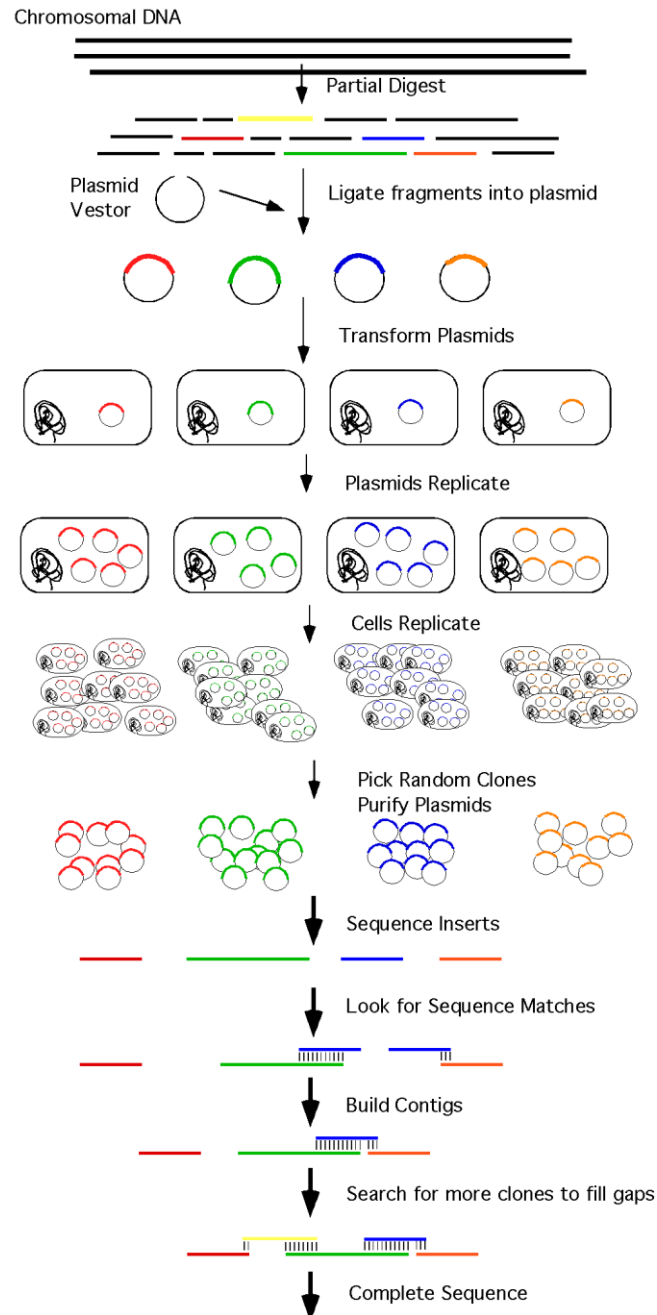


# **Lecture 3: Genomic and cDNA Libraries**

# DNA Library

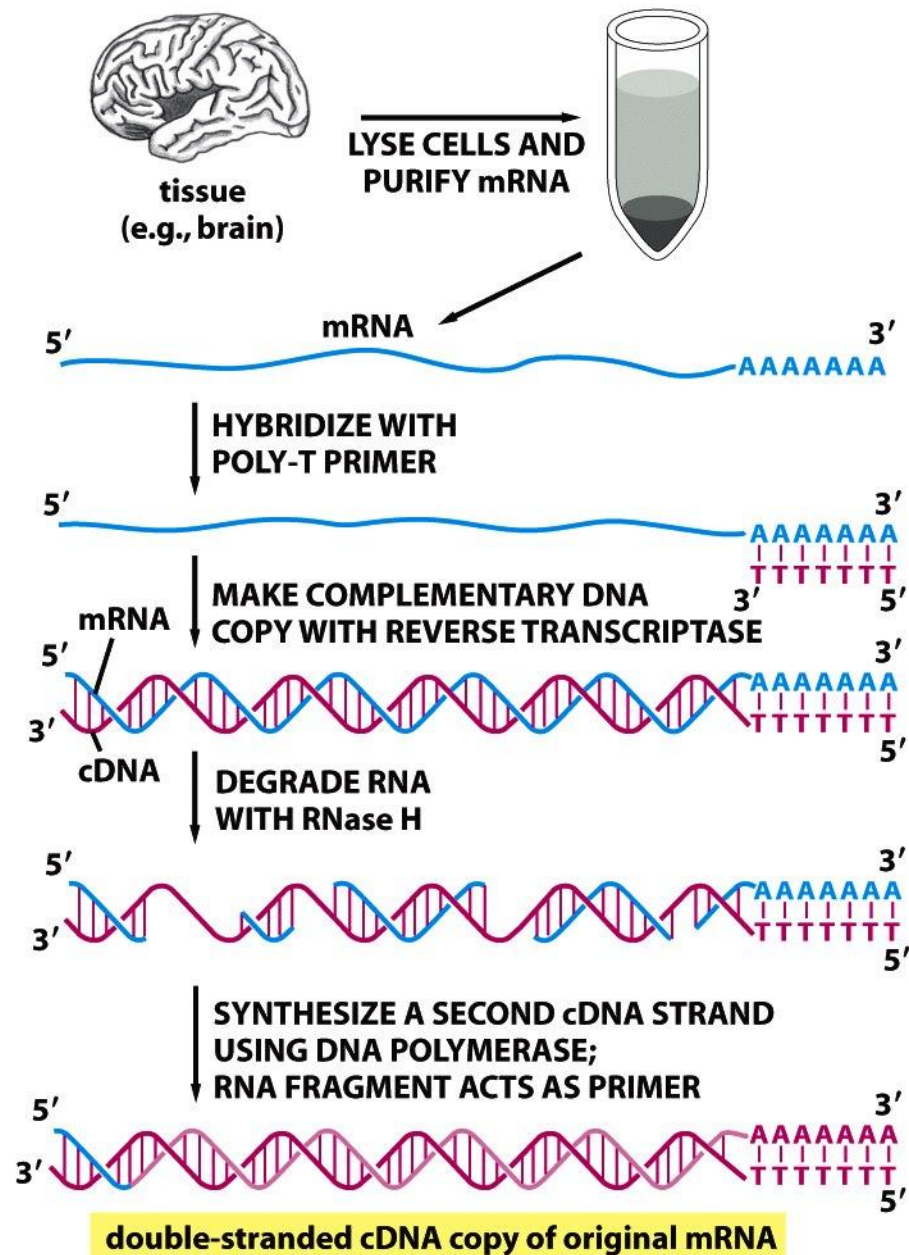
- A DNA library is a **collection** of DNA fragments that have been **cloned** into **vectors** so that DNA fragments of interest can be **identified** and **isolated** for further study.
- There basically are two kinds of libraries: **genomic DNA** and **cDNA** libraries.
- Genomic DNA libraries contain **large** fragments of DNA in either bacteriophages or bacterial or P1-derived artificial chromosomes (BACs and PACs).
- cDNA libraries are made with cloned, **reverse-transcribed mRNA**, and, therefore, **lack** DNA sequences corresponding to genomic regions that are **not expressed**, such as introns and 5'- and 3'-noncoding regions.
- cDNA libraries generally contain much **smaller** fragments than genomic DNA libraries, and are usually cloned into **plasmid** vectors.

# Construction and Analysis of a Genomic DNA Library

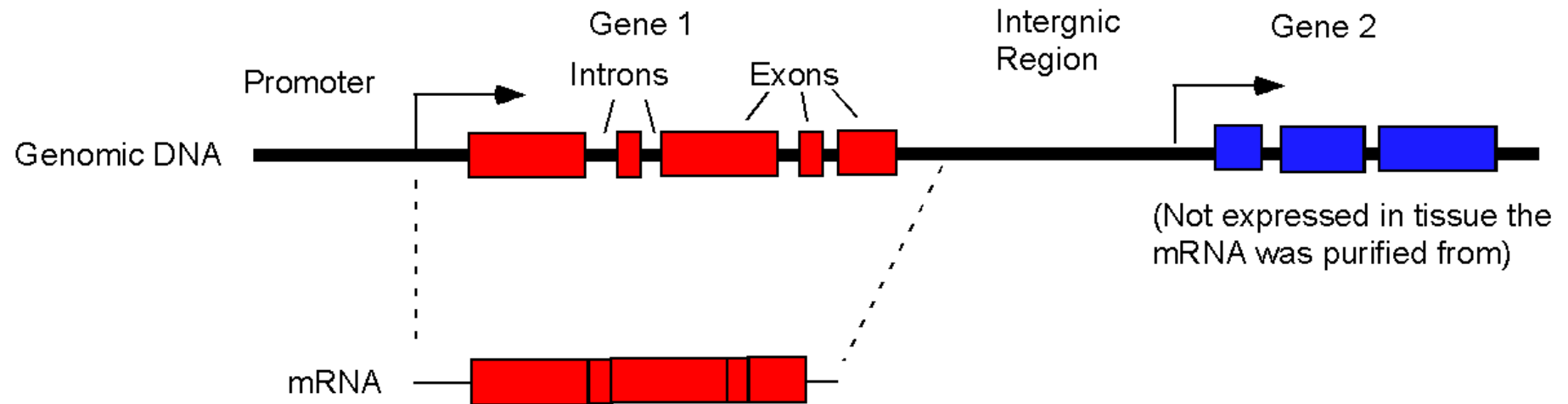


# Synthesis of cDNA

- **Total** mRNA is extracted from a tissue and **reverse transcriptase** produces DNA copies (**cDNA**) of the mRNA molecules.
- A short oligonucleotide complementary to the **poly-A tail** at the 3' end of the mRNA is first hybridized to the RNA into a cDNA chain, thereby forming a **DNA/RNA hybrid** helix.
- Treating the DNA/RNA hybrid with **RNase H** creates nicks and gaps in the RNA strand.
- RNase H is a **non-specific** endonuclease that cleaves the 3'-O-P-bond of RNA in a DNA/RNA duplex to produce 3'-hydroxyl and 5'-phosphate terminated products.
- **DNA polymerase** then copies the remaining single-stranded cDNA into **double-stranded** cDNA.
- The fragment of the original mRNA is the **primer** for this synthesis reaction.
- Because the DNA polymerase used to synthesize the second DNA strand can synthesize through the bound RNA molecules, the RNA fragment that is **base-paired to the 3' end of the first DNA strand** usually acts as the **primer** for the **final** product of the **second strand** synthesis.



# Differences Between a Genomic and cDNA Library



## Genomic Library

Promoters

Introns

Intergenic

Non-expressed genes

## cDNA Library

Expressed genes

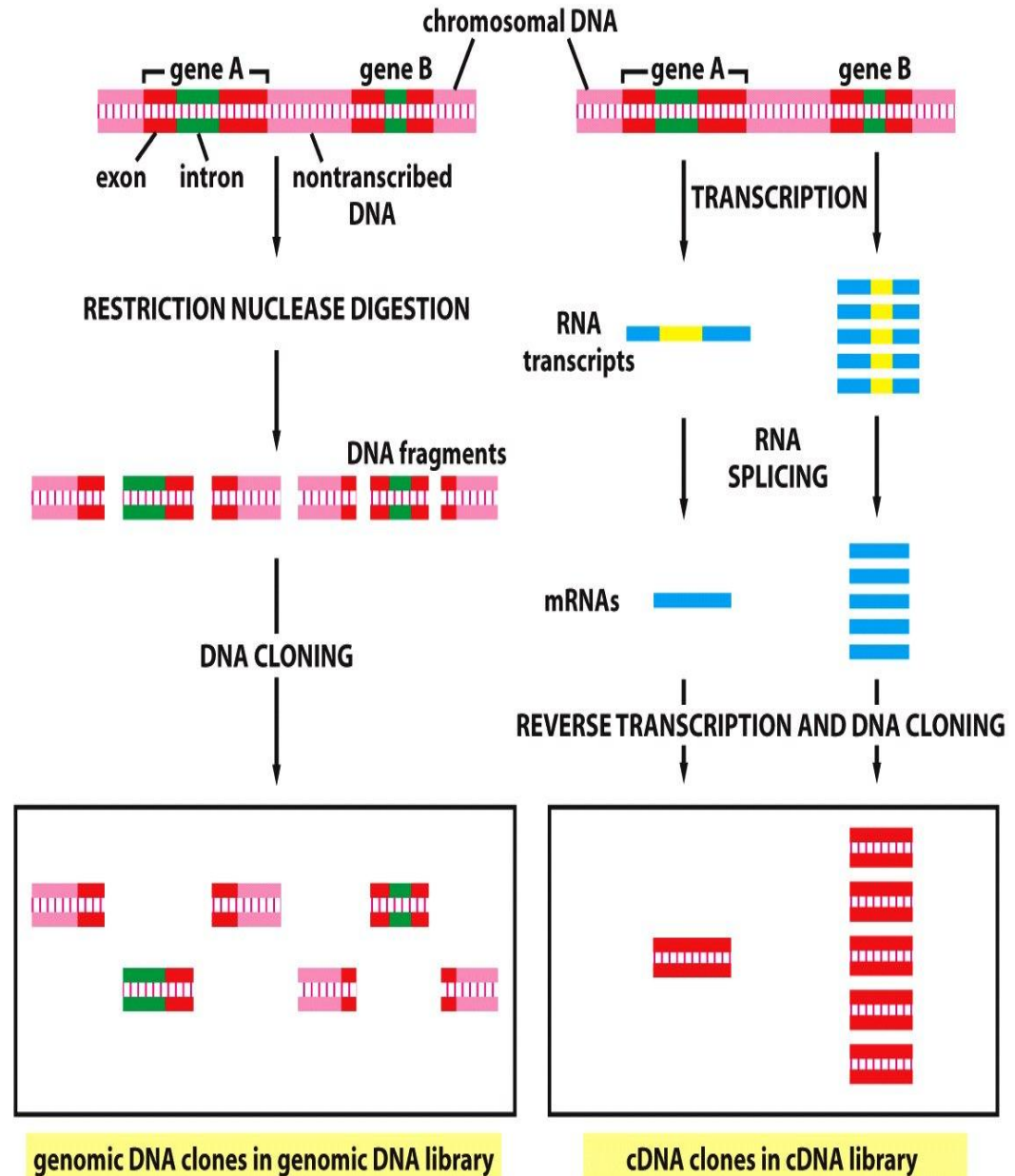
Transcription start sites

Open reading frames (ORFs)

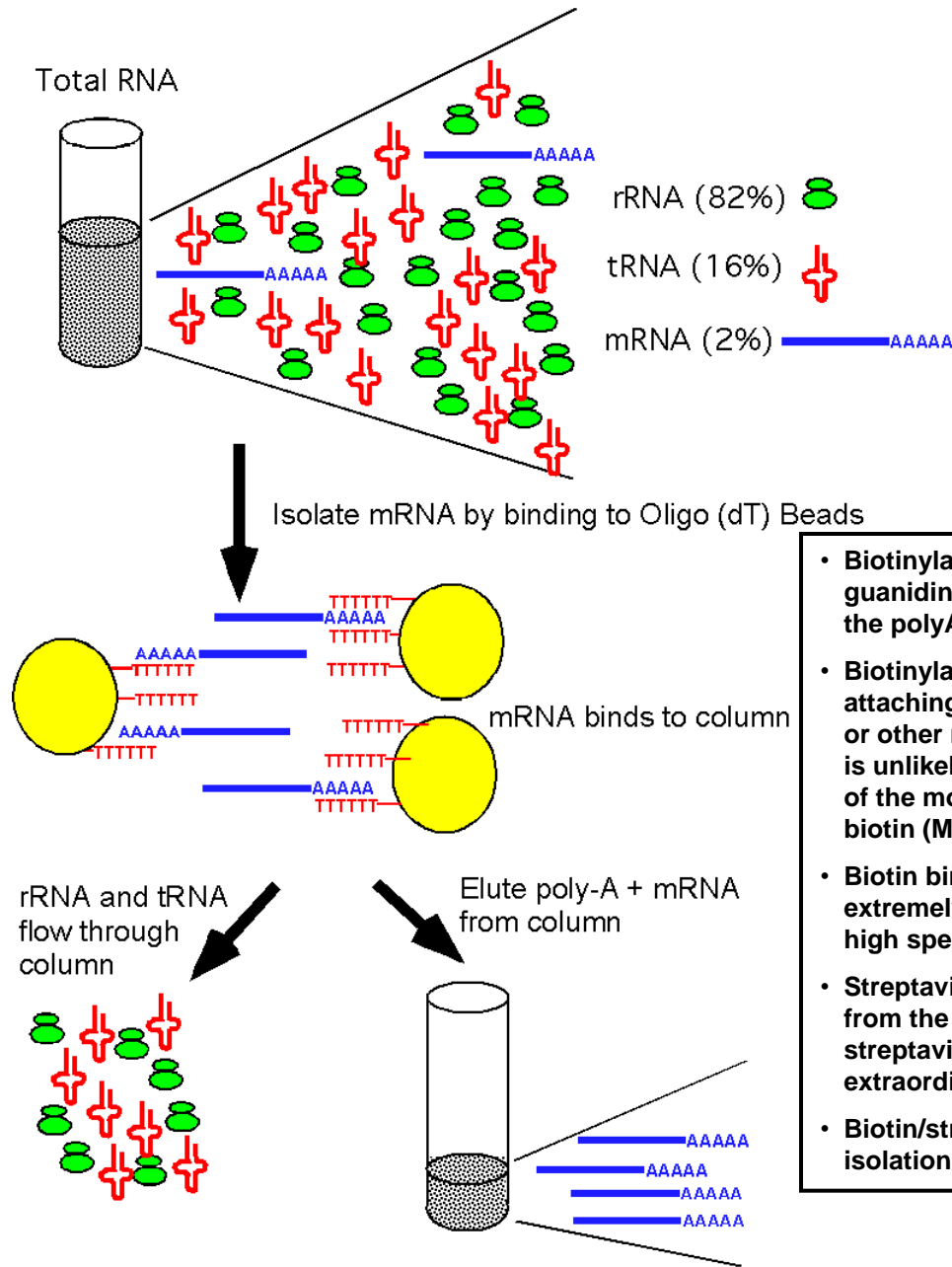
Splice points

# Genomic DNA and cDNA Clones

- Gene A is **infrequently** transcribed, whereas gene B is **frequently** transcribed, and **both** genes contain **introns** (green).
- In a genomic DNA library, both the **introns** and the **non-transcribed DNA** (pink) are included in the clones, and most clones contain, at most, **only part** of the coding sequence of a gene (red).
- In the cDNA clones, the intron sequences (yellow) have been removed by **RNA splicing** during the formation of the mRNA (blue), and a **continuous** coding sequence is, therefore, present in each clone.
- Because gene B is transcribed **more frequently** than gene A in the cells from which the cDNA library was made, it is represented much **more frequently** than A in the **cDNA library**.
- In contrast, A and B are in principle represented **equally** in the genomic **DNA library**.



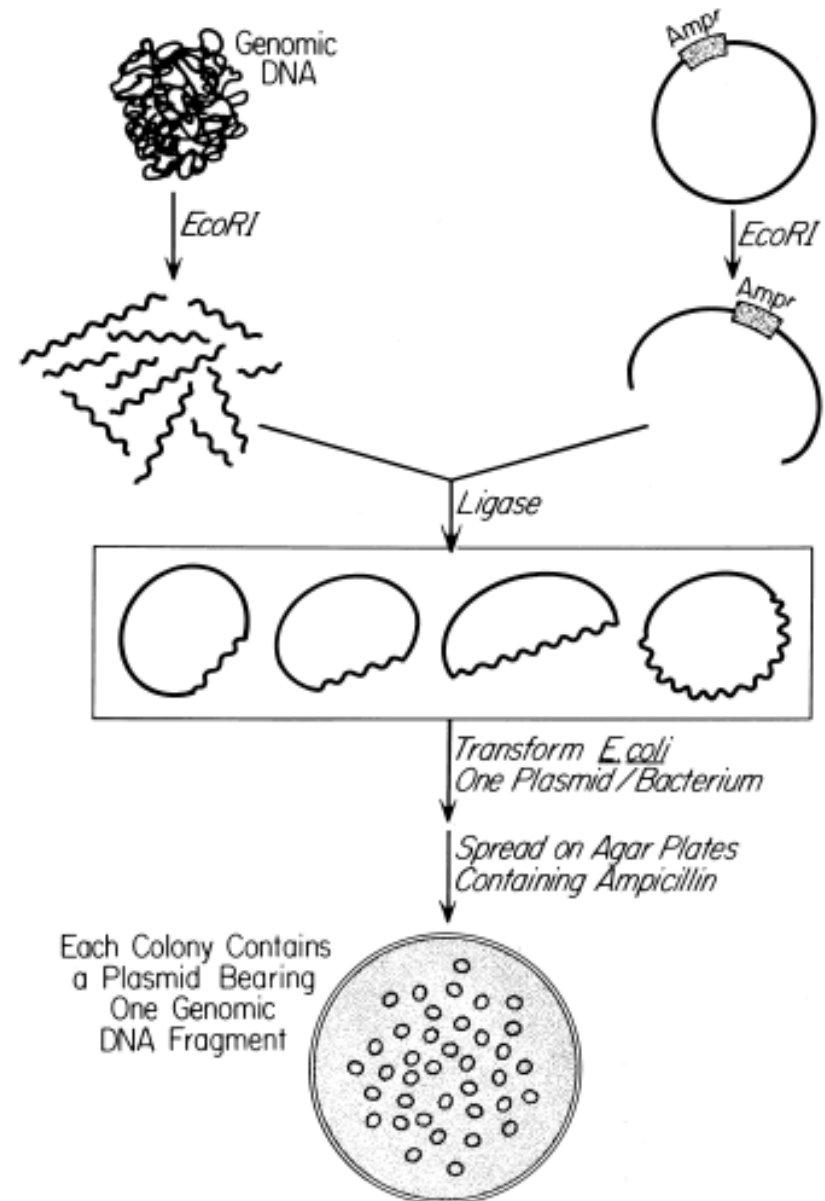
# Purification of mRNA



- Biotinylated oligo dT is added to guanidinium-treated cells, and anneals to the polyA tail of mRNAs.
- Biotinylation is the process of covalently attaching biotin to a protein, nucleic acid or other molecule; it is rapid, specific and is unlikely to perturb the natural function of the molecule due to the small size of biotin (MW = 244.31 g/mol).
- Biotin binds to streptavidin and avidin with extremely high affinity, fast on-rate and high specificity,
- Streptavidin is a 52.8 kDa-protein purified from the bacterium *Streptomyces avidinii*; streptavidin homo-tetramers have an extraordinarily high affinity for biotin.
- Biotin/streptavidin interactions permit isolation of the mRNA/oligo dT complexes.

# Amplification of Genomic Library

- A few nanograms of foreign DNA is digested with EcoRI.
- The DNA must contain the **same** restriction endonuclease recognition sites as the vector.
- Plasmid vector is also digested with EcoRI to create a linear DNA molecule.
- The “**sticky**” single-stranded ends of the foreign DNA align and base-pair with the **complementary** “sticky ends” of the plasmid, after which DNA ligase **covalently bonds** foreign DNA to plasmid DNA.
- This recombinant DNA is introduced into *E. coli* by **transformation**.
- The plasmid contains a bacterial **origin of replication** so that as the bacterial culture grows, plasmids replicate resulting in **several** copies in each bacterium.
- When the culture has grown to sufficient size, plasmid DNA is isolated and **foreign DNA** is **cut** from the plasmid using EcoRI.
- The resulting yield will often be **milligrams** of DNA, i.e., **>100-fold** amplification.



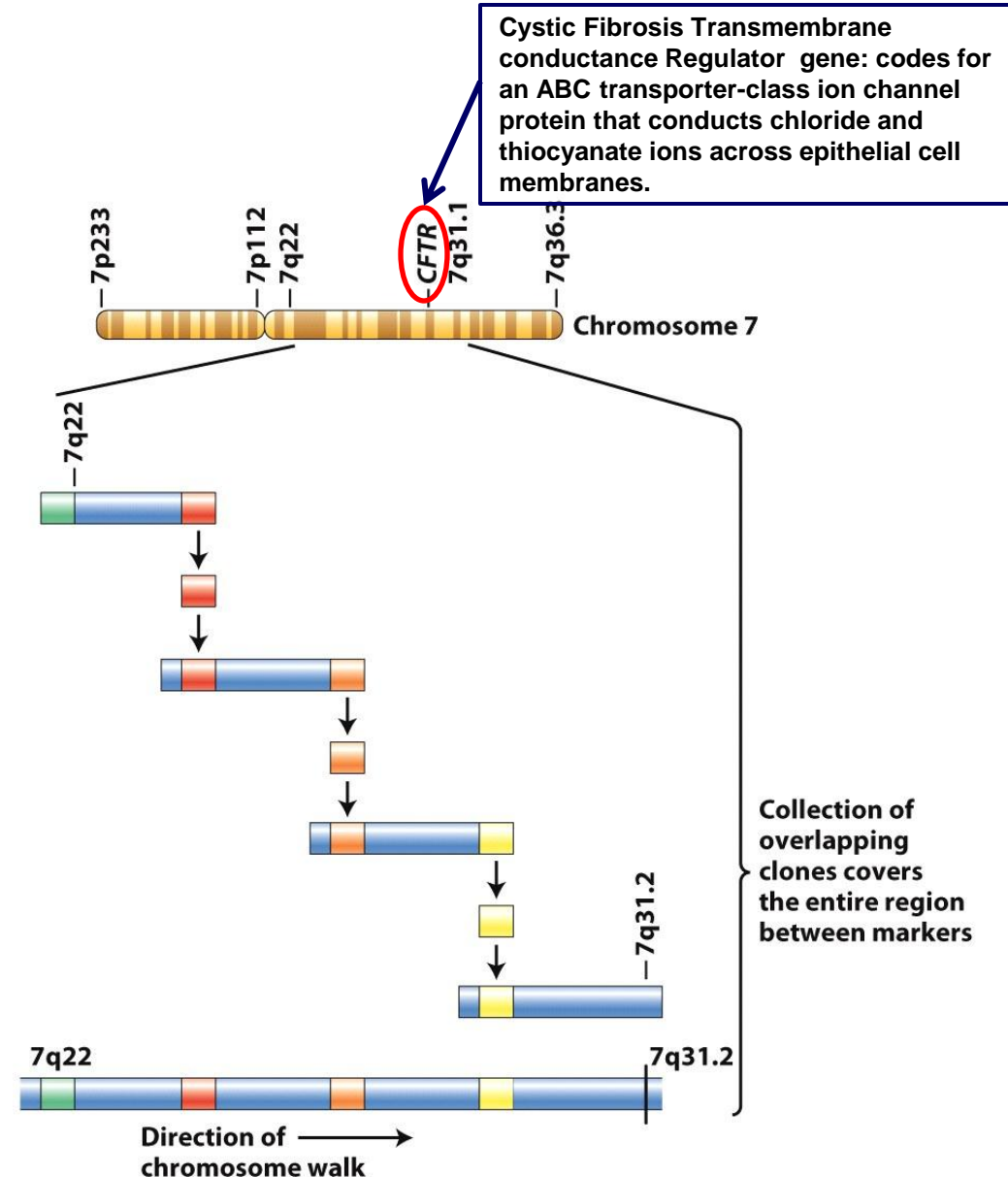


# Overlapping and Non-overlapping Fragments

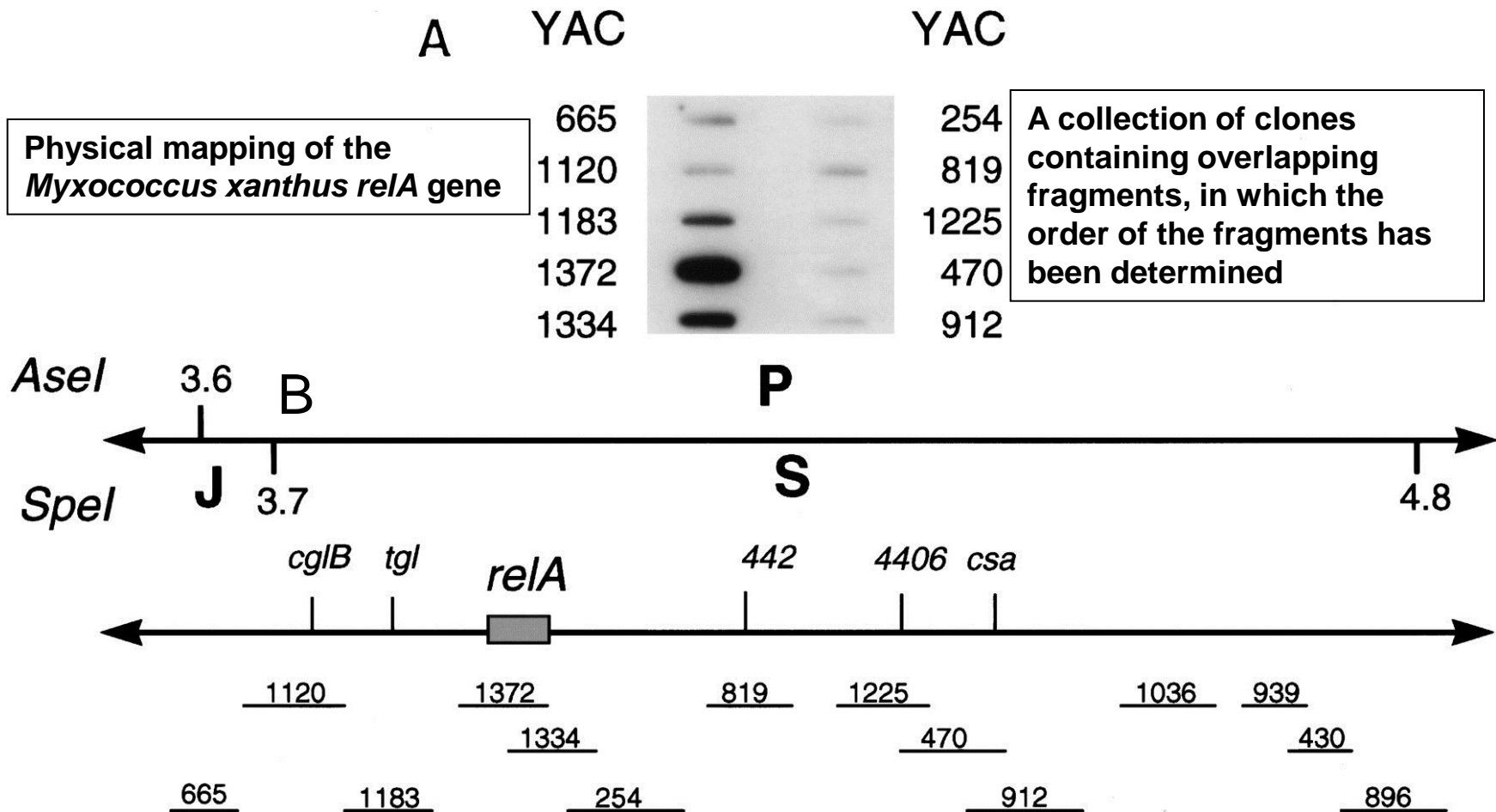
- **Incomplete** digestion with an **endonuclease** will result in a library containing **overlapping** fragments.
- **Complete** digestion will result in a library containing **no overlapping** fragments.
- Using incomplete digestion, sequence information obtained from one clone will allow the **isolation** of clones containing neighboring (**overlapping**) sequence information.
- This process can allow **large contiguous** stretches of sequence information to be obtained by “**chromosome walking**”.

# Using a Chromosomal Walk to Order a Set of Clones

- **Chromosomal walking** is a method of **positional cloning** used to find, isolate and clone a **particular** allele in a gene library.
- It involves **mapping** of the **position** of a DNA site or a gene by using **overlapping** restriction fragments.
- This chromosomal walk begins with a **recombinant** phage or BAC clone obtained from a library that contains large inserts representing an **entire** eukaryotic genome.
- The **molecular marker** 7q22 was used to **probe** a human genomic library.
- **Only** the insert DNAs are shown.
- The insert DNA selected by the probe is then used to **isolate** another recombinant phage or BAC containing a **neighboring segment** of eukaryotic DNA.
- This walk illustrates how to start at molecular landmark 7q22 and get to marker 7q31.1, which is on the other side of the CF gene.



# Ordered Library



- **A**, Slot blots using total YAC DNA from an ordered YAC library, probed with DNA fragment harboring the *relA* gene (*relA* gene product is responsible for the synthesis of guanosine 3',5'-bispyrophosphate (ppGpp) during the stringent response to amino acid starvation).
- **B**, Schematic of the 3.6- to 4.8-Mb region of the physical map of *M. xanthus*. YACs covering this region are indicated and labeled at the *bottom*. The position of *relA* is designated by the shaded box.
- The top line represents *Asel* and *SpeI* fragments as obtained from the literature; the second line represents the position of previously mapped markers also obtained from the literature.

# Random Library

Genome DNA  
of the target organism



↓ Fragmentation



↓ Ligation

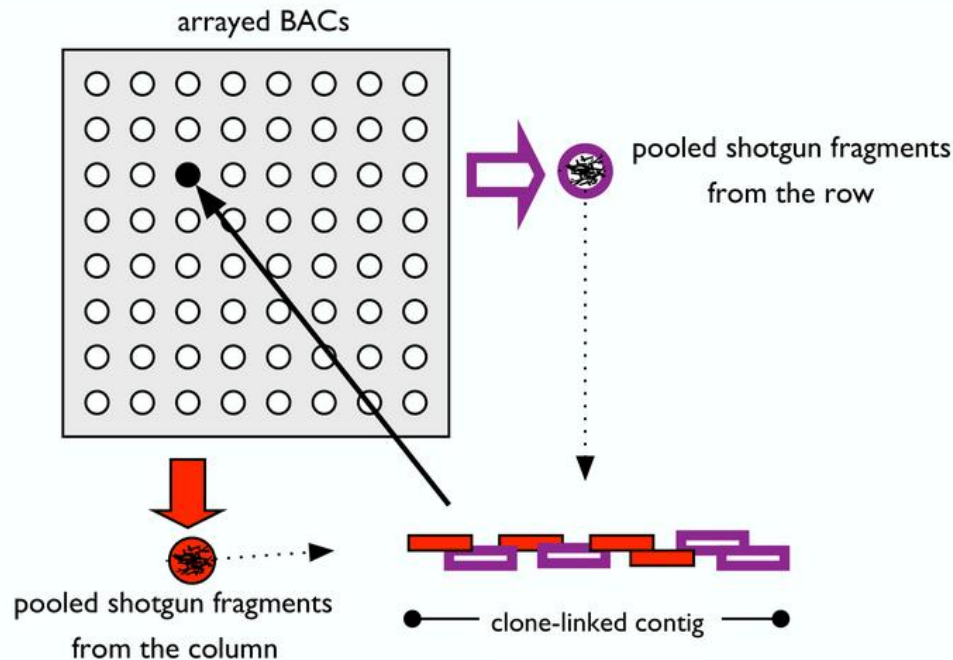


↓ PCR



Random DNA pool

# Arrayed Library



- DNA extracted from each clone is **pooled** together with other clones in the **same** row and column.
- **Subclone libraries** are prepared from the pools, and **shotgun** sequences are collected from the sub-libraries.
- Sequences are **assembled** into **contigs**.
- If a contig contains sequences from a row and a column pool's sub-library, the contig is assigned to the BAC at the **intersection** of the row and the column.