




# Altered expression of apoptosis-related genes in rheumatoid arthritis peripheral blood mononuclear cell and related miRNA regulation

Hamidreza Ebrahimian<sup>1</sup> | Maryam Akhtari<sup>2</sup>  | Maassoumeh Akhlaghi<sup>3</sup> | Elham Farhadi<sup>3,4</sup>  | Ahmadreza Jamshidi<sup>3</sup> | Gholam Hossein Alishiri<sup>5,6</sup> | Mahdi Mahmoudi<sup>3,4</sup>  | Mahmood Tavallaie<sup>1</sup>

<sup>1</sup>Human Genetic Research Center, Baqiyatallah University of Medical Sciences, Tehran, Iran

<sup>2</sup>Tobacco Prevention and Control Research Center (TPCRC), National Research Institute of Tuberculosis and Lung Diseases (NRITLD), Shahid Beheshti University of Medical Sciences, Tehran, Iran

<sup>3</sup>Rheumatology Research Center, Tehran University of Medical Sciences, Tehran, Iran

<sup>4</sup>Inflammation Research Center, Tehran University of Medical Sciences, Tehran, Iran

<sup>5</sup>Chemical Injuries Research Center, Systems Biology and Poisonings Institute, Baqiyatallah University of Medical Sciences, Tehran, Iran

<sup>6</sup>Department of Rheumatology, Faculty of Medicine, Baqiyatallah University of Medical Sciences, Tehran, Iran

## Correspondence

Mahdi Mahmoudi, Rheumatology Research Center, Shariati Hospital, Tehran University of Medical Sciences, Kargar Ave., Tehran 1411713137, Iran.  
Email: [mahmoudim@tums.ac.ir](mailto:mahmoudim@tums.ac.ir)

Mahmood Tavallaie, Human Genetic Research Center, Baqiyatallah University of Medical Sciences, Tehran 19395-5487, Iran.  
Email: [tavalla.mah@gmail.com](mailto:tavalla.mah@gmail.com)

## Abstract

**Aim:** Impaired apoptosis and proliferation resulted in autoreactive lymphocyte development and inflammation in Rheumatoid arthritis (RA). *TP53*, *BAX*, *FOXO1*, and *RB1* are related genes in cell survival, proliferation, and inflammation which could be important in RA development and disease severity. Here we investigated their expression in peripheral blood mononuclear cells (PBMCs) from RA patients in comparison to healthy controls.

**Methods:** Fifty healthy controls and 50 RA patients were selected. The quantitative real-time polymerase chain reaction was used to assess the gene expression level in PBMCs.

**Results:** The mRNA expression of *TP53* (FC = 0.65,  $p = .000$ ), *BAX* (FC = 0.76,  $p = .008$ ), *FOXO1* (FC = 0.59,  $p = .000$ ) and *RB1* (FC = 0.50,  $p = .000$ ) were significantly reduced in RA PBMCs. *TP53* expression was negatively correlated with miR-16-5p ( $p = .032$ ) and *FOXO1* expression was negatively correlated with miR-335-5p ( $p = .005$ ) and miR-34a-5p ( $p = .014$ ). A positive correlation was seen between *TP53* expression and its downstream gene, *BAX* ( $p = .001$ ). *FOXO1* expression was also negatively correlated with disease activity, DAS28 ( $p = .021$ ).

**Conclusion:** All selected genes have downregulated expression in RA PBMCs which could be correlated with RA pathogenesis by regulating apoptosis, cell survival, inflammatory mediator production, and proliferation. Due to the correlation of miR-16-5p, miR-34a-5p, and miR-335-5p with *TP53* and *FOXO1* expression in RA PBMCs, they could be used as future therapeutic targets.

## KEYWORDS

apoptosis, autoimmunity, BAX, FOXO1, miRNAs, P53, RB1, rheumatoid arthritis (RA)

## 1 | INTRODUCTION

Rheumatoid arthritis (RA) is a chronic inflammatory disease that primarily affects joints, but can also cause extra-articular complications.<sup>1,2</sup> In general, both genetic and environmental factors (epigenetic) may contribute to the development of RA, including sex hormones, exposure to ultraviolet light, smoking, changes in the oral, lung, or intestinal microbiota, microbial infections, indeed the infections or tissue injury may be capable of reducing tolerance and inducing autoreactivity.<sup>3–6</sup> Synovial inflammation is closely related to changes in the function of RA fibroblast-like synoviocytes (RAFLS). The FLSS' nonstop proliferation, cell survival, and production of pro-inflammatory actors resulted in joint damage in RA.<sup>7</sup>

Programmed cell death also known as “controlled” cell death or apoptosis, is an important mechanism in maintaining organ homeostasis.<sup>8</sup> It has been shown that cell proliferation and apoptosis were dysregulated in RA.<sup>9–11</sup> Decreased FLS and lymphocyte apoptosis with aggressive proliferation and activation, interrupting joint and immune balance and function in RA patients.<sup>12,13</sup> Altered gene expression and genetic polymorphisms in apoptotic or cell cycle-related genes have been reported in RA.

Approximately 70% of the human genome encodes the non-protein-coding genes including miRNA which play an essential role in genome expression and autoimmune pathogenesis as an epigenetic factor.<sup>14</sup> miRNAs are evolutionarily conserved with 18–25 nucleotides, which have an essential function in gene expression.<sup>15</sup> Many studies reported the critical role of miRNAs in autoimmune diseases such as RA.<sup>15–18</sup>

Based on our last experiment miR-335-5p, miR-150-5p, miR-34a-5p, and miR-16-5p were upregulated in peripheral blood mononuclear cells (PBMCs) of RA patients. In the miRTaRBase database ([https://mirtaRBase.cuhk.edu.cn/~miRTaRBase/miRTaRBase\\_2022/php/index.php](https://mirtaRBase.cuhk.edu.cn/~miRTaRBase/miRTaRBase_2022/php/index.php)), these miRNAs target the expression of important genes contributing to the cell-cycle and apoptosis regulation. The miR-335-5p targeted *RB1*, and *FOXO1*, the miR-150-5p targeted *TP53*, the miR-34a-5p directly targeted *BAX*, and *TP53*, and indirectly targeted *FOXO1*, and the miR-16-5p targeted *TP53*.<sup>19</sup>

P53 tumor suppressor protein which is referred to as the “Guardian of the Genome” is encoded by the *TP53* gene and is one of the most important proteins that contributes to cell-cycle regulation.<sup>20</sup> P53 protein protects the DNA integrity in cells, however, it is also involved in other functions such as aging, cell differentiation, and development.<sup>21</sup> P53 abnormality resulted in cell survival and nonstop proliferation alongside DNA damage.<sup>22</sup> P53 abnormal function or dysregulation was reported in RA synovial cells or lymphocytes which led to DNA damage

and resistance to apoptosis.<sup>23,24</sup> Forkhead box protein O1 (FOXO1), is a protein encoded by the *FOXO1* gene. FOXO1 is also involved in important pathways including apoptosis, cell cycle, oxidative stress response, autophagy, immune system regulation, DNA repair, cell differentiation, and metabolism.<sup>25–28</sup> *FOXO1* dysregulation is reported in different diseases including RA.<sup>29,30</sup> The *RB1* is encoded by the *RB1* gene which was discovered in the malignant tumor of the retina known as the retinoblastoma.<sup>31</sup> The tumor suppression activity of *RB1* was well-defined in the inhibition of cell cycle progression.<sup>32,33</sup> Alongside cancers, *RB1* dysregulation was reported in RA pathogenesis.<sup>34,35</sup> Apoptosis regulator BAX, BCL2 associated X, was encoded by the *BAX* gene. The main function of this protein is initiating the apoptosis pathway by releasing proapoptotic factors, such as cytochrome c from mitochondria.<sup>36</sup> The importance of autoreactive lymphocyte survival and apoptosis resistance was one of the main factors in autoimmunity pathogenesis. The low expression level of *BAX* was reported in RA lymphocytes which reduces the apoptosis in these cells.<sup>11</sup> The correlation of the expression of mentioned miRNAs with the expression of targeted genes has previously been confirmed in targeting these genes by strong evidence.

Herein, we investigate the expression of *RB1*, *BAX*, *FOXO1*, and *TP53* which are targets of mentioned miRNA and are essential genes in cell cycle progression and apoptosis in PBMCs from Iranian RA patients and also analyze the correlation between the expression of these genes with their corresponding miRNAs.

## 2 | METHODS

### 2.1 | Study subjects

Fifty patients with RA diagnosis were selected according to the American College of Hematology criteria<sup>37</sup> and recruited to the Rheumatology Research Center, Shariati Hospital, Tehran, Iran. Fifty matched controls in age and gender were also selected with not any autoimmunity diagnosis even in their relatives. The human research ethics committee of Tehran University of Medical Science reviewed and approved the current study (Approval No: IR.TUMS.VCR.REC.1398.885).

### 2.2 | Measuring transcription levels of *RB1*, *TP53*, *BAX*, and *FOXO1*

For mRNA expression analysis first-strand cDNA was synthesized by RT-ROSET (REF: EB983019, ROJE

Technologies). The Real Q Plus Master Mix Green High ROX (AMPLIQON) was used for quantitative real-time polymerase chain reaction (PCR) (Step One Plus, Applied Biosystems). The test reaction included 10  $\mu$ L 2 $\times$  SYBR Green PCR master mix, 8  $\mu$ L cDNA (1/8 dilution), and 1  $\mu$ L each primer for reaction. The primer details are listed in Table 1. Real-time PCR was conducted as follows: 95°C for 15 min, 95°C for 15 s, 56°C for 30 s, and 72°C for 1 min. For normalizing the relative quantification levels of the target mRNAs, *GAPDH* mRNA transcript level was used. Livak and Schmittgen's method was used in relative mRNA expression calculation.<sup>38</sup>

## 2.3 | Statistical analysis

SPSS v.28 was used to calculate data. The Kolmogorov-Smirnov test was applied for normality evaluation. Spearman's rho test or Pearson's correlation coefficient was carried out for correlation analysis. For gene expression analysis Unpaired *T* test or Mann-Whitney were performed. The GraphPad Prism v.8 software was used for plotting.

## 3 | RESULTS

### 3.1 | Study groups

Clinical details of healthy control and RA patients are listed in Table 2. The disease-modifying antirheumatic drugs (DMARDs), including methotrexate (MTX), hydroxychloroquine, prednisone (steroid), sulfasalazine, and alendronate were in RA patients' treatment program. Six patients are in early diagnosis and do not take any treatment.

### 3.2 | Evaluation of *TP53*, *BAX*, *FOXO1*, and *RB1* expression

The mRNA expression of *TP53* (FC = 0.65,  $p = .000$ ), *BAX* (FC = 0.76,  $p = .008$ ), *FOXO1* (FC = 0.59,  $p = .000$ ), and *RB1* (FC = 0.50,  $p = .000$ ) were significantly reduced in PBMCs of RA patients in comparison to controls (Figure 1). No correlation was found between prednisone, methotrexate, hydroxychloroquine, and sulfasalazine with all gene expression in RA patients. In RA patients with alendronate treatment *TP53* expression was significantly upregulated ( $p = .039$ ) and *RB1* expression downregulated ( $p = .032$ ). however, the mRNA expression of *TP53* and *RB1* was also significantly decreased ( $p = .000$  and  $p = .001$ , respectively) in RA patients who did not receive alendronate treatment compared to controls.

### 3.3 | Correlation analysis

Based on our recent experiment,<sup>42</sup> the miRNA expression levels of miR-335-5p, miR-16-5p, miR-150-5p, and miR-34a-5p were considerably higher in patient-derived PBMCs than in control samples. In the present work, we explored the relationship between the expression of the analyzed genes and these four miRNAs in RA patients' PBMCs. *TP53* expression was negatively correlated with miR-16-5p expression ( $p = .032$ ) and *FOXO1* expression was also negatively correlated with miR-335-5p expression ( $p = .005$ ). The expression of miR-34a-5p was negatively correlated with *FOXO1* ( $p = .014$ ). A positive correlation was seen between the expression of *TP53* and its downstream gene, *BAX* ( $p = .001$ ). *FOXO1* expression was also negatively correlated with disease activity DAS28 ( $p = .021$ ). All of the miRNAs' data were listed in Table 3.

TABLE 1 Primer sequences and details.

Genes	Primer set sequence	Tm (°C)	Amplicon size (kb)	References
<i>FOXO1</i>	5'-GCAGATCTACGAGTGGATGGTC-3' (forward primer)	60.29	325	[30]
	5'-AAACTGTGATCCAGGGCTGTC-3' (reverse primer)	60.27		
<i>RB1</i>	5'-ACCAGATCATGTCAAGAGA-3' (forward primer)	54.97	340	[39]
	5'-ACTGCTGGGTTGTGTCAAAT-3' (reverse primer)	57.93		
<i>BAX</i>	5'-GGACGAACTGGACAGTAACATGG-3' (forward primer)	61.17	150	[40]
	5'-GCAAAGTAGAAAAGGGCGACAAC-3' (reverse primer)	60.85		
<i>TP53</i>	5'-TCAACAAGATGTTTTGCCAACTG-3' (forward primer)	58.57	118	[41]
	5'-ATGTGCTGTGACTGCTTGTAGATG-3' (reverse primer)	61.39		
<i>GAPDH</i>	5'-AAGGTCGGAGTCAACGGATT-3' (forward primer)	59.65	66	[42]
	5'-ACCAGAGTTAAAGCAGCCCTG-3' (reverse primer)	60.82		

TABLE 2 Clinical characterization of RA patients and control.

Item	RA patients (N = 50)	Control (N = 50)
Age	48.54 ± 11.54	47.24 ± 9.85
Gender (N)	Female (43)/male (7)	Female (43)/male (7)
Treatment duration	38.30	
Drug	Prednisone (36), methotrexate (29), hydroxychloroquine (16), sulfasalazine (13), alendronate (13)	No drugs
DAS28	5.10 ± 1.2	-
Family history	RA (9), DM (10), SLE (1), MS (2), thyroid diseases (3)	No diseases
Concurrent syndrome	thyroid diseases (6), DM (4)	No diseases
RF (%)	33 (66)	-
ACCP (%)	43 (86)	-

Abbreviations: ACCP, anti-cyclic citrullinated peptide; DAS28, Disease Activity Score of 28 joints; DM, diabetes mellitus; MS, multiple sclerosis; RA, rheumatoid arthritis; RF, rheumatoid factor; SLE, systemic lupus erythematosus.

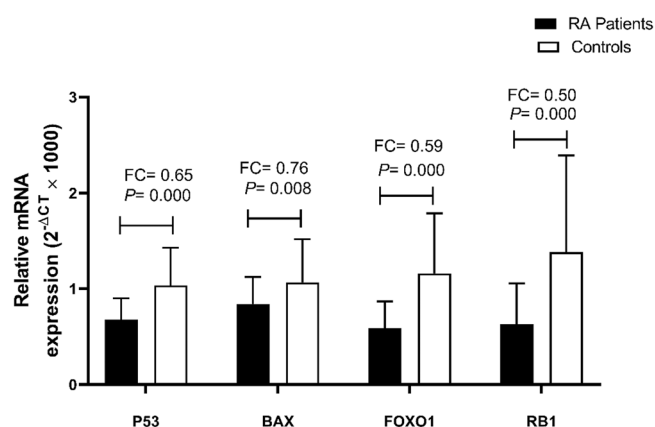


FIGURE 1 *TP53*, *BAX*, *FOXO1*, and *RB1* expression in RA patients and control. The expression was assessed by real-time PCR. The groups were compared by the Mann–Whitney test. All expressions show a significant reduction in RA patients. RA, rheumatoid arthritis.

## 4 | DISCUSSION

RA is a systemic inflammatory autoimmune disease marked by the deterioration of joints.<sup>43</sup> Apoptosis resistance and hyperproliferation in synovium tissue, RAFLS, and immune cells result in the creation of pannus and the inflammation and degeneration of joints.<sup>44</sup> The development and selection of lymphocytes are closely regulated mechanisms that eliminate autoreactive cells by apoptosis. Due to autoreactive lymphocyte activity, any interruption of the cell cycle and survival leads to a variety of autoimmunity.<sup>45,46</sup> This notion is supported by *lpr* or *gld* mutations (mutations in the Fas and Fas ligand genes) in mice that generated autoantibodies<sup>47,48</sup> FasL was the inducer of apoptosis which induce clustering of Fas to trigger apoptosis signaling.<sup>49</sup>

In our previous report,<sup>42</sup> we found a significant upregulation of some miRNAs including miR-335-5p, miR-16-5p, miR-150-5p, and miR-34a-5p in PBMCs of Iranian RA patients. Herein, the expression of their important target genes including *TP53*, *BAX*, *FOXO1*, and *RB1* which contribute to the cell-cycle regulation was investigated in the same RA patients' samples. As we discussed later, the P53 and its downstream proteins including BAX, FOXO1, and RB1 were necessary for cellular apoptosis and proliferation which are essential in autoimmune disorders including RA. P53 was reported to be downregulated in mRNA and protein levels in different cell types including PBMCs<sup>50</sup> and FLS<sup>51</sup> in RA. Dysregulated expression or mutation of *TP53* which finally leads to abnormal cell function could enhance the aggressive nature of RAFLS or autoreactivity in lymphocytes and leads to the inflammatory phenotype in RA joints.<sup>52</sup> *TP53* knockdown in RAFLS resulted in the production of inflammatory cytokines and T-cell differentiation which is important in RA pathogenesis.<sup>53</sup> *TP53* knockout mice were also reported to have higher arthritis scores compared with *TP53*<sup>+</sup> mice.<sup>54</sup> *TP53* downregulation can cause dendritic cells (DCs) maturation and activation, and reduce the expression of *FOXP3* in T cells, which is a master regulator of regulatory T cells (Tregs) and Th17 polarization. All these changes were important in RA pathogenesis.<sup>23</sup>

Our study showed a significant reduction in *TP53* transcription in RA patients' samples that might be correlated with RA pathogenesis. One of the important regulators of this protein is miR-16-5p which significantly increased<sup>42</sup> and negatively correlated with *TP53* expression in RA PBMCs. As much as we know, these findings are the first report in RA PBMCs. P53 also has the ability to induce the expression of proapoptotic genes



**TABLE 3** Correlation of the expression of miRNAs and *TP53*, *BAX*, *FOXO1*, and *RBI* in RA patients.

Genes	miR-335-5p	miR-16-5p	miR-34a-5p	miR-150-5p
<i>TP53</i> (p Value)	-0.196 (0.18)	-0.316 (0.032)*	-0.33 (0.82)	0.132 (0.38)
<i>BAX</i> (p Value)	-0.454 (0.001)*	-0.251 (0.088)	0.003 (0.98)	0.015 (0.92)
<i>FOXO1</i> (p Value)	-0.402 (0.005)*	-0.268 (0.049)*	-0.364 (0.014)*	0.241 (0.11)
<i>RBI</i> (p Value)	-0.260 (0.078)	-0.015 (0.92)	-0.075 (0.62)	-0.102 (0.51)

Note: Significant correlations were tagged by \*.

such as *BAX*.<sup>55,56</sup> Herein, we reported the positive correlation between the expression of *TP53* and *BAX* in RA patients as one of the *BAX* regulatory mechanisms. In this study, a significantly lower expression of *BAX* was reported in RA PBMCs. It was previously reported that T lymphocytes in RA patients were resistant to apoptosis which could be correlated with reduced expression of some related genes including *BAX* in these cells.<sup>11</sup> The induction of *TP53* and *BAX* expression was reported in another study which could help control normal apoptosis as a new strategy for the treatment of RA.<sup>57</sup>

*FOXO1* is an important mediator in apoptosis, autophagy, antioxidative stress, cell cycle arrest, metabolism, and immune system regulation, which was important in the pathogenesis of different diseases.<sup>58</sup> Previous studies reported a lower expression level of *FOXO1* in PBMCs of RA patients which could be correlated with disease pathogenesis and severity.<sup>30</sup> The other study reported that lower expression of *FOXO1* in patients' peripheral blood and synovial tissue is correlated with disease pathogenesis, clinical parameters, and inflammatory mediators such as IL-6 expression.<sup>29</sup> Another study revealed that *FOXO1*-silenced RAFLSs have increased survival with reduced apoptosis due to a reduction in caspase 3/7 activity.<sup>59</sup> *FOXO1*-silenced RAFLSs also have reduced inflammatory cytokines including IL-6, IL-8, TNF- $\alpha$ , and IL-1 $\beta$  which were essential in RA pathogenesis.<sup>59</sup> Herein, we observed the lower expression of *FOXO1* in PBMCs of RA patients which also negatively correlated with disease activity, DAS28 in patients. Our previous study reported the significant upregulation of miR-34a-5p and miR-335-5p.<sup>42</sup> It was reported that miR-34a-5p and miR-335-5p were negatively correlated with *FOXO1* expression in the RA background.

In general, the cell cycle is progressed with cyclin-cyclin-dependent kinase (cyclin-CDK). RB1 is inactivated with CDK kinase activity. Cyclin-dependent kinase inhibitors (CDKIs) activated the RB1 by inhibiting CDK activity. It was reported that CDKI gene delivery into arthritic joints could arrest RAFLS proliferation and reduce the expression of pro-inflammatory cytokine.<sup>60</sup> Active RB1 transfer to RAFLS suppresses the matrix metalloproteinase-3 (MMP-3)

and monocyte chemoattractant protein-1 (MCP-1) production which was crucial in inflammation and joint destruction in RA.<sup>60</sup> *TP53* induction in RAFLS resulted in P21 overexpression, a potent inhibitor of CDK, and could activate RB1 and cell cycle progression in these cells.<sup>51</sup> Herein, the low expression level of *RBI* was reported in PBMCs of RA patients which might be important in the pathogenesis of this disorder.

Based on our analysis the reduction in the expression of studied genes in patients was not associated with their received treatment and we did not find any significant association between patients' treatment and the expression of selected genes, except for alendronate. Alendronate was able to alter *TP53* and *RBI* gene expression to some extent in PBMCs of RA patients that did not influence the statistical analysis.

In conclusion, the expression of apoptosis and cell survival-related genes, such as *TP53*, *BAX*, *FOXO1*, and *RBI*, was reduced in RA patients' PBMCs. This reduction in gene expressions may result in cell survival, resistance to apoptosis, and expression of inflammatory mediators, which were crucial for RA joint destruction or auto-reactive lymphocyte selection. Here, the association between the expression of these genes and the miRNAs previously identified to target them was explored in RA PBMCs. Based on our results, miR-16-5p may negatively influence the expression of *TP53*, while miR-335-5p and miR-34a-5p negatively regulate the expression of *FOXO1* in RA PBMCs. Due to the fact that there is no specific and effective treatment for RA patients, these regulatory miRNAs and their target genes or molecular pathways might be used as future therapeutic targets which require additional research.

## AUTHOR CONTRIBUTIONS

*Acquisition of data, drafting the article, analysis and interpretation of data, and final approval of the article:* Hamidreza Ebrahimiyan, Maryam Akhtari, Maassoumeh Akhlaghi, and Gholam Hossein Alishiri. *The conception and design of the study, revising the article critically, interpretation of data, and final approval of the article:* Elham Farhadi, Ahmadreza Jamshidi, Mahdi Mahmoudi, and Mahmood Tavallaie.

## ACKNOWLEDGMENTS

This study was supported by a grant from the Tehran University of Medical Sciences (Grant No: 98-3-96-46136).

## CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

## DATA AVAILABILITY STATEMENT

The data generated and/or analyzed during the current study are available from the corresponding author upon reasonable request.

## ETHICS STATEMENT

This study was performed based on the Declaration of Helsinki guidelines and was approved by the Ethics Committee of Tehran University of Medical Sciences (Approval No: IR.TUMS.VCR.REC.1398.885). The written informed consent was signed by all participants before enrolling in the study.

## ORCID

Maryam Akhtari  <http://orcid.org/0000-0002-5342-3440>

Elham Farhadi  <http://orcid.org/0000-0002-4324-2697>

Mahdi Mahmoudi  <http://orcid.org/0000-0002-8164-8831>

## REFERENCES

- Littlejohn EA, Monrad SU. Early diagnosis and treatment of rheumatoid arthritis. *Prim Care-Clin Off Pract*. 2018;45(2):237-255.
- Aletaha D, Neogi T, Silman AJ, et al. 2010 rheumatoid arthritis classification criteria: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. *Arthritis & Rheumatism*. 2010;62(9):2569-2581.
- Myasoedova E, Crowson CS, Kremers HM, Therneau TM, Gabriel SE. Is the incidence of rheumatoid arthritis rising?: results from Olmsted County, Minnesota, 1955-2007. *Arthritis Rheum*. 2010;62(6):1576-1582.
- Aletaha D, Smolen JS. Diagnosis and management of rheumatoid arthritis: a review. *JAMA*. 2018;320(13):1360-1372.
- Tan EM, Smolen JS. Historical observations contributing insights on etiopathogenesis of rheumatoid arthritis and role of rheumatoid factor. *J Exp Med*. 2016;213(10):1937-1950.
- Deane KD, Demoruelle MK, Kelmenson LB, Kuhn KA, Norris JM, Holers VM. Genetic and environmental risk factors for rheumatoid arthritis. *Best Pract Res Clin Rheumatol*. 2017;31(1):3-18.
- Wu Z, Ma D, Yang H, et al. Fibroblast-like synoviocytes in rheumatoid arthritis: surface markers and phenotypes. *Int Immunopharmacol*. 2021;93:107392.
- Malemud C, Gillespie H. The role of apoptosis in arthritis. *Curr Rheumatol Rev*. 2005;1(2):131-142.
- Henson PM, Bratton DL, Fadok VA. Apoptotic cell removal. *Curr Biol*. 2001;11(19):R795-R805.
- Harris Perlman P, Lisa J. Pagliari P, Michael V. Volin V. Regulation of apoptosis and cell cycle activity in rheumatoid arthritis. *Curr Mol Med*. 2001;1(5):597-608.
- Cao Y, Liu J. Impaired apoptosis of peripheral blood CD4+T cells in patients with rheumatoid arthritis. *Xi Bao Yu Fen Zi Mian Yi Xue Za Zhi*. 2015;31(5):682-685.
- Zamani MR, Aslani S, Salmaninejad A, Javan MR, Rezaei N. PD-1/PD-L and autoimmunity: a growing relationship. *Cell Immunol*. 2016;310:27-41.
- Schiotis RE, Buzoianu AD, Mureşanu DF, Suci S. New pharmacological strategies in rheumatic diseases. *J Med Life*. 2016;9(3):227-234.
- Khatun J, Risk B, Giddings M. Long noncoding RNAs are rarely translated in two human cell lines. *Genome Res*. 2012;22(9):1646-1657.
- Tavasolian F, Abdollahi E, Rezaei R, Momtazi-borojeni AA, Henrotin Y, Sahebkar A. Altered expression of microRNAs in rheumatoid arthritis. *JCB*. 2018;119(1):478-487.
- Xiao C, Nemazee D, Gonzalez-Martin A. MicroRNA control of B cell tolerance, autoimmunity and cancer. *Sem Cancer Biol*. 2020;64:102-107.
- Nakasa T, Nagata Y, Yamasaki K, Ochi M. A mini-review: microRNA in arthritis. *Physiol Genomics*. 2011;43(10):566-570.
- Ebrahimiyan H, Ahmadzadeh A. miRNAs and rheumatoid arthritis: new update in expression pattern and pathogenicity. *Rheumatol Res*. 2021.
- Wang G, Yao J, Li Z, et al. miR-34a-5p inhibition alleviates intestinal ischemia/reperfusion-induced reactive oxygen species accumulation and apoptosis via activation of SIRT1 signaling. *Antioxid Redox Signal*. 2016;24(17):961-973.
- Lane DP. p53, guardian of the genome. *Nature*. 1992;358(6381):15-16.
- Jain AK, Barton MC. p53: emerging roles in stem cells, development and beyond. *Development*. 2018;145(8):dev158360.
- Hernández Borrero LJ, El-Deiry WS. Tumor suppressor p53: biology, signaling pathways, and therapeutic targeting. *Biophys Acta Rev Cancer*. 2021;1876(1):188556.
- Taghadosi M, Adib M, Jamshidi A, Mahmoudi M, Farhadi E. The p53 status in rheumatoid arthritis with focus on fibroblast-like synoviocytes. *Immunol Res*. 2021;69(3):225-238.
- Abou-Shousha SA, Salah E, Wagdy E. Study of P53 in peripheral blood and synovial mononuclear cells of rheumatoid arthritis and osteoarthritis patients and its relation to the degree of disease activity. *Egypt J Immunol*. 2005;12(2):61-70.
- Murtaza G, Khan AK, Rashid R, Muneer S, Hasan SMF, Chen J. FOXO transcriptional factors and long-term living. *Oxid Med Cell Longevity*. 2017;2017:1-8.
- Wang S, Xia P, Huang G, et al. FoxO1-mediated autophagy is required for NK cell development and innate immunity. *Nat Commun*. 2016;7(1):11023.
- Gross DN, Wan M, Birnbaum MJ. The role of FOXO in the regulation of metabolism. *Curr Diab Rep*. 2009;9(3):208-214.
- Tia N, Singh AK, Pandey P, Azad CS, Chaudhary P, Gambhir IS. Role of Forkhead Box O (FOXO) transcription factor in aging and diseases. *Gene*. 2018;648:97-105.
- Grabiec AM, Angiolilli C, Hartkamp LM, van Baarsen LGM, Tak PP, Reedquist KA. JNK-dependent downregulation of FoxO1 is required to promote the survival of fibroblast-like synoviocytes in rheumatoid arthritis. *Ann Rheum Dis*. 2015;74(9):1763-1771.

30. Kuo CC, Lin SC. Altered FOXO1 transcript levels in peripheral blood mononuclear cells of systemic lupus erythematosus and rheumatoid arthritis patients. *Mol Med*. 2007;13(11-12):561-566.
31. Knudson AG Jr. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci*. 1971;68(4):820-823.
32. Weinberg RA. The retinoblastoma protein and cell cycle control. *Cell*. 1995;81(3):323-330.
33. Dyson N. The regulation of E2F by pRB-family proteins. *Genes Dev*. 1998;12(15):2245-2262.
34. Bradley K, Scatizzi JC, Fiore S, et al. Retinoblastoma suppression of matrix metalloproteinase 1, but not interleukin-6, through a p38-dependent pathway in rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum*. 2004;50(1):78-87.
35. Wangyang Y, Yi L, Wang T, et al. MiR-199a-3p inhibits proliferation and induces apoptosis in rheumatoid arthritis fibroblast-like synoviocytes via suppressing retinoblastoma 1. *Biosci Rep*. 2018;38(6):BSR20180982.
36. Dadsena S, King LE, García-Sáez AJ. Apoptosis regulation at the mitochondria membrane level. *Biochim Biophys Acta-Biomembr*. 2021;1863(12):183716.
37. Arnett FC, Edworthy SM, Bloch DA, et al. The American rheumatism association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum*. 1988;31(3):315-324.
38. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative CT method. *Nat Protoc*. 2008;3(6):1101-1108.
39. Sauerbrey A, Stammer G, Zintl F, Volm M. Expression and prognostic value of the retinoblastoma tumour suppressor gene (RB-1) in childhood acute lymphoblastic leukaemia. *Br J Haematol*. 1996;94(1):99-104.
40. Wilson RL, Owens K, Sumser EK, et al. Nanoparticle mediated increased insulin-like growth factor 1 expression enhances human placenta syncytium function. *Placenta*. 2020;93:1-7.
41. Pei S-G, Wang J-X, Wang X-L, Zhang Q-J, Zhang H. Correlation of survivin, p53 and Ki-67 in laryngeal cancer Hep-2 cell proliferation and invasion. *Asian Pac J Trop Med*. 2015;8(8):636-642.
42. Ebrahimiyan H, Rezaei N, Vojdani M, Aslani S, Jamshidi A, Mahmoudi M. microRNA involvement in the regulation of survivin in peripheral blood mononuclear cells from rheumatoid arthritis patients. *Int J Rheum Dis*. 2019;22(6):1107-1114.
43. Smolen JS, Aletaha D, Barton A, et al. Rheumatoid arthritis. *Nat Rev Dis Primers*. 2018;4:18001.
44. Ogata A, Kato Y, Higa S, Yoshizaki K. IL-6 inhibitor for the treatment of rheumatoid arthritis: a comprehensive review. *Mod Rheumatol*. 2019;29(2):258-267.
45. Kuhlreiber W, Hayashi T, Dale E, Faustman D. Central role of defective apoptosis in autoimmunity. *J Mol Endocrinol*. 2003;31(3):373-399.
46. Gámez-Díaz L, Grimbacher B. Immune checkpoint deficiencies and autoimmune lymphoproliferative syndromes. *Biomed J*. 2021;44(4):400-411.
47. Fisher GH, Rosenberg FJ, Straus SE, et al. Dominant interfering Fas gene mutations impair apoptosis in a human autoimmune lymphoproliferative syndrome. *Cell*. 1995;81(6):935-946.
48. McNally J, Yoo DH, Drappa J, et al. Fas ligand expression and function in systemic lupus erythematosus. *J Immunol*. 1997;159(9):4628-4636.
49. Eguchi K. Apoptosis in autoimmune diseases. *Intern Med*. 2001;40(4):275-284.
50. Maas K, Westfall M, Pietenpol J, Olsen NJ, Aune T. Reduced p53 in peripheral blood mononuclear cells from patients with rheumatoid arthritis is associated with loss of radiation-induced apoptosis. *Arthritis Rheum*. 2005;52(4):1047-1057.
51. Migita K, Tanaka F, Yamasaki S, et al. Regulation of rheumatoid synoviocyte proliferation by endogenous p53 induction. *Clin Exp Immunol*. 2001;126(2):334-338.
52. Aupperle KR, Boyle DL, Hendrix M, et al. Regulation of synoviocyte proliferation, apoptosis, and invasion by the p53 tumor suppressor gene. *Am J Pathol*. 1998;152(4):1091-1098.
53. Tang BX, You X, Zhao LD, et al. p53 in fibroblast-like synoviocytes can regulate T helper cell functions in patients with active rheumatoid arthritis. *Chin Med J*. 2011;124(3):364-368.
54. Yamanishi Y, Boyle D, Pinkoski M, eds. *p53 Regulates Onset and Severity of Murine Collagen-Induced Arthritis. Arthritis and Rheumatism*. Lippincott Williams & Wilkins; 2000.
55. Jeffers JR, Parganas E, Lee Y, et al. Puma is an essential mediator of p53-dependent and-independent apoptotic pathways. *Cancer Cell*. 2003;4(4):321-328.
56. Aubrey BJ, Kelly GL, Janic A, Herold MJ, Strasser A. How does p53 induce apoptosis and how does this relate to p53-mediated tumour suppression? *Cell Death Differ*. 2018;25(1):104-113.
57. Que W, Liu H, Yang Q, Xu S. NK4 inhibits the proliferation and induces apoptosis of human rheumatoid arthritis synovial cells. *Cell Biochem Funct*. 2018;36(5):273-279.
58. Xing Y, Li A, Yang Y, Li X, Zhang L, Guo H. The regulation of FOXO1 and its role in disease progression. *Life Sci*. 2018;193:124-131.
59. Liu Y, Yan X. Eriodictyol inhibits survival and inflammatory responses and promotes apoptosis in rheumatoid arthritis fibroblast-like synoviocytes through AKT/FOXO1 signaling. *JCB*. 2019;120(9):14628-14635.
60. Nonomura Y, Nagasaka K, Hagiyaama H, et al. Direct modulation of rheumatoid inflammatory mediator expression in retinoblastoma protein-dependent and -independent pathways by cyclin-dependent kinase 4/6. *Arthritis Rheum*. 2006;54(7):2074-2083.

**How to cite this article:** Ebrahimian H, Akhtari M, Tavallaie M, et al. Altered expression of apoptosis-related genes in rheumatoid arthritis peripheral blood mononuclear cell and related miRNA regulation. *Immun Inflamm Dis*. 2023;11:e914. doi:10.1002/iid3.914