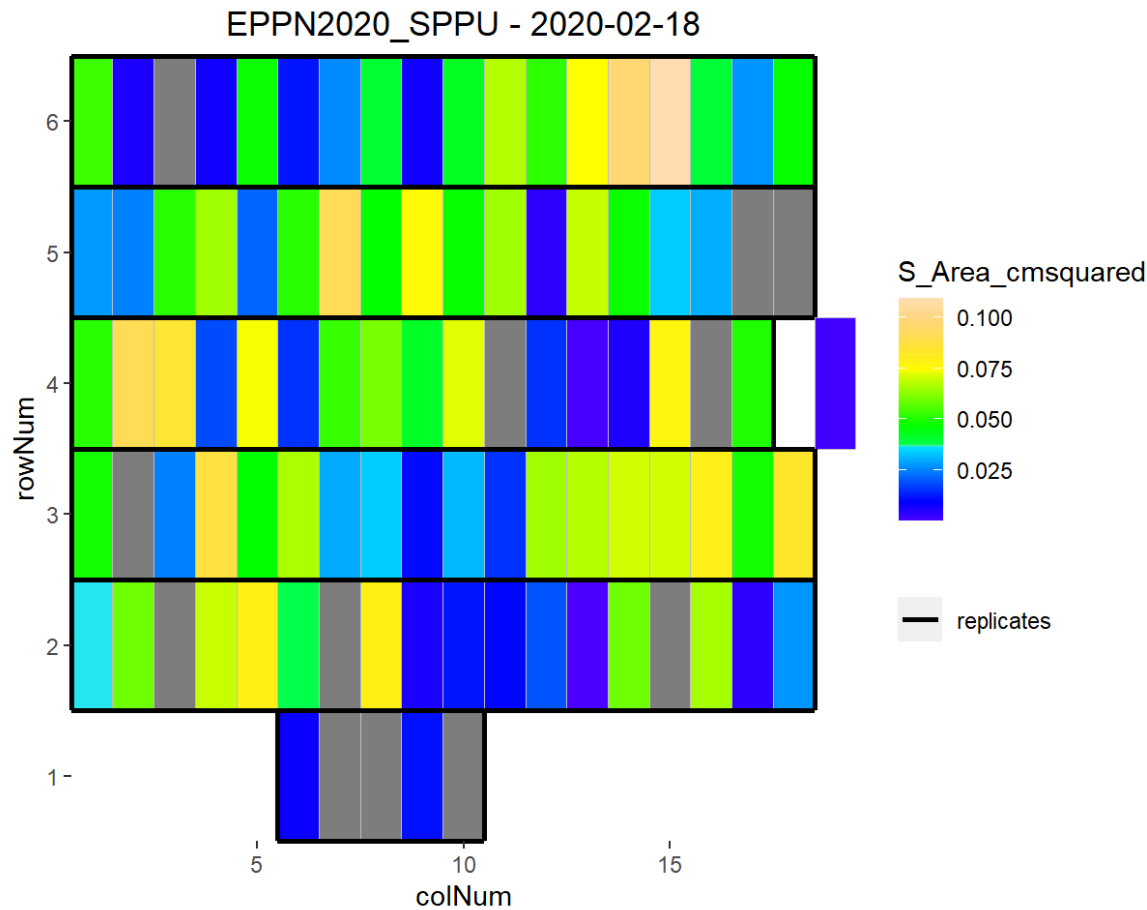


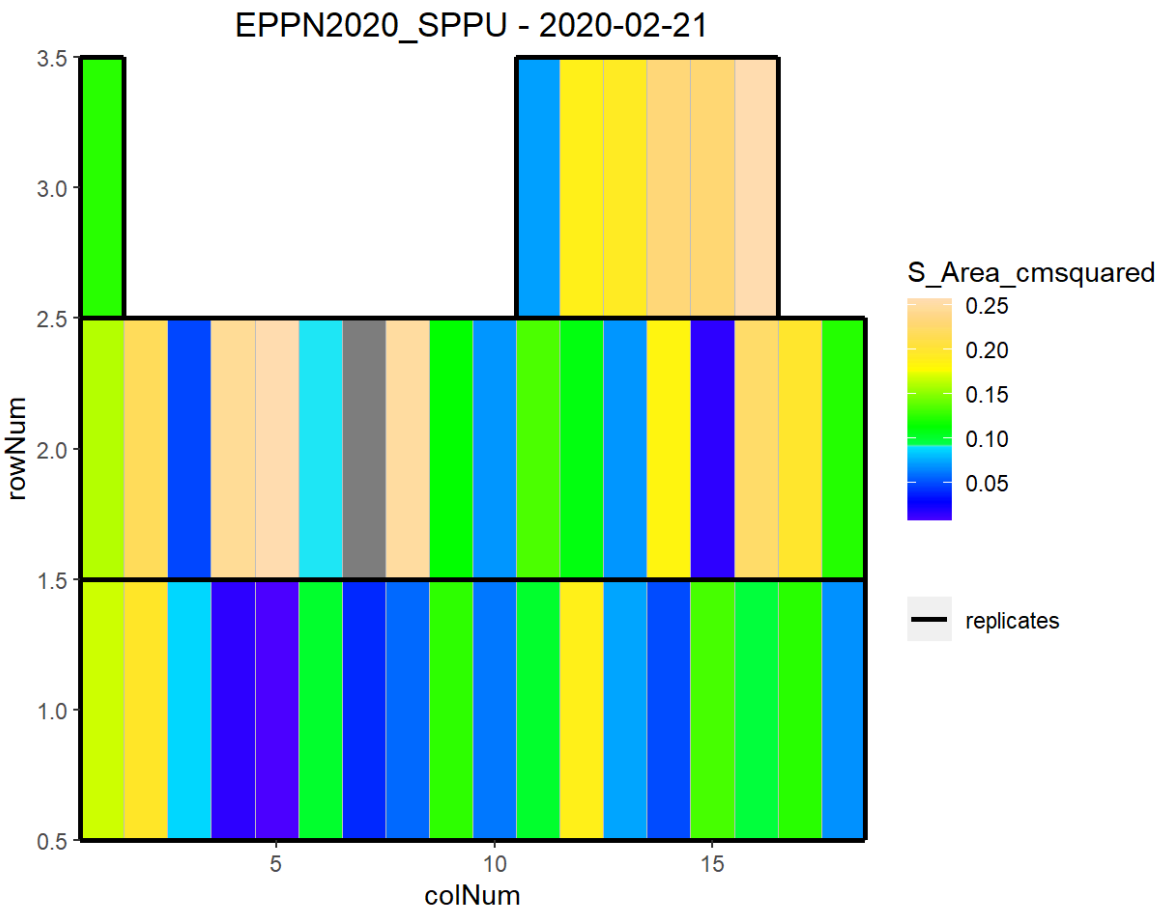
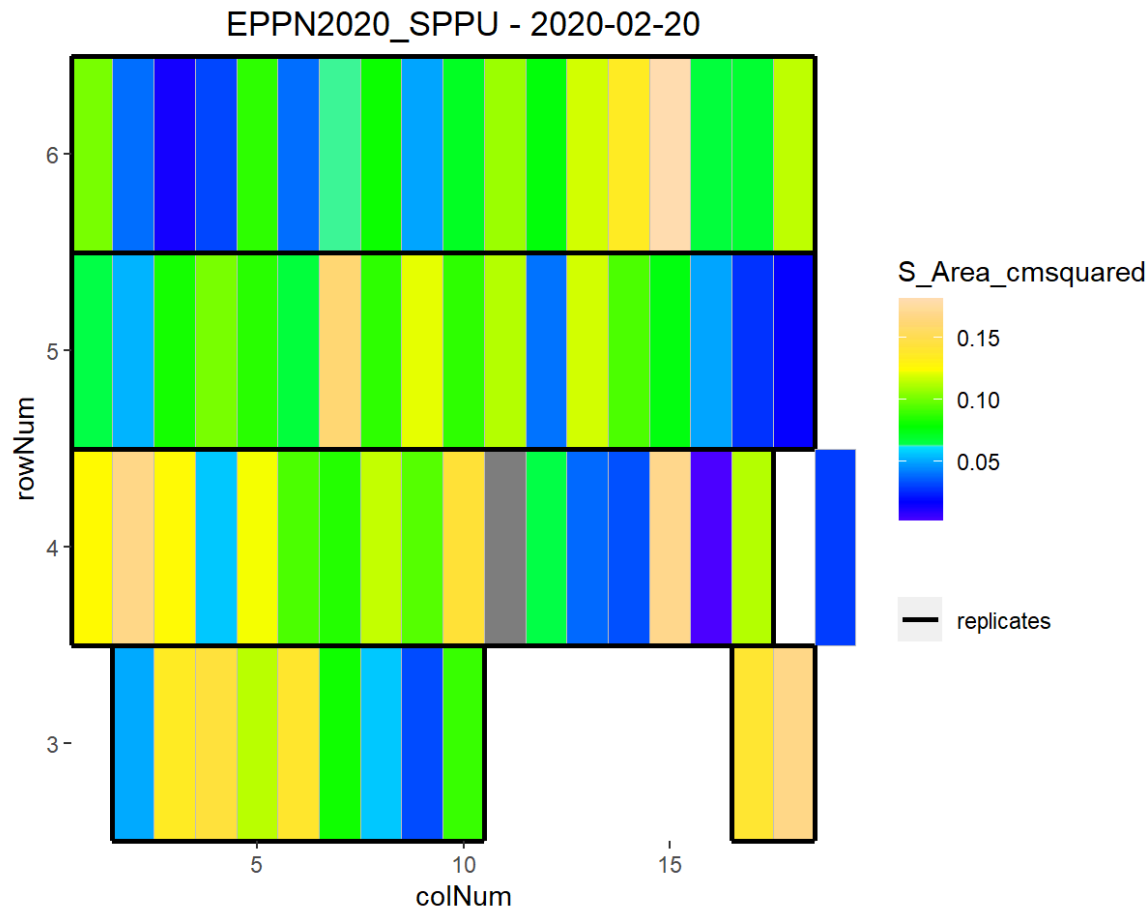


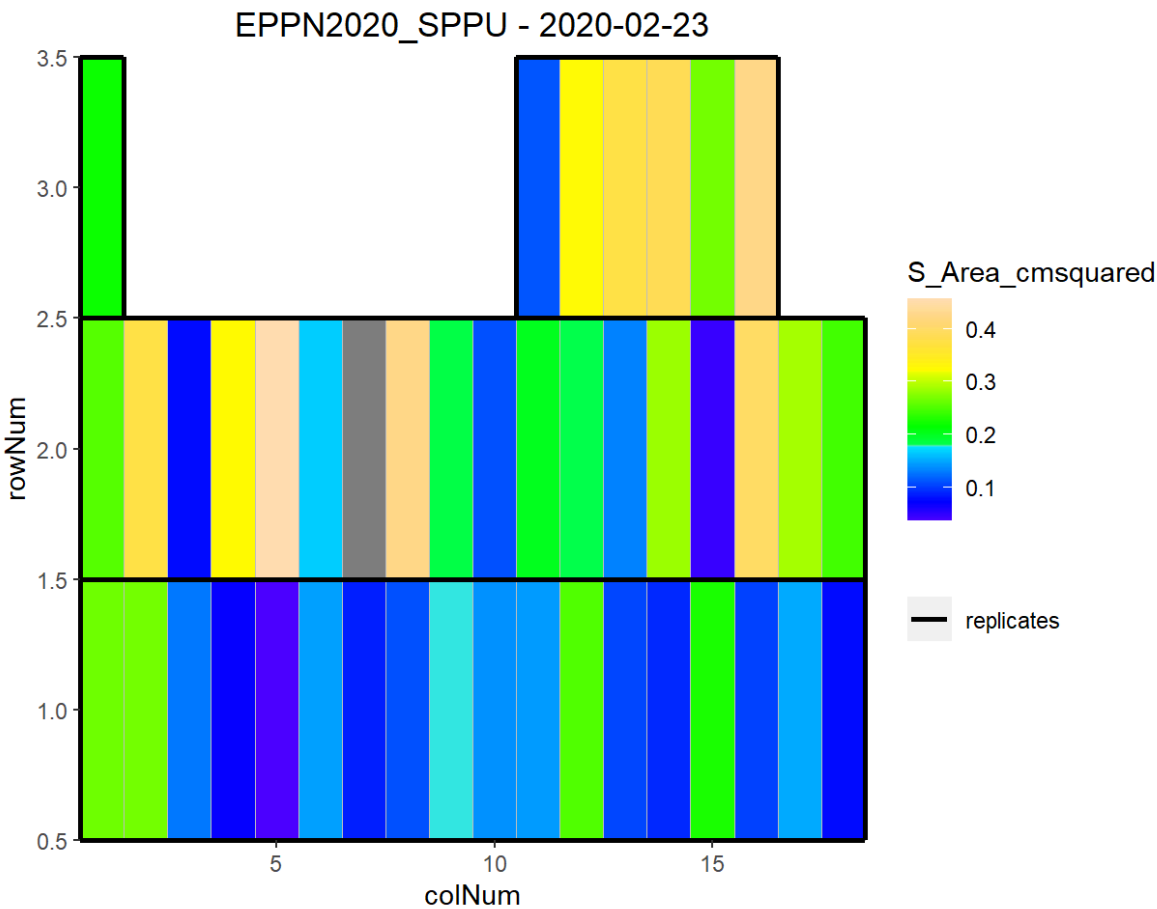
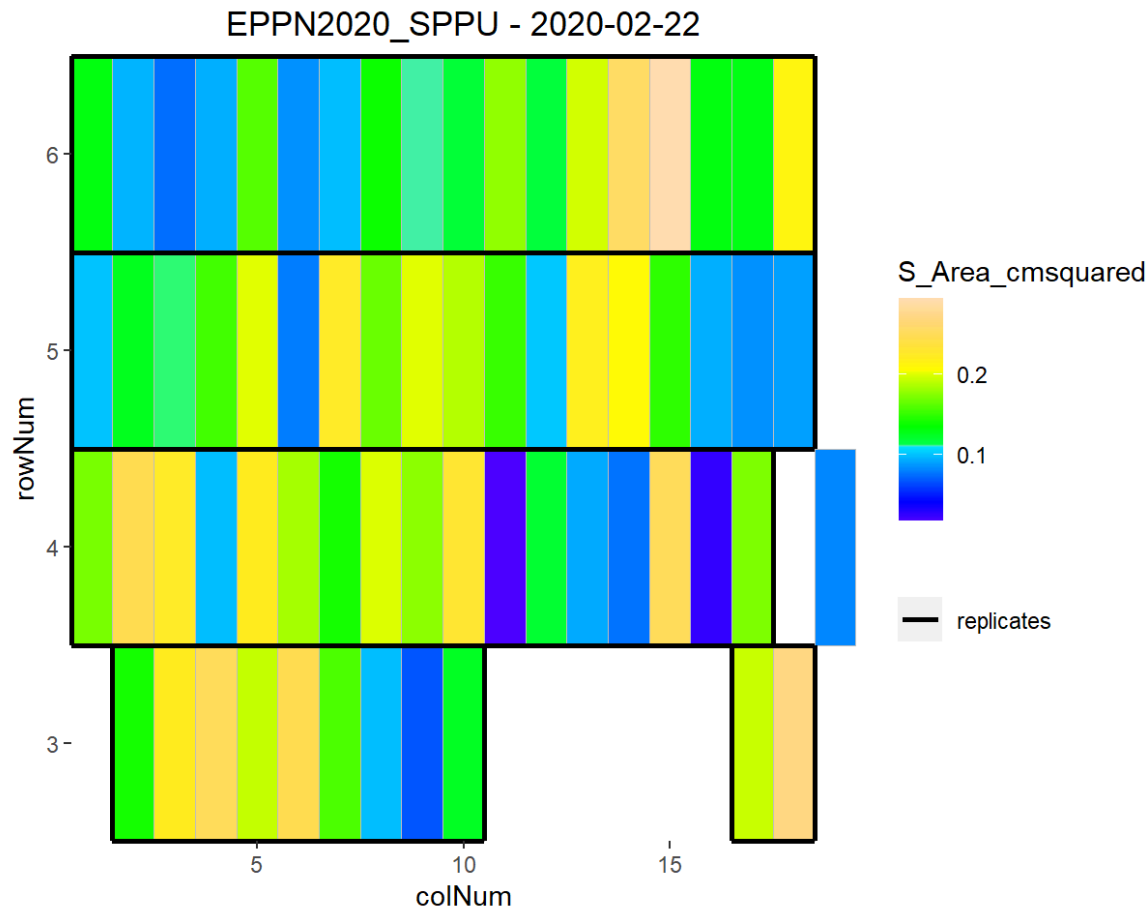




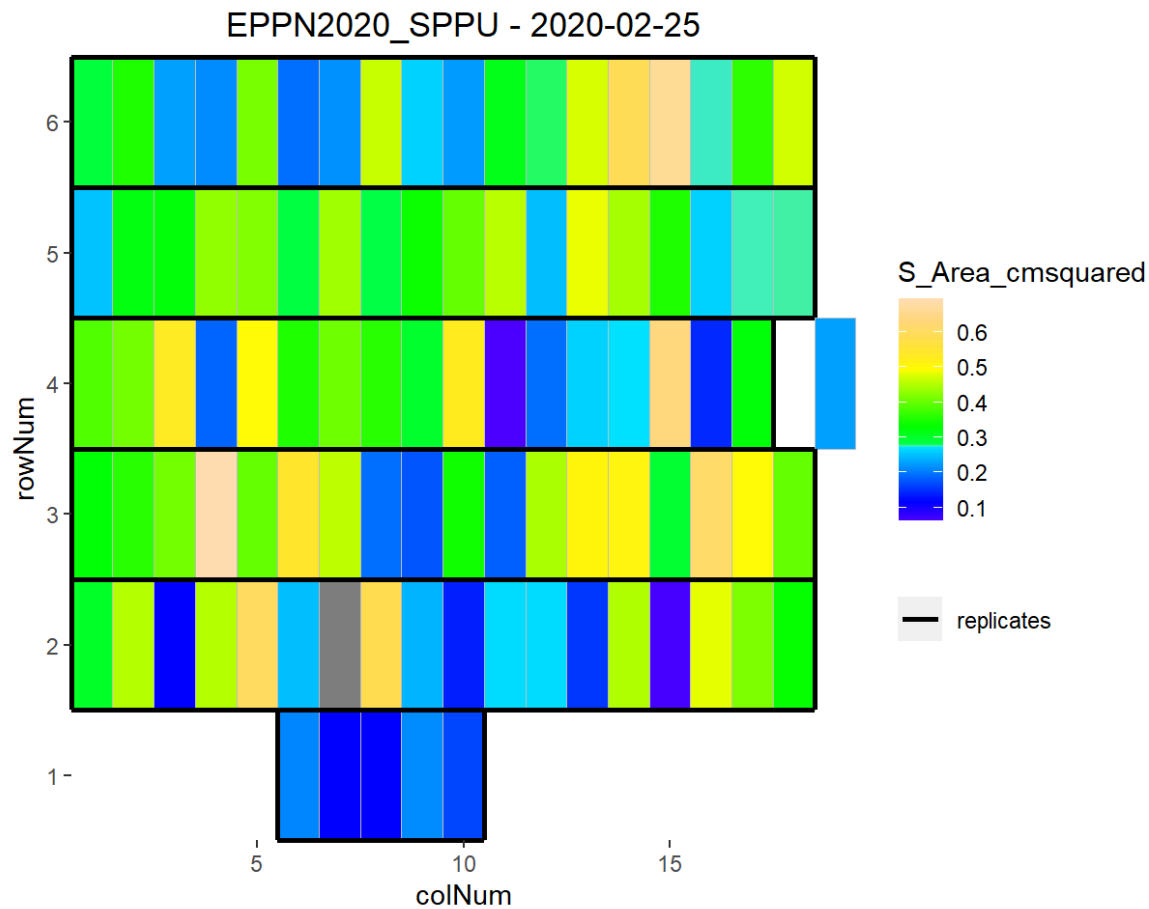
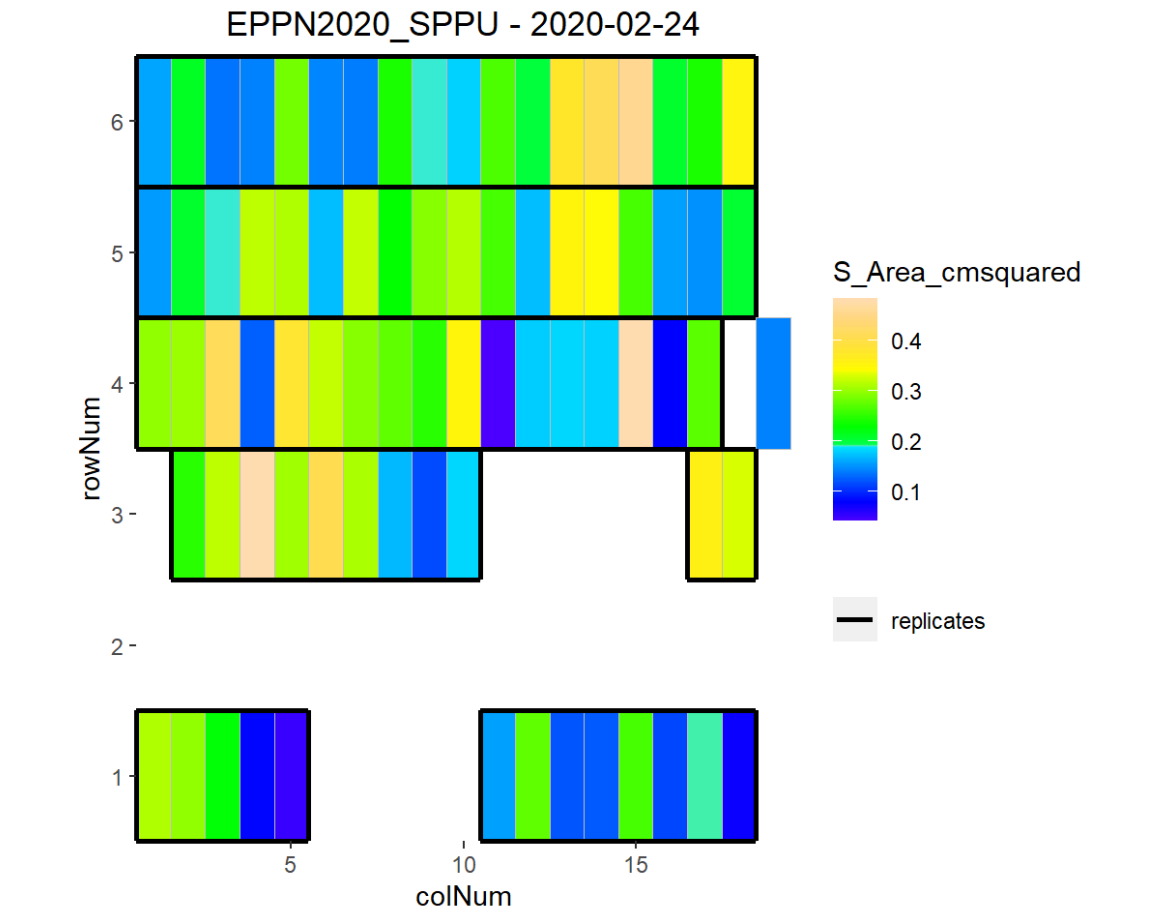


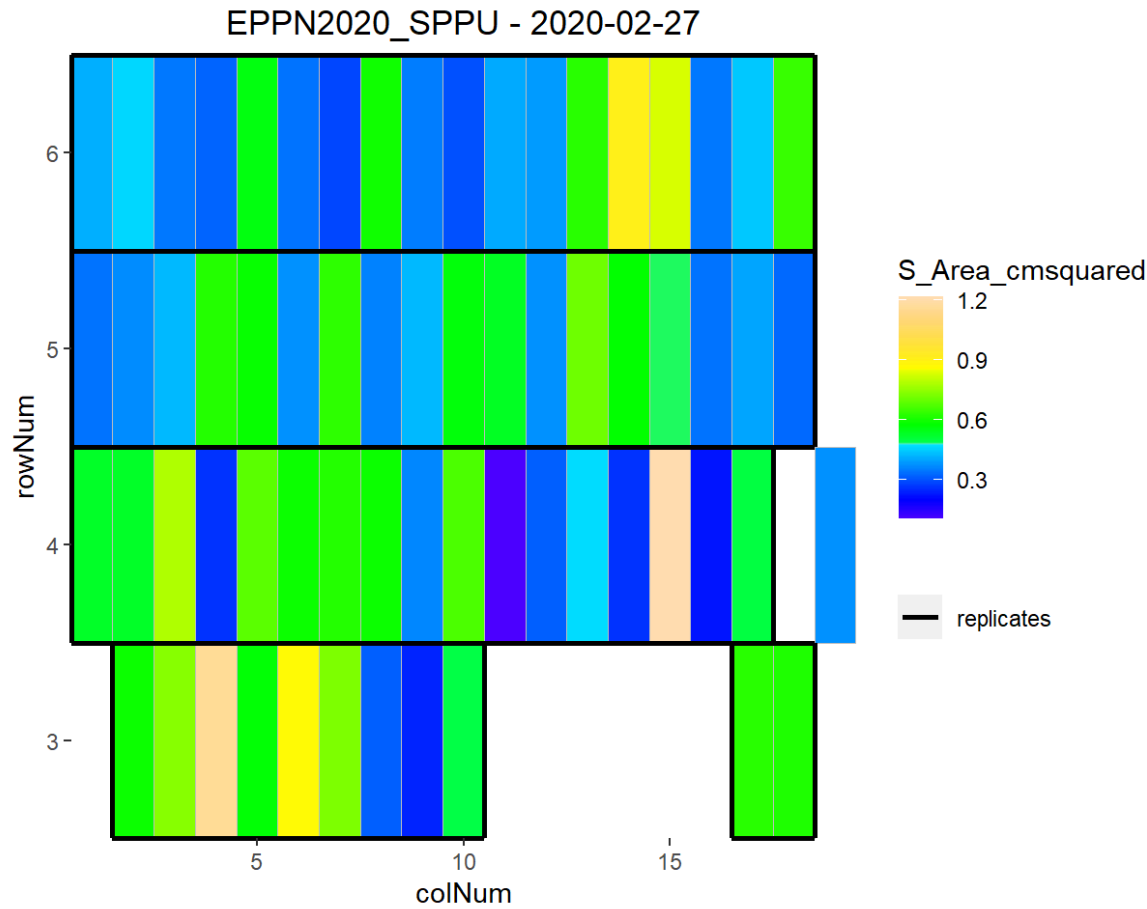
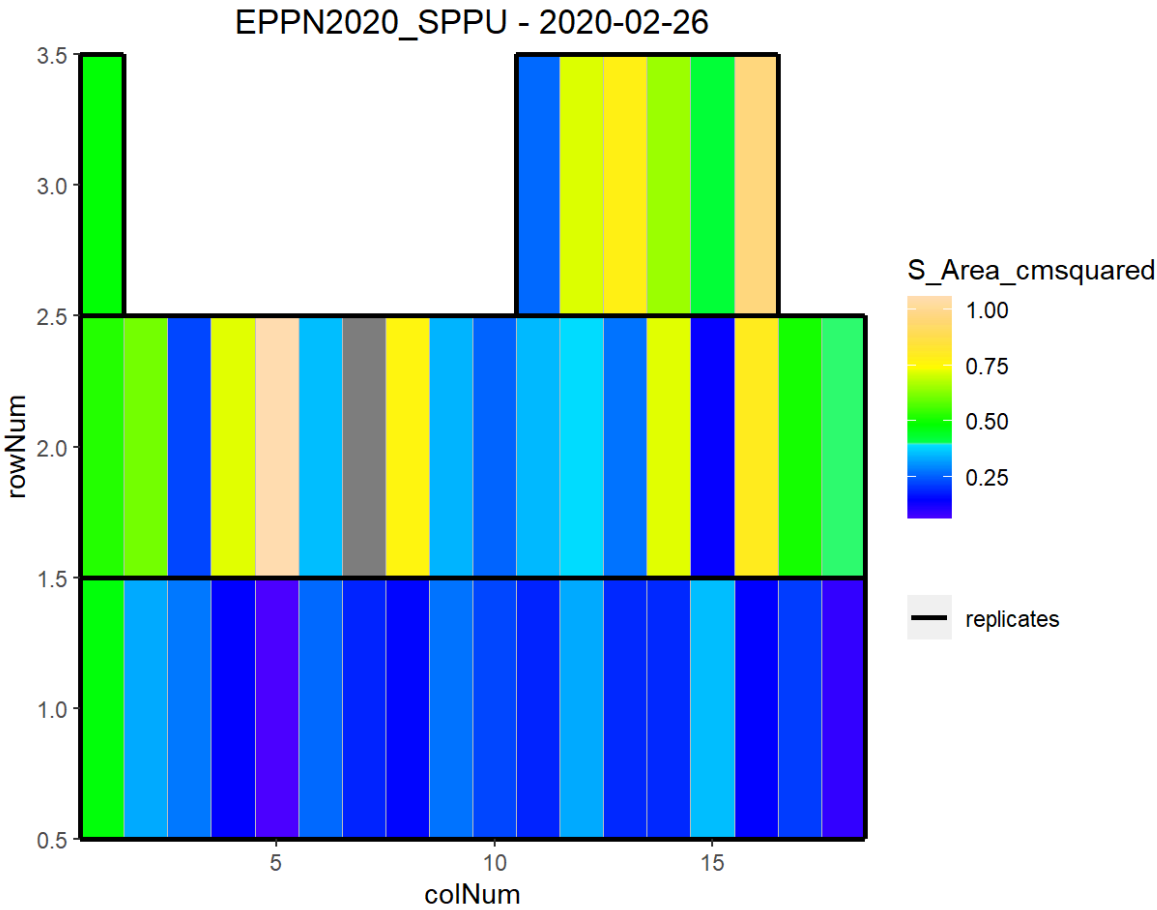


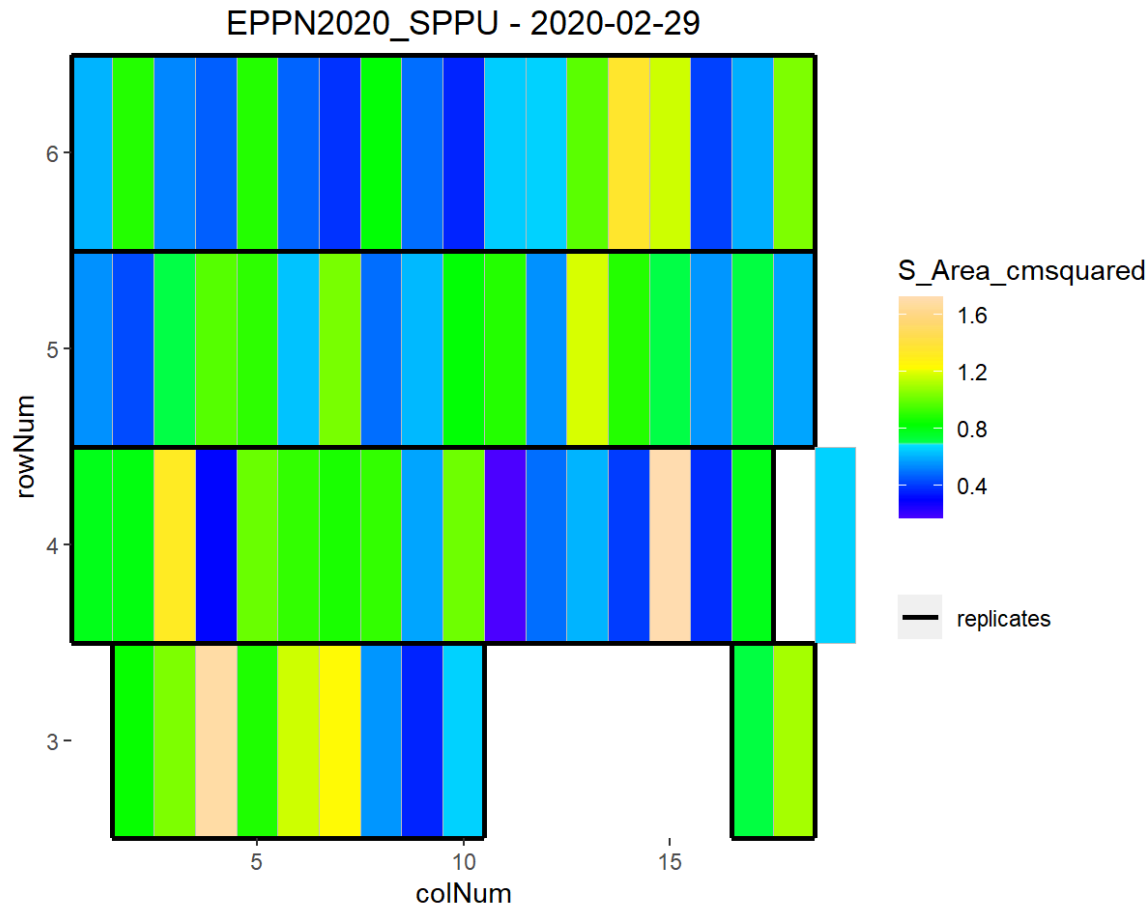
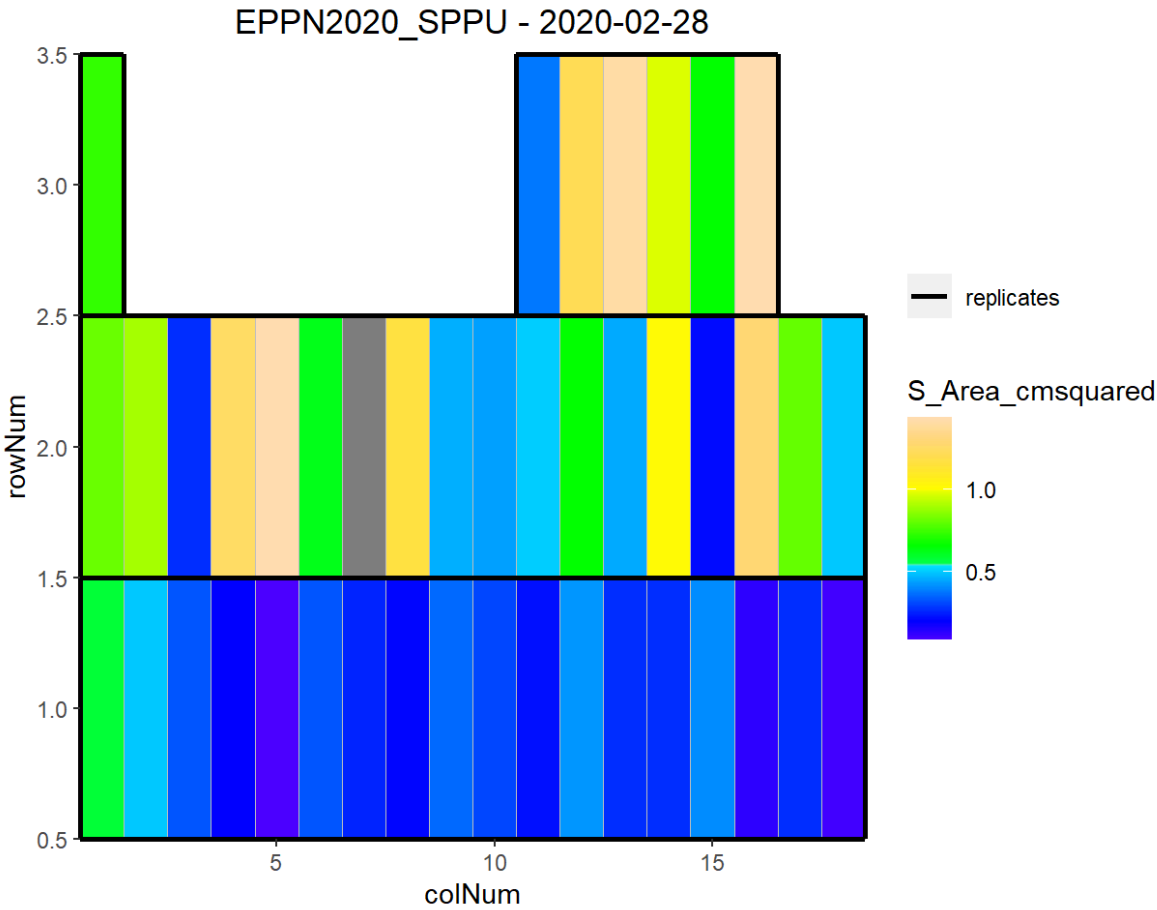


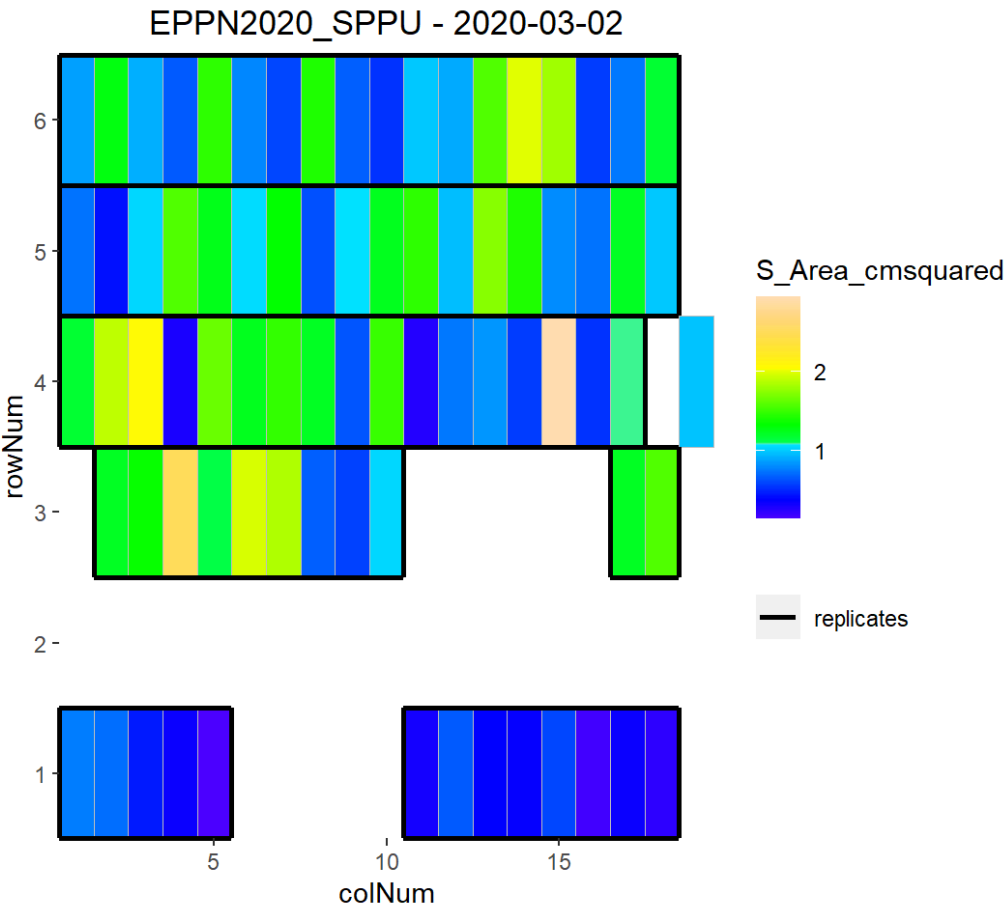
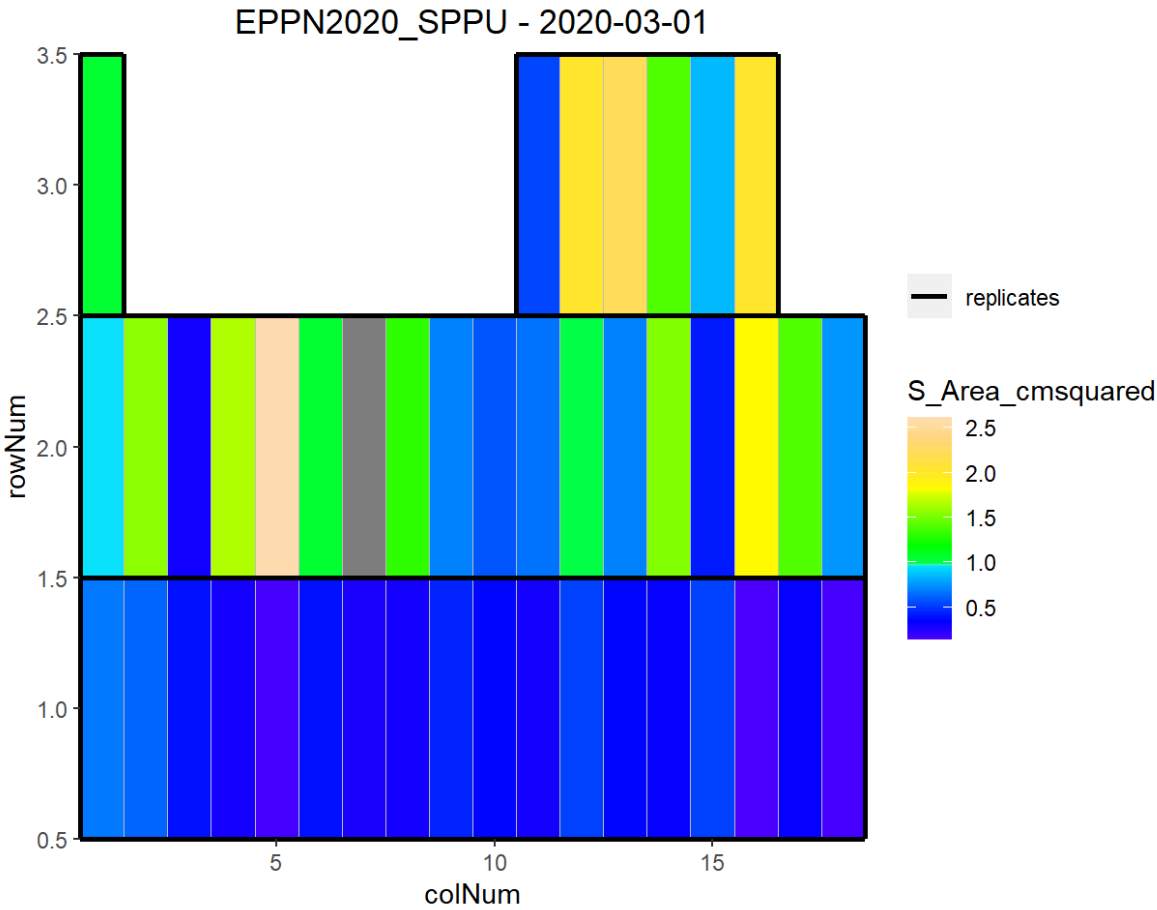


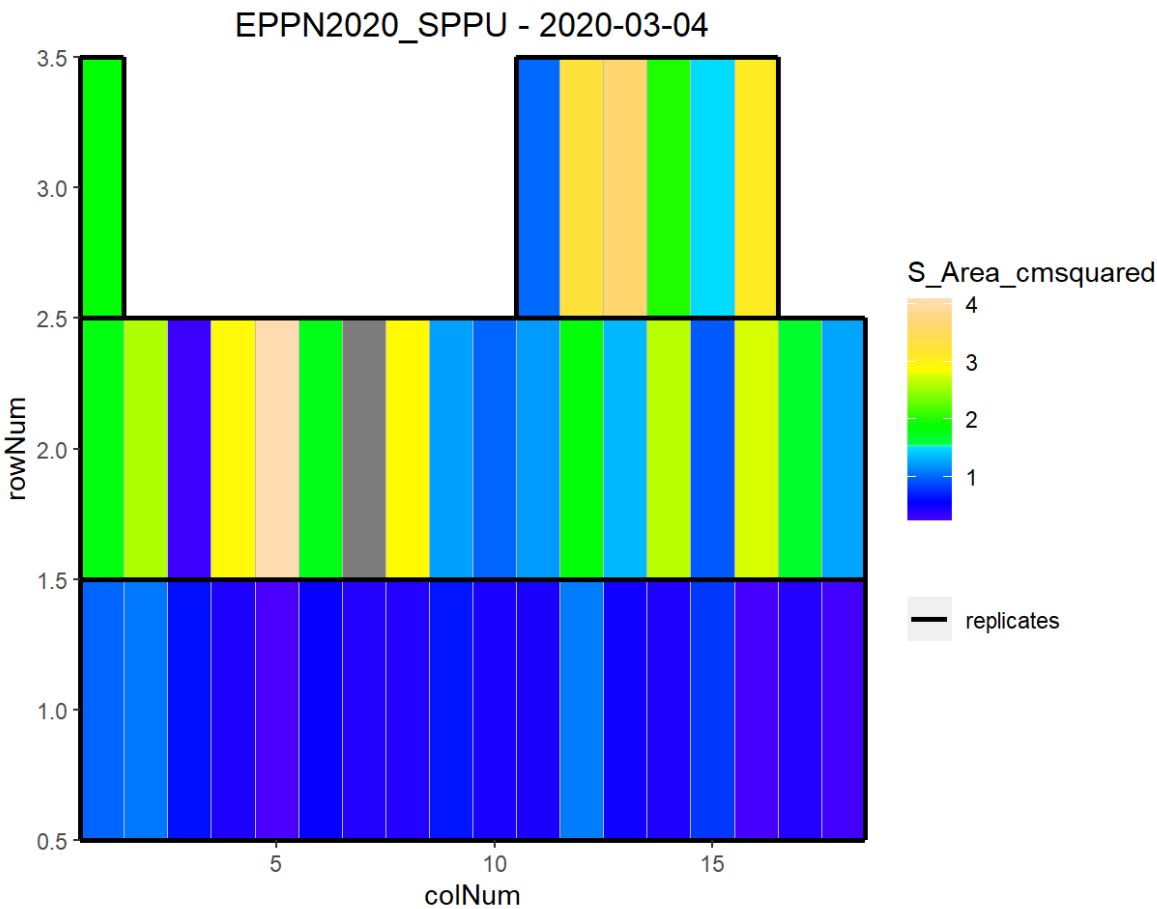
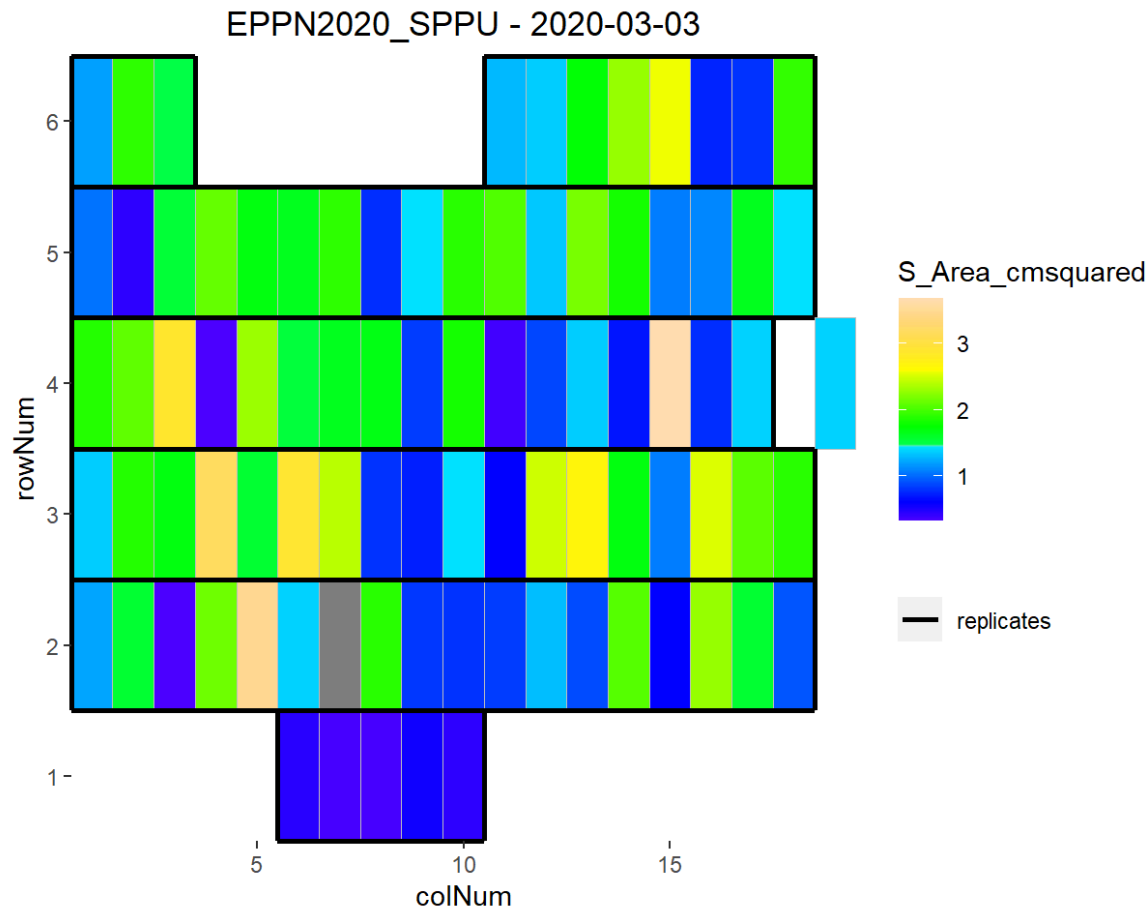


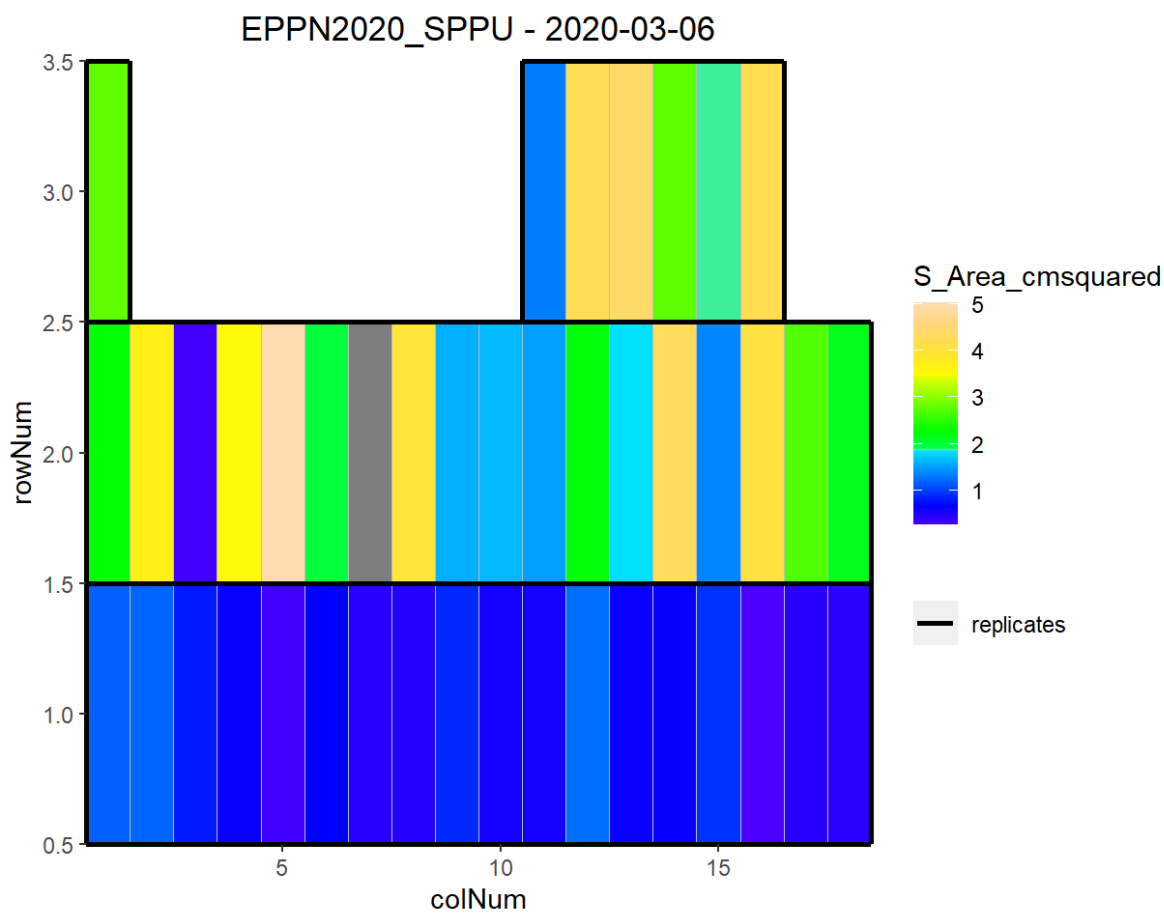
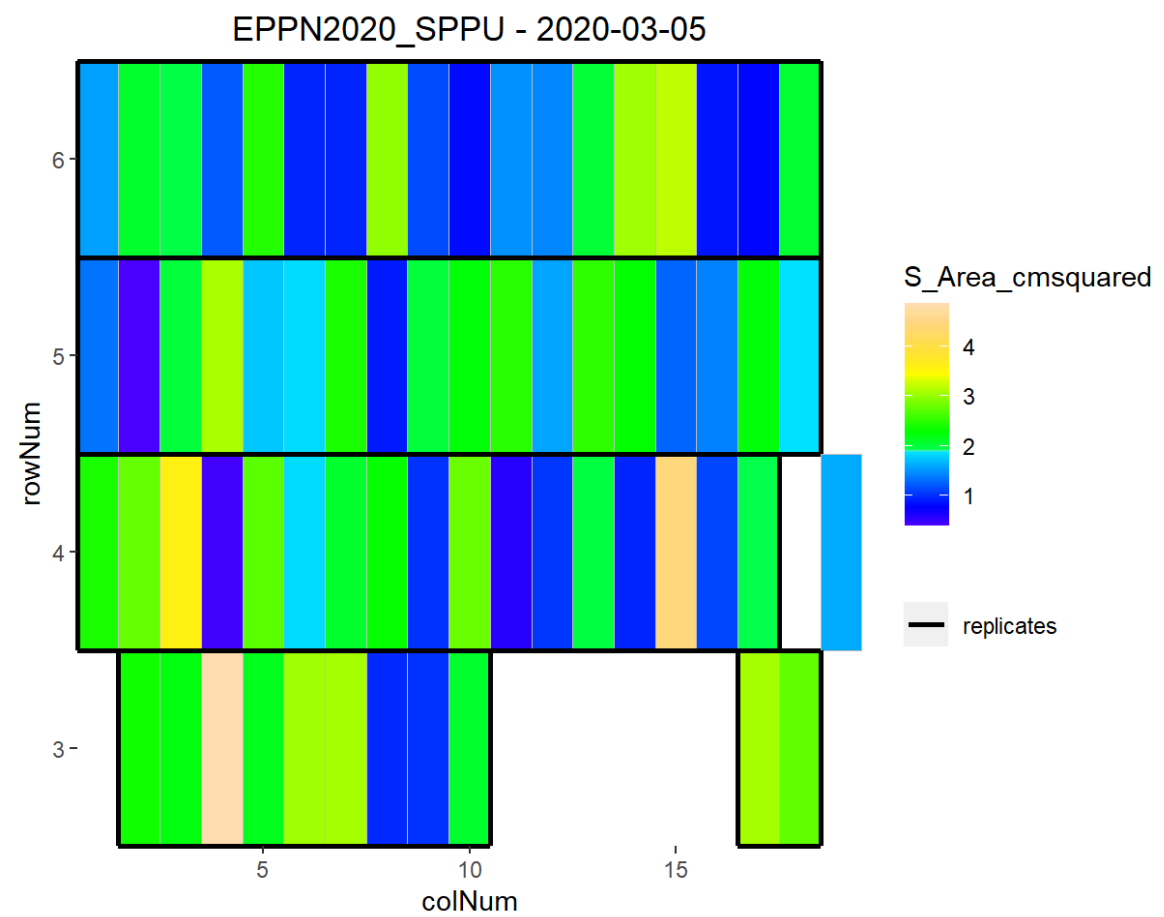


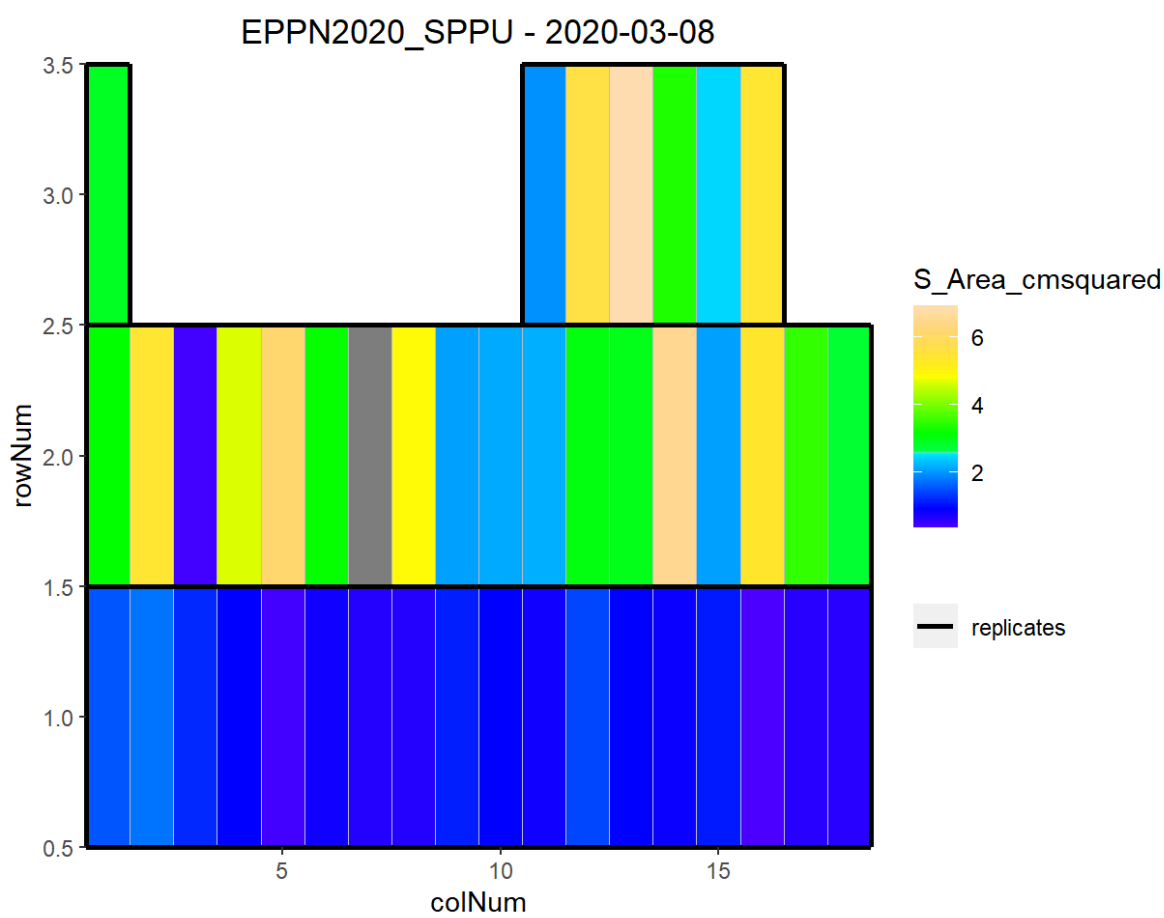
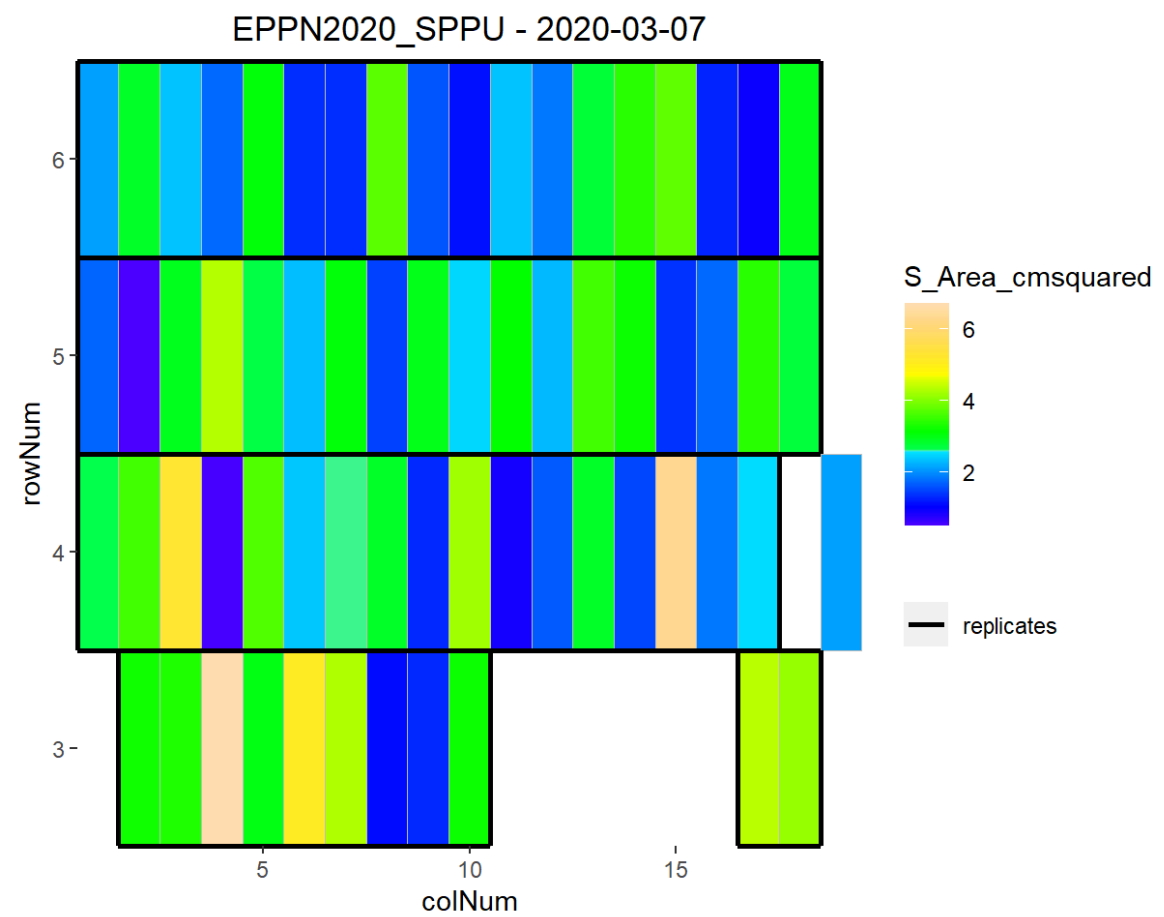


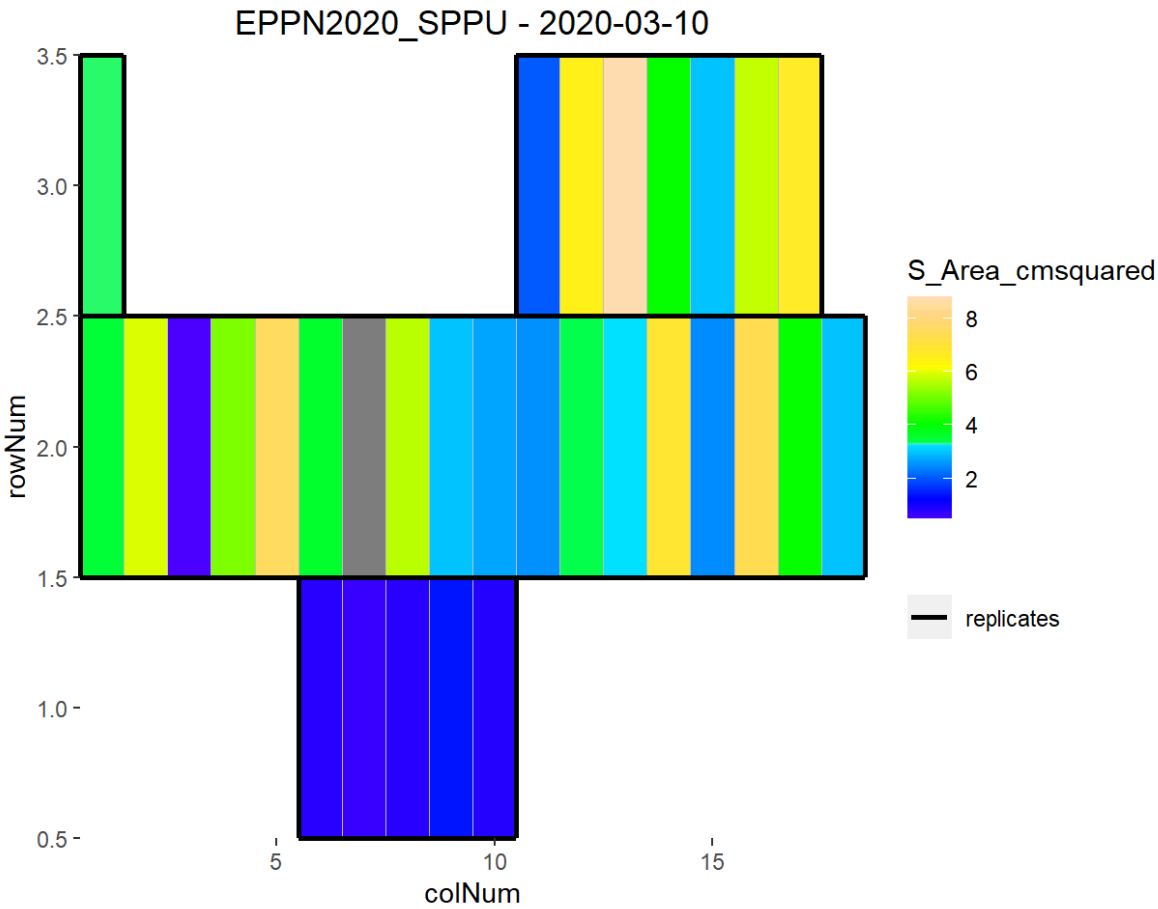
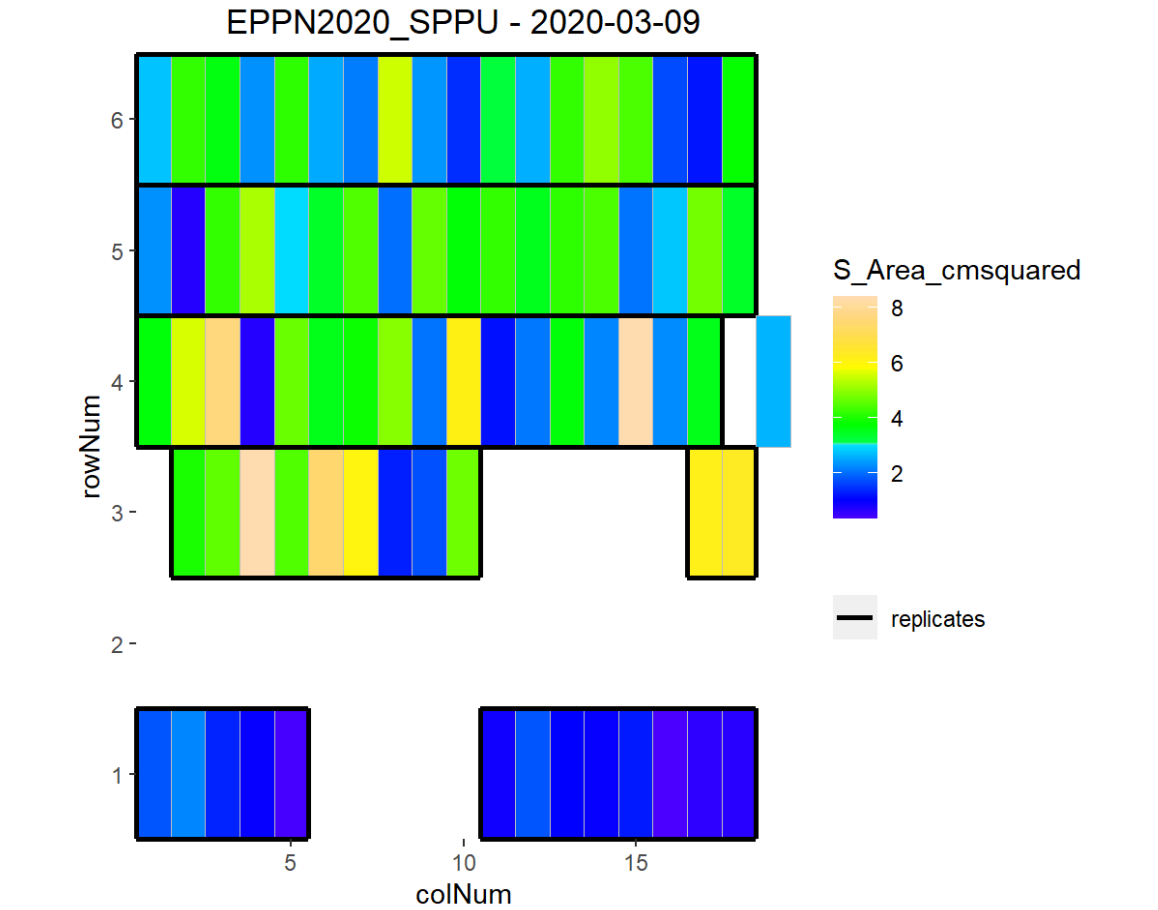










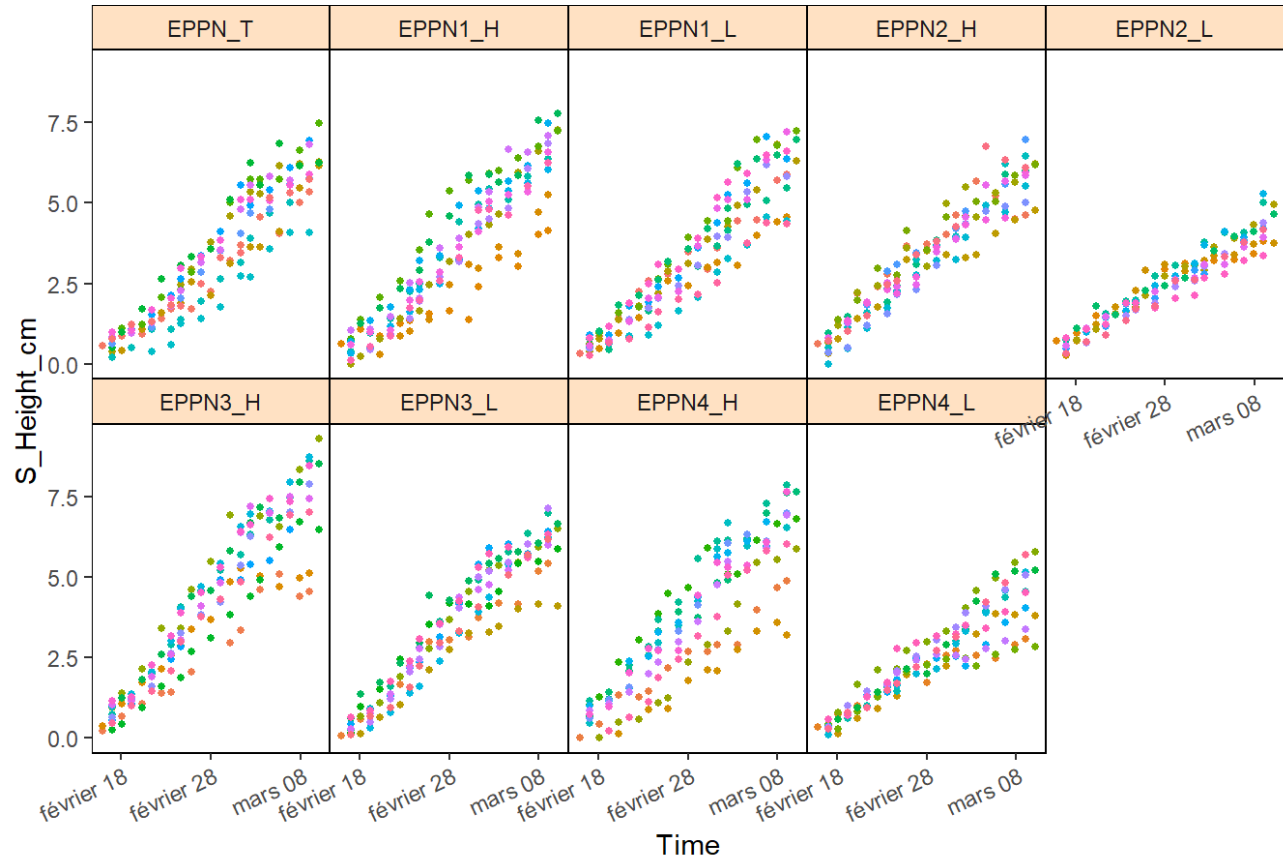


Check time course of raw data per time point

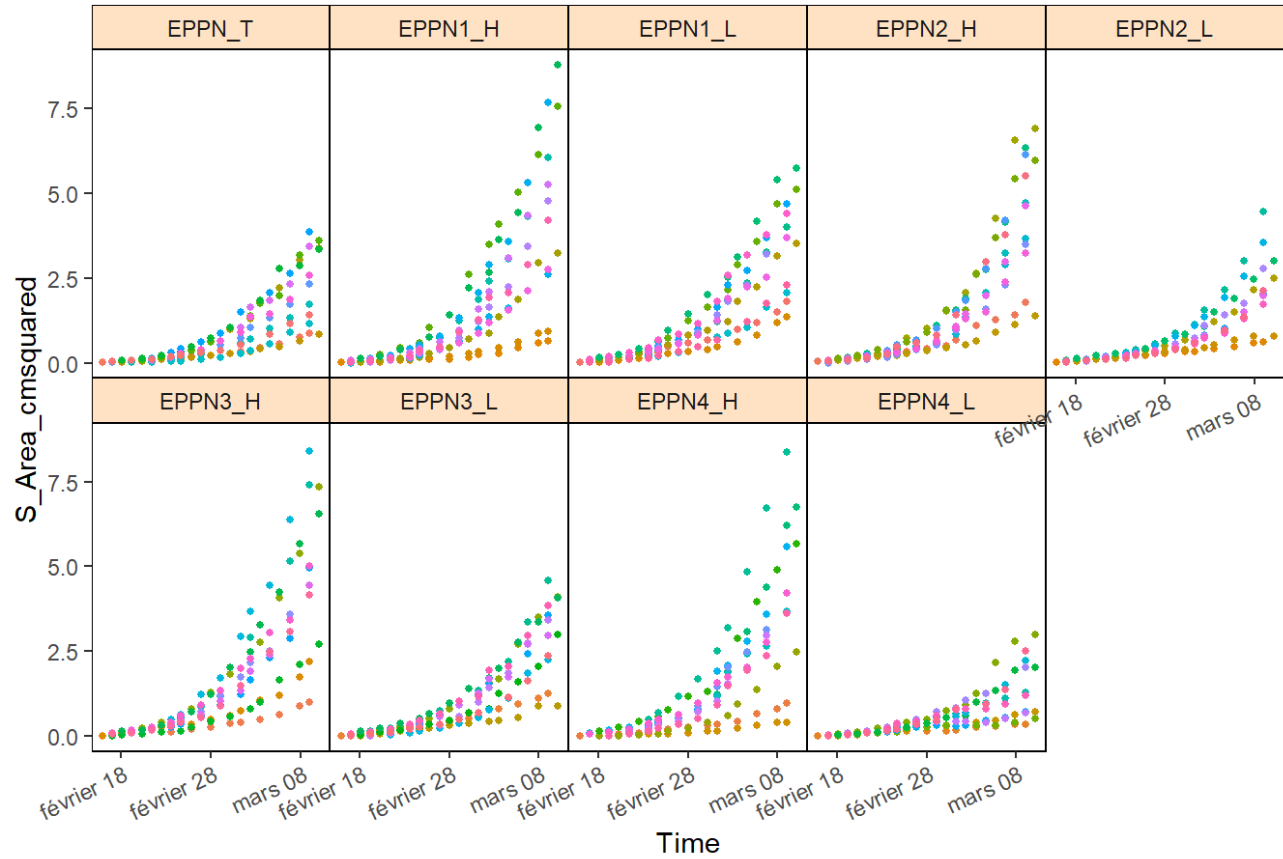


```
for (trait_name in traits) {  
  plot(timePoint_S,  
        traits = trait_name,  
        plotType = "raw")  
}
```

EPPN2020\_SPPU - S\_Height\_cm - raw data



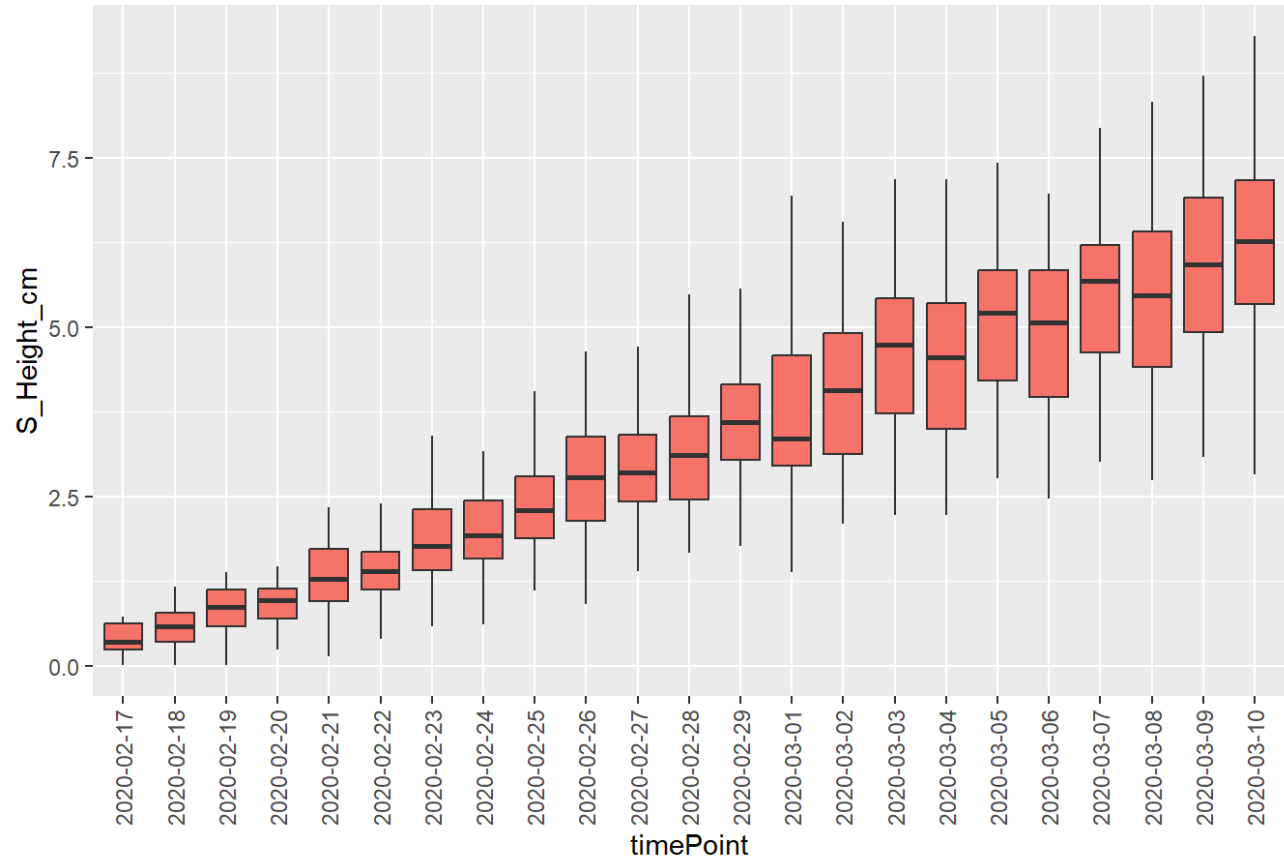
EPPN2020\_SPPU - S\_Area\_cmsquared - raw data



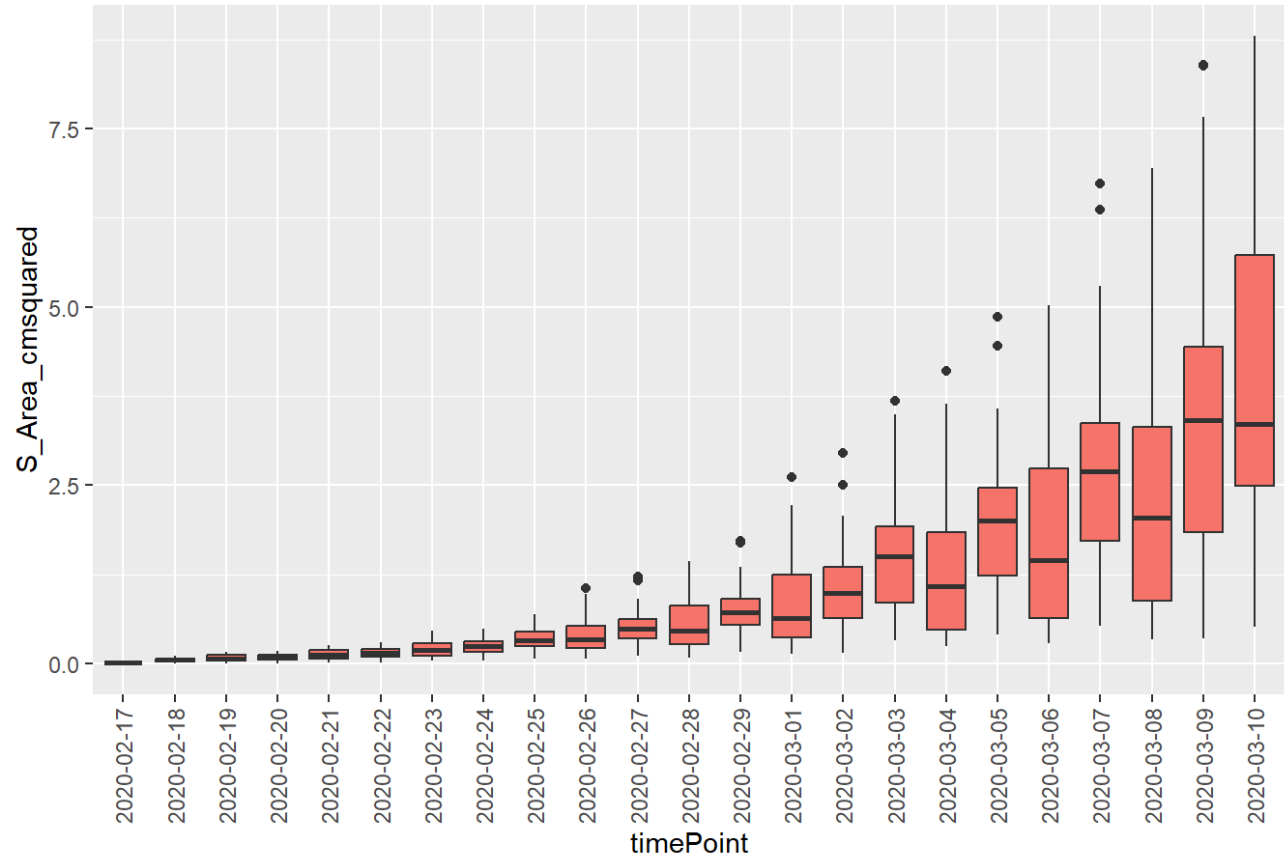
Check the boxplots of raw data per time point

```
for (trait_name in traits) {  
  plot(timePoint_S,  
        plotType = "box",  
        traits = trait_name)  
}
```

EPPN2020\_SPPU - S\_Height\_cm



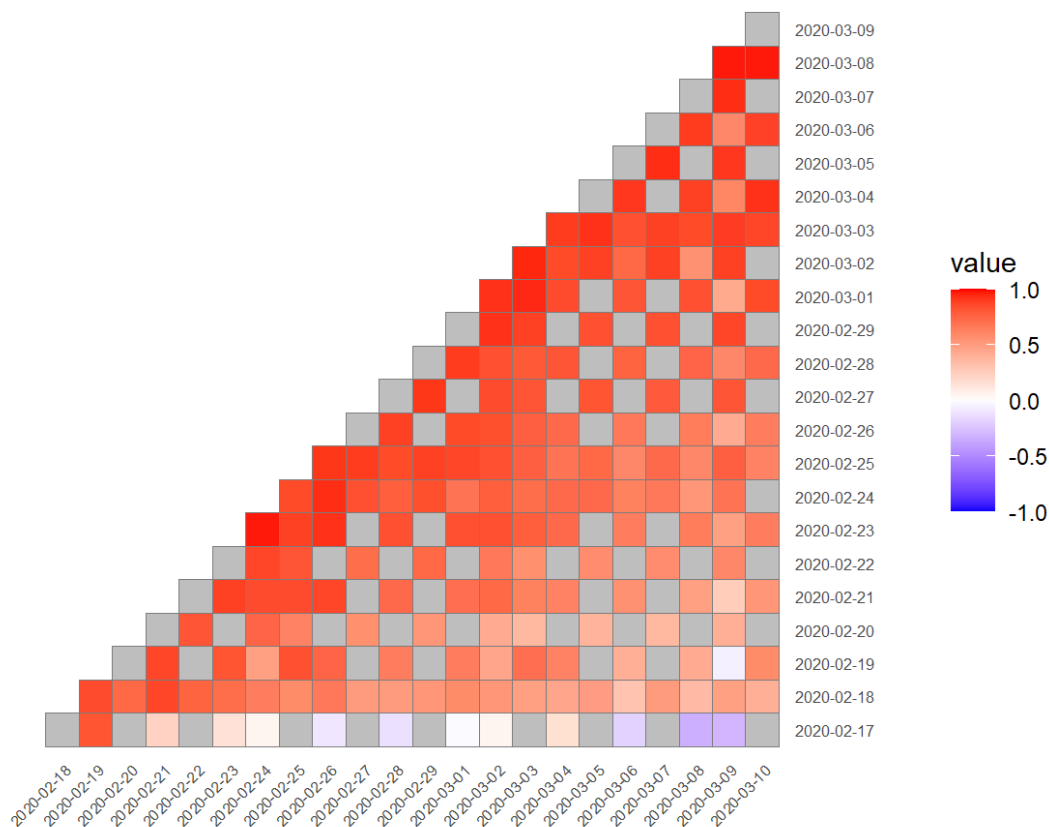
EPPN2020\_SPPU - S\_Area\_cmsquared



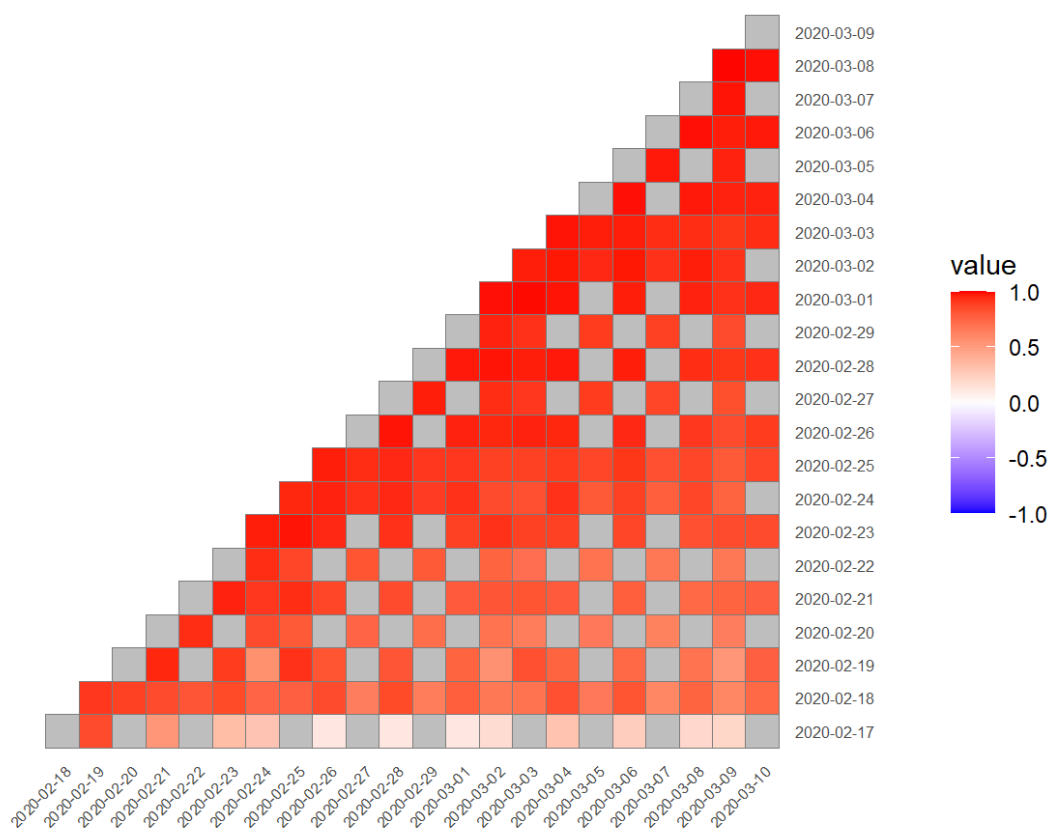
Check the correlation plots of raw data per time point

```
for (trait_name in traits) {  
  plot(timePoint_S,  
        plotType = "cor",  
        traits = trait_name)  
}
```

## EPPN2020\_SPPU - Correlations of timepoints for S\_Height\_cm



## EPPN2020\_SPPU - Correlations of timepoints for S\_Area\_cmsquared



## 1. Detection of outliers for single observations

Using the SingleOut detect and single functions. We select a subset of plants to adjust the settings for the confIntSize and nnLocfit.

```
plantSel<- c(1,2,3,4,5,6,7,8,9,10)
```

```
ci <- 5 # confidence interval
```

```
nn <- 0.8 # nearest neighbor
```

```
ce <- FALSE
```

```
for (trait_name in traits) {
```

```
  variable_name <- paste0("Single_test_", trait_name)
```

```
  single_test <- detectSingleOut(
```

```
    TP = timePoint_S,
```

```
    trait = trait_name,
```

```
    plotIds = plantSel,
```

```
    confIntSize = ci,
```

```
    nnLocfit = nn,
```

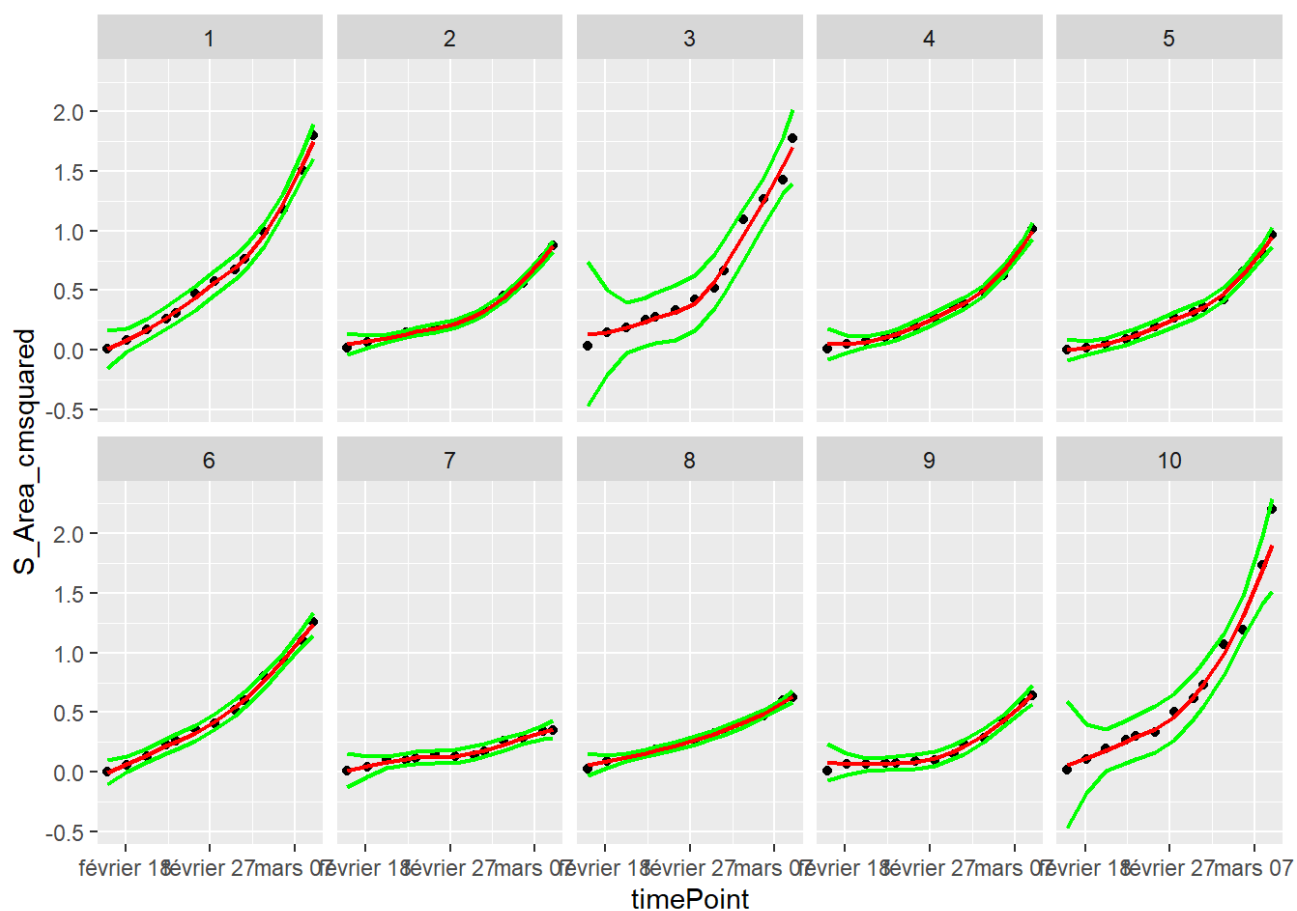
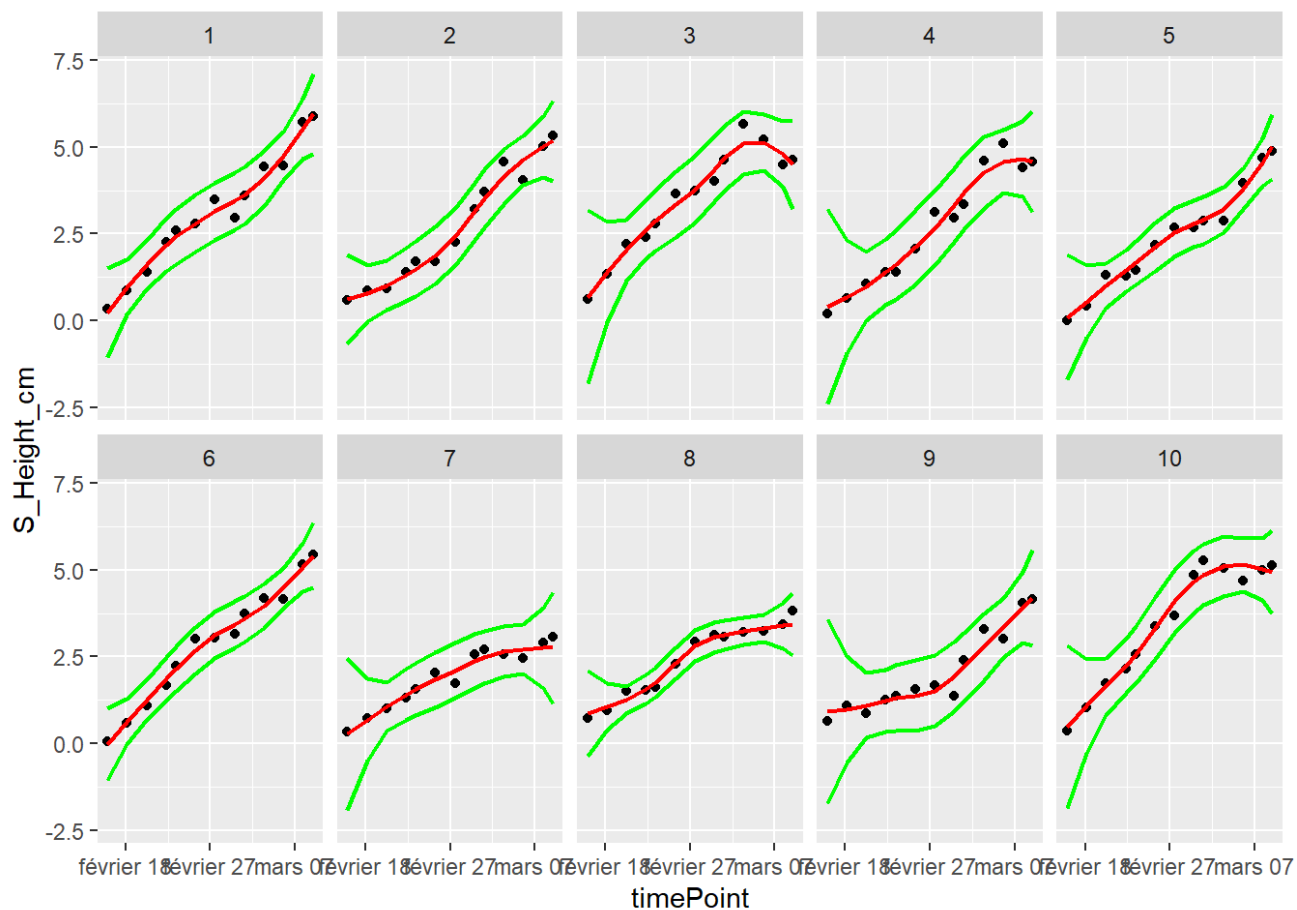
```
    checkEdges = TRUE # check for outlier values in start and end of experiment
```

```
  )
```

```
  assign(variable_name, single_test)
```

```
  plot(single_test, outOnly = FALSE)
```

```
}
```



We can then run on all plants of the data set.



```

for (trait_name in traits) {
  single_test_object_name <- paste0("Single_test_", trait_name)
  Single_test <- get(single_test_object_name)
  if (any(Single_test$outlier == 1)) {
    outliers_count <- with(Single_test[Single_test$outlier == 1,], table(timePoint))
    print(trait_name)
    print(outliers_count)

    Single_outliers <- removeSingleOut(timePoint_S, Single_test)
    assign(paste0("Single_outliers_", trait_name), Single_outliers)

    readr::write_tsv(Single_test, sprintf("%s/single_outliers_%s.tsv", datadir, trait_name))
  } else {
    cat("No outlier for", trait_name, "\n")
  }
}

```

```

## No outlier for S_Height_cm
## No outlier for S_Area_cmsquared

```

## 2. Correction for spatial trends

Fit a model for all time points with no extra fixed effects.

```

#for (trait_name in traits) {
#  single_outliers_name <- paste0("Single_outliers_", trait_name)
#
#  if (exists(single_outliers_name)) {
#    Single_outliers <- get(single_outliers_name)
#
#    assign(paste0("modTP_", trait_name),
#           fitModels(TP = Single_outliers,
#                     trait = trait_name,
#                     geno.decomp = "Plant_type"))
#  } else {
#    assign(paste0("modTP_", trait_name),
#           fitModels(TP = timePoint_S,
#                     trait = trait_name,
#                     geno.decomp = "Plant_type"))
#  }
#}

print("Erreur dans `contrasts<-`(`*tmp*`, value = contr.funs[1 + isOF[nn]]) :
      les contrastes ne peuvent être appliqués qu'aux facteurs ayant au moins deux niveau
x")

```

```

## [1] "Erreur dans `contrasts<-`(`*tmp*`, value = contr.funs[1 + isOF[nn]]) : \n les
      contrastes ne peuvent être appliqués qu'aux facteurs ayant au moins deux niveaux"

```

## Model visualisation

### 3. Outlier detection for series of observations

By using the splines.

fitModels

```
for (trait_name in traits) {  
  Spatial_Corrected_name <- paste0("Spatial_Corrected_", trait_name)  
  modTP_name <- paste0("modTP_", trait_name)  
}
```

Plot the splines for a plant selection

detectSerieOut

removeSerieOut

### 4. With the cleaned data, re-do the spatial correction

This is used to compare the values before and after.

Need to write a for loop for all the variables.

For S\_Height\_cm

Estimation of parameter from time series