



Assignment 1

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Question 1

Part (a) Plotting to understand the relationship among the variables.

Loading necessary packages

```
library(tidyuesdayR)
```

```
library(dplyr)
```

```
library(ggplot2)
```

```
library(corrplot)
```

- First, we will load the data from the CSV file using the **read.csv()** function and store it in a data frame called **DB**, excluding 'X' as it is not a variable.

Load the dataset

```
starbucks <- read.csv("C:/Users/demir/OneDrive/Desktop/Assignment/starbucks.csv")
```

Exclude column 'X'

```
DB <- select(starbucks, -X)
```

- Next, we can inspect the first few rows of the data frame using the **head()** function to get a sense of the variables and their values.

```
head(DB)
```

	calories	total_fat_g	cholesterol_mg	sodium_mg	total_carbs_g	sugar_g	caffeine_mg
1	-1.637189	-1.020169	-0.8481098	-1.446475	-1.621355	-1.558173	0.4883517
2	-1.629925	-1.020169	-0.8481098	-1.392764	-1.621355	-1.558173	1.2949157
3	-1.622661	-1.020169	-0.8481098	-1.392764	-1.621355	-1.558173	2.1526900
4	-1.622661	-1.020169	-0.8481098	-1.392764	-1.621355	-1.558173	3.1768982
5	-1.637189	-1.020169	-0.8481098	-1.446475	-1.621355	-1.558173	-0.9839475
6	-1.629925	-1.020169	-0.8481098	-1.392764	-1.621355	-1.558173	-0.9199345

- Displaying the structure of the dataset.

```
str(DB)
```

```
'data.frame': 1147 obs. of 7 variables:
 $ calories      : num -1.64 -1.63 -1.62 -1.62 -1.64 ...
 $ total_fat_g   : num -1.02 -1.02 -1.02 -1.02 -1.02 ...
 $ cholesterol_mg: num -0.848 -0.848 -0.848 -0.848 -0.848 ...
 $ sodium_mg     : num -1.45 -1.39 -1.39 -1.39 -1.45 ...
 $ total_carbs_g : num -1.62 -1.62 -1.62 -1.62 -1.62 ...
 $ sugar_g       : num -1.56 -1.56 -1.56 -1.56 -1.56 ...
 $ caffeine_mg   : num 0.488 1.295 2.153 3.177 -0.984 ...
```

- Descriptive Statistics: You can use the **summary()** function in R to get a summary of your data.

Get descriptive statistics for each variable

```
summary(DB)
```

calories		total_fat_g		cholesterol_mg		sodium_mg	
Min.	:-1.65898	Min.	:-1.0369	Min.	:-0.8481	Min.	:-1.50019
1st Qu.	:-0.71470	1st Qu.	:-0.8693	1st Qu.	:-0.8481	1st Qu.	:-0.74823
Median	:-0.06097	Median	:-0.2827	Median	:-0.5699	Median	:-0.04999
Mean	: 0.00000	Mean	: 0.00000	Mean	: 0.00000	Mean	: 0.00000
3rd Qu.	: 0.66540	3rd Qu.	: 0.6392	3rd Qu.	: 0.8211	3rd Qu.	: 0.64825
Max.	: 2.98979	Max.	: 3.6563	Max.	: 3.3250	Max.	: 2.47442
total_carbs_g		sugar_g		caffeine_mg			
Min.	:-1.62136	Min.	:-1.55817	Min.	:-1.1760		
1st Qu.	:-0.76160	1st Qu.	:-0.75671	1st Qu.	:-0.7919		
Median	:-0.03081	Median	:-0.04429	Median	:-0.2158		
Mean	: 0.00000	Mean	: 0.00000	Mean	: 0.00000		
3rd Qu.	: 0.65700	3rd Qu.	: 0.62360	3rd Qu.	: 0.7444		
Max.	: 2.50547	Max.	: 2.40463	Max.	: 4.9052		

- Calculating the correlation coefficients using the `cor()` function to get a numerical measure of the strength and direction of these relationships.

```
cor_matrix <- cor(DB)
cor_matrix
```

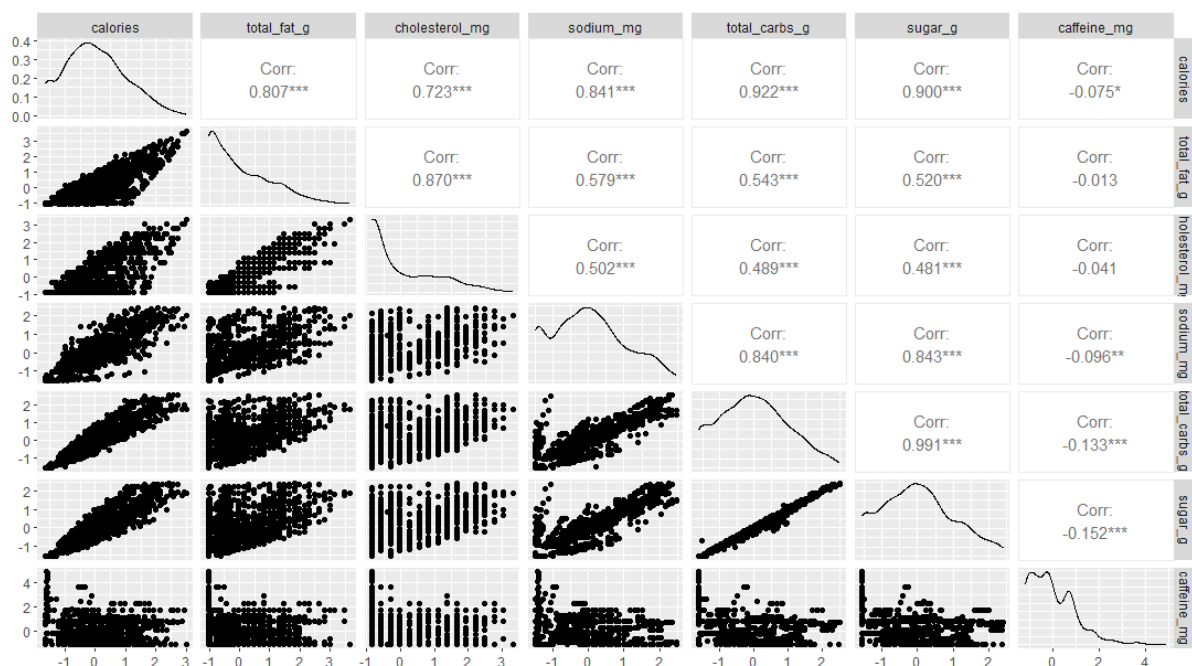
	calories	total_fat_g	cholesterol_mg	sodium_mg
calories	1.00000000	0.80731381	0.72298837	0.84080546
total_fat_g	0.80731381	1.00000000	0.87040512	0.57938004
cholesterol_mg	0.72298837	0.87040512	1.00000000	0.50222940
sodium_mg	0.84080546	0.57938004	0.50222940	1.00000000
total_carbs_g	0.92159269	0.54302297	0.48856249	0.83953671
sugar_g	0.90028087	0.51958432	0.48118993	0.84318545
caffeine_mg	-0.07455882	-0.01307082	-0.04112263	-0.09575978

	total_carbs_g	sugar_g	caffeine_mg
calories	0.9215927	0.9002809	-0.07455882
total_fat_g	0.5430230	0.5195843	-0.01307082
cholesterol_mg	0.4885625	0.4811899	-0.04112263
sodium_mg	0.8395367	0.8431855	-0.09575978
total_carbs_g	1.0000000	0.9912846	-0.13310151
sugar_g	0.9912846	1.0000000	-0.15198468
caffeine_mg	-0.1331015	-0.1519847	1.00000000

The correlation coefficients range from -1 to 1, where -1 indicates a perfect negative correlation, 0 indicates no correlation, and 1 indicates a perfect positive correlation. Overall, these results suggest that there are complex relationships between different nutrition levels, and that a high calorie item is likely to be high in other nutrients as well, except caffeine.

- Pair Plots: Pair plots are a fantastic way to visualize the distributions of each variable and the relationships between pairs of variables. The `pairs()` function in R can be used to create these.

```
# Create pair plots to visualize relationships between all variables
pairs(DB)
# Load GGally package
library(GGally)
# Create a pair plot
ggpairs(DB)
```



Exploring relationships

From the correlation plot plots, I can see that there are some correlations between the variables:

The correlation plot matrix shows that there are positive correlations between calories and total fat, total carbs and sugar. Additionally, there is a positive correlation between total fat and cholesterol, as well as sodium, and a positive correlation between total carbs and sodium. Sugar and total carbs have a positive correlation, while there is no strong correlation between sugar and sodium or caffeine and calories. The highest correlation is between Total carbs and sugar as 0.991. These relationships suggest that high calorie items are likely to be high in other nutrients. Caffeine, on the other hand, seems like uncorrelated with the other variables.

Part (b) Checking whether the data has already been centred and standardized Providing the supporting R output.

```
# Calculate the mean and standard deviation of each column in DB
DB_mean <- apply(DB, 2, mean)
DB_sd <- apply(DB, 2, sd)
# Display the mean and standard deviation of each column in DB
data.frame(mean = DB_mean, sd = DB_sd)
```

	mean	sd
calories	-4.713415e-17	1
total_fat_g	1.887117e-17	1
cholesterol_mg	-5.479299e-16	1
sodium_mg	3.597547e-17	1
total_carbs_g	-7.194017e-17	1
sugar_g	3.509718e-16	1
caffeine_mg	-4.600359e-16	1

The standard deviations of variables are one, and the means can be considered zero, this indicates that the data has been standardized, and centred.

Part (c) Performing a principal component analysis on the variables. Providing the proportion of the variance explained by the principal components and the cumulative variance percentage.

```
# Perform PCA on the centered data
pca <- prcomp(DB)

# Calculate proportion of variance explained by each principal component
pca_prop_var <- round(100 * pca$sdev^2 / sum(pca$sdev^2), 2)

pca_prop_var

[1] 66.59 15.48 12.73  3.01  1.88  0.26  0.06

# Calculate cumulative variance percentage
cum_var <- cumsum(pca_prop_var)
cum_var

[1] 66.59 82.07 94.80 97.81 99.69 99.95 100.01

# Combine proportion of variance and cumulative variance percentage into a data frame
Principal_Component = paste0("PC", 1:length(pca_prop_var))
df_pca_results <- data.frame(Principal_Component = Principal_Component,
                             Proportion_of_Variance = pca_prop_var,
                             Cumulative_Variance_Percentage = cum_var)
# Display data frame of PCA results
df_pca_results
```



Principal_Component	Proportion_of_Variance	Cumulative_Variance_Percentag
PC1	66.59	66.59
PC2	15.48	82.07
PC3	12.73	94.80
PC4	3.01	97.81
PC5	1.88	99.69
PC6	0.26	99.95
PC7	0.06	100.01

These outputs allow us to understand the importance of each principal component in capturing the variability in the data set.

Part (d) Identifying the number of principal components that are needed to explain at least 95% of the variance in the starbucks data.

```
# Identify principal components that explain at least 95% of the variance
pc_95 <- which(cum_var >= 95)[1]
pc_subset <- 1:pc_95
pc_subset
[1] 1 2 3 4
```

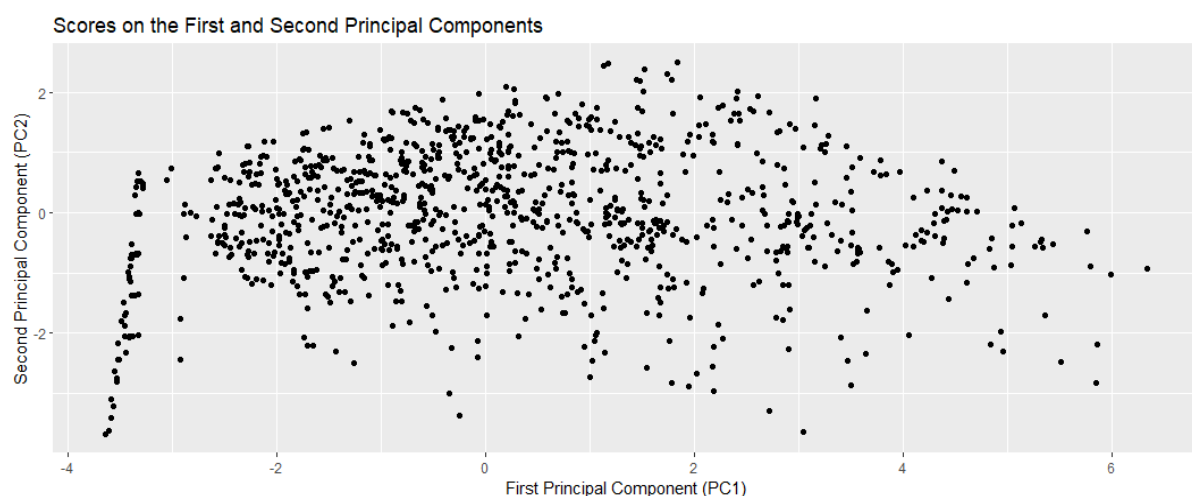
The resulting pc_subset vector indicates the principal components that explain at least 95% of the variance in the starbucks data. So, the first 4 components cover 97.81% of the variance in dataset.

Part (e) Create a plot comparing the first and second principal components. Comment on this plot.

```
# Perform PCA
pca_results <- prcomp(DB, scale. = FALSE)

# Create a data frame for the first two principal components
pca_df <- data.frame(pca_results$x[,1:2])

ggplot(pca_df, aes(x = PC1, y = PC2)) +
  geom_point() +
  xlab("First Principal Component (PC1)") +
  ylab("Second Principal Component (PC2)") +
  ggtitle("Scores on the First and Second Principal Components")
```



Comments on the Plot:

Looking at the scatter plot of the first and second principal components, I notice a broad dispersion of data points. This suggests a wide range of variability captured by these components, but without obvious clustering. This might mean that the underlying structure of the data is complex and not easily separable by these two components alone. The variance captured by each axis would give more insights, and it would be interesting to see how the original variables contribute to these components.

Question 2

Part (a) R to simulate both X and Y

```
# Set seed for reproducibility
set.seed(123)

# Define parameters
n <- 200
m <- 300
mu <- 5
sigma_sq <- 12
sigma <- sqrt(sigma_sq)

# Simulate independent random samples X and Y
X <- rnorm(n, mean = mu, sd = sigma)
Y <- rnorm(m, mean = mu, sd = sigma)
```

Part (b) Using your simulated data, calculate the value of each estimator; T_1 and T_2 .

```
# Calculate sample means
X_bar <- mean(X)
Y_bar <- mean(Y)

# Compute estimators
T1 <- (n * X_bar + m * Y_bar) / (n + m)
T2 <- 0.5 * (X_bar + Y_bar)

# Print results
cat("X_bar:", X_bar, "\n")
cat("Y_bar:", Y_bar, "\n")
cat("T1:", T1, "\n")
cat("T2:", T2, "\n")
```

Outcome:

```
X_bar: 4.970311
Y_bar: 5.219501
T1: 5.119825
T2: 5.094906
```



Part (c) Using nonparametric bootstrapping (sampling with replacement), generate $b = 100$ bootstrapped values for each estimator. Construct histograms for each of the estimators.

```
# Required library for ggplot2
library(ggplot2)

# Set the number of bootstraps
b <- 100

# Initialize vectors to store bootstrap estimates
T1_boot <- numeric(b)
T2_boot <- numeric(b)

# Perform bootstrapping
for (i in 1:b) {
  # Sample with replacement
  X_boot <- sample(X, size = n, replace = TRUE)
  Y_boot <- sample(Y, size = m, replace = TRUE)

  # Calculate sample means
  X_bar_boot <- mean(X_boot)
  Y_bar_boot <- mean(Y_boot)

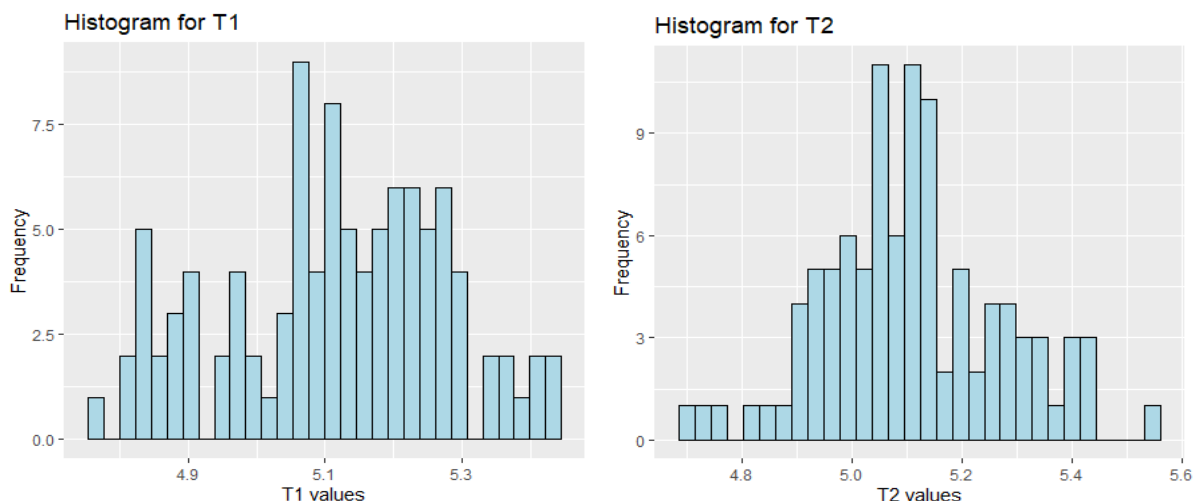
  # Compute estimators
  T1_boot[i] <- (n * X_bar_boot + m * Y_bar_boot) / (n + m)
  T2_boot[i] <- 0.5 * (X_bar_boot + Y_bar_boot)
}

# Create data frame for plotting
boot_estimates <- data.frame(T1 = T1_boot, T2 = T2_boot)

# Construct histograms
ggplot(boot_estimates, aes(x = T1)) +
  geom_histogram(color = "black", fill = "lightblue", bins = 30) +
  labs(title = "Histogram for T1",
       x = "T1 values",
       y = "Frequency")

ggplot(boot_estimates, aes(x = T2)) +
  geom_histogram(color = "black", fill = "lightblue", bins = 30) +
  labs(title = "Histogram for T2",
       x = "T2 values",
       y = "Frequency")
```

Outcome:



Part (d) Using the bootstrapped estimators, comment on each estimator. Which estimator would you prefer? Provide reasoning.

Both estimators are quite close to each other and to the true population mean ($\mu = 5$). This is expected because both estimators are unbiased for the population mean.

However, the values of T1 and T2 are slightly different from each other. The distribution of T1 appears to be slightly wider than the distribution of T2, meaning T1 has a bit more variability. This makes sense because T1 is a weighted average of X and Y with weights proportional to the sample sizes, so it tends to be influenced more by larger samples and thus exhibits more variability.

On the other hand, T2 is a simple average of the sample means, so it doesn't give more weight to the larger sample and therefore has less variability.

Overall, in this case, I would prefer estimator T2 because it has less variability as indicated by the narrower distribution of bootstrap estimates.

Part (e) Construct a 95% confidence interval for both estimators using the bootstrapped values and the Empirical Distribution Function (ECDF)

```
# Function to calculate ECDF-based confidence intervals
ecdf_conf_interval <- function(boot_values, alpha = 0.05) {
  lower_quantile <- quantile(boot_values, probs = alpha / 2)
  upper_quantile <- quantile(boot_values, probs = 1 - alpha / 2)
  return(c(lower_quantile, upper_quantile))
}

# Calculate 95% confidence intervals for T1 and T2
T1_conf_interval <- ecdf_conf_interval(T1_boot, alpha = 0.05)
T2_conf_interval <- ecdf_conf_interval(T2_boot, alpha = 0.05)

# Print results
cat("95% confidence interval for T1: (", T1_conf_interval[1], ", ", T1_conf_interval[2], ")\n")
cat("95% confidence interval for T2: (", T2_conf_interval[1], ", ", T2_conf_interval[2], ")\n")
```

Outcome:

95% confidence interval for T1: (4.827335 , 5.432221)

95% confidence interval for T2: (4.786906 , 5.424964)

Part (f) (Without using R) Provide the theoretical expectation for each estimator? Does this agree with your empirical results in part (d)?

Let's derive the theoretical expectation for each estimator $T1$ and $T2$.

The theoretical expectation of T1 can be derived as follows:

$$\begin{aligned}
 E[T1] &= E[(\bar{n}X + \bar{m}Y)/(n + m)] \\
 &= (\bar{n}E[X] + \bar{m}E[Y])/(n + m) \\
 &= (\bar{n}\mu + \bar{m}\mu)/(n + m)
 \end{aligned}$$

$$= (\bar{n} + \bar{m})\mu / (n + m)$$

$$= \mu$$

Therefore, the theoretical expectation of $T1$ is equal to μ , which is 5 in this case.

The theoretical expectation of $T2$ can be derived as follows:

$$E[T2] = E[1/2(\bar{X} + \bar{Y})]$$

$$= 1/2(E[\bar{X}] + E[\bar{Y}])$$

$$= 1/2(\mu + \mu)$$

$$= \mu$$

Again, the theoretical expectation of $T2$ is equal to μ , which is 5 in this case.

Yes, the theoretical expectations derived in part (f) agree with the empirical results obtained in part (d). Both indicate that the expected values of the estimators $T1$ and $T2$ are equal to the population mean μ , which is 5 in this case.

In conclusion, the theoretical expectations for both estimators $T1$ and $T2$ are equal to the population mean μ

Question 3

Part (a) Likelihood and log-likelihood function for θ :

Given a sequence of i.i.d. random variables X_i with the given distribution, the likelihood function is the product of the probability mass functions of each X_i :

$$L(\theta | X) = \prod_i f_X(x_i) = \prod_i [\theta^{1-x_i} * (1 - \theta)^{x_i}] = \theta^{\sum_i (1 - x_i)} * (1 - \theta)^{\sum_i x_i}$$

The log-likelihood function is the natural logarithm of the likelihood function, which turns the product into a sum:

$$l(\theta | X) = \log(L(\theta | X)) = \sum_i [(1 - x_i) \log(\theta) + x_i \log(1 - \theta)]$$

Part (b) Derive the maximum likelihood estimator (MLE) $\hat{\theta}$ MLE of θ :

To obtain the MLE, I need to take the derivative of the log-likelihood with respect to θ , set it equal to zero and solve for θ . This gives me the value of θ that maximizes the log-likelihood function:

- $l'(\theta | X) = \sum_i [(1 - x_i) / \theta - x_i / (1 - \theta)]$
- $0 = \sum_i (1 - x_i) / \theta - \sum_i x_i / (1 - \theta)$
- $\theta_{mle} = \sum_i x_i / n$



Part (c) Calculate the Cramér-Rao lower bound (CRLB) for the variances of unbiased estimators of θ .

$$I(\theta) = E\left[\left(\frac{d}{d\theta} \log(f(x|\theta))\right)^2\right]$$

$$= E\left[\left(\frac{1-x}{\theta} - \frac{x}{1-\theta}\right)^2\right]$$

$$= n/[\theta(1-\theta)]$$

Therefore, the CRLB is:

$$\text{CRLB}(\theta) = 1 / I(\theta) = \theta(1-\theta) / n$$

Part (d) Find the variance of the MLE, $\hat{\theta}_{\text{mle}}$ and compare to the CRLB.

- $\text{Var}(\hat{\theta}_{\text{mle}}) = \text{Var}(\sum X_i / n) = \theta(1-\theta)/n$
- The Fisher Information, $\text{CRLB} = 1/I(\theta)$.

$$\text{So, } \text{Var}(\hat{\theta}_{\text{MLE}}) = \text{CRLB} = \theta(1-\theta)/n$$

This indicates that the sample mean is an efficient estimator of θ in a Bernoulli distribution, because it achieves the lower bound of the variance.