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M.Sc. BIOINFORMATICS AND THEORETICAL SYSTEMS BIOLOGY

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# bgwas3: a pipeline for kmer based association testing in bacteria

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## Acknowledgements

# Abstract

Genome-wide association studies (GWAS) are now applied more often to microbes. However, bacteria genomes are significantly more variable in both content and sequence than eukaryotic genomes, meaning these association studies have naturally different, more complex considerations. These studies utilise short unique DNA patterns, referred to as Kmers, as units of genetic variation as opposed to single nucleotide polymorphisms. Various software solutions are now available which can perform the multiple association tests of kmers, notable the python based *pyseer*. However, there is not at present a succinct tool which pipelines the necessary pre and post data processing and analysis steps such as associating the Kmers to genes, that make an association study whole.

In metabolomic studies of bacteria there are thousands of possible measures that can be made which can be considered as traits of that bacteria at a that point in time. Coupling genomics and metabolomics may provide a powerful technique to understanding how a bacteria has adapted, and in the scope of disease, may aid in the understanding of the function of drug resistance and other virulence features. However conducting individual association tests for each trait may prove time and computationally intensive.

I developed a tool, *bgwas2*, which wraps the Kmer association function of *pyseer* with other open source software tools into a single installable package. When run, *bgwas3* takes the simplest of input files and a configuration file, and performs gene annotation, phylogeny estimation, kmer association testing, kmer-gene mapping and finally visualisation with a automatically generated web report.

In creating the tool, best practices for computational biology were exercised; resulting in a final tool that expresses appropriate testing, documentation and packaging; meaning it can easily be utilised by others in the future. CGAT-core is implemented in Python 3 and installable via Conda and PyPI with minimal dependencies. We have successfully deployed and tested the code on OSX, Red Hat and Ubuntu. We have made CGAT-core and associated repositories open-source under the MIT licence, allowing full and free use for both commercial and non-commercial purposes. Our software is fully documented (<https://pypi.org>), version controlled and has extensive testing using continuous integration (<https://travis-ci.org/cgat-developers>.) We welcome community participation in code development and issue reporting through GitHub

The tool was also used to test the association of 26 traits - 18 relating corresponding to a metabolomic measurement, and 8 corresponding to either a measure of antibiotic resistance or bacterial motility. In 14 of the traits, between 1 and 16450 significant Kmers were found, and were mapped to Genes.

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## Abbreviations

**Foo** description of foo

**Bar** descirption of bar

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# 1 Introduction

## 1.1 Pangenome wide association studies

Genome wide association studies have been successfully used to identify genetic and phenotype associations in a hypothesis free manner. Characteristics of microbes suggest they would be ideal candidates for GWAS: their genomes are small which limits the study size; they exude much less phenotypic flexibility; and, generally, phenotypes that are of interest such as virulence and drug resistance tend to be the result of strong selective pressures. There is also good reason to perform association studies with bacteria. As identifying the genetic basis of traits that correspond with its pathogenic ability could lead to better understanding of disease and lead to new targets for pharmaceutical intervention.

Commonly, GWAS have focused on single nucleotide polymorphisms (SNPs) as units of genetic variation. SNPs are useful in organisms where recombination is reliable and genomes are relatively constant among populations. Because of linkage disequilibrium, identifying a highly associated SNP often means that the true associated loci is nearby.

However, the nature of bacteria replication makes the use of SNPs in association studies less useful. For one, significant regions of the genome can be in linkage disequilibrium; and though some species experience high rates of recombination, the recombination is not as reliable or consistent as in eukaryotes to reliably reduce linkage.

Secondly, bacteria experience much higher trans- during recombination. Therefore, bacteria of the same species can vary considerably in both order of gene content, and in the accessory genome - regions of the genome which differ.

Therefore, first finding SNPs becomes a tricky process which involves multiple alignments. Such tools such as MUMmer (and other?) do meet this problem, but even still, SNPs associated with only the core genome can be retrieved and does not consider the huge variable accessory genome in bacteria/ will not encapsulate the full

As an alternative, some recent studies have instead utilised K-mers.

Kmers are simply variable length sequences of DNA. Initially a concept of genome assembly. Can capture multiple genetic variations SNPs, longer deletions/ insertions and recombination. The size of kmers affects the genetic variation captured. Longer more specific. Shorter are more sensitive.

Getting kmers is alignment free, releases the burden of full alignment. But more specifically utilising kmers



means the core and accessory genome can be tested.

*Seer* (Lees et al. 2016) and its python reimplementation *pyseer* (Lees et al. 2018) *dbgwas*

## 1.2 The Need for Multiple Association Tests - Metabolomics

Metabolomics provides a detailed snapshot of an organism's physiological state through the quantification of hundreds to thousands of small molecules (Zampieri et al., 2017).

Coupling genomics and metabolomics to study *Pseudomonas aeruginosa* in unique clinical samples can allow us to understand how bacteria adapt to the lung environment.

The nature of metabolomics means that multiple molecules abundance may be measured and of interest to investigate.

Determine the genetic basis of metabolomics of *P.* in the context of chronic infections

## 1.3 Scope of work

The foundation of this project was to create a tool, *bgwas3*, an integration of multiple open source genomics tools and custom scripts written in python, R and bash into a single installable package that facilitates conducting multiple pangenome wide association studies in a single user step.

The tool aims to be comprehensive and robust enough to convert only the most basic required input files into valuable and interpretable results including static and interactive visualisations.

When run on a node within a computer cluster, tasks that are computationally intensive or long are distributed among the cluster, and can run simultaneously.

Through the additional configuration file, the specifics of multiple intermediary steps can be altered, which can have variable effects on the final results.

In light of the necessity to perform multiple.. (1)

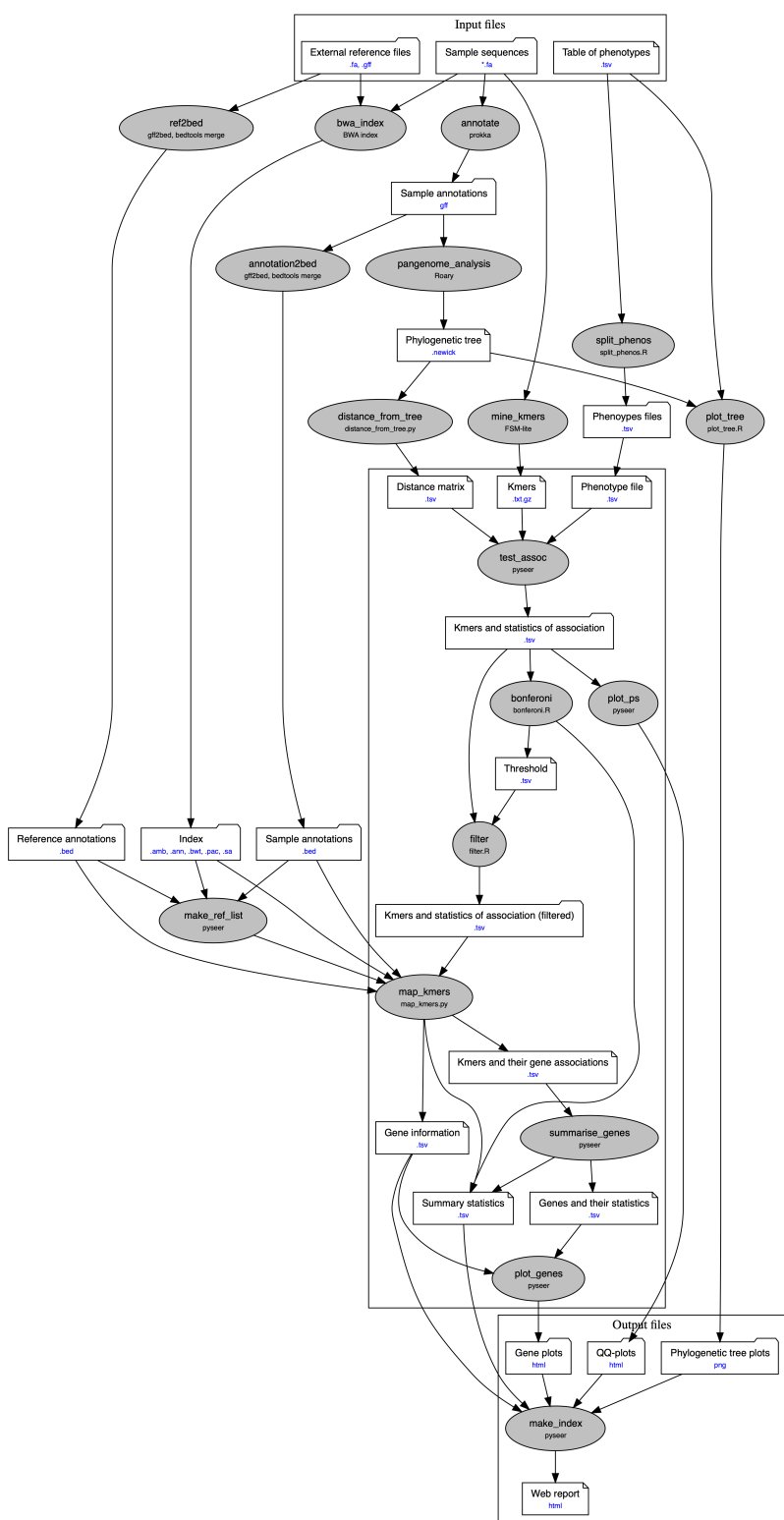


Figure 1: Pipeline of bgwas

## 2 Implementation

### 2.1 Pipelining with CGAT-Core

I wrote *bgwas3* primarily in python, utilising the pipelining framework CGATCore, (Cribbs et al. 2019) The CGATCore is extremely portable package coming as a dependency free Python package, but has been used to construct many complex pipelines that are scalable to large amounts of data ?ref

With the framework, individual pipelining steps, referred to as ‘tasks’, are defined as python functions whose arguments include the necessary input files and the expected output files.

With the use of python decorators, the output files of one task can be taken as the input arguments of later tasks. Allowing multiple data analysis to be strung together and run in the correct order. The framework utilises a system of checking the modification date of files, so only runs tasks in which their are either new input files, or input files have been modified. This mechanism is exceedingly useful, as a directory of project files may be moved from one computer to another, and the framework will still recognise which tasks and steps are out of date.

CGATCore introduces another significant feature which is an interface to control a distributed resource management application API (DRMAA) such as PBS-Pro used by Imperial College London's resource computing service. Therefore, tasks which are deemed large are need to be run at batch, parallel processes can be distributed to computers in a cluster, and the system resource (eg RAM) can be set on a per task-basis. This makes executing pipeline that involve computationally intensive intermediary steps faster.

*bgwas3* was written as 15 ‘tasks’ which are visualised in 1.

The

### 2.2 Genome Annotation

In the *bgwas* task “annotatate” all sample sequences are annotated by Prokka (Seemann 2014) The tool functions as follows: It first makes predictions on features using a number of external tools including Prodigal [1] for the annotation of coding region, RNAmmer for ribosomal RNA genes, Aragorn for transfer RNA genes, SignalP for signal leader peptides and Infernal for non-coding RNA. After feature prediction, the speculative features are queried against a number of databases such as UniProt [2], Refseq [3] and Pfam.

The function of annotating input sequences in the *bgwas3* pipeline is twofold. First, gene annotations are later used to estimate the phylogenetic tree of the sample, and secondly, significantly associated Kmers are

later mapped to the annotated genomes when identifying the Kmer's genetic identity.

## 2.3 Kmer mining and counting

*bgwas3* integrates the tool fsm-lite (Välimäki 2018) to count 'mine' and count Kmers. of user defined variable length kmers in the all samples. fsm-lite benefits first by being able to run on a single core, but also because of its ability to read and count Kmers not of a single length, but of a range of lengths, unlike similar tools such as DSK? As mentioned before, the length of Kmers effects the specificity and ? of the association testing. Therefore *bgwas3* allows the Kmers length to be changed by editing the configuration yml file ?

## 2.4 Phylogeny estimation and covariate estimation

*bgwas3* currently takes a pangenomic approach to phylogeny estimation; as in it considers the relative presence and absence of genes in the samples as An estimated phylogenetic tree is estimated with *bgwas3* primarily utilizing the tool Roary (Page et al. 2015). In summary, Roary determines genes which fall within the core genome, and then performs clustering of isolates are clustered based on gene presence in the accessory genome. A newick tree output of roary, is then converted into a distance matrix using a python script from pyseer.

The single reasoning for predicting phylogeny in the *bgwas3* pipeline is to use the distances defined in the distance matrix as covariates in the association testing.

This is different to the standard method of multi-dimensional scaling often used in human gwas and as part of the standard seer workflow. In general, if a phylogeny is accurate, th

## 2.5 Kmer association testing

For linear mixed model association testing, a *bgwas3* constructs a distance matrix based on a phylogenetic tree. Association between kmers and phenotypes is implemented with pyseer (Lees et al. 2018).

Specifically, pyseer is used to perform a linear mixed model using the Fast-LMM algorithm (Lippert et al. 2011).

LMM tackle confounders in association tests by using a measure of similarity (in this case a distance defined from a estimated phylogenetic tree) as a random effect in a linear model.

TODO equation

Given a means of determining the hierarchical relatedness between samples, the mixed model is generally preferred, and has been shown in past studies to control the inflation of p-values better (Lees et al. 2017).

## 2.6 Bonferroni correction

The output of `pyseer`, a list of all kmers and statistics relating to their association to the given phenotype, are then filtered by their p-value through Bonferroni correction. The Bonferroni correction is a multiple-comparison correction used when several dependent or independent statistical tests, utilised to limit the number of spurious positive tests. If multiple hypotheses are tested, the chance of a rare event increases, and therefore, the likelihood of incorrectly rejecting a null hypothesis (i.e., making a Type I error) increases. An R script `bonf.R` calculates a threshold p-value of which to accept an associated kmer as significant based on its p-value.

## 2.7 Significant Kmer mapping

A new script, written in R `map_kmers.R`, and which calls on the external tools BWA-Mem [1] and Bedtools [2] attempts to assign each associated kmer to a gene found in either a supplied reference genome, or one of the sample genomes which has previously been annotated. The script works as follows, and demonstrated in `fig:make_kmers`.

1. Generate a Multi-Fasta file whereby all Kmers are represented as a single fasta entry.
- 2.

## 2.8 Visualisation

An important feature of *bgwas3* is the automated generation of multiple figures, integrated into a final web-based report.

## 2.9 Scientific computing practices

A significant goal of *bgwas3* was to implement a useful and reusable tool that may be used outside the scope of this singular project.

As such, various software development principles and concepts were applied to the project.

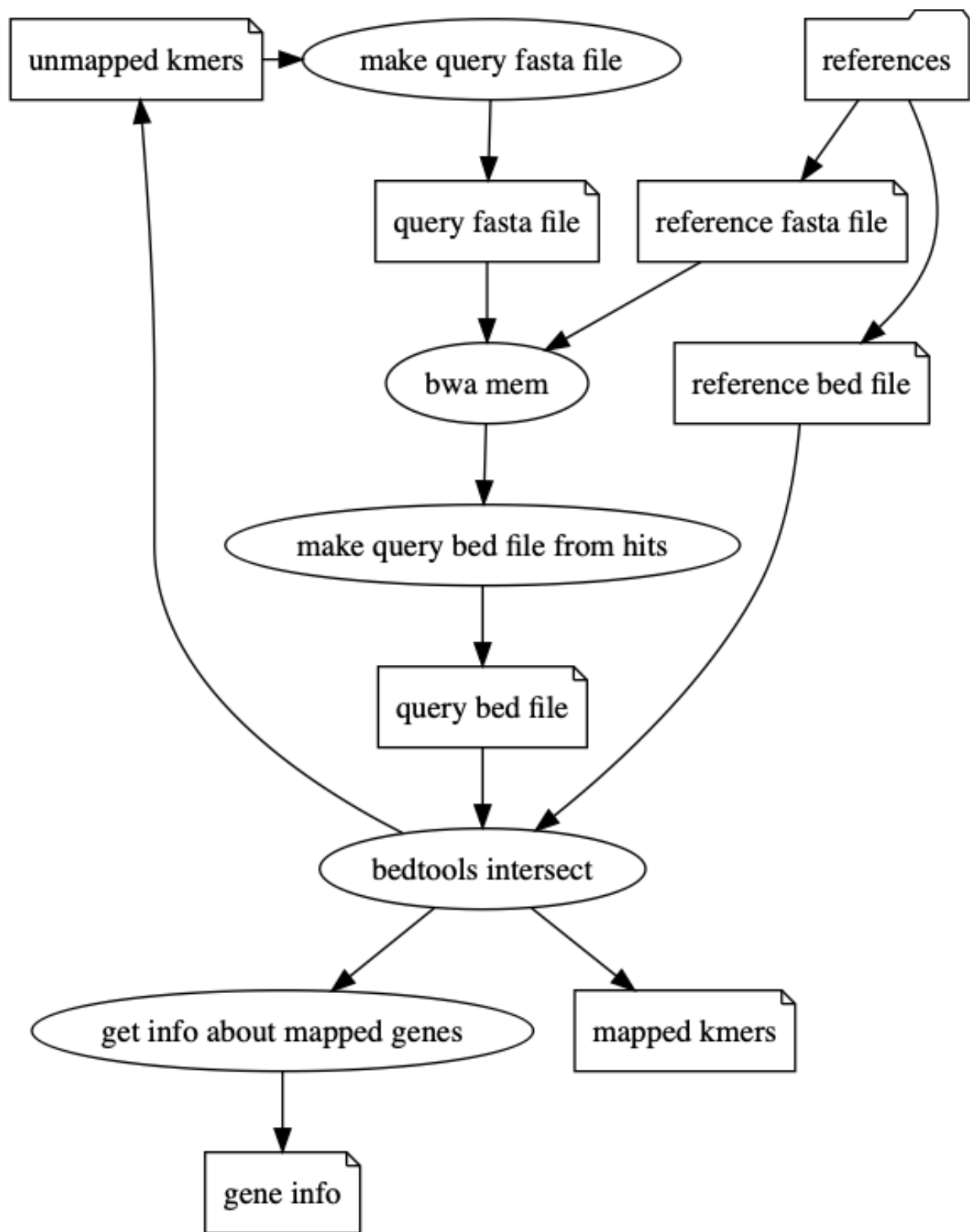


Figure 2: Graphical representation of algorithm to map Kmers to reference genes

### 2.9.1 Testing with PyTest and Travis

Pytest and

### 2.9.2 Documentation with Sphinx

### 2.9.3 Packaging with Conda

## 3 Results

For the development, testing, and evaluation of *bgwas3* a dataset of

Cystic fibrosis is an autosomal recessive disorder that, due to a single gene mutation, limits the ability to *Pseudomonas aeruginosa* is one of the primary bacteria present in the lungs of late stage and terminal patients with cystic fibrosis. ref(Schaedel et al., 2002; Hauser, 2011) (becomes chronic). when the lung begins to exhibit decreased function and signs of failure For this reason, understanding the genetic adaptations *pseudomonas* experience when in chronic state is significantly important in talking this disease

This dataset involves  $n$  strains collected over a period of 24 years from 18 patients. ref

In paper () it was shown that there was ‘clear evidence’ of the metabolic lung environment surrounding the bacteria. Specifically acetate production was negatively associated with length of infection and others?

To test the usability and application of *bgwas3*,

The strong LD caused by the clonal reproduction of bacterial populations means that non-causal  $k$ -mers may also appear to be associated.?

Confirmation of known resistance

Table 1: Table of non-metabolite phenotypes

Name	Description
Tobromycin	Resistance to inhalant antibiotic Tobromycin
Imipenem	Resistance to intravenous antibiotic Imipenem
Aztreonam	Resistance to intravenous/intramuscular antibiotic Aztreonam
Ciprofloxacin	Resistance to oral antibiotic Ciprofloxacin
Colistin	Resistance to ‘last-resort’ antibiotic Colistin

Name	Description
Swim	Measure of cell surface bacteria movement by flagella
Swarm	Measure of rapid surface movement by multiple bacteria with rotating flagella
Twitch	Measure of slow bacteria movement powered by pili

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Chemical

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Hydrogen Cyanide

Cyanide

2-Furoate

3-Hydroxyisovalerate

3-Methylthiopropionic acid

Anthranilate

Betaine

Cystine

Formate

Fumarate

Histidine

Isoleucine

Leucine

Methanol

Methionine

Tryptophan

Uracil

Valine

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### 3.1 Processing of phenotypes

When association testing a qualitative variable with linear regression, it is generally assumed that the variable follows a normal distribution. Non-normal variables may fail to control for type-1 errors.

Quantitative (continuous) traits are preferred because they contain more information. the linear regression



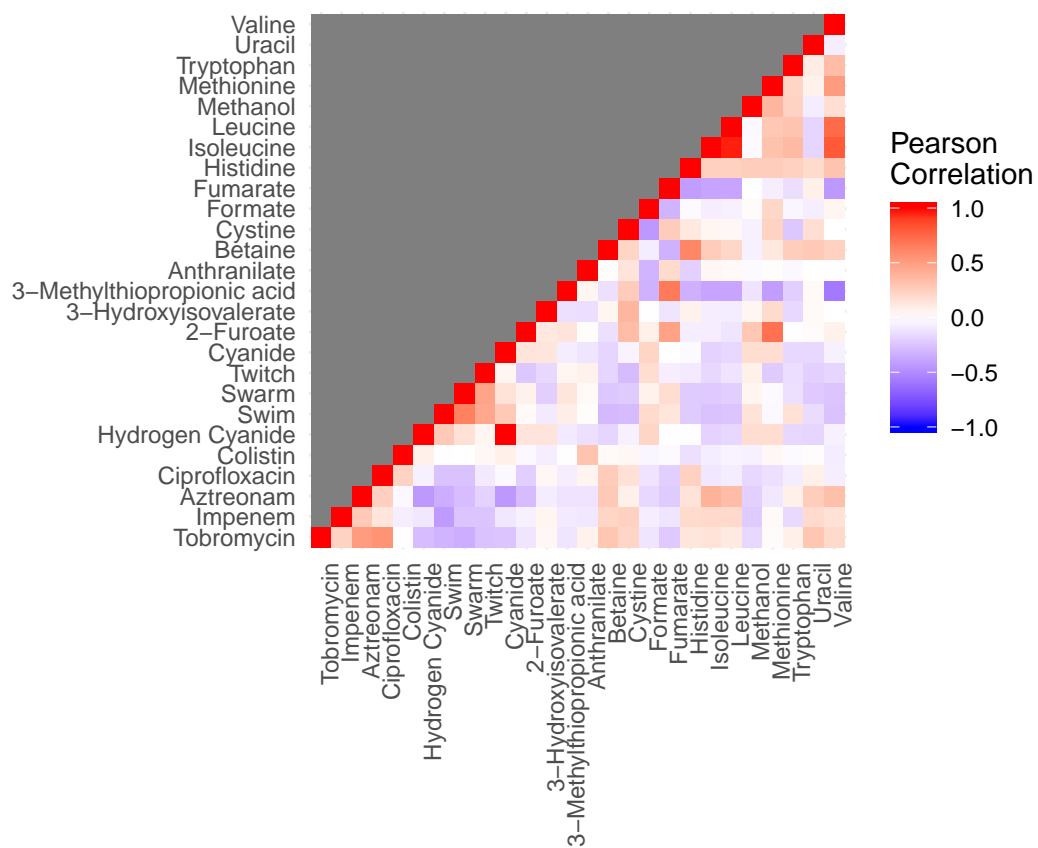


Figure 3: Correlation matrix of phenotypes

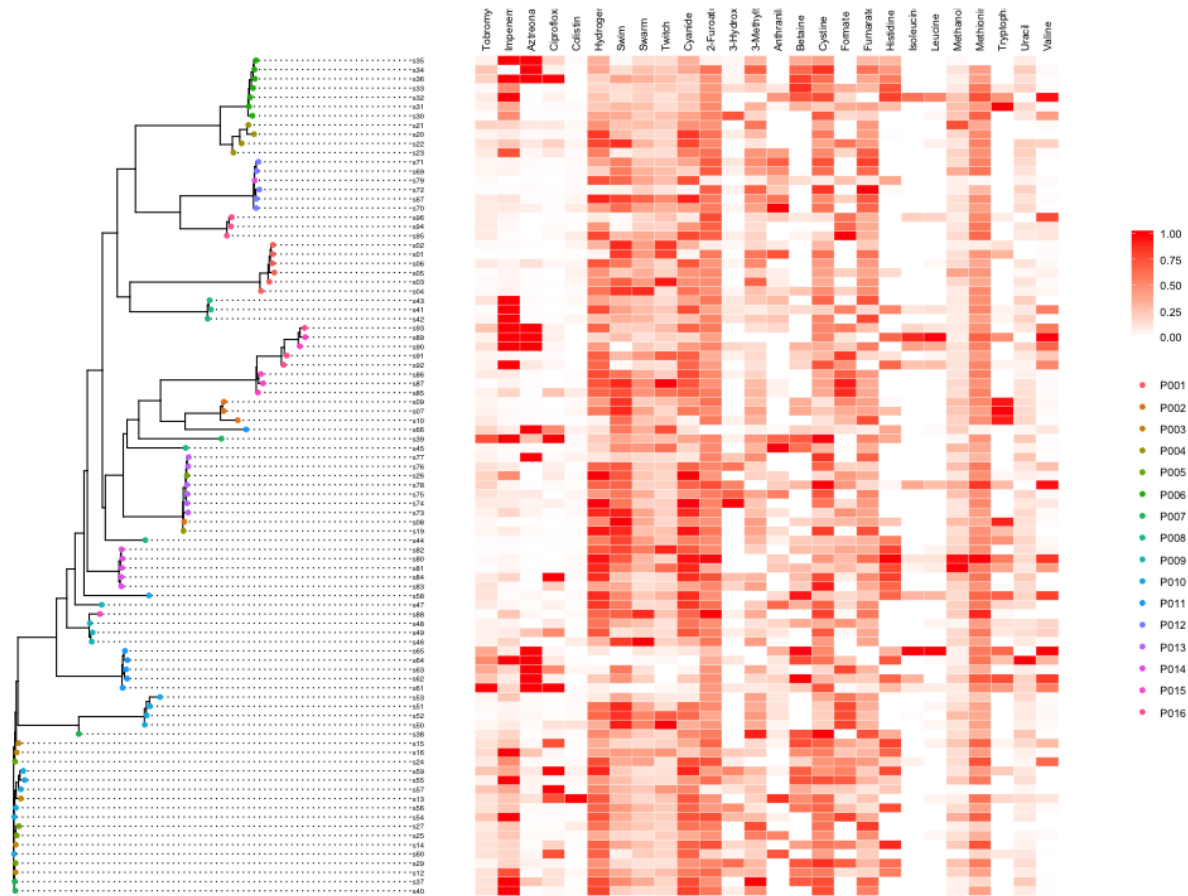


Figure 4: Density plots

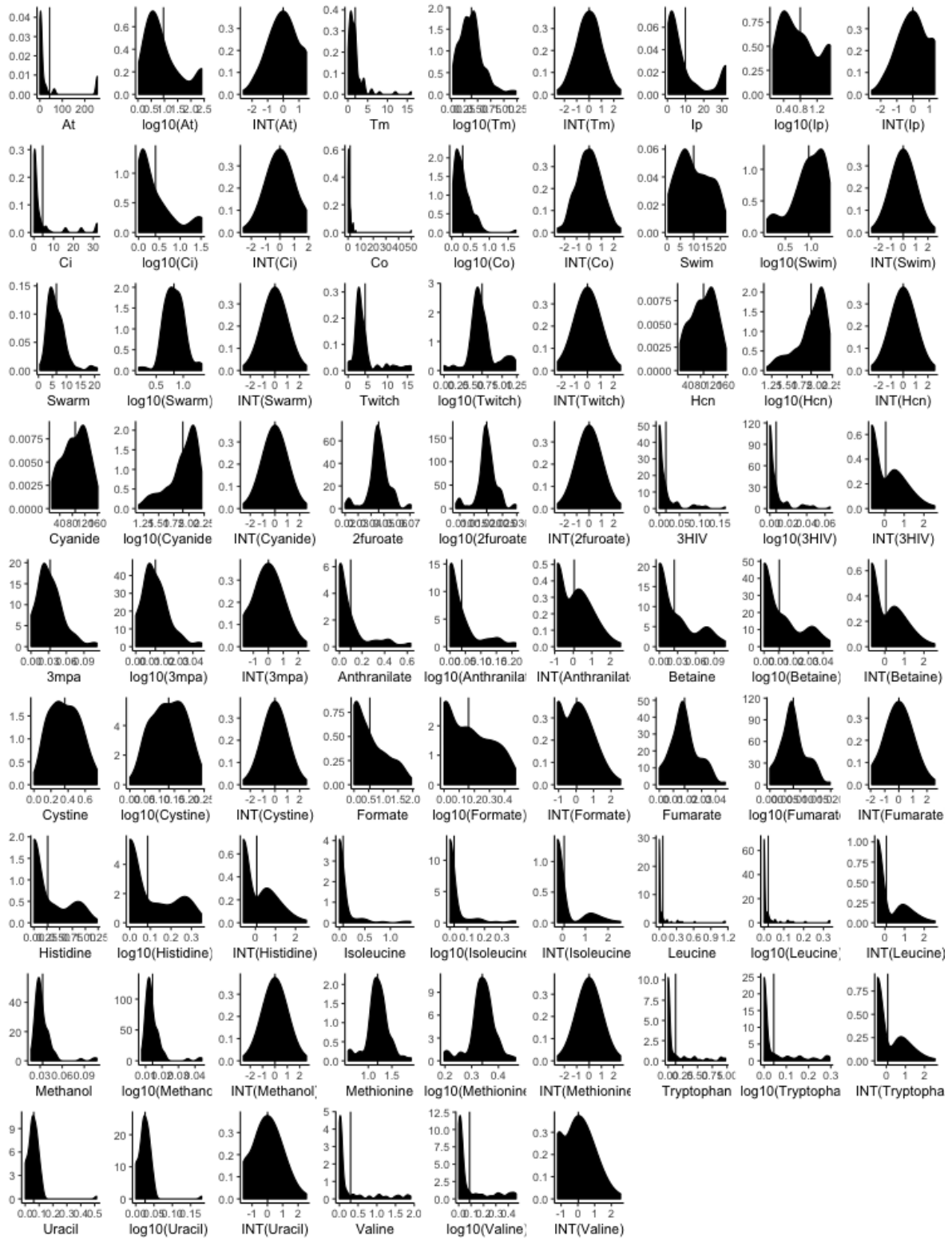


Figure 5: Density plots

analysis requires all variables to be multivariate normal.

A popular statistical technique to bring normality to a continuous trait in association studies involves performing log transformation, or a rank based normal transformation.

Prior to running *bgwas*, the  $n$  phenotypes were separately log transformed and INT, essentially tripling the number of phenotypes tested.

## 3.2 Annotation

All 91 genomes were annotated and 14643 unique genes were identified using Prokka.

## 3.3 Kmer mining

Number etc

## 3.4 Association results

Phenotype	Significant Kmers	Genes
Leucine	16450	289
log(Colistin)	9811	233
Isoleucine	5097	132
log(Isoleucine)	4985	128
log(Leucine)	4913	125
Uracil	2707	75
log(Uracil)	2470	72
Tobromycin	1848	46
log(Aztreonam)	900	19
Ciprofloxacin	285	14
3-Hydroxyisovalerate	1814	13
log(3-Hydroxyisovalerate)	1814	13
Anthranilate	210	7
log(Anthranilate)	210	7
Methanol	216	7

Phenotype	Significant Kmers	Genes
log(Methanol)	216	7
Swarm	229	7
log(Swim)	2	1
Tryptophan	20	1
log(Tryptophan)	1	1
Aztreonam	26891	0
log(Ciprofloxacin)	1	0
log(Hydrogen Cyanide)	1	0
isoleucine_int	1	0

## 4 Discussion

In this work... ## Genome annotation

### 4.1 Kmer mining

The tool fsm

Prokka is a good tool ?Settings ## Phylogeny prediction Currently, *bgwas3* implements only a pangenomic approach approach of distance estimation. There are other tools which involve alignment of the core genome and snps... May or may not be better Reintroduce the problem of a large multiple alignment.

## References

Cribbs, Adam P., Sebastian Luna-Valero, Charlotte George, Ian M. Sudbery, Antonio J. Berlanga-Taylor, Stephen N. Sansom, Tom Smith, et al. 2019. “CGAT-Core: A Python Framework for Building Scalable, Reproducible Computational Biology Workflows.” *F1000Research* 8 (April): 377. <https://doi.org/10.12688/f1000research.18674.1>.

Lees, John A, Nicholas J Croucher, David Goldblatt, François Nosten, Julian Parkhill, Claudia Turner, Paul Turner, and Stephen D Bentley. 2017. “Genome-Wide Identification of Lineage and Locus Specific

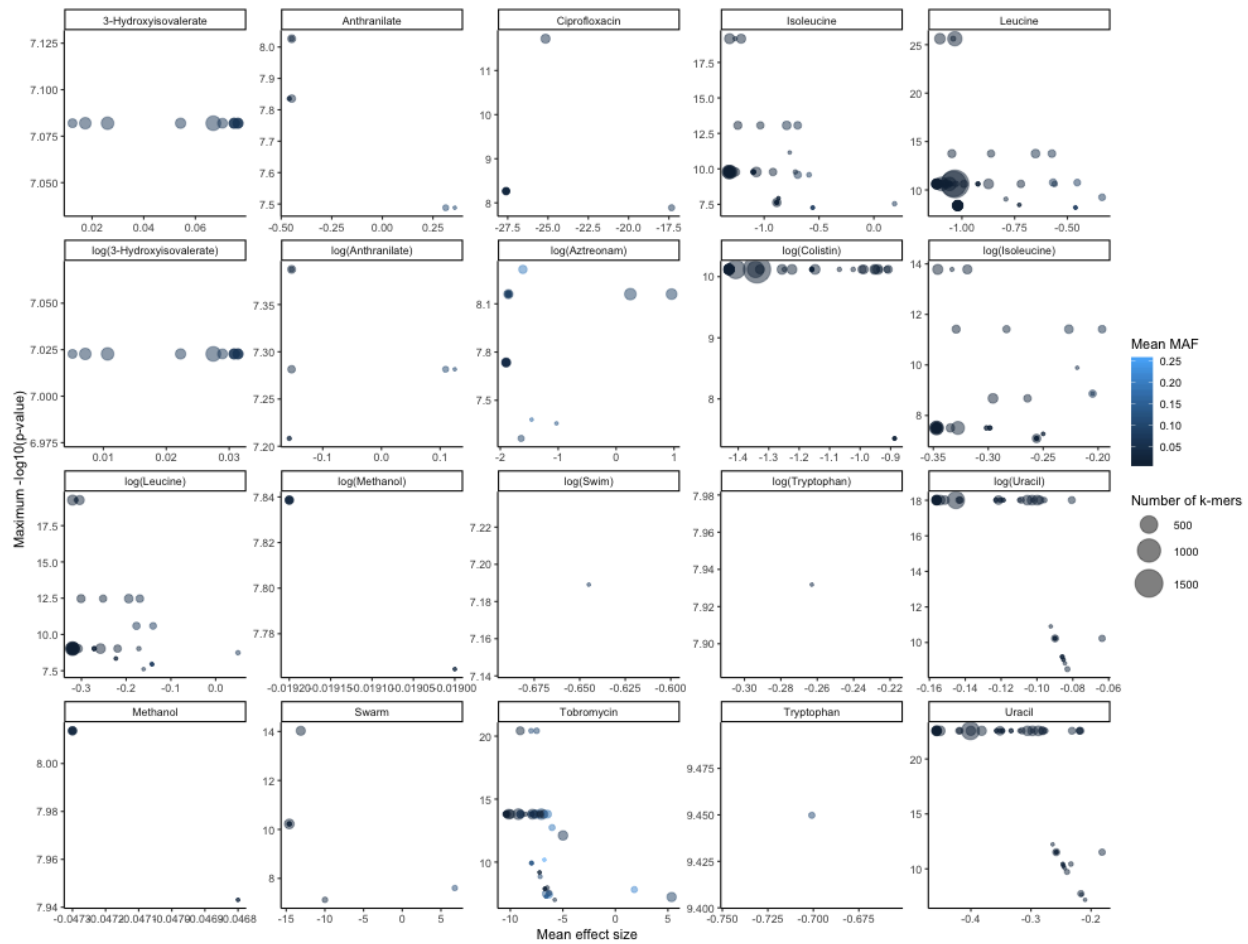


Figure 6: Gens

- Variation Associated with Pneumococcal Carriage Duration.” Edited by Sarah Cobey. *eLife* 6 (July): e26255. <https://doi.org/10.7554/eLife.26255>.
- Lees, John A, Marco Galardini, Stephen D Bentley, Jeffrey N Weiser, and Jukka Corander. 2018. “Py-seer: A Comprehensive Tool for Microbial Pangenome-Wide Association Studies.” Edited by Oliver Stegle. *Bioinformatics* 34 (24): 4310–2. <https://doi.org/10.1093/bioinformatics/bty539>.
- Lees, John A., Minna Vehkala, Niko Välimäki, Simon R. Harris, Claire Chewapreecha, Nicholas J. Croucher, Pekka Marttinen, et al. 2016. “Sequence Element Enrichment Analysis to Determine the Genetic Basis of Bacterial Phenotypes.” *Nature Communications* 7 (1): 12797. <https://doi.org/10.1038/ncomms12797>.
- Lippert, Christoph, Jennifer Listgarten, Ying Liu, Carl M Kadie, Robert I Davidson, and David Heckerman. 2011. “FaST Linear Mixed Models for Genome-Wide Association Studies.” *Nature Methods* 8 (10): 833–35. <https://doi.org/10.1038/nmeth.1681>.
- Page, Andrew J., Carla A. Cummins, Martin Hunt, Vanessa K. Wong, Sandra Reuter, Matthew T. G. Holden, Maria Fookes, Daniel Falush, Jacqueline A. Keane, and Julian Parkhill. 2015. “Roary: Rapid Large-Scale Prokaryote Pan Genome Analysis.” *Bioinformatics* 31 (22): 3691–3. <https://doi.org/10.1093/bioinformatics/btv421>.
- Seemann, Torsten. 2014. “Prokka: Rapid Prokaryotic Genome Annotation.” *Bioinformatics* 30 (14): 2068–9. <https://doi.org/10.1093/bioinformatics/btu153>.
- Välimäki, Niko. 2018. “Fsm-Lite.” <https://github.com/nvalimak/fsm-lite>.