

2023 EBS F/V AKk GAP Special Collections Tally Sheet

<div>Observer Specimen Collection</div> <div>ADRIANA MYERS</div> <div>freeze ❄️</div> <div>Select specimens of typical adult size, in good condition and uncut. Bag individually or in small groups of 3-5, include a label in each bag.</div>	<div>EBS gear redesign</div> <div>LUKAS DEFILIPPO</div> <div></div> <div>Follow standard catch/haul collection for the EBS shelf surveys</div>	<div>Shark genetics and age structure sampling</div> <div>CINDY TRIBUZIO</div> <div>freeze or EtOH ❄️</div> <div>Clip 1cm x 1cm piece of tissue from any fin. place in either EtOH or baggie and freeze. EtOH is preferred.</div> <div>Sleeper shark only</div> <div>Morphometric measurements</div> <div>Collect eyes and vertebrae from OBVIOUSLY DECEASED. The eyes can be removed whole from the orbitals, by the easiest means available. A section of 4-5 vertebrae from the area approximately dorsal of the pectoral fin. If possible, bag separately and label left/right eyes separately. Eye bags can be placed within vertebrae bag. Store frozen.</div> <div>Take a picture of the whole fish with the associated specimen number label</div> <div>Record sex and pre-caudal length</div> <div>Extra credit: weight, maturity, pictures ovaries and testes.</div>	<div>EBS Slope Tows</div> <div>LUKAS DEFILIPPO</div> <div></div> <div>Process catch and haul data according to existing EBS slope survey protocol</div>
<div>Arctic and saffron cod growth</div> <div>ESTHER GOLDSTEIN</div> <div>freeze ❄️</div> <div>Collect fish by species, by haul, and by the two length categories. Such that fish are placed in bags in the above categories. Collect at any haul when either of the two species is caught. Label each bag with: cruise, species, haul, size category. The fish/bags should be frozen. Haul level data can be collected as is normal for any otolith collections.</div>	<div>Fish Condition Index-Pollock/Cod</div> <div>BIANCA PROHASKA</div> <div>freeze ❄️</div> <div>1) Choose 5 F adult pollock or P. cod from a given tow (pref. 1 species once/day) ideally already being kept for otolith collection</div> <div>2) Record weight (total, liver, stomach content, ovary), length, otolith #, core body temp</div> <div>3) Record average Distell and Yamato fat meter readings from muscle tissue just below dorsal fin and liver tissue (meter will provide mean reading from 3 measurements)</div> <div>4) Freeze a chunk of muscle tissue from the same area scanned and complete liver (bagged separately) with specimen labels: 25-50 of each species in both the EBS and NBS</div> <div>5) After 50 of each species/survey have been collected, continue recording scanned measurements from steps 1-3</div>	<div>Suryan/Copeman/Stowell RWP Shrimp Lipids</div> <div>MICHELLE STOWELL</div> <div>freeze ❄️</div> <div>Sample 5 individuals each of Crangon sp., Pandalus sp., and Argis sp., wearing gloves if possible and using forceps as necessary to prevent damage. Place individuals grouped by genus and station in plastic bags labeled with Event/Haul #, Station #, and taxa (genus, species if known). Record station/haul, genera, and number of individuals collected on the data sheet provided. Freeze samples immediately.</div>	<div>Collecting dissolved oxygen and pH with CTDs</div> <div>SEAN ROHAN</div> <div></div> <div>Collection to be carried out by Sean and/or Nicole.</div>
<div>Observer collections</div> <div>SARAH FRIEDMAN</div> <div>freeze ❄️</div> <div>Collect up to 5 specimens per species (<70cm) and freeze</div>	<div>Genetic identification of larval sandlance</div> <div>MELANIE PAQUIN</div> <div>freeze ❄️</div> <div>Up to 10 specimens from each geographic region: EBS and NBS. In the EBS collect fish from near Nunivak Island or within 8 km of the island. If 10 fish are not collected there then collect sandlance north in the NBS. Collect fish opportunistically whenever encountered. Collect 10 sandlance from any one location (haul) and geographic area described; 30 fish maximum. Freeze whole fish and label field collection tags with vessel, cruise, haul, and my name. Bag individual fish from a single haul together in a freezer bag but place flat (side-by-side), not stacked.</div> <div>-Ammodytes hexapterus</div>	<div>Juvenile Prowfish, Zaprora silenus, as prey fish</div> <div>MELANIE PAQUIN</div> <div>freeze ❄️</div> <div>Collect up to 10 juvenile prowfish (Zaprora silenus). Fish can be collected opportunistically when they are present (up to 10 specimens per haul, both legs). Sampling from any area of the EBS shelf trawl surveys will be adequate (i.e. opportunistic sampling). Please collect 5 males and 5 females. Adult specimens < 80 cm in length are preferred. Freeze whole fish and label field collection tags with vessel, cruise, haul, and my name. Bag individual fish from a single haul together in a freezer bag but place flat (side-by-side), not stacked.</div>	<div>IPHC sampling on the NOAA trawl surveys</div> <div>KAYLA UALESİ</div> <div></div> <div>after sorting, measure the halibut using a NOAA Fisheries measuring board; weigh the halibut on a scale provided by IPHC, assess sex and maturity by making a small incision, removing the gonads, and making visual maturity assessments based on pre-determined criteria; remove the left side otolith by cutting either through the opercular opening or through the top of the head; check for prior hooking wounds through a simple external examination; and take a small fin clip for genetic sampling.</div>