

User manual for e3SIM:
Epidemiological-ecological-evolutionary simulation
framework for genomic epidemiology

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Glossary

drug resistance	The trait value affecting the probability of pathogen survival. A higher value of drug resistance is associated with a higher probability of survival at each tick. 25 , 36
effective contact	An effective contact involves a pair of hosts connected by an edge, where one host is infected (the infector) and the other is capable of being infected (the infectee). When superinfection is not activated, only susceptible state hosts are capable of being infected, while hosts in susceptible, exposed and infected states are capable of being infected when superinfection is activated. 35
epoch	A range of consecutive ticks. Within each epoch, the base rates for compartmental transitions and the genetic architecture for each trait remain constant. 35 , 38
host	The larger organism that harbors one or more pathogens. 5 , 34
initial sequences	The pathogen genome(s) that infect a fully susceptible host population when an outbreak simulation starts. 14
pathogen	The organism whose transmission between hosts and genome evolution are explicitly modeled. 5
tick	Analogous to the definition of tick in population genetics simulation framework SLiM. It is the time unit of epidemiological events and mutation, and the unit of branch length in the genealogy output. 15 , 32 , 35 , 38
transmissibility	The trait affecting the probability of pathogen transmission per contact. A higher value of transmissibility is associated with a higher probability of transmitting the pathogen to a connected host at each tick. 25 , 35

Preface

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0.1 An overview of e3SIM

e3SIM (Epidemiological-ecological-evolutionary **simulation** framework for genomic epidemiology) is an agent-based, discrete, forward-in-time outbreak simulator that models pathogen evolution and transmission dynamics featuring host population contact structures. The tool consists of three components: pre-simulation, main simulation, and post-simulation, all accessible through the command line. Additionally, we offer a graphical user interface (GUI) to let users interactively run all the pre-simulation modules and generate a configuration for the main module. The GUI is structured to guide users step-by-step through the process of configuring and initiating simulations, providing clear and intuitive interaction with the software's features.

0.1.1 Introduction

e3SIM implements an agent-based, discrete and forward-in-time approach to simultaneously simulate the transmission dynamics and molecular evolution of a haploid transmissible **pathogen** population within a static host population structure. It highlights the coupling of evolutionary and epidemiological processes and explicitly models transmission on top of a **host** contact network. Utilizing a SEIRS compartmental model, e3SIM balances the simplification of real-world processes with the theoretical constructs inherent in both epidemiological and quantitative genetic models. For a thorough introduction to the background and principles on which e3SIM is built, please refer to our manuscript.

The package integrates Python, R, and SLiM scripts. Its first component, the pre-simulation modules, generates essential input files for the main module to run. The second part executes the main simulation using SLiM—a script-based simulator designed for population genetics—as the backend engine. We allow various permissible transitions between compartments, where transmissions and treatments are affected by genetically based trait values. The third component, executed together with the main module, includes analysis and visualizations for the simulation output, including raw whole-genome sequences, summary statistics, and graphic representations of results, such as the phylogeny of the transmission tree and the compartments' trajectories averaged over all simulation replicates.

0.1.2 A quick summary of usage

e3SIM contains five modules. Four modules belong to the pre-simulation component: *NetworkGenerator* (Section 2.1), *SeedGenerator* (Section 2.2), *GeneticEffectGenerator* (Section 2.3), and *HostSeedMatcher* (Section 2.4).

One module belongs to the main component: *OutbreakSimulator* (Chapter 3), which also executes the post-simulation modules. The software can be run in one of the following ways.

- **Sequential:** Run each module individually and sequentially on the command line.
- **Streamlined:** Run all pre-simulation, main, and post-processing modules together from the command line using a complete configuration file. However, this is not recommended when users are still adjusting parameters, as e3SIM does not currently support checkpointing/caching and resuming from an intermediate time point.
- **GUI:** Run pre-simulation modules sequentially in the GUI and then execute *OutbreakSimulator* on the command line, using the configuration file generated by the GUI.

The pre-simulation modules generate several specific input files necessary for the main simulation. Users can also create these input files themselves, but the files should follow a specific format. The required files for the main simulation include:

1. **Host contact network:** a space-delimited adjacency list file defining the host contact network, where each row represents a host in order and lists its direct connections. This file is generated by the `NetworkGenerator` module (Section 2.1).
2. **Seeding pathogen sequences:** One file for each seed in the Variant Call Format (VCF) containing the mutation profiles of the seeding pathogen sequences, which are introduced to the host population at the start of the simulation. These files are generated by the `SeedGenerator` module (Section 2.2) through stochastic simulation from the reference genome, unless users choose to seed the host population with the reference genome. The phylogeny of the seeding sequences can also be provided or generated in the Newick (NWK) format.
3. **Genetic architecture specification:** A CSV file detailing the genetic architecture of pathogen genomes, specifying the effect sizes for traits such as transmissibility and drug resistance associated with mutations in designated genetic elements. This file is generated by the `GeneticEffectGenerator` module (Section 2.3).
4. **Host-seed assignment:** A CSV file that maps each seeding pathogen sequence to a specific host within the contact network. This file is generated by the `HostSeedMatcher` module (Section 2.4).
5. **Main module configuration:** A JSON file that provides the epi-eco-evo parameters used in the main module. If the GUI is used to run the pre-simulation modules, this file will be generated by the GUI. For users who prefer directly interacting with the configuration file, a template of the configuration is provided in our GitHub repository for reference.

Templates for all required input files are available in our GitHub repository. If users choose to create these input files manually rather than using the e3SIM pre-simulation modules, we still recommend running the relevant pre-simulation modules in `user_input` mode (adding the `-method user_input` flag to each pre-simulation module). This will validate the user-provided input files and make a copy of the files with fixed filenames in the specified working directory.

0.2 Installation

Please follow the instructions in `README.md`.

Part I

Pre-simulation modules

Chapter 1

Overview

Before diving into how to navigate the software, let us take a look at the prerequisites for running the pre-simulation modules:

- A **reference genome** in FASTA format
- A path to a **working directory**. All e3SIM pre-simulation modules, when running on the command line, require a consistent path to the working directory (provided by the `-wkdir` option), so that all input files will be generated in the same directory.

Other inputs may be required depending on specific user needs. For example, in order to randomly generate a genetic architecture for the genome, a GFF (general feature format) file is required when running the **GeneticEffectGenerator** program in random generation mode. We will introduce all required and optional inputs below and include more details in the respective chapters.

Each pre-simulation module should be run in order to ensure the proper functioning of dependent programs. Below is a typical workflow of running pre-simulation modules:

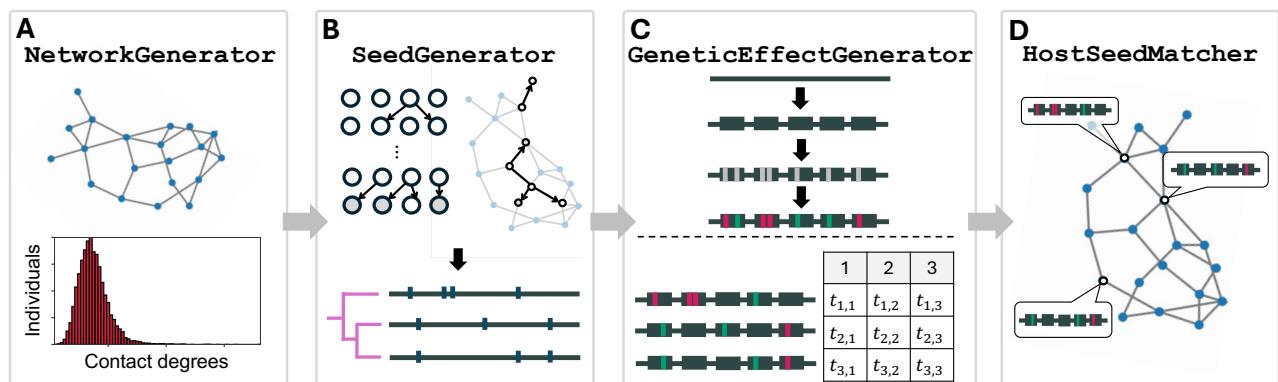


Figure 1.1: Pre-simulation modules.

1. Run **NetworkGenerator** to create the host population's contact network. The network is required by **OutbreakSimulator**, but also needed for **SeedGenerator** if you want to generate seed sequences using a burn-in with an epidemiological model, for example.
2. Run **SeedGenerator** if you want your seed sequences to be different from the provided reference genome. If you want to use the unchanged reference genome as the initial sequence for all your seeds, this step could be skipped.

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3. Run ***GeneticEffectGenerator*** if you want to enable non-neutral mutations in your simulation. When this step is skipped, mutations are still modeled but do not affect the trait values of the pathogens.
4. Run ***HostSeedMatcher*** after ***NetworkGenerator*** and ***SeedGenerator*** to match each initial sequence to a specific host.

In the following chapters, different options to interact with the pre-simulation modules are introduced.

Chapter 2

Run each pre-simulation module

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2.1 NetworkGenerator

e3SIM models the host population and its contact network explicitly. Hosts are modeled as “subpopulations” in SLIM. The host contact network is modeled as an undirected, unweighted graph, with nodes representing hosts and edges indicating contacts between them. The network structure remains static throughout the main simulation. In the main module, only connected hosts can transmit pathogens to each other.

NetworkGenerator is used to generate an adjacency list in the working directory that stores the host contact network. To run *NetworkGenerator*, the required inputs are path to the working directory (`-wkdir`), host population size (`-popsize`), and method of specifying the contact network (`-method`), which could be either random generation (`-method randomly_generate`) or user-defined contact network (`-method user_input`). In the user-input mode, users must specify the path to a customized adjacency list rather than use *NetworkGenerator* to randomly generate one; however, it is still recommended to execute this program to validate your network file.

Here is how you can run *NetworkGenerator*:

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize ${HOST_SIZE} \
-method ${METHOD} \
[Other params]
```

Listing 2.1: NetworkGenerator Usage

We introduce all the options:

- `wkdir: [*REQUIRED] (string)`
Absolute path to the working directory, which should be consistent in one workflow for running each module.
- `popsize: [*REQUIRED] (integer)`
Size of the host population, i.e. number of nodes in the network graph.

- `method`: [*REQUIRED] (string) `user_input` or `randomly_generate`

The method of generating the network, which can only be one of the two methods above. By specifying `user_input`, users are required to provide their own contact network file by `-path_network`. By specifying `randomly_generate`, users are required to provide a random network model by `-model` and corresponding parameters.

- `path_network`: [*OPTIONAL] (string)

Required if `method` is specified as `user_input`. Absolute path to the customized contact network file. The file has to be in an [adjacency list format](#).

- `model`: [*OPTIONAL] (string) `ER` or `BA` or `RP`

Required if `method` is specified as `randomly_generate`. The random graph model you want to apply for generating the contact network. The choices are ER (Erdős–Rényi network), BA (Barabási–Albert network) or RP (Random partition network that has two partitions).

- `p_ER`: [*OPTIONAL] (float)

Required if `model` is specified as `ER`. It is the parameter for generating an Erdős–Rényi network, specifying the probability of presence of an edge between two host individuals.

- `rp_size`: [*OPTIONAL] (list of ints)

Required if `model` is specified as `RP`. It is the parameter for generating a random partition network, specifying the size of each partition. As of now, we only support a two-partition model, thus this option has to be in the format of an integer list of size 2, which sums up to be the size of the host population provided by `-popsize`.

- `p_within`: [*OPTIONAL] (list of floats)

Required if `model` is specified as `RP`. It is the parameter for generating a random partition network, specifying the probability of edge creation between host individuals within the same partition. As of now, we only support a two-partition model, thus this option has to be a float list of size 2, each representing a contact probability between individuals within each partition, corresponding to the order of population specified in `rp_size`.

- `p_between`: [*OPTIONAL] (float)

Required if `model` is specified as `RP`. It is the parameter for generating a random partition network, specifying the probability of edge creation between individuals from different partitions.

- `m`: [*OPTIONAL] (int)

Required if `model` is specified as `BA`. It is the parameter for generating a Barabási–Albert network (number of contacts to attach from a new individual to existing individuals when creating this network).

- `random_seed`: [*OPTIONAL] (int)

The random seed used to generate random numbers in the module. Using the same `random_seed` ensures identical output when the module is rerun with the same parameters.

In the upcoming sections, we introduce details and examples about the generation of various host contact networks.

2.1.1 User input

If the user opts to supply their own contact network, they need to make sure the contact network file is in an [adjacency list format](#), which is a space-delimited file. Lines starting with # will be marked as comments. A typical command to run this mode is:

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize ${HOST_SIZE} \
-method user_input \
-path_network ${NETWORK_PATH}
```

Listing 2.2: NetworkGenerator user_input mode

NetworkGenerator will read the network from the user-provided file path and check its format. If the file satisfies the format requirements, the network will be rewritten into a file named `contact_network.adjlist` in the working directory. Note that *OutbreakSimulator* cannot be run without a properly formatted and named `contact_network.adjlist` in the working directory. This step is recommended even if you are providing your own contact network. However, you can modify `contact_network.adjlist` by adding/deleting new contact edges manually as well, but we recommend rerunning *NetworkGenerator* by taking the absolute path of `${WKDIR}/contact_network.adjlist` as input for `-path_network` after your manual modification to make sure it is in a proper format except for non-canonical usage.

Non-canonical usage includes using a weighted graph; by default the contact network is unweighted. Any connection between two hosts will only appear once in the file `contact_network.adjlist`, since the network is undirected and unweighted by default. In the main module of e3SIM, each connection will only be evaluated once for transmission per tick. Users can add weight to an edge by duplicating the specific node IDs in the relevant lines of `contact_network.adjlist`, so that the same contact will be evaluated multiple times per tick, imitating adding weight to an edge. Do not rerun the program; otherwise, the weight of edges will not be retained. Users should pay attention to maintaining proper formatting when making this edit.

2.1.2 Well-mixed Host population

To generate a contact network for a specified host population size and a well-mixed host population, where all hosts have the same number of expected contacts, users can utilize the Erdős–Rényi (ER) random graph model. The ER model is parameterized by `p_ER`, which represents the probability of any edge being present in the network. The degrees of any node of the generated network will follow a binomial distribution, with the expected number of edges per individual calculated as $p_ER \times \text{popsize}$.

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize ${HOST_SIZE} \
-method randomly_generate \
-model ER \
-p_ER ${PROB_ER}
```

Listing 2.3: NetworkGenerator ER network

NetworkGenerator will use the parameters provided to generate an Erdős–Rényi graph and write the network into a file named `contact_network.adjlist` in the working directory. For example, to generate a uniform host population of 10,000 hosts where each host has an expected degree of contact being 10 ($= 10000 \times 0.001$), we run:

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize 10000 \
-method randomly_generate \
-model ER \
-p_ER 0.001
```

Listing 2.4: NetworkGenerator ER network example

2.1.3 Superspreaders

Using this option will generate a host contact network using the Barabási–Albert (BA) random network model that features unbalanced connectivity, where a few potential “superspreaders” have high connectivity and the rest of the population have low connectivity, representative of a real-world social network for sexually transmitted disease, like HIV. The BA model has a single parameter, `-m`, specifying the number of edges to attach from a new node to existing nodes in a BA graph.

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize ${HOST_SIZE} \
-method randomly_generate \
-model BA \
-m ${m}
```

Listing 2.5: NetworkGenerator BA network

NetworkGenerator will use the parameters provided to generate a Barabási–Albert random network model and write the network into a file named `contact_network.adjlist` in the working directory. For examples, to generate a host population of 10,000 hosts with $m = 2$, we run:

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize 10000 \
-method randomly_generate \
-model BA \
-m 2
```

Listing 2.6: NetworkGenerator BA network example

2.1.4 Two Subpopulations

This option is designed for a host contact network with subpopulation structures, where there are two partitions in the whole graph. Each partition has their own intra-partition contact probability and there is an inter-partition contact probability for nodes between these two partitions. This subpopulation configuration can represent separate rural and urban areas, two countries, an ecological barrier in natural world, and so on. A typical command for this option is:

```
python network_generator.py \
-wkdir ${WKDIR} \
-popsize ${HOST_SIZE} \
-method randomly_generate \
-model RP \
-rp_size ${POP1_SIZE} ${POP2_SIZE} \
-p_within ${POP1_PROB} ${POP2_PROB} \
-p_between ${PROB_BETWEEN_POP1_POP2}
```

Listing 2.7: NetworkGenerator RP network

NetworkGenerator will use the parameters provided to generate a random partition graph and write down the network into a file named `contact_network.adjlist` in the working directory. For example, the following command generates a host population of 10,000 individuals, of which 6,000 hosts come from an urban area where expected connectivity for each host is as high as 12 ($= 6000 \times 0.002$), and the rest 4,000 come from a rural area where expected connectivity for each host is as low as 4 ($= 4000 \times 0.001$). Meanwhile, there are a few connections between the two areas so that hosts from one partition are expected to be connected with 3 ($= 6000 \times 0.0005$) hosts from the other partition:

```
python network_generator.py \
    -wkdir ${WKDIR} \
    -popsize 10000 \
    -method randomly_generate \
    -model RP \
    -rp_size 4000 6000 \
    -p_within 0.001 0.002 \
    -p_between 0.0005
```

Listing 2.8: NetworkGenerator RP network example

2.2 SeedGenerator

Initial sequences (a.k.a. seeds) are pathogen genomes that are planted in the host population at the start of the simulation of the main module. They can have distinct genomic sequences and different trait values such as transmissibility and drug-resistance. These trait values are directly affected by their genomic sequences (Section 2.3). If the user wants simulations to seed the main module with the reference genome(s), instead of mutated genome(s) for each [initial sequences](#) compared to the reference genome, this module can be skipped. Note that using the reference genome as the initial sequence has to be specified when running the *OutbreakSimulator* (Chapter 3). Otherwise, users can run *SeedGenerator* in random generation mode to generate mutated pathogen genomes by forward-in-time simulation, which serve as initial sequences in the main simulator. If users choose to provide their own initial sequences, it is recommended to run *SeedGenerator* in user-input mode to check the format of the user-provided genomic sequences (in VCF format).

SeedGenerator will create a new folder named `originalvcfs` under the provided working directory and store one [VCF](#) file for each initial sequence, named `seed.X.vcf` in the `originalvcfs` folder (X ranges from 0 to $\$ \{NUM_INIT_SEQ\} - 1$). Please note that any existing `originalvcfs` folder in the working directory will be overwritten when running *SeedGenerator*.

SeedGenerator also stores the genealogy of the initial sequences in the working directory if applicable. If random generation mode is being used, it is not guaranteed that the generated seeds have a single common ancestor. If there is a single common ancestor, the genealogy will be stored in a NWK format file named `seeds.nwk` in the working directory. Otherwise, *SeedGenerator* will create a folder named `seeds_phylogeny_uncoalesced` in the working directory and write a NWK file for each common ancestor for the seeds. In the NWK file(s), tip labels of the tree are integers from 0 to $\$ \{NUM_INIT_SEQ\} - 1$, corresponding to the initial sequence IDs in the `originalvcfs` folder. The `seeds.nwk` file, if present in the working directory, will be used by the post-simulation modules to generate a complete genealogy of the sampled pathogens during the main simulation. Please see more details in Chapter 3.

```
python seed_generator.py \
    -wkdir ${WKDIR} \
    -num_init_seq ${NUM_INIT_SEQ} \
    -method ${METHOD} \
    [Other params]
```

Listing 2.9: SeedGenerator Usage

We now introduce all the options:

- `wkdir: [*REQUIRED] (string)`

Absolute path to the working directory, which should be consistent for each module in one workflow.

- `num_init_seq: [*REQUIRED] (integer)`

Number of initial sequences for the main simulator, i.e. number of infected hosts at the start of the main simulation. Note that this quantity is for the main simulator, rather than the number of starting sequences for running the current module.

- **method:** [*REQUIRED] (string) `user_input` or `SLiM_burnin_WF` or `SLiM_burnin_epi`
 Method used for generating the initial sequences, which can only be one of the three choices: `user_input` or `SLiM_burnin_WF` or `SLiM_burnin_epi`. By specifying `user_input`, users are required to provide their own VCF file for the initial sequences by `-init_seq_vcf`. By specifying one of the SLiM-burnin methods, you are required to provide a reference genome by `-ref_path` and other corresponding parameters for the method you choose.
- **init_seq_vcf:** [*OPTIONAL] (string)
 Required if `method` is specified as `user_input`. Absolute path to a VCF file containing the mutation profiles of the initial sequences.
- **path_init_seq_phylogeny:** [*OPTIONAL] (string)
 Optional if `method` is specified as `user_input`, not required otherwise. Absolute path to the initial sequences' phylogeny in NWK format.
- **ref_path:** [*OPTIONAL] (string)
 Required if `method` is specified as `SLiM_burnin_WF` or `SLiM_burnin_epi`. Absolute path to the reference genome of the pathogen you want to simulate with.
- **use_subst_matrix:** [*OPTIONAL] (bool)
 Optional if `method` is specified as `SLiM_burnin_WF` or `SLiM_burnin_epi`. Whether to use a substitution probability matrix to parametrize mutation probability, default is `False`. When `False`, the substitution model will default to a Jukes–Cantor model.
- **mu:** [*OPTIONAL] (float)
 Required if `method` is specified as `SLiM_burnin_WF` or `SLiM_burnin_epi`, and that `-use_subst_matrix` is specified to be `False`. Mutation probability of the genomes during burn-in, which is the expected number of mutations per site for each genome in one `tick`, which is the parameter α for a Jukes–Cantor model (3.2.2). `mu` need not have the same value when running *OutbreakSimulator*, but note that this will cause the branch lengths of the generated phylogenies to be measured in different units, which may be misleading if directly compared. Manual rescaling of the branch length of the initial sequences' phylogeny might be of interest after running *SeedGenerator* in this case.
- **mu_matrix:** [*OPTIONAL] (str)


```
'{"A": [0, p_ac, p_ag, p_at], "C": [p_ca, 0, p_cg, p_ct],  
 "G": [p_ga, p_gc, 0, p_gt], "T": [p_ta, p_tc, p_tg, 0]}'
```

 Required if `method` is specified as `SLiM_burnin_WF` or `SLiM_burnin_epi`, and that `-use_subst_matrix` is specified to be `True`. A JSON string having the format shown above, where `p_ij` gives the probability of one site in allele i mutating to allele j in one tick, thus they have to be within the range of $[0, 1]$. Following the tradition of SLiM, $p_{ii}=0$ for $i \in \{A,C,G,T\}$, but in practice the probability for a site in allele i staying in allele i during one tick will be calculated as $1 - \sum_{j \neq i} p_{ij}$. For details of the evolution model, please refer to Section 3.2.2.
- **n_gen:** [*OPTIONAL] (int)
 Required if `method` is specified as `SLiM_burnin_WF` or `SLiM_burnin_epi`. Number of ticks/generations that will be run during the seed generation.
- **Ne:** [*OPTIONAL] (int)
 Required if `method` is specified as `SLiM_burnin_WF`. The effective population size of a Wright–Fisher model.

- `host_size`: [*OPTIONAL] (int)

Required if `method` is specified as `SLiM_burnin_epi`. Host population size, which must be consistent with the size of the network in the `contact_network.adjlist` file in the working directory provided by `-wkdir`.

- `seeded_host_id`: [*OPTIONAL] (list of ints)

Required if `method` is specified as `SLiM_burnin_epi`. ID(s) of the host(s) that will be infected with a pathogen whose genome is identical to the reference genome at the first tick of the seed generation process. The host IDs have to be chosen from 0 to `host_size` - 1.

- `S_IE_prob`: [*OPTIONAL] (float)

Required if `method` is specified as `SLiM_burnin_epi`. The probability of a successful transmission for one effective contact (an infected host and a susceptible host that are connected to each other) per tick. The default value is 0.

- `E_I_prob`: [*OPTIONAL] (float)

Required if `method` is specified as `SLiM_burnin_epi` and `-latency_prob` is also greater than 0. The probability of an exposed host becoming infected per tick. The default value is 0.

- `E_R_prob`: [*OPTIONAL] (float)

Could be supplied if `method` is specified as `SLiM_burnin_epi` and `-latency_prob` is greater than 0. The probability of an exposed host recovering per tick. The default value is 0.

- `latency_prob`: [*OPTIONAL] (float)

Could be supplied if `method` is specified as `SLiM_burnin_epi`. The probability of a susceptible host becoming exposed upon transmission per tick. The default value is 0, meaning that all susceptible hosts become infected upon a relevant transmission.

- `I_R_prob`: [*OPTIONAL] (float)

Required if `method` is specified as `SLiM_burnin_epi`. The probability of an infected host recovering per tick. The default value is 0.

- `I_E_prob`: [*OPTIONAL] (float)

Required if `method` is specified as `SLiM_burnin_epi`. The probability of an infected host deactivating (transitioning to exposed state) per tick. The default value is 0.

- `R_S_prob`: [*OPTIONAL] (float)

Required if `method` is specified as `SLiM_burnin_epi`. The probability of a recovered host losing immunity (going back to susceptible) per tick. The default value is 0, meaning that recovered hosts will not return to the susceptible state. Note that if this variable is zero, the outbreak will eventually diminish, which might cause insufficient number of seeding sequences to sample from.

- `random_seed`: [*OPTIONAL] (int)

The value that helps generate random numbers in the module. Using the same `random_seed` ensures identical output when the module is rerun with the same parameters.

2.2.1 User input

If the user wants to provide their own initial sequences' mutation information, they need to make sure the provided file is in [VCF](#) format. The `.vcf` file should contain the exact number of samples as specified by `-num_init_seq`, and sample names do not matter, as initial sequences will be renamed based on the order in the provided `.vcf` file from 0 to `NUM_INIT_SEQ` - 1. If the provided `.vcf` file has more individuals than

what is specified by `-num_init_seq`, *SeedGenerator* will take the first `NUM_INIT_SEQ` columns for the initial sequences' genetic information, and the subsequent columns will be ignored. Please note that since e3SIM only supports haploid pathogens for now, the phasing won't be informative in the user-provided `.vcf` file; i.e. 0/1 and 1/0 in the sample columns will all be rewritten as 1 during validation. To run this mode:

```
python seed_generator.py \
-wkdir ${WKDIR} \
-num_init_seq ${NUM_INIT_SEQ} \
-method user_input \
-init_seq_vcf ${PATH_TO_INIT_SEQ_VCF}
```

Listing 2.10: SeedGenerator Usage user_input

2.2.2 Burn-in using Wright-Fisher model

The user can generate initial pathogen genomes with desired genetic diversity using the option to run a forward-in-time simulation using a Wright–Fisher model with SLiM. We run the WF model with `-Ne` constant population size for `-n_gen` generations with a Jukes–Cantor substitution model. Here is a template for how to initiate the seed generation process:

```
python seed_generator.py \
-wkdir ${WKDIR} \
-num_init_seq ${NUM_INIT_SEQ} \
-method SLiM_burnin_WF \
-ref_path ${REF_PATH} \
-use_subst_matrix F \
-mu ${MU} \
-Ne ${Ne} \
-n_gen ${NUM_GENS_BURNIN}
```

Listing 2.11: SeedGenerator Usage SLiM.burnin_WF

At the last generation/tick in *SeedGenerator*, initial sequences will be sampled from the current population. *SeedGenerator* will create a new folder named `originalvcfs` in the provided working directory and write one `.vcf` file for each initial sequence in the `WKDIR/originalvcfs` directory named `seed.X.vcf` where $X \in \{0, \dots, \text{NUM_INIT_SEQ} - 1\}$. *SeedGenerator* records the phylogeny of the sequences sampled, but sometimes not all sampled seeding sequences can be traced back to a single common ancestor. If single common ancestor exists, the initial sequences' phylogeny will be written to the working directory as `seeds.nwk` in `NWK` format. If multiple ancestors exist, *SeedGenerator* will create a folder named `seeds_phylogeny_uncoalesced` in the working directory and write a `.nwk` file for each separate ancestor.

For example, if we want to run a burn-in process for COVID-19 genomes using the WF model, we download SARS-CoV-2 genome from the GISAID website into the current directory, retrieving a file named `EPI_ISL_402124.fasta`. We specify the mutation rate as 1×10^{-6} , which corresponds to the mutation rate of COVID-19 per day in real-time scale. We can run the burn-in for 3650 ticks to simulate 10 years' burn-in period in real-time scale. Let us use an effective population size of 1000 and sample 5 initial sequences:

```
python seed_generator.py \
-wkdir ${WKDIR} \
-num_init_seq 5 \
-method SLiM_burnin_WF \
-ref_path EPI_ISL_402124.fasta \
-mu 1e-6 \
-Ne 1000 \
-n_gen 3650
```

Listing 2.12: SeedGenerator Usage SLiM.burnin_WF example

2.2.3 Burn-in using epidemiological model

Sometimes a burn-in process using the epidemiological model (which can be the same model and parameters as the real simulation) is desirable. In this case, the user must run *NetworkGenerator* program before proceeding to this step. This mode can be viewed as a minimal version of the *OutbreakSimulator* with sampling only at the last generation/tick and with no genetic effects being simulated. In this example, we use a customized substitution probability matrix for the evolution model by specifying `-use_subst_matrix T`.

```
python seed_generator.py \
-wkdir ${WKDIR} \
-num_init_seq ${NUM_INIT_SEQ} \
-method SLiM_burnin_epi \
-ref_path ${REF_PATH} \
-use_subst_matrix T \
-mu_matrix '["A": [0, , ,], "C": [, 0, ,], "G": [ , , 0], "T": [ , , 0] ]' \
-n_gen ${NUM_GENS_BURNIN} \
-host_size ${HOST_SIZE} \
-seeded_host_id ${SEEDED_HOST_ID} \
-S_IE_prob ${S_IE_prob} \
-E_I_prob ${E_I_prob} \
-E_R_prob ${E_R_prob} \
-latency_prob ${latency_prob} \
-I_R_prob ${I_R_prob} \
-I_E_prob ${I_E_prob} \
-R_S_prob ${R_S_prob}
```

Listing 2.13: SeedGenerator Usage SLiM_burnin.epi

Note that the transition probabilities all have a default value of 0 and are thus not strictly required to be specified otherwise. However, not providing non-zero probabilities for some transitions can lead to undesired effects. You often want to avoid making any state an absorbing dead end since that can lead to an epidemic dying out before any sequence gets sampled. If the number of existing pathogens is less than the desired quantity of initial sequences, the program will instruct you to rerun the burn-in process or modify the parameters. The SEIR trajectory during the burn-in process is recorded and written as a compressed .csv file in the working directory provided by `-wkdir`, so users are welcome to inspect the file (named `burn_in_SEIR_trajectory.csv.gz`) to get an impression of the dynamics produced by your current rate parameter settings. If sufficient pathogens are present, *SeedGenerator* will create a new folder named `originalvcfs` under the provided working directory and write one `seed.X.vcf` file for each initial sequence.

As with the `SLiM_burnin_WF` option, *SeedGenerator* records the phylogeny of the sequences sampled, but sometimes not all sampled seed sequences can be traced back to a single common ancestor. If single common ancestor exists, the initial sequences' phylogeny will be written to the working directory as `seeds.nwk` in `NWK` format.

For example, if we want to run a burn-in process for COVID-19 genomes using an epidemiological model, we download COVID-19 genome from the NCBI website into the current directory, retrieving a file named `EPI_ISL_402124.fasta`. We want to specify a mutation probability matrix as such:

	A	C	G	T
A	–	7.16×10^{-8}	2.84×10^{-7}	5.45×10^{-8}
C	1.17×10^{-7}	–	3.51×10^{-8}	1.5×10^{-6}
G	4.32×10^{-7}	3.27×10^{-8}	–	2.32×10^{-7}
T	5.05×10^{-8}	8.54×10^{-7}	1.42×10^{-7}	–

We can run the burn-in for 3650 ticks to simulate 10 years' burn-in period in real-time scale. Say that we used *NetworkGenerator* to generate a contact network with 10000 hosts and decide that we want to start seed

simulation at individual 256. We want to specify an epidemiological model where each infected host infects each of its connections with a probability of 0.003, and all infected hosts will go through a latent stage upon infection. Since the latency period for COVID-19 is usually 6 days after infection, we choose an activation probability 0.16 ($\approx 1/6$). Since the recovery period for COVID-19 is usually 28 days after infection, we choose a recovery probability of 0.035 ($\approx 1/28$). Post-recovery immunity to COVID-19 lasts for up to 6 months but is highly protective only for the first 2 months, so we set 2 months as the expected time for losing immunity by choosing the probability of immunity loss to be 0.018 ($\approx 1/56$). We do not want to allow infected hosts to go back to latency, or let latent hosts recover. Finally, we want to sample 5 initial sequences. To realize the specification, we run the following:

```
python seed_generator.py \
    -wkdir ${WKDIR} \
    -num_init_seq 5 \
    -method SLiM_burnin_epi \
    -ref_path EPI_ISL_402124.fasta \
    -use_subst_matrix T \
    -mu_matrix '{"A": [0,7.16e-08,2.848e-07,5.45e-08], "C": [1.17e-07,0,3.51e-08,1.5e-06], "G": [4.32e-07,3.27e-08,0,2.32e-07], "T": [5.05e-08,8.54e-07,1.42e-07,0]}' \
    \
    -n_gen 3650 \
    -host_size 10000 \
    -seeded_host_id 256 \
    -S_I_E_prob 0.003 \
    -E_I_prob 0.16 \
    -E_R_prob 0 \
    -latency_prob 1 \
    -I_R_prob 0.035 \
    -I_E_prob 0 \
    -R_S_prob 0.018
```

Listing 2.14: SeedGenerator Usage SLiM_burnin.epi example

For `-seeded_host_id`, you can manually check your contact network file to find a host that you think is good, or you can utilize *HostSeedMatcher* (Section 2.4) to find a specific host that satisfies your needs.

2.3 GeneticEffectGenerator

The *GeneticEffectGenerator* module in e3SIM is designed to configure the causal genomic elements for the main simulator module. *GeneticEffectGenerator* will create `causal_gene_info.csv` in the working directory, where each row represents one causal site. The first column gives the position on the genome, and the other columns represent the effect sizes for mutations at each causal site for each trait. Effect size columns for transmissibility and drug resistance are named `transmissibility_X` or `drug_resistance_X`, where X is the 1-indexed trait ID within each trait type.

Code block 2.15 shows an example of a `causal_gene_info.csv` generated by *GeneticEffectGenerator*, where three causal sites with additive effects on three different traits are specified.

```
Sites,transmissibility_1,drug_resistance_1,drug_resistance_2
450,0.0,0.3,0.0
651,-0.2,0.0,0.0
4608,0.0,0.0,1.3
```

Listing 2.15: Example of an output file of GeneticEffectGenerator

For details on how the causal sites affect relevant event probabilities, please refer to Section 4.3. As shown in the example above, *GeneticEffectGenerator* supports more than one trait for transmissibility and/or drug resistance, like `drug_resistance_1` and `drug_resistance_2`. e3SIM operates in an epoch-based manner

(Section 4.1), so that within one epoch, only one of the traits for the same trait type could be utilized. In this example, during one epoch, users could activate drug treatment by using either `drug_resistance_1` or `drug_resistance_2`, which could differ among epochs. For details about how to specify this in the main module, please refer to Section 4.1.

The causal genomic element configuration file generated by *GeneticEffectGenerator* could come from two modes: user-defined or randomly generated. For a user-defined architecture, users are recommended to run the user-defined mode to check the format of the file they provided, which should be in the same format as Code block 2.15. To randomly generate a genetic architecture, a CSV file specifying genomic regions that contain candidate causal sites for each trait should be provided through the `-csv` option. In the candidate causal regions file, each row represents a genomic region. The first two columns specify the start and end position of the region on the genome, and the other columns specify the causal status of this region to each trait. For example, Code block 2.16 specifies three genomic regions, with the first region (position 1–50000) causal to the transmissibility trait, the second region (position 50001–100000) causal to the drug resistance trait, and the third region (position 200000–300000) causal to both traits. The causal sites for each trait will be drawn from these causal regions. Please note that though one genomic element can be causal to multiple traits, since e3SIM does not provide random generation of pleiotropic effect size structures, the drawn causal sites won't overlap across traits; i.e., one site will only be causal to at most one trait.

```
Start,End,transmissibility_1,drug_resistance_1
1,50000,1,0
50001,100000,0,1
200000,300000,1,1
```

Listing 2.16: Example of an input file of *GeneticEffectGenerator* random generation mode

In this random generation mode, *GeneticEffectGenerator* generates the genetic architecture in a 4-step process:

Step 1. Choose causal sites from the specified causal regions for each trait

For trait i , let $S_0^{(i)}$ be the trait-specific candidate set of genomic sites (the union of causal regions for trait i specified in the provided genomic region file), and $n_i = |S_0^{(i)}|$ be the cardinality of $S_0^{(i)}$. Let $c_{ij} \in \{0, 1\}$ for the causal indicator that site $j \in S_0^{(i)}$ is causal for trait i , and define the number of causal sites for trait i as $K_i = \sum_{j \in S_0^{(i)}} c_{ij}$.

We model heterogeneity in polygenicity across traits by placing a Beta–Binomial prior on K_i :

$$\pi_i \sim \text{Beta}(a_i, b_i), \quad K_i | \pi_i \sim \text{Binomial}(n_i, \pi_i).$$

To make hyperparameters interpretable, we re-parameterize the Beta prior for per-trait polygenicity by its mean $\mu_i = \mathbb{E}[\pi_i]$ and concentration ξ , such that $a_i = \mu_i \xi$ and $b_i = (1 - \mu_i) \xi$. Under this parameterization,

$$\mathbb{E}[K_i] = n_i \mu_i, \quad \text{Var}(K_i) = n_i \mu_i (1 - \mu_i) \frac{\xi + n_i}{\xi + 1}.$$

Users specify an expected fraction of causal sites μ_i with default at 0.003 through option `-site_frac`, and an optional prior strength ξ controlling dispersion around μ_i through option `-site_disp`. We default to a moderately informative choice $\xi = 100$.

Given K_i , causal sites are sampled uniformly without replacement from the available candidate set for that trait. When more than one trait is simulated, to avoid spurious pleiotropy, we enforce trait exclusivity so that each site can be causal for at most one trait by requiring $\sum_j c_{ij} \leq 1$ for all genomic sites j . We process traits in a random permutation, and then iterate over the traits in that order to minimize order effects. For trait i , define the available candidate site set $A^{(i)} = S_0^{(i)} \setminus \bigcup_{\ell < i} S^{(\ell)}$ and draw $S^{(i)} \subseteq A^{(i)}$ uniformly without replacement with $|S^{(i)}| = K_i$ and set $c_{ij} = 1$ for $j \in S^{(i)}$ and 0 otherwise. If $|A^{(i)}| < K_i$, we set $S^{(i)} = A^{(i)}$ (thus effectively capping $K_i^* = |A^{(i)}|$) and issue a warning.

Step 2. Draw non-zero effect sizes for each causal site

Conditional on $c_{ij} = 1$, we draw q_{ij} on the link scale from one of the following parametric families based on user's input:

1. Normal (point-normal): $q_{ij} \sim \mathcal{N}(0, \tau_i^2)$ with sparsity handled in Step 1. Users can specify the variance of the point normal distribution τ_i^2 for each trait via option `-taus`, default to 1.0.
2. Laplace (Bayesian lasso): $q_{ij} \sim \text{Laplace}(0, \eta_i)$ (heavy-tailed with shrinkage near zero). Users can specify the scale of the Laplace distribution η_i for each trait via option `-bs`, default to 1.0.
3. Student-t: $q_{ij} \sim t_\nu(0, s_i)$ (heavy-tailed to allow a few large effects). Users can specify the scale s_i for each trait (default = 1.0) via option `-s` and a degree of freedom value ν across all traits (default = 3) via option `-nv`.

We require $\eta_i > 0$, $s_i > 0$, and recommend $\nu > 2$ for a finite slab variance $\text{Var}(q_{ij}) = s_i^2 \nu / (\nu - 2)$.

Step 3. Standardize genetic values and effect sizes

If the calibration option is activated (`-calibration T`), `GeneticEffectGenerator` standardizes the genetic architecture effect sizes based on the genetic values of the seeding sequences. This option defaults to True in random generation mode and to False in user-input mode. Given additive link-scale genetic values $G_i = \sum_{j \in S^{(i)}} q_{ij} x_j$ where $x_j \in \{0, 1\}$ indicates the presence of the effect allele (any alternative alleles different from the reference allele are defined as effect allele in e3SIM) at site j in the haploid pathogen genome for seeds and trait i , we standardize G_i to mean zero and a user-specified variance (specified via option `var_target`, default to 1.0) on the link scale.

We adopt allele-frequency centering, which is standard in quantitative genetics. Let m be the number of seed genomes, and let $G_{i,k}$ be the raw link-scale genetic value for trait i in seed genome $k = 1, \dots, m$. Let $x_{j,k} \in \{0, 1\}$ denote the derived allele indicator at site j in seed k . Define the allele frequency of mutation j in the seeding population and the centered genotype as:

$$\hat{p}_j = \frac{1}{m} \sum_{k=1}^m x_{j,k}, \quad z_{j,k} = x_{j,k} - \hat{p}_j.$$

The centered genetic value is then

$$G'_{i,k} = \sum_{j \in S^{(i)}} q_{ij} z_{j,k}.$$

Because $\mathbb{E}[z_{j,k}] = 0$ by construction, $\frac{1}{m} \sum_{k=1}^m G'_{i,k} = 0$ for any fixed effect vector (q_{ij}) .

The across-seed dispersion of genetic values is then fixed by moment matching. We first compute the empirical mean and variance of $G_{i,k}$ across seeds:

$$\hat{\mu}_i = \frac{1}{m} \sum_{k=1}^m G'_{i,k} = 0, \quad \hat{V}_i = \frac{1}{m-1} \sum_{k=1}^m (G'_{i,k} - \hat{\mu}_i)^2.$$

Given a user-chosen target variance $V_{i,\text{target}} > 0$, define the standardized trait value and scaled effect sizes as:

$$\tilde{G}_{i,k} = r_i (G'_{i,k} - \hat{\mu}_i), \quad \tilde{q}_{ij} = r_i q_{ij}, \quad \text{where } r_i = \sqrt{\frac{V_{i,\text{target}}}{\hat{V}_i}},$$

By construction, $\frac{1}{m} \sum_{k=1}^m \tilde{G}_{i,k} = 0$ and $\frac{1}{m-1} \sum_{k=1}^m \tilde{G}_{i,k}^2 = V_{i,\text{target}}$. If seeds are monomorphic at all causal sites for trait i , $\hat{V}_i = 0$, so empirical per-SD (standard deviation) calibration is undefined. We therefore

standardize using the expectation of additive genetic variance under a Beta prior on allele frequencies p_j rather than the empirical seed variance:

$$V_i^* = \sum_{j \in S^{(i)}} q_{ij}^2 \mathbb{E}[p_j(1 - p_j)].$$

By default, we assume loci are in linkage equilibrium and use a uniform Beta prior to set a reference scale, for which $\mathbb{E}[p_j(1 - p_j)] = 1/6$. We then rescale effects by $\tilde{q}_{ij} = r_i q_{ij}$ with:

$$r_i = \sqrt{\frac{V_{i,\text{target}}}{V_i^*}},$$

giving a fixed reference unit $\text{SD}_i = \sqrt{V_{i,\text{target}}}$.

The standardized (or unstandardized if `-calibration F`) effect sizes of causal sites will be stored in the working directory as the genetic architecture file used in the main simulation, named `causal_gene_info.csv`, with the format shown in Code block 2.15. Using this final genetic architecture, the trait values for all seeding sequences, based on their mutation profiles will be calculated and stored as a file named `seeds_trait_values.csv` in the working directory, which could be inspected by the user manually before running the next module. If using the GUI, this trait information will be shown in the HostSeedMatcher tab for users to inspect. The format is as follows:

```
Seed_ID,transmissibility_1,drug_resistance_1
0,0.72,0.0
1,0.2,-0.34
2,0.72,-0.8
```

Listing 2.17: Example of seeds_trait_values.csv format

Step 4. Calibrate the link scale slope

If the calibration option for link scale slope is activated (`calibration_link T`), `GeneticEffectGenerator` calibrates the link scale slope α by specifying the per-SD effect multipliers via option (`-Rs`).

- Logit link: user supplies per-SD odds ratio $R_{i,\text{OR}} > 0$ for trait i ; set:

$$\alpha_i = \frac{\log R_{i,\text{OR}}}{\text{SD}_i}.$$

- Cloglog link transmissibility: user supplies per-SD transmission hazard ratio $R_{\text{trans},\text{HR}} > 0$; set:

$$\alpha_{\text{trans}} = \frac{\log R_{\text{trans},\text{HR}}}{\text{SD}_{\text{trans}}}.$$

- Cloglog link drug resistance: user supplies per-SD clearance hazard ratio $R_{\text{drug},\text{clr}} > 0$; set:

$$\alpha_{\text{drug}} = -\frac{\log R_{\text{drug},\text{clr}}}{\text{SD}_{\text{drug}}}.$$

A typical command to run `GeneticEffectGenerator` is:

```
python genetic_effect_generator.py \
    -wkdir ${WKDIR} \
    -method ${METHOD} \
    [Other params]
```

Listing 2.18: GeneticEffectGenerator Usage

We now introduce all the options:

- **wkdir:** [*REQUIRED] (string)
Absolute path to the working directory, which should be the same for all modules.
- **method:** [*REQUIRED] (string) `user_input` or `randomly_generate`
The method of setting the genetic architecture, which can only be `user_input` or `randomly_generate`. By specifying `user_input`, users are required to provide their own effect size file via the option `-csv`, that has the format shown in code block 2.15. In this mode, *GeneticEffectGenerator* will not perform the random generation of causal sites and effect sizes, but can do effect size calibration if required. It will mandatorily check the format of provided files and calculate the trait values of the seeding sequences. By specifying `randomly_generate`, you are required to provide the relevant parameters and options to randomly generate causal sites and associated effect sizes from a candidate genomic region file (See the format from Code block 2.16) through the option `-csv`.
- **trait_n:** [*REQUIRED] (JSON string) `'{"transmissibility": x, "drug_resistance": y}'`
A JSON string specifying the number of transmissibility and drug resistance traits. x and y are integer numbers specifying the number of traits for each trait type.
- **num_init_seq:** [*REQUIRED] (integer)
Number of initial sequences for the main simulator, i.e. number of infected hosts at the start of the main simulation. This quantity will be used when calculating the trait values for each initial seeding sequences, and should be the same as specified when running the previous pre-simulation programs. If more sequences than specified are detected in the working directory, only the first `num_init_seq` sequences will be used. The program will issue an error and terminate if fewer sequences than the specified number are detected.
- **csv:** [*REQUIRED] (string)
Absolute path to the user-provided effect size file when `method` is specified as `user_input` (See the format from Code block 2.15); Absolute path to the user-provided candidate causal regions file when `method` is specified as `randomly_generate` (See the format from Code block 2.16).
- **site_frac:** [*OPTIONAL] (list of floats)
Could be specified if `method` is specified as `randomly_generate`. The expected fraction of causal sites to be sampled for each trait from the candidate causal regions. Should be a float list with the same length of the number of traits in total. Each element of the list should be a float within the range (0,1]. Defaults to 0.003. See the details in the Step 1 of *GeneticEffectGenerator*, μ_i in the section.
- **site_disp:** [*OPTIONAL] (float)
Could be specified if `method` is specified as `randomly_generate`. Modeling the dispersion of the expected fraction of causal sites in the candidate causal regions. Defaults to 100. See the details in the Step 1 of *GeneticEffectGenerator*, ξ in the section.
- **func:** [*OPTIONAL] (string) `n` or `l` or `st`
Required if `method` is specified as `randomly_generate`. The distribution to sample effect sizes from given selected causal sites. By specifying `n`, the distribution will be a normal distribution, where users have to provide the variance of each trait's causal effect sizes via `-taus` option. By specifying `l`, the distribution will be a laplace distribution, where users have to provide the scales of each trait's causal effect sizes via `-bs` option. By specifying `st`, the distribution will be a Student's t distribution, where users have to provide the scales of each trait's causal effect sizes via `-s` option, and optionally the degree of freedom via `-nv` option.

- `taus`: [*OPTIONAL] (list of floats)

Could be specified if `method` is specified as `randomly_generate` and `func` is specified as `n`. The variance parameter of the normal distribution for the effect sizes. Should be a float list with the same length of the number of traits in total. Each element of the list should be a number within the range $(0, \infty)$. Default is 1.

- `bs`: [*OPTIONAL] (list of floats)

Could be specified if `method` is specified as `randomly_generate` and `func` is specified as `l`. The scale parameter of the Laplace distribution for the effect sizes. Should be a float list with the same length of the number of traits in total. Each element of the list should be a number within the range $(0, \infty)$. Default is 1.

- `s`: [*OPTIONAL] (list of floats)

Could be specified if `method` is specified as `randomly_generate` and `func` is specified as `st`. The scale parameter of the Student's t distribution for the effect sizes. Should be a float list with the same length of the number of traits in total. Each element of the list should be a number within the range $(0, \infty)$. Default is 1.

- `nv`: [*OPTIONAL] (float)

Could be specified if `method` is specified as `randomly_generate` and `func` is specified as `st`. The degree of freedom parameter of the Student's t distribution for the effect sizes across all traits. Should be a positive float. Default is 3. It's recommended to have the value bigger than 2.

- `calibration`: [*OPTIONAL] (bool)

Optional. Whether to calibrate the sampled effect sizes in order to match the initial seeding population's trait values to the variance specified by the users. Default is `False` when `method` is specified as `user_input`, and default is `True` when `method` is specified as `randomly_generate`. If specified as `True`, the users have to specify the target trait variance via `var_target` option.

- `var_target`: [*OPTIONAL] (list of floats)

Could be specified if `calibration` is specified as `True`. The target variance of the trait values in the seeding populations, will be used for calibrating the effect sizes. Default is 1.

- `calibration_link`: [*OPTIONAL] (bool)

Optional. Whether to calibrate and provide a link slope α for each trait, according to the seeding population trait variances. Default is `False`. If specified as `True`, the users have to specify the link type of the link function via `link` option, and the odds ratio for the transmission/survival per SD of trait values under logit, or the hazard ratio per SD under cloglog via `Rs` option.

- `link`: [*OPTIONAL] (string) `logit` or `cloglog`

Required if `calibration_link` is specified as `True`. The link function to map the trait values to event type. If specified as `logit`, the option `Rs` will represent the odds ratio for the transmission/survival per SD of trait values. If specified as `cloglog`, the option `Rs` will represent hazard ratio per SD of the trait values.

- `Rs`: [*OPTIONAL] (list of floats)

Required if `calibration` is specified as `True`. The odds ratio for the transmission/survival per SD of trait values or the hazard ratio per SD of the trait values. Should be a positive float list with the same length of the number of traits in total. Default is 1.5 for all traits with logit link or transmissibility trait with cloglog link; default is 0.67 (1/1.5) for drug resistance trait with cloglog link.

- `random_seed`: [*OPTIONAL] (int)

The value that helps generate random numbers in the module. Using the same `random_seed` ensures identical output when the module is rerun with the same parameters.

2.3.1 User input

By specifying `-method user_input`, user could provide a customized genetic architecture file. *GeneticEffectGenerator* will read the user-provided effect size file, check the format and entry of the file, and then rewrite it to `$WKDIR/causal_gene_info.csv`. *GeneticEffectGenerator* also calculates the trait values for each seed based on the genetic architecture file provided, and stores the trait values in `$WKDIR/seeds_trait_values.csv`. The provided genetic architecture file has to be in the format shown in Code block 2.15 with correct format of column.

```
python genetic_effect_generator.py \
    -wkdir ${WKDIR} \
    -method user_input \
    -trait_n
    '{"transmissibility": ${NUM_SET_TRANSMISSIBILITY}, "drug_resistance": ${NUM_SET_DRUGRESIST}}' \
    -num_init_seq ${NUM_SEED} \
    -csv ${EFF_SIZE_CSPATH}
```

Listing 2.19: GeneticEffectGenerator usage in the user input mode

2.3.2 Generate from candidate regions

The user can also randomly generate an effect size file using a candidate region file as input. A candidate genomic region file has the format like Code block 2.16.

The genetic architecture for one trait is a set of causal sites along with their effect sizes selected for the trait. Several traits for one trait type could be specified. When executing *GeneticEffectGenerator* in the random generation mode, users are required to specify numbers of traits for the two different trait types: `transmissibility` and `drug resistance` according to the needs in the main simulator. For example, two traits for drug resistance with different causal genomic regions and effect sizes could be generated, representing resistance for two different types of treatments. In *OutbreakSimulator*, the user could specify two epochs with those two different treatments by activating different drug resistance traits for different epochs.

GeneticEffectGenerator contains two main steps to randomly generate the genetic architecture, as discussed earlier in the section. First, the causal sites are drawn from the candidate regions in the csv file by independent draws. Given the causal sites, effect sizes for each site will be drawn from a normal distribution (`-func n`), a Laplace distribution (`-func l`) or a Student's t distribution (`-func st`). All the distributions assume the mean value being 0, and the users specify the dispersion. For normal distribution, users specify the variance. For Laplace or Student's t distribution, users specify the scale.

Moreover, if `-calibration T` is specified, the effect sizes will be rescaled according to the existing mutations of the seeding populations. The trait values of seeding sequences will be calculated and the variance of the seeding sequences population will be anchored to rescale the effect sizes such that rescaled trait variance matches a user-specified target variance (`-var_target`). To run genetic effect generators without effect size calibration:

```
python genetic_effect_generator.py \
    -wkdir ${WKDIR} \
    -method randomly_generate \
    -num_init_seq ${NUM_SEED} \
    -csv ${CSV_PATH} \
    -trait_n
```

```
'{"transmissibility": ${NUM_SET_TRANSMISSIBILITY}, "drug_resistance": ${NUM_SET_DRUGRESIST}}' \
-site_disp ${DISPERSION} \
-site_frac ${FRAC_CAUSALTRAIT_1} ... ${FRAC_CAUSALTRAIT_N} \
-func n -taus ${VAR_EFFSIZE_TRAIT_1} ... ${VAR_EFFSIZE_TRAIT_N}
```

Listing 2.20: GeneticEffectGenerator Usage randomly generate

If the user would like to add calibration of the effect sizes they will have to specify a few more options:

```
... \
-calibration T \
-var_target ${VAR_TARGET_TRAIT_1} ... ${VAR_TARGET_TRAIT_N}
```

Listing 2.21: Calibration of effect sizes

Except for the generation of genetic architecture, `GeneticEffectGenerator` can also be used to calibrate the link scale slope according to the seeding population trait values. This action can be activated by specifying the following options:

```
... \
-calibration_link T \
-link logit \
-Rs ${R_TRAIT_1} ... ${R_TRAIT_N}
```

Listing 2.22: Calibration of the link scale slope

For example, now we want to generate the genetic architecture for simulating a *Mtb* outbreak. *Mtb* has a 4.4 megabase genome, among which we would like to select from the candidate regions specified in Code block 2.16, and the file is named `candidate_regions.csv`. We want to have two sets of drug-resistance traits, so that we can simulate two different treatment stages later. We want to generate the effect sizes for these traits from normal distributions. For the drug resistance traits, we want to generate effect sizes from normal distributions $\mathcal{N}(0, 2)$ and $\mathcal{N}(0, 3)$, respectively. For the first drug resistance trait, we want to select an expected number of 0.1% sites from the candidate regions, while for the second drug resistance trait, we would like the expected fraction being 0.2%. We would like a slightly more dispersed prior than the default value, so we choose $\zeta = 50$. In this case, we have already generated 10 seeding sequences from `SeedGenerator`. We want to calibrate our effect sizes such that the variance of drug-resistance traits are 2 and 3, respectively, among the initial sequences. We then execute:

```
python genetic_effect_generator.py \
-wkdir ${WKDIR} \
-num_init_seq 10 \
-method randomly_generate \
-csv candidate_regions.csv \
-trait_n '[{"transmissibility": 0, "drug_resistance": 2}]' \
-site_frac 0.001 0.002 \
-site_disp 50 \
-func n \
-taus 3 2 1 \
-calibration_link T \
-Rs 2 3
```

Listing 2.23: GeneticEffectGenerator Usage randomly generate example

The generated causal gene information file can be inspected and manually modified by users as well. It is recommended to rerun by `user_input` mode after modifications, so that trait values for each seed can be recalculated and centered. In the random generation mode of `GeneticEffectGenerator`, it is prevented that any causal site has non-zero effect sizes for more than one trait, essentially preventing the program from generating **pleiotropic** genetic architecture. However, pleiotropy can be supported in the user-input mode.

For example, the user can define a site where alternative alleles induce lower transmissibility and higher drug-resistance at the same time.

Please note that e3SIM is not intended to be a comprehensive quantitative trait simulator. Users who require biologically calibrated genetic architectures should either derive them from empirical studies or generate them with dedicated architecture/phenotype simulator.

2.4 HostSeedMatcher

After configuring the genetic architectures of the pathogen genomes, the last step before running the main simulator module is to determine the start of the simulation, which includes determining which host(s) are the patient zero(s) of the simulated outbreak and which seeding sequence each patient zero is infected with. A typical command to run *HostSeedMatcher* is:

```
python seed_host_matcher.py \
    -wkdir ${WKDIR} \
    -method ${METHOD} \
    -num_init_seq ${NUMBER_OF_INITIAL_SEQUENCES} \
    [Other params]
```

Listing 2.24: HostSeedMatcher Usage

Upon successful execution of this module, one csv file named `seed_host_match.csv` is produced in the working directory. It has two columns named `host_id` and `seed`, and each row represents one seed–host match. All seeds must be matched to one host, and the rows are ordered by host ID. One example of the output file is:

```
seed,host_id
4,1188
3,1290
1,4059
0,4446
2,7100
```

Listing 2.25: Example of an output file of HostSeedMatcher

Five seeding sequences are provided in this file, each of which is matched to one of the hosts. The seeds are ordered as 4, 3, 1, 0, 2 due to the ID of the hosts they are matched to. The host IDs that are smaller appear earlier, which is required for the correct execution of the main simulator module.

The available options for running *HostSeedMatcher* are as follows:

- `wkdir: [*REQUIRED] (string)`

Absolute path to the working directory, which should be consistent with the directory used for running previous modules. It is assumed that *NetworkGenerator* has been run, so that `contact_network.adjlist` file is in this directory.

- `method: [*REQUIRED] (string) user_input or randomly_generate`

The method used for matching, which can only be `user_input` or `randomly_generate`. When specifying `user_input`, users are required to provide their own matching file using the `-path_matching` option. On the other hand, specifying `randomly_generate` necessitates providing a match scheme and matching parameters. (`-match_scheme` and `-match_scheme_param`).

- `num_init_seq: [*REQUIRED] (int)`

Total number of seeding sequences to be matched to the hosts.

- `path_matching`: [*OPTIONAL] (string)

Required when `method` is set to `user_input`. Absolute path to the user-provided matching file, which should have the format shown in Code block 2.25.

- `match_scheme`: [*OPTIONAL] (JSON string) `'{"0": M0, "1": M1 ...}'`

Required when `method` is set to `randomly_generate`. A dictionary specifying the matching scheme for each initial sequence, having the format shown above. The keys are the IDs of the seeds, and the values (`M0, M1...`) are the matching scheme for each seeding sequence, respectively. The values (`M0, M1...`) should be one of `["ranking", "percentile", "random"]`. If all seed sequences are to be matched randomly, the dictionary can be replaced by just a single `"random"`. For seeding sequences that were not specified with a matching scheme, `"random"` will be the default.

- `match_scheme_param`: [*OPTIONAL] (JSON string) `'{"0": P0, "1": P1 ...}'`

Required when `method` is set to `randomly_generate`. A dictionary containing parameters for the matching scheme of each seeding sequences. For example, if for seeding sequence 0, `"ranking"` is specified in `-match_scheme`, then `P0` here should be an integer number specifying the rank of the host. For detailed introduction, please refer to the sections below for the format of parameters corresponding to each matching method.

- `random_seed`: [*OPTIONAL] (int)

The value that help generate random numbers in the module. Using the same `random_seed` ensures identical output when the module is rerun with the same parameters.

2.4.1 User-provided matching file input

If users would like to match host(s) with exact ID(s) to the seeding sequences, one customized seed-host matching file could be provided and checked by `HostSeedMatcher` module in the `user_input` mode. The provided matching file has to be in the specific format described by Code block 2.25. An example for running `HostSeedMatcher` in this mode is:

```
python seed_host_matcher.py \
    -wkdir ${WKDIR} \
    -method user_input \
    -num_init_seq 10 \
    -path_matching ${PATH_TO_MATCHING_CSV_FILE}
```

Listing 2.26: HostSeedMatcher user input example

2.4.2 Generate matching based on user-specified methods & parameters

e3SIM provides three different matching schemes: `ranking`, `percentile` and `random`. Different matching scheme can be chosen for each seeding sequence, specified by the `-match_scheme` option. Separate matching scheme parameters have to be provided for each seeding sequence. All three schemes are explained below with an example:

Ranking Method ('ranking')

- This scheme assigns seeds to specific hosts based on the host's contact degree ranking. The required parameter is one integer number denoting the host ranking. For example, the host with the highest number of contacts receives the first seed, the second-highest receives the second seed, and so on. Each seed can be assigned to a host with any rank.

- Lower ranks (smaller numbers) correspond to hosts with higher connectivity. For example, rank 1 corresponds to the host with the highest connectivity in the contact network.
- EXAMPLE: To assign seeding sequence 1 to a host that ranks 100 in connectivity: `-match_scheme '{"...": "ranking", ...}', -match_scheme_param '{"...": {"1": 100, ...}}`

Percentile Method ('percentile')

- This scheme assigns seeds to hosts within a connectivity range, specified by percentiles of the host contact degree distribution. The required parameter is an integer list of length two, specifying one percentile range to choose from. Thus the integer numbers in the list have to be within the closed interval [0, 100], with the first number smaller than the second number.
- All hosts are assigned to different percentiles based on the contact degree distribution. By specifying a percentile range, one host within that range is uniformly drawn to be the matched host.
- EXAMPLE: To assign seeding sequence 1 to a host whose connectivity is in the top 25%: `-match_scheme '{"...": "percentile", ...}', -match_scheme_param '{"...": {"1": [0, 25], ...}}`

Random Method ('random')

- This scheme randomly selects one host from the available host lists. No parameter is required for this scheme. It is also the default method for all seeding sequences. For each seeding sequence, if no specific matching scheme is provided, it is matched by this scheme.
- EXAMPLE: If users want to match all seeding sequences randomly, they do not need to provide `-match_scheme` and `-match_scheme_param` options.

We hereby show a practical example for executing HostSeeder: Consider the following scenario that matches 3 seeding sequences by three different schemes. We want to match the first sequence to a host completely at random. We want to match the second sequence to the host possessing the highest number of contacts within the network. Finally, we want to match the third sequence to a host that is less connected than the median value. To achieve this customized instance of matching, the following command would be executed:

```
python seed_host_matcher.py \
-wkdir ${WKDIR} \
-num_init_seq 3 \
-method randomly_generate \
-match_scheme '{"0": "random", "1": "ranking", "2": "percentile"}' \
-match_scheme_param '{"1": 1, "2": [50, 100]}'
```

Listing 2.27: HostSeeder user-specified matching method(s) example

Part II

Simulator

Chapter 3

Overview of the usage

Contents

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3.1 Run OutbreakSimulator

In this chapter, we introduce the main simulator module of e3SIM: *OutbreakSimulator*. To run this module, users should ensure that they have already run the necessary pre-simulation modules. *OutbreakSimulator* requires several files in a specific format to be present **in the specified working directory** (e.g., \${WKDIR}); these files are generated by the pre-simulation modules. These files are listed below:

- `contact_network.adjlist`: Required. Generated by *NetworkGenerator*, an adjacency list specifying the contact network of the host population (Section 2.1).
- `originalvcfs`: Required if the user would like to use seeding sequences different from the reference genome. Generated by *SeedGenerator*; a folder where the VCF files for all seeding sequences are located. The VCF files are named `seed.X.vcf`, where X is the ID of the seeding sequences starting from 0 (Section 2.2).
- `seed_host_match.csv`: Required. Generated by *HostSeedMatcher*, a CSV file specifying the matching of seeding sequences to the hosts (Section 2.4).
- `seeds.nwk`: Optional. Generated by *SeedGenerator* (Section 2.2), a NWK file specifying the genealogy of the seeding sequences. If the file exists, in the post-simulation module, *OutbreakSimulator* could generate the full pathogen genealogy by binding the genealogy of the progeny of each seeding sequence to the genealogy of the seeding sequences.
- `causal_gene_info.csv`: Required if the user would like to incorporate the coupling of evolutionary process and epidemiological processes in the simulation. Generated by *GeneticEffectGenerator*, a CSV file specifying the positions of causal sites and their effect sizes for modeled traits (Section 2.3).
- A configuration file in JSON format: Required. If the user used the GUI to perform the pre-simulation modules, the configuration file will be generated in the working directory. Otherwise, users are required to provide a customized configuration file for *OutbreakSimulator*, and the template configuration file is available in our repository.

To execute `OutbreakSimulator`, only one option is required, which is the absolute path to the configuration file that specifies all the parameters of the simulation. An example to run `OutbreakSimulator` is:

```
python outbreak_simulator.py \
    -config ${YOUR_SIM_CONFIG_PATH}
```

Listing 3.1: OutbreakSimulator usage

- `config` [*REQUIRED] (string)

Absolute path to the configuration file for running *OutbreakSimulator* in a specific format.

We will explain the structure and contents of the configuration file in this chapter. However, it is recommended to go through Chapter 4, which breaks down the components of `OutbreakSimulator` and explains how the simulation works in detail, to fully understand the meaning of the parameters. In Chapter 5, we will walk through some realistic examples of how to specify and simulate your own model.

3.2 Configuration file

There are several sections in the configuration file: "BasicRunConfiguration", "EvolutionModel", "SeedsConfiguration", "GenomeElement", "NetworkModelParameters", "EpidemiologyModel" and "Postprocessing_options". In the following paragraphs, we introduce the format and meaning for all options within the configuration file in detail.

3.2.1 BasicRunConfiguration

This section specifies basic configurations to execute the simulator.

- `"cwd": (string)`

Absolute path to the working directory. This should be the same path used to run all the pre-simulation programs, and the necessary files are expected to be present in this directory.

- `"n_replicates": (int)`

The number of replicates to run with the configuration specified in this file.

(`OutbreakSimulator` will create subdirectories for the output of each replicate in the working directory specified by `"cwd"`).

3.2.2 EvolutionModel

This section specifies the evolution model used in the simulation.

- `"n_generation": (int)`

The number of ticks the user wants the simulation to run.

- `"subst_model_parameterization": (string) mut_rate_matrix or mut_rate`

How to parameterize the molecular evolution model. If `mut_rate` is specified, "`mut_rate`" needs to be specified in the configuration file. If `mut_rate_matrix` is specified, "`mut_rate_matrix`" option needs to be specified in the configuration file.

- `"mut_rate": (float)`

Needed to be specified when "`subst_model_parameterization`" is set to `mut_rate`. This is the per-tick mutation probability for any site in the pathogen genome. Under the "`mut_rate`" option, `OutbreakSimulator`

uses the Jukes–Cantor model in SLiM; thus, the probability of one site mutating into each of the alternative allele (for example, A to C) per tick is `mut_rate`/3, and the overall mutation probability per site per tick is `mut_rate`.

- "mut_rate_matrix": (list)

```
[[0,p_AC,p_AG,p_AT],[p_CA,0,p(CG,p_CT],[p_GA,p_GC,0,p_GT],[p_TA,p_TC,p_TG,0]]]
```

Needed to be specified when "subst_model_parameterization" is set to be `mut_rate_matrix`. A numerical list of length 4 showing the mutation probability matrix to be used in the simulation. The four sublists in the list represent the per-tick mutation probabilities for a site that currently has allele A, C, G, or T. `p_uv` represents the probability of allele u mutating to allele v per tick. All probabilities should be numerical values in the closed interval [0, 1], and the sum of the probabilities in each sublist should not exceed 1.

- "within_host_reproduction": (bool) `true` or `false`

Whether to activate within-host reproduction. If activated, pathogens are permitted to reproduce within the same host with a probability specified by "within_host_reproduction_rate".

- "within_host_reproduction_rate": (float)

Per-tick probability of reproducing within-host for each existing pathogen genome. This parameter is applied only when "within_host_reproduction" is set to True.

- "cap_withinhost": (int)

The maximum number of pathogens that are allowed to exist in one host. i.e., if the cap is reached for a host, then no within-host reproduction will occur in that host, and the host cannot be an infectee in a transmission event. This parameter defaults to 1. However, if within-host reproduction and/or superinfection is set to true, it is recommended to set a different value for "cap_withinhost".

3.2.3 SeedsConfiguration

This section specifies the initial sequences.

- "seed_size": (int)

Number of seed pathogen sequences. Should be the same as what is used for *SeedGenerator* and *Host-SeedMatcher*. For example, the `originalvcfs` folder in the working directory is expected to contain the same number of VCF files (if applicable).

- "use_reference": (bool) `true` or `false`

Whether to use the reference genome as the seeding sequence(s). If `true`, then it is not required to run *SeedGenerator* in the pre-simulation modules.

3.2.4 GenomeElement

This section specifies the causal genomic element settings that will be used in the simulation.

- "use_genetic_model": (bool) `true` or `false`

Whether to allow mutation profiles to affect trait values such as transmissibility and drug resistance. If `True`, then it is required to run *GeneticEffectGenerator* in the pre-simulation modules.

- "ref_path": (string)

Absolute path to the reference genome in FASTA format. It should be the same as the one used for *SeedGenerator* (if applicable). The FASTA file should contain only one sequence and should be composed only of the four alleles A, C, G, and T.

- "traits_num": (JSON object) {"transmissibility": x, "drug_resistance": y}
A JSON object specifying the number of traits (the genetic architectures) for 'transmissibility' and 'drug_resistance', which should be the same as what is used in *GeneticEffectGenerator*'s -trait_n option. e.g., "traits_num": {"transmissibility": 1, "drug_resistance": 2} represents 1 set for transmissibility and 2 sets for drug-resistance.
- "trait_prob_link":
This subsection specifies the link function of the genetic architecture, i.e., how to map genetic values to event probabilities. Please see more details about the genetic architecture and link function in Section 4.3.
 - "link": (string) logit or cloglog
The link function that will be used to map trait values to the probability scale. If "logit" is specified here, then "alpha_trans" and "alpha_drug" under the "logit" section has to be specified, and OutbreakSimulator will use the logit link function (Eq. 4.2). If "cloglog" is specified here, then "alpha_trans" and "alpha_drug" under the "cloglog" section has to be specified, and OutbreakSimulator will use the complementary log-log link function (Eq. 4.3).
 - "logit" or "cloglog": This subsection specifies the link scale slope for each trait. Only one of the sections needs to be filled in depending on which link function type is used. The link function type stays the same across trait types and simulation epochs.
 - * "alpha_trans": (list of floats)
The link scale slope α_{trans} for each transmissibility trait. Has to be a list with the same length as the number of transmissibility traits specified in traits_num.
 - * "alpha_drug": (list of floats)
The link scale slope α_{drug} for each drug-resistance trait. Has to be a list with the same length as the number of drug-resistance traits specified in traits_num.

3.2.5 NetworkModelParameters

This section specifies information about the host population.

- "host_size": (int)
host population size, which should be the same as what was specified in *NetworkGenerator*.

3.2.6 EpidemiologyModel

This section specifies the epidemiological model, which includes the compartmental model and the transition probabilities between compartments.

- "slim_replicate_seed_file_path": (string)
Absolute path to a .csv file with a single column named random_number_seed that stores the random number generator ("seeds" in computational science generally; different from the seeding sequences in e3SIM) for each replicate. If not specified, the random number generator will be chosen by SLiM and can be checked afterwards in the log file of SLiM.
- "model": (string) SIR or SEIR
The compartmental model that will be applied. The user can choose from "SIR" or "SEIR". See Section 4.4 for the permitted compartments and transitions. The transitions involving the exposed state will be disabled if "SIR" is chosen.

- "epoch_changing":

This subsection specifies the `epoch` setting of the simulation. Ticks, the time units in e3SIM, are organized into *epochs*. Each epoch comprises a sequence of consecutive ticks. Within each epoch, one set of epidemiological parameters and genetic architecture is used. Please see more details about epochs in Section 4.1.

- "n_epoch": (int)

Number of epochs used in the simulation. By specifying this parameter, all configurations in "genetic_architecture" and "transition_prob" section will be lists with the length of `n_epoch`, as one set of parameters needs to be specified for each epoch.

- "epoch_changing_generation": (list of ints)

The `tick`(s) at which the simulation switches to the next epoch and its corresponding set of parameters. It should be an integer list with length `n_epoch` - 1. For example, if "n_epoch": 3 is specified, then you should specify "epoch_changing_generation": [t1, t2], where $t1, t2 \in \{1, \dots, n_generation\}$ ("n_generation" was specified in Section 3.2.2).

- "genetic_architecture":

This subsection specifies whether to invoke genetic architecture, and which trait to use in each `epoch`. Each parameter listed in this subsection should take the format of a numerical list whose length is the same as `n_epoch`.

- "transmissibility": (list of ints)

An integer list that specifies which transmissibility trait should be used for each epoch. Should be a list of length `n_epoch`, and each element is an integer representing the trait ID. For example, if "traits_num" is specified to be '{"transmissibility": 2, "drug_resistance": 3}', meaning that there are 2 traits for transmissibility, then here each element in "transmissibility" should be one of {0, 1, 2}, where 0 represents not using genetic effect on transmissibility in this epoch, and 1 or 2 denotes that the first or second transmissibility genetic architecture will be used in an epoch.

- "drug_resistance": (list of int)

An integer list that specifies which drug resistance trait should be used for each epoch. Should be a list of length `n_epoch`, where each element is an integer representing the trait id. For example, if "traits_num" is specified to be

'{"transmissibility": 2, "drug_resistance": 3}', meaning that you have 3 effect size sets for drug-resistance, then here each element in "drug_resistance" should be one of {0, 1, 2, 3}, where 0 represents not using genetic effect on drug-resistance in an epoch, and 1 or 2 or 3 denotes that the first, second or third drug resistance genetic architecture will be used in an epoch.

- "transition_prob":

This subsection specifies the (base) transition probability between compartments in each `epoch`. Each parameter listed in this subsection should take the format of a numerical list whose length is the same to `n_epoch`.

- "S_IE_prob": (list of floats)

The base probability of a successful transmission for an `effective contact` in one tick. The actual probability of transmission for one contact can be modified by `transmissibility` trait of the pathogen from the infector if "transmissibility" is not 0 in this epoch.

- "I_R_prob": (list of float)

The base probability of an infected host recovering for each epoch. In epochs where "drug_resistance" is not 0, it represents the probability of an infected host being evaluated for recovery per tick, and

the actual recovery probability is modified by the `drug_resistance` trait values of pathogens in the host.

- `"R_S_prob": (list of float)`

The probability of a recovered host losing immunity per tick (going back to susceptible state) during each epoch.

- `"latency_prob": (list of float)`

The probability of a susceptible host becoming exposed upon undergoing a successful transmission for each epoch.

- `"E_I_prob": (list of float)`

The probability of an exposed host activating (going to the infected state) per tick for each epoch.

- `"I_E_prob": (list of float)`

The probability of an infected host deactivating (going to the exposed state) for each epoch.

- `"E_R_prob": (list of float)`

The probability of an exposed host recovering per tick for each epoch.

- `"sample_prob": (list of float)`

The sequential sampling probability of an infected host per tick for each epoch.

- `"recovery_prob_after_sampling": (list of float)`

The probability of a host recovering upon being sequentially sampled for each epoch.

- `"massive_sampling"`: A subsection that specifies concerted sampling events (if any). Please refer to Section 4.4 for how these events work.

- `"event_num": (int)`

Number of concerted sampling events to happen.

- `"generation": (list of int)`

Ticks when each concerted sampling event happens. Should be a list of length `event_num`.

- `"sampling_prob": (list of float)`

Probability of infected hosts being sampled in each concerted sampling event. Should be a list of length `event_num`.

- `"recovery_prob_after_sampling": (list of float)`

Probability of infected hosts recovering upon being sampled in each concerted sampling event. Should be a list of length `event_num`.

- `"super_infection": (bool) true or false`

Whether to permit superinfection. If `true`, the host can be an infectee regardless of existing infection, as long as the number of pathogens within the host does not exceed the specified maximum capacity (`"cap_withinhost"`), and all successful transmissions to this host within a single tick are retained.

3.2.7 Postprocessing options

This subsection specifies whether to do post-simulation data processing and how to visualize the simulated data. Please refer to Part III for all post-processing details.

- `"do_postprocess": (bool) true or false`

Whether to do post-simulation processing. If `true`, then some plots and metadata files will be generated. If `false`, then for each replicate, only logging files for epidemiological events and `treesequence` files of the sampled pathogen genomes will be generated.

- "tree_plotting":

This sub-subsection specifies how to produce the plotted genealogy of sampled pathogen genomes.

- "branch_color_trait": (int)

How to color the branches in the generated tree plot, given that "do_postprocess": true. If specified as 0, branches will be colored by seed ID. If specified as IDs of other traits (both transmissibility and drug resistance traits are ordered together), branches will be colored by relative trait value (lowest in blue, highest in red). For example, if "traits_num" is specified to be '{"transmissibility": 2, "drug_resistance": 3}', then specifying "branch_color_trait": 2 will generate a tree plot with branches colored by the second transmissibility trait, while specifying "branch_color_trait": 4 will generate a tree plot with branches colored by the second drug resistance trait.

- "heatmap": (string) `none` or `transmissibility` or `drug_resistance`

Whether and which trait to plot as a heatmap for each sample in the transmission tree plot, given that "do_postprocess": true is specified.

- "sequence_output":

This sub-subsection specifies how to export the sequences of the sampled pathogen genomes.

- "vcf": (bool) `true` or `false`

Whether to output the sampled sequences in vcf format.

- "fasta": (bool) `true` or `false`

Whether to output the sampled sequences in fasta format.

Chapter 4

Modules of the simulator

Contents

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4.1 Epochs and ticks

(A) The configuration file

```
...
"transition_prob": {
    "S_IE_prob": [ $\beta_1, \beta_2, \dots, \beta_N$ ],
    "I_R_prob": [ $\gamma_1, \gamma_2, \dots, \gamma_N$ ],
    "R_S_prob": [ $\omega_1, \omega_2, \dots, \omega_N$ ],
}
...
...
```

(B) Ticks and epochs in e3SIM

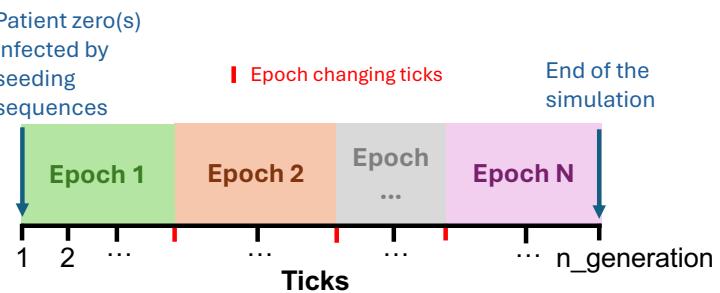


Figure 4.1: **Ticks and epochs in e3SIM.** (A) The configuration file for this simulation. (B) All the ticks are assigned to N epochs, and each epoch uses one set of epidemiological parameters, as shown in (A).

OutbreakSimulator operates in discrete time, where each step, or **tick**, represents the unit of time during which actions occur for each pathogen and host. The simulation concludes when a predefined number of ticks is reached. In every tick, decisions are made for all possible reproduction and compartmental transitions. Ticks are organized into **epochs**, each comprising a sequence of consecutive ticks. Within each epoch, the base rates for compartmental transitions and the genetic architecture for each trait remain constant. These parameters can change to new values when the simulation transitions to a new epoch. The epoch structure allows the simulation to incorporate time-dependent environmental changes, such as implementing new intervention strategies at specific time points after the outbreak starts. For example, in the following configuration file (only part of it is shown), two epochs are specified in the configuration file:

```
...
"epoch_changing": {
```

```

    "n_epoch": 2,
    "epoch_changing_generation": [100]
},
"genetic_architecture": {
    "transmissibility": [0, 1],
    "drug_resistance": [1, 2],
    ...
},
"transition_prob": {
    "S_IE_prob": [0.1, 0.2],
    "I_R_prob": [0.3, 0.3],
    ...
}
...

```

Listing 4.1: Config file for multiple epochs

In the first epoch (tick 1 through tick 99), no genetic architecture is specified for the transmissibility trait, and the first drug resistance trait is used. The base transmission probability and base recovery probability are 0.1 and 0.3, respectively. In the second epoch (tick 100 through the end of the simulation), the first transmissibility trait is used, and the second drug resistance trait is used. The base transmission probability and base recovery probability are 0.2 and 0.3, respectively.

4.2 Evolutionary model

In e3SIM, mutations accumulate in all existing pathogen genomes at each time tick according to a user-specified substitution model. The transition probability matrix \mathbf{P} for nucleotides is a 4×4 matrix where P_{ij} ($i \neq j$) represents the probability of a nucleotide in state i transitioning to state j in one time tick ($i, j \in \{\text{A, C, G, T}\}$). The overall probability of a site in state i acquiring a mutation in each tick is given by $\sum_{j \neq i} P_{ij}$. If no transition probability matrix is provided, users must specify a mutation rate, defaulting the molecular evolution model to the Jukes–Cantor model ("subst_model_parameterization": "mut_rate").

Otherwise, a mutation probability matrix should be provided ("subst_model_parameterization": "mut_rate_matrix"), specifying P_{ij} ($i \neq j$). Following the format of the mutation probability matrix in SLiM, the diagonal values of the mutation probability matrix should be zero (while in practice, the probability of not mutating for allele i is $1 - \sum_{j \neq i} P_{ij}$). Note that the provided mutation probability matrix is essentially a transition **probability** matrix in a discrete-time Markov chain, where each time unit is one tick. To simulate the evolution of some specific pathogen, the mutation probability matrix should be scaled to the real time t_0 that each tick represents (unit: real time/tick). For example, a transition rate matrix \mathbf{Q} estimated by popular maximum likelihood inference methods such as RAxML or IQ-TREE, represents instantaneous rate of transition, in the time unit of the time needed for 1 mutation to happen per site. To specify an appropriate transition probability matrix for e3SIM, the molecular clock μ_0 (unit: mutation/site/real time) of this pathogen should be used to calculate the transition probability matrix. The calculated transition probability matrix should be

$$\mathbf{P} = e^{\mathbf{Qt}}$$

where $t = t_0 \times \mu_0$. After changing all the diagonal values to 0, this matrix can be supplied to e3SIM.

Mutations occur stochastically at a constant probability each tick, as defined by the mutation probability matrix, regardless of whether the pathogen reproduces. Trait values, influenced by the mutation profiles of the pathogens, are recalculated before transmission events. These recalculated trait values subsequently alter the probabilities of relevant transitions, such as transmission and recovery. This mechanism captures the crucial interaction between epi-eco-evo processes, which is often overlooked in classical epidemiological simulators.

4.3 Genetic effects on trait values

We model traits via standard quantitative-genetic additive effects. For trait i , let $S^{(i)}$ denote the set of causal sites, set up by `GeneticEffectGenerator` (For details, see Section 2.3). We define the additive genetic value of trait i for a pathogen genome as:

$$G_i = \sum_{j \in S^{(i)}} q_{ij} x_j, \quad (4.1)$$

where $x_j \in \{0, 1\}$ indicates the presence of the effect allele at site j in the haploid pathogen genome, and q_{ij} is the additive effect on the link scale of site j on trait i .

We map the genetic value G_i of trait i to the corresponding event probability for each pathogen using standard GLM links: the logit link or the complementary log-log (cloglog) link. Here, we illustrate two traits, transmissibility and drug resistance, but e3SIM supports any user-defined pathogen genetic traits.

Logit link (log-odds scale).

Let $\beta \in (0, 1)$ be the baseline per-contact transmission probability at $G_{\text{trans}} = 0$, and $s \in (0, 1)$ the baseline per-tick pathogen survival probability under treatment at $G_{\text{drug}} = 0$. We map the additive genetic values to probabilities via:

$$\begin{aligned} P(\text{transmission}) &= \text{logit}^{-1} (\text{logit}(\beta) + \alpha_{\text{trans}} G_{\text{trans}}), \\ P(\text{survival} \mid \text{treatment}) &= \text{logit}^{-1} (\text{logit}(s) + \alpha_{\text{drug}} G_{\text{drug}}), \end{aligned} \quad (4.2)$$

where $\text{logit}(p) = \log(p/(1-p))$, $\text{logit}^{-1}(z) = 1/(1+e^{-z})$, and $\alpha_{\text{trans}}, \alpha_{\text{drug}} > 0$ are slope parameters. Thus, under this link, a unit increase in G_{trans} multiplies the odds of transmission by $e^{\alpha_{\text{trans}}}$, and a unit increase in G_{drug} multiplies the odds of survival by $e^{\alpha_{\text{drug}}}$.

Complementary log-log (cloglog) link (discrete-time proportional hazards).

Let $\lambda = -\log(1-\beta) > 0$ be the baseline transmission hazard per contact and $\kappa = -\log(s) > 0$ the baseline pathogen clearance hazard per tick under treatment (so baseline pathogen survival equals $e^{-\kappa}$). Then,

$$\begin{aligned} P(\text{transmission}) &= 1 - \exp(-\lambda e^{\alpha_{\text{trans}} G_{\text{trans}}}), \\ P(\text{survival} \mid \text{treatment}) &= \exp(-\kappa e^{-\alpha_{\text{drug}} G_{\text{drug}}}). \end{aligned} \quad (4.3)$$

Thus, $e^{\alpha_{\text{trans}}}$ is the transmission hazard ratio per +1 unit of G_{trans} , and $e^{-\alpha_{\text{drug}}}$ is the clearance hazard ratio per +1 unit of G_{drug} (so larger G_{drug} increases survival by reducing clearance hazard). These expressions correspond to standard discrete-time proportional hazards models.

4.4 Compartmental model

e3SIM employs a classical compartmental epidemiological model by dividing all hosts into four compartments: susceptible (S), exposed (E), infected (I), and recovered (R). Transitions between compartments have base rates that are specified in the configuration file. e3SIM supports a wide range of transitions:

In every tick, **reproduction** happens first. There are two types of reproduction events in e3SIM: transmission and within-host replication.

Transmission occurs when a pathogen infects a new host by producing offspring within the new host, which can only happen when the hosts are in direct contact. During each tick, all effective contacts are evaluated for potential transmissions. An effective contact involves a pair of hosts connected by an edge, where one host is infected (the infector) and the other is capable of being infected (the infectee). For each effective contact, a single pathogen is randomly selected from the infector's pathogen population. The transmission success of the chosen pathogen is determined by a Bernoulli trial with the success probability calculated from both the base transmission probability β and the transmissibility trait (Equations 4.2 and 4.3)

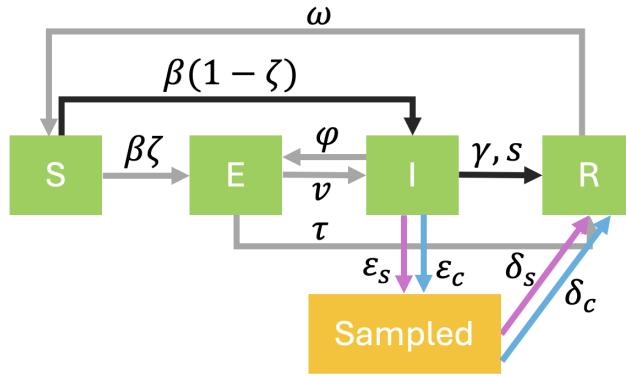


Figure 4.2: **The compartmental model in e3SIM.** Green blocks are the compartments of hosts. The grey arrows show optional transitions whose probabilities can be specified as constants; the red arrows show transitions whose probabilities are affected by the relevant trait values of pathogens. The Greek letters denote the corresponding base transition probabilities, which are fixed within one epoch.

Table 4.1: List of symbols for epidemiological parameters. All probabilities on a per-tick basis.

Symbol	Description	Keys
ω	Probability of immunity loss for a recovered host	R_S_prob
β	Base probability of successful infection event for one effective contact	S_IE_prob
ζ	Probability of latent infection for each successful infection event	latency_prob
ϕ	Probability of deactivation for an infected host	I_E_prob
ν	Probability of activation for an exposed host	E_I_prob
τ	Probability of transitioning to recovered state for an exposed host	E_R_prob
γ	In treatment-free epochs: Base probability of recovery for an infected host; In treatment epochs: Probability of recovery evaluation for an infected host.	I_R_prob
s	Base probability of pathogen survival under treatment per tick.	surviv_prob
ϵ_s	Probability of being sampled in a sequential sampling event for an infected host	sample_prob
δ_s	Probability of recovery for an infected host following a sequential sampling event	recovery_prob_after_sampling
ϵ_c	Probability of being sampled in a concerted sampling event for an infected host	sampling_prob
δ_c	Probability of recovery for an infected host following a concerted sampling event	recovery_prob_after_sampling

Successful trials result in pathogen transmission to the infectee. e3SIM assumes a transmission bottleneck, where only one pathogen genome from the infector is transmitted to the infectee per successful transmission.

“Superinfection” refers to a scenario in which a host can be infected by multiple sources, either within the same tick or across different ticks. If disabled, a host can only be infected if it is in the susceptible state. In this case, if multiple successful transmissions target the same host in a tick, only one infection event, selected at random, is retained and all other infection events are omitted. If superinfection is enabled, the host can be an infectee regardless of existing infection, as long as the number of pathogens within the host does not exceed the specified maximum capacity. All successful transmissions to this host within a single tick are retained.

Within-host replication occurs when a pathogen reproduces within a single host. This process is enabled only if the within-host replication mechanism is active and the total number of pathogens within the host does not exceed the user-defined maximum capacity. Under these conditions, each pathogen has a probability of producing exactly one offspring within the same host during each simulation tick. Mutations accumulate over time independently of reproduction events (Section 3.2.2). Therefore, all pathogens, regardless of their reproduction status, have the same probability of mutation per tick.

State changes occur after reproduction events. The transitions for hosts in different states are listed below:

- A newly infected host that is in **susceptible** state before being infected enters the exposed state with a probability ζ and the infected state with a probability $1 - \zeta$. If `superinfection=false`, for hosts that are infected more than once in this tick, only one of the infection events, selected at random, will be kept and all other infection events are omitted. Susceptible hosts that are not involved in transmission events in the current tick remain in the susceptible state.
- For **exposed** hosts, new states are decided by a random draw from a multinoulli distribution with event probabilities v , τ and $1 - \tau - v$, which represent the probability of activation (`E_I_prob`), recovery (`E_R_prob`), and staying exposed.
- For **infected** hosts, in non-treatment epochs (when "drug_resistance" in the "genetic_architecture" section is specified as 0 for the current epoch) new states are decided by a random draw from a multinoulli distribution with event probabilities γ , ϵ , ϕ and $1 - \gamma - \epsilon - \phi$, which represent the probability of recovery evaluation (`I_R_prob`), being sequentially sampled (`sample_prob`), deactivation (`I_E_prob`), and staying infected. For those chosen to be sequentially sampled, an additional Bernoulli trial with parameter δ (`recovery_prob_after_sampling`) determines whether the host ends in the recovered or infected state. If drug resistance is inactive in the current epoch, a successful recovery evaluation transitions the host into the recovered state. However, in treatment epochs (when "drug_resistance" in the "genetic_architecture" section is specified as a drug-resistance trait set different than 0 for the current epoch), hosts are first evaluated for recovery before undergoing other events. We first evaluate the innate recovery event with probability γ ; on success, the host is set to recovered, and all within-host pathogens are cleared. If the immune trial fails, we then evaluate drug-induced clearance at the pathogen level: each pathogen k present at the start of the tick survives treatment independently with probability $p_k = P(\text{survival} \mid \text{treatment})$, mapped from its drug-resistance genetic value (Equation 4.1) via the specified link (Equation 4.2 and 4.3) and baseline survival s at $G_{\text{drug}} = 0$. The host recovers if and only if all within-host pathogens fail their survival trials in that tick; otherwise, the host remains in its current infected state with the surviving pathogens. After the recovery evaluation, the remaining hosts in the infected state undergo sequential sampling event with probability ϵ_s , deactivation event with probability ϕ , and stay unchanged with probability $1 - (\phi + \epsilon_s)$, modeled by random draws from a multinomial distribution.
- For **recovered** hosts, new states are decided by a Bernoulli trial with probability ω (`R_S_prob`) to see whether they lose their immunity and become susceptible again.

After these sequential state changing events, concerted sampling events happen if specified (specified in the `massive_sampling` section of the configuration file). Concerted sampling happens only at specific ticks, and each event has to be specified by its timing, sampling probability (ϵ_c) and recovery probability after sampling (δ_c).

Chapter 5

Specifying your own configuration

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This chapter provides examples of different scenarios to demonstrate how the configuration file and *OutbreakSimulator* work. To avoid syntax errors when copying and pasting from this PDF, all code is provided in the `run.sh` script located in the respective vignette directory (“`tutorial/vignettes/5.1_test_minimal_model`” or “`tutorial/vignettes/5.2_test_drugresist`”), which also serves as the working directory (`$(WKDIR)`). Run bash `run.sh` from within that vignette directory.

Before running the tutorials, set both `${e3SIM}` and `${WKDIR}` in `run.sh` to **absolute paths** so that all required inputs are found regardless of where the script is invoked from. Do not use a relative path (for example, `./vignettes/...`).

```
# Set to the ABSOLUTE path where your e3SIM source code is located
e3SIM="/absolute/path/to/e3SIM-main/e3SIM_codes"

# Set to the ABSOLUTE path of the current vignette folder
# TIP: Run 'pwd' in your terminal and paste the output here.
WKDIR="/absolute/path/to/tutorial/vignettes/5.1_test_minimal_model"
# or "/absolute/path/to/tutorial/vignettes/5.2_test_drugresist"
# If you are already in the vignette folder: WKDIR="$(pwd)"
```

Listing 5.1: `run.sh` setup

5.1 A minimal model for transmissibility

In this section, we will simulate a *Mycobacterium tuberculosis* (*Mtb*) outbreak in a host population of size 10,000 for 10 years. We set each tick to represent 1 day, so we simulate 3,650 ticks. The mutation rate for *Mtb* genome is 0.5 SNPs per genome per year, which scales to be 3.12×10^{-10} SNPs/bp/day. You can set a random seed for this pipeline for reproducibility by setting `RANDOMSEED=1100` before running the pipeline, which should reproduce the results we provide in the repository.

1. The first step is to run *NetworkGenerator*. We want to simulate a well-mixed population with every individual having 10 contacts. We thus run:

```
python ${e3SIM}/network_generator.py \
    -popsize 10000 \
    -wkdir ${WKDIR} \
```

```
-method randomly_generate \
-model ER \
-p_ER 0.001 \
-random_seed ${RANDOMSEED}
```

Listing 5.2: NetworkGenerator model 1

2. We then run *SeedGenerator* to generate the seeds. We want to run a Wright-Fisher model of $N_e = 1000$ and run for 4000 generations. During the burn-in, we want to let one generation scale to one year, and thus use mutation rate 1.1×10^{-7} . We want to generate 5 seeds. The reference genome we use here is *Mtb* genome, which we put in our repository as well.

```
python -u ${e3SIM}/seed_generator.py \
-wkdir ${WKDIR} \
-num_init_seq 5 \
-method SLiM_burnin_WF \
-Ne 1000 \
-ref_path GCF_000195955.2_ASM19595v2_genomic.fna \
-mu 1.1e-7 \
-n_gen 4000 \
-random_seed ${RANDOMSEED}
```

Listing 5.3: SeedGenerator model 1

If all seeds have a single common ancestor, *seeds.nwk* will appear in the working directory, and we can manually scale the branch lengths by a factor of 365 to scale the tree based on the daily mutation rate. For now, we leave *seeds.nwk* unchanged. It is fine if there is not a *seeds.nwk* file, but we will not be able to generate a full phylogeny tree after *OutbreakSimulator*; rerun this program until all seeds coalesce if desired.

3. Next, we would like to run *GeneticEffectGenerator* to generate causal sites from candidate regions. A CSV file named *candidate_regions.csv* that contains 100 genes from the TB genome annotation is provided in our repository. We would like to use the default parameters to select the causal sites, and use a normal distribution with variance 1.5 to generate effect sizes. By default, the effect sizes will be calibrated by seeding sequences' mutation profiles. We also would like to get the calibrated link scale slope α , by setting the transmission odds ratio as 2. We thus run:

```
python ${e3SIM}/genetic_effect_generator.py \
-wkdir ${WKDIR} \
-method randomly_generate \
-num_init_seq 5 \
-trait_n '{"transmissibility": 1, "drug_resistance": 0}' \
-csv ${WKDIR}/candidate_regions.csv \
-func n \
-taus 1.5 \
-calibration_link T \
-Rs 2 \
-random_seed ${RANDOMSEED}
```

Listing 5.4: GeneticEffectGenerator model 1

Running *GeneticEffectGenerator* should output *causal_gene_info.csv* and *seeds_trait_values.csv*. *seeds_trait_values.csv* shows the transmissibility value for each seed based on the genetic architecture that was generated and stored in *causal_gene_info.csv*.

Using the provided random seed, all seeding sequences resulted in zero trait values because no mutations occurred at the designated causal sites. To generate polymorphic trait values, you can rerun the program with a different random seed or manually modify the file `causal_gene_info.csv` and rerun the program using the `-method user_input` option to adjust the genetic effects.

You will also see the calibrated link scale slope in the terminal. Using the random seed we provided, you should see the following information in terminal:

```
The calibrated link-scale slopes under the logit link are as follows.
transmissibility_1: 0.6648
Please write the following part into your configuration file under the "trait_prob_link" key:
{
  "link": "logit",
  "logit": {
    "alpha_trans": [
      0.6648
    ],
    "alpha_drug": []
  }
}
```

Listing 5.5: Printout message for GeneticEffectGenerator's link scale slope calibration

This gives the calibrated α for all the traits; in this case, there is only one $\alpha_{\text{trans}} = 0.6648$, since transmissibility is the only trait generated. The value will not be automatically written to the configuration file, so you will need to manually add it, as shown in Configuration file 5.7.

4. The last step before running *OutbreakSimulator* is matching the seeds to hosts using *HostSeedMatcher*. This time, we want all the seeds to be matched randomly. We can run:

```
python ${e3SIM}/seed_host_matcher.py \
  -wkdir ${WKDIR} \
  -num_init_seq 5 \
  -method randomly_generate \
  -random_seed ${RANDOMSEED}
```

Listing 5.6: HostSeedMatcher model 1

`seed_host_match.csv` should appear in the working directory.

5. Now, we can run *OutbreakSimulator*. We first download the template configuration file from the repository, then make the desired modifications. According to the CDC, latency period for TB can be around 3 months, and recovery time can be 4 months for a rifapentine-moxifloxacin treatment regimen. We want to model all new infections initially leading to latency. Though it is controversial, several studies indicate that immunity does not exist in TB. We thus assume that immunity loss occurs at around 20 days. For base transmission probability, we assume an arbitrary probability of 0.004, meaning that for each contact pair, the per-day infection probability is 0.4%. We do not want to model other transitions between compartments, and we do not want to model massive sampling events. This is the configuration file we specify using these parameters:

```
{
  "BasicRunConfiguration": {
    "cwdir": "5.1_test_minimal_model",
    "n_replicates": 3
  },
```

```

"EvolutionModel": {
    "subst_model_parameterization": "mut_rate",
    "n_generation": 3650,
    "mut_rate": 3.12e-10,
    "within_host_reproduction": false,
    "within_host_reproduction_rate": 0,
    "cap_withinhost": 1
},
"SeedsConfiguration": {
    "seed_size": 5,
    "use_reference": false
},
"GenomeElement": {
    "use_genetic_model": true,
    "ref_path": "GCF_000195955.2_ASM19595v2_genomic.fna",
    "traits_num": {
        "transmissibility": 1,
        "drug_resistance": 0
    },
    "trait_prob_link": {
        "link": "logit",
        "logit": {
            "alpha_trans": [
                0.6648
            ],
            "alpha_drug": []
        }
    }
},
"NetworkModelParameters": {
    "host_size": 10000
},
"EpidemiologyModel": {
    "model": "SEIR",
    "slim_replicate_seed_file_path": "slimseeds.csv",
    "epoch_changing": {
        "n_epoch": 1,
        "epoch_changing_generation": []
    },
    "genetic_architecture": {
        "transmissibility": [
            1
        ],
        "drug_resistance": [
            0
        ]
    },
    "transition_prob": {
        "S_IE_prob": [
            0.004
        ],
        "I_R_prob": [
            0.008
        ],
        "R_S_prob": [
            0.05
        ],
        "latency_prob": [
            1
        ],
    }
}

```

```

    "E_I_prob": [
        0.01
    ],
    "I_E_prob": [
        0
    ],
    "E_R_prob": [
        0
    ],
    "sample_prob": [
        1e-05
    ],
    "recovery_prob_after_sampling": [
        0
    ]
},
"massive_sampling": {
    "event_num": 0,
    "generation": [],
    "sampling_prob": [],
    "recovery_prob_after_sampling": []
},
"super_infection": false
},
"Postprocessing_options": {
    "do_postprocess": true,
    "tree_plotting": {
        "branch_color_trait": 1,
        "heatmap": "transmissibility"
    },
    "sequence_output": {
        "vcf": true,
        "fasta": false
    }
}
}

```

Listing 5.7: config model 1

We thus use the absolute path to this configuration file to run *OutbreakSimulator* by:

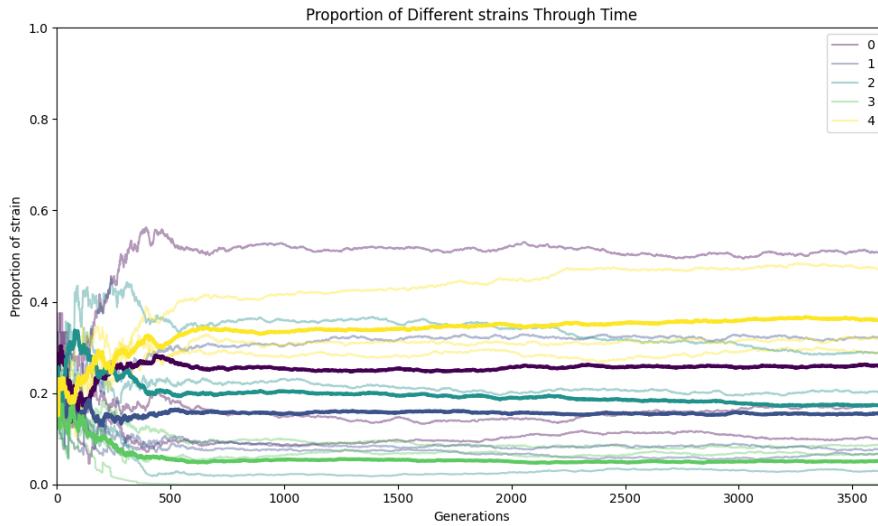
```
python -u outbreak_simulator.py \
    -config ${PATH_TO_THIS_CONFIG}
```

Listing 5.8: OutbreakSimulator model 1

OutbreakSimulator will print logging information to standard output as it progresses through the simulation. When the program finishes, you can look in the working directory and in each replicate's subdirectory for the results. Please note that the file paths in the configuration have to be changed to the full paths of your own directories before running.

6. For detailed instructions on how to analyze the output files, please refer to Chapter 6. Using the current setup and random seed, you will encounter a terminal warning message: “WARNING: There are no mutations related to any trait in samples from this replication.” This occurs because *Mtb* has an extremely low mutation rate, and none of the sampled genomes carry mutations at the causal sites defined in Step 3. Consequently, the resulting plotted phylogenetic trees will appear entirely gray, as they are colored by the transmissibility trait, which remains identical across all sampled genomes. Similarly, the

strain-frequency plot over time will appear largely homogeneous, since no mutation confers a selective advantage—most strains spread at approximately equal probabilities.



5.2 Multi-stage drug treatment

In this section, we will simulate a COVID-19 outbreak in a host population of size 10,000 for 2 years. We want each tick to represent 1 day, so we want to simulate 730 ticks. The mutation rate of coronavirus is 6.67×10^{-4} SNPs/bp/year, which is 1.8×10^{-6} SNPs/bp/day. In this simulation, we model realistic host population structure and different treatment stages. You can set a random seed for this pipeline for reproducibility by setting RANDOMSEED=1100 before running the pipeline, which should reproduce the results we provide in the repository.

1. The first step is to run *NetworkGenerator*. We want to simulate a realistic host population structure using a scale-free graph. We thus run:

```
python ${e3SIM}/network_generator.py \
    -wkdir ${WKDIR} \
    -popsize 10000 \
    -method randomly_generate \
    -model BA \
    -m 2 \
    -random_seed ${RANDOMSEED}
```

Listing 5.9: NetworkGenerator model 2

2. In this example, we are starting from the reference genome, so we do not need to run *SeedGenerator*.
3. We then run *GeneticEffectGenerator*. In this example, we are using a predefined effect size file `causal_gene_info.csv` in the working directory. We do not want to do calibration, since we are using the reference genome as the seed sequences.

```
python ${e3SIM}/genetic_effect_generator.py \
    -wkdir ${WKDIR} \
    -method user_input \
    -num_init_seq 1 \
    -csv ${WKDIR}/causal_gene_info.csv \
    -trait_n '{"transmissibility": 1, "drug_resistance": 2}' \
```

```
-calibration F \
-calibration_link T \
-random_seed ${RANDOMSEED}
```

Listing 5.10: GeneticEffectGenerator model 2

Here `-trait_n` tells e3SIM that the provided effect-size file contains one transmissibility trait set and two drug-resistance trait sets. You should see `causal_gene_info.csv` appearing in your working directory, along with a `seeds_trait_values.csv` file where all the traits of the single seeding sequence are shown as 0s. We calibrated the link scale slope here as well, but we decided to use different link scale slope in the simulation, as shown in the configuration file 5.12.

4. We then run *HostSeedMatcher*. In this example, we are using only one seed, and we want to match it to a host that has median contact degree. We thus utilize the ranking method to match the seed and host.

```
python ${e3SIM}/seed_host_matcher.py \
-wkdir ${WKDIR} \
-method randomly_generate \
-num_init_seq 1 \
-match_scheme '{"0": "ranking"}' \
-match_scheme_param '{"0": 5000}' \
-random_seed ${RANDOMSEED}
```

Listing 5.11: SeedHostMatcher model 2

You should see `seed_host_match.csv` in your working directory.

5. Now, we can run *OutbreakSimulator*. We first download the template configuration file from our GitHub repository, then make the desired modifications. According to the CDC, latency period for COVID is around 3 days, and recovery time is around 2 weeks. We want to model all new infections initially leading to latency. Though it is controversial, immunity can be lost within 30 days. For basic transmission probability, we assume an arbitrary probability of 0.03, meaning that for each contact pair, the per-day infection probability is 3%. We do not want to model other transitions between compartments, and we do not want to model massive sampling events. We conduct post-processing, asking *OutbreakSimulator* to plot genealogies of the sampled pathogens and include a drug resistance heatmap. This is the configuration file we specify using these parameters:

```
{
  "BasicRunConfiguration": {
    "cwdir": "5.2_test_drugresist",
    "n_replicates": 3
  },
  "EvolutionModel": {
    "subst_model_parameterization": "mut_rate",
    "n_generation": 730,
    "mut_rate": 1.8e-06,
    "within_host_reproduction": false,
    "within_host_reproduction_rate": 0,
    "cap_withinhost": 1
  },
  "SeedsConfiguration": {
    "seed_size": 1,
    "use_reference": true
  },
  "GenomeElement": {
    "use_genetic_model": true,
```

```

    "ref_path": "COVID/EPI_ISL_402124.fasta",
    "traits_num": {
        "transmissibility": 1,
        "drug_resistance": 2
    },
    "trait_prob_link": {
        "link": "logit",
        "logit": {
            "alpha_trans": [
                0.405
            ],
            "alpha_drug": [
                0.0116,
                0.0273
            ]
        }
    }
},
"NetworkModelParameters": {
    "host_size": 10000
},
"EpidemiologyModel": {
    "model": "SEIR",
    "slim_replicate_seed_file_path": "slimseeds.csv",
    "epoch_changing": {
        "n_epoch": 3,
        "epoch_changing_generation": [
            250,
            500
        ]
    },
    "genetic_architecture": {
        "transmissibility": [
            1,
            1,
            1
        ],
        "drug_resistance": [
            0,
            1,
            2
        ]
    },
    "transition_prob": {
        "S_IE_prob": [
            0.04,
            0.02,
            0.02
        ],
        "I_R_prob": [
            0.03,
            0.06,
            0.05
        ],
        "R_S_prob": [
            0.05,
            0.1,
            0.1
        ],
        "latency_prob": [

```

```

        1,
        1,
        1
    ],
    "E_I_prob": [
        0.3,
        0.3,
        0.3
    ],
    "I_E_prob": [
        0,
        0,
        0
    ],
    "E_R_prob": [
        0,
        0,
        0
    ],
    "sample_prob": [
        0.0002,
        0.0003,
        0.0003
    ],
    "recovery_prob_after_sampling": [
        0,
        0,
        0
    ],
    "surviv_prob": [
        0.01,
        0.96,
        0.96
    ]
},
"massive_sampling": {
    "event_num": 0,
    "generation": [],
    "sampling_prob": [],
    "recovery_prob_after_sampling": []
},
"super_infection": false
},
"Postprocessing_options": {
    "do_postprocess": true,
    "tree_plotting": {
        "branch_color_trait": 1,
        "heatmap": "drug_resistance"
    },
    "sequence_output": {
        "vcf": true,
        "fasta": false
    }
}
}

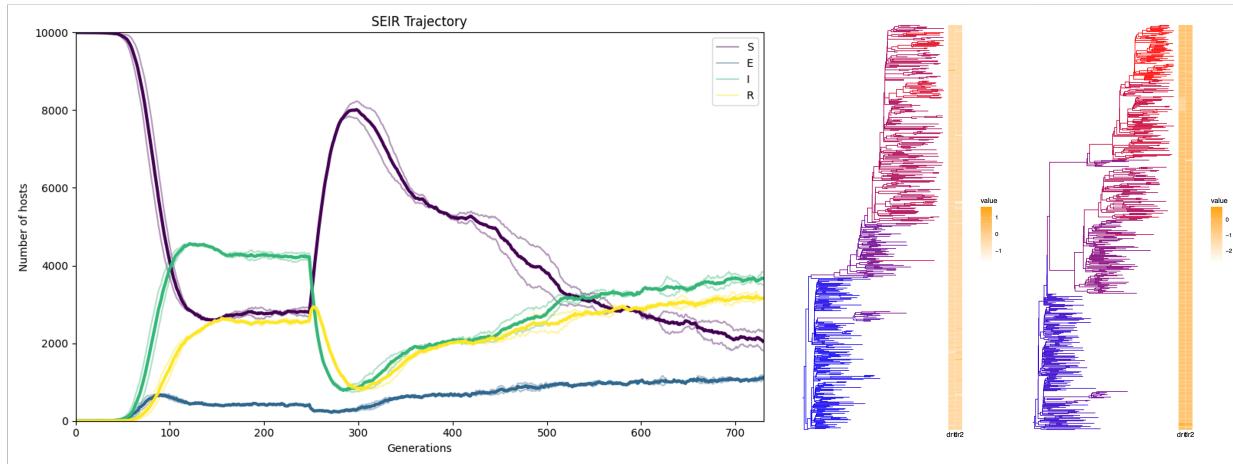
```

Listing 5.12: config model 2

OutbreakSimulator will print logging information to standard output as it progresses through the simu-

lation. Since this is a stochastic simulation, it is not guaranteed that the outbreak will reach an endemic stage; i.e., the outbreak might end quickly enough that there are not many samples retrieved. If no samples are retrieved at all, *OutbreakSimulator* will give a warning and there will be no post-simulation processing steps. With this setup, a replicate may have the epidemic die out early and produce no output. In that case, the terminal prints: “WARNING: Replicate X failed or produced no output”. Even with the same seed, the exact replicate index and outputs can vary slightly across platforms and software versions.

- When the program finishes, you can look in the working directory and each replicate’s subdirectory for the results. For detailed instructions on how to analyze the output files, please refer to Chapter 6. The results and configurations of one test run are in our repository for comparison. Below, we show the aggregated SEIR trajectory for two replicates (since one of the replicates fails to produce samples) from one simulation run. The different behavior of the outbreak in different epochs can be clearly seen. We also show examples of the transmission trees for two replicates; in both replicates, the population accumulates higher transmissibility through mutations over time.



Part III

Post-simulation processing

Chapter 6

Output

In this chapter, we describe the structure of the output of e3SIM and how to read the outputs.

6.1 Output structure

For each replicate, e3SIM creates a folder named with the replicate ID (starting at 1) in the working directory. Each replicate folder contains the following output files:

- `infection_raw.csv.gz`: Compressed CSV file that stores all raw transmission events (including events that were omitted when superinfection is deactivated and therefore do not appear in `infection.csv.gz`). Every row in the file describes one infection event. The first column gives the tick number, and the second and third columns give the infector's and infectee's host IDs, respectively.
- `infection.csv.gz`: Compressed CSV file that stores all preserved transmission events. For example, when superinfection is deactivated, only one transmission event per infectee is preserved per tick; all other transmissions are unsuccessful and thus are not stored in `infection.csv.gz`. Every row in the file describes one infection event. The first column gives the tick number, and the second and third columns give the infector's and infectee's host IDs, respectively.
- `recovery.csv.gz`: Compressed CSV file that stores all recovery events that occurred. Every row in the file describes one recovery event. The first column gives the tick number, and the second column gives the recovering host ID.
- `sample.csv.gz`: Compressed CSV file that stores all sampling events that occurred. Every row in the file describes one sampling event. The first column gives the tick number, the second column gives the sampled host ID, and the third column gives the ID of the seeding sequence for each sampled sequence.

Tick	Infector	Infectee
2	7108	3085
4	2735	5423
5	6646	4344
5	9408	1738
9	9408	1554

infection_raw.csv

Tick	Infector	Infectee
2	7108	3085
4	2735	5423
5	6646	4344
5	9408	1738
9	9408	1554

infection.csv

Tick	Recovering host
160	2214
168	7109
169	6040
181	7612
182	3890

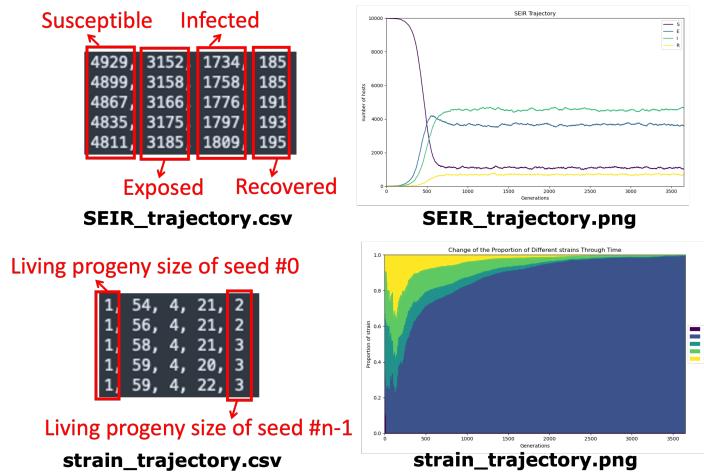
recovery.csv

Tick	seed id
803	9357
818	8061
820	3972
822	4311
837	7770

sample.csv

- `SEIR_trajectory.csv.gz`: Compressed CSV file that stores the SEIR compartment sizes for each tick. It has a number of rows equal to the total number of ticks. The four columns represent the sizes of susceptible, exposed, infected, and recovered, respectively.

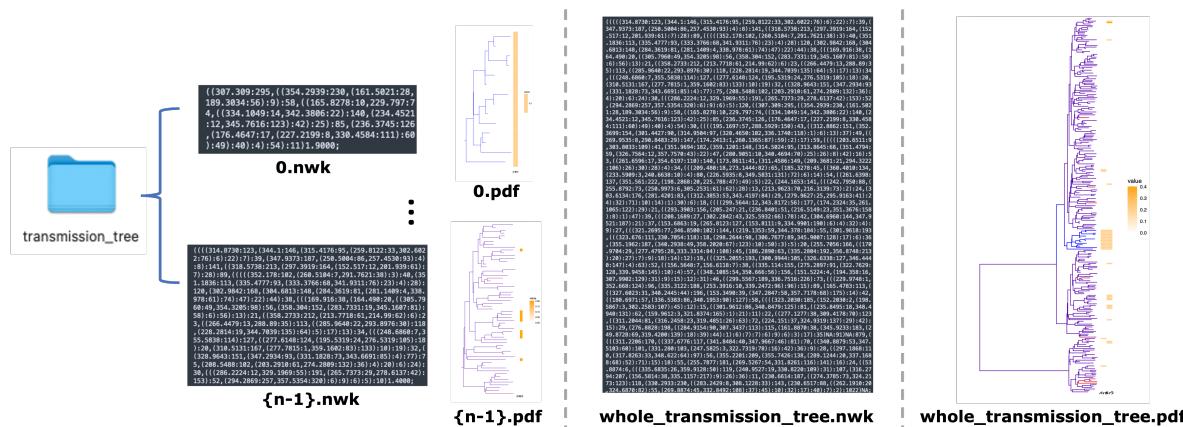
- **strain_trajectory.csv.gz**: Compressed CSV file that stores the size of the progeny of each strain for each tick. It has a number of rows equal to the total number of ticks. Each column represents the progeny size of one seed (strain), in the order of seed IDs.
- **SEIR_trajectory.png**: Plot of **SEIR_trajectory.csv.gz**.
- **strain_trajectory.png**: Plot of **strain_trajectory.csv.gz** in proportion.



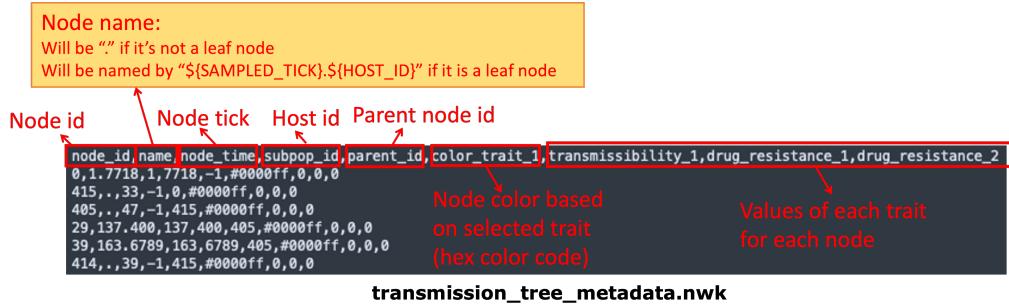
- **transmission_tree**: A folder that stores the transmission tree for each seed's progeny; it contains **x.nwk** for each seed ID **x**. For seeds that have more than one sampled offspring, an **tree.x.pdf** file (a plotted tree) will also be present.

The leaf labels in the **x.nwk** file is in the format of **\${SAMPLED_TICK} . \${SAMPLED_HOST_ID}**, which corresponds to the sample names in the **.vcf** file

- **whole_transmission_tree.pdf**: Plotted transmission tree, concatenating the seed's phylogeny and each seed's transmission tree. This file will be generated only if the seed phylogeny is provided in the working directory.
- **whole_transmission_tree.nwk**: Transmission tree, concatenating the seed's phylogeny and each seed's transmission tree. This file will be generated only if the seed phylogeny is provided in the working directory.



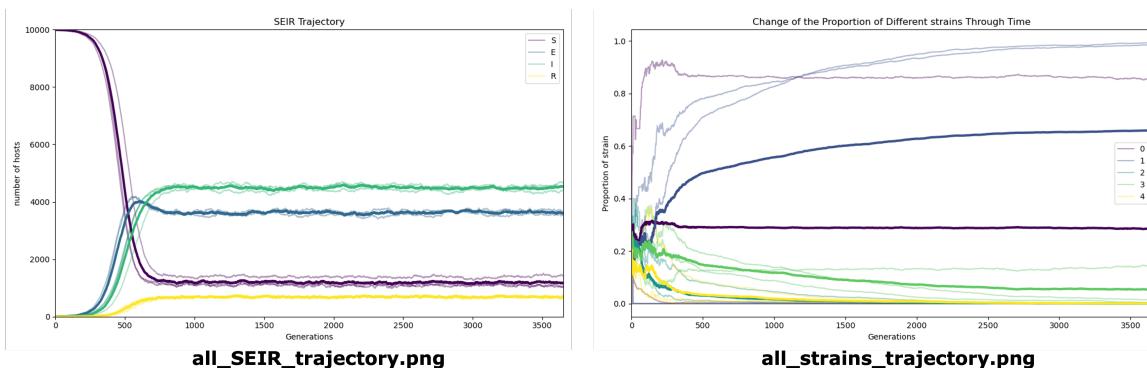
- `sampled_genomes.trees`: Tree sequence of the sampled genomes, output by SLiM in OutbreakSimulator.
- `transmission_tree_metadata.csv`: Metadata for the transmission tree, storing the trait values for each node in the transmission tree and the coloring of each node.



- `sampled_pathogen_sequences.vcf`: The mutation profiles of the sampled pathogen sequences in VCF format, where sample names correspond to the naming convention in `whole_transmission_tree.nwk`.
- `sample.SNPs_only.fasta`: A FASTA file showing the concatenated SNPs for each sampled pathogen, where sample names correspond to the naming convention in `whole_transmission_tree.nwk`.
- `final_samples_snp_pos.csv`: A CSV file showing the positions of each site corresponding to the reference genome in the concatenated SNPs for each sampled pathogen.
- `sample.wholegenome.fasta`: A FASTA file showing the sampled pathogens' whole genome sequences, where sample names correspond to the naming criteria in `whole_transmission_tree.nwk`.

In the working directory, there will be a folder called `output_trajectories`, containing:

- `all_SEIR_trajectory.png`: Plot the aggregated results from `SEIR_trajectory.csv.gz` for each replicate, with the average trajectory overlaid.
- `all_strains_trajectory.png`: Plot the aggregated results from `strain_trajectory.csv.gz` for each replicate, with the average trajectory overlaid.



Part IV

GUI

Chapter 7

Introduction to the GUI

e3SIM provides a graphical user interface for users who prefer visual interactions over command-line tools. The GUI can be used to execute all the pre-simulation modules and generate the configuration file for the main simulator module. In each tab, there will be a “Next” button. Clicking this button saves the information entered in the current tab to the backend, so it is mandatory to click “Next” before navigating to the next tab.
If the “Next” button does not appear in the interface, please switch to full-screen mode.

To launch the GUI, first locate the GUI subdirectory within the e3SIM_codes folder of your e3SIM installation. Assuming the path to the e3SIM_codes folder is defined as \${e3SIM}, the GUI can be started with the following command:

```
python ${e3SIM}/gui
```

Listing 7.1: Launch the GUI

It is recommended to launch the program from the working directory where you intend to run your simulation, so that the GUI will automatically use the current directory as the default for file and path selections. Please note that when pre-simulation modules are run in the backend, the GUI will freeze until the step finishes.

The GUI screenshots and settings shown in this chapter correspond to the Section 5.2 tutorial vignette (`tutorial/vignettes/5.2_test_drugresist`); i.e., the GUI should generate a configuration file that can be used to run the same simulation shown in Configuration file 5.12.

A GUI window pops up with several tabs. We describe them in detail below.

Chapter 7 Introduction to the GUI

A GUI window pops up with several tabs:

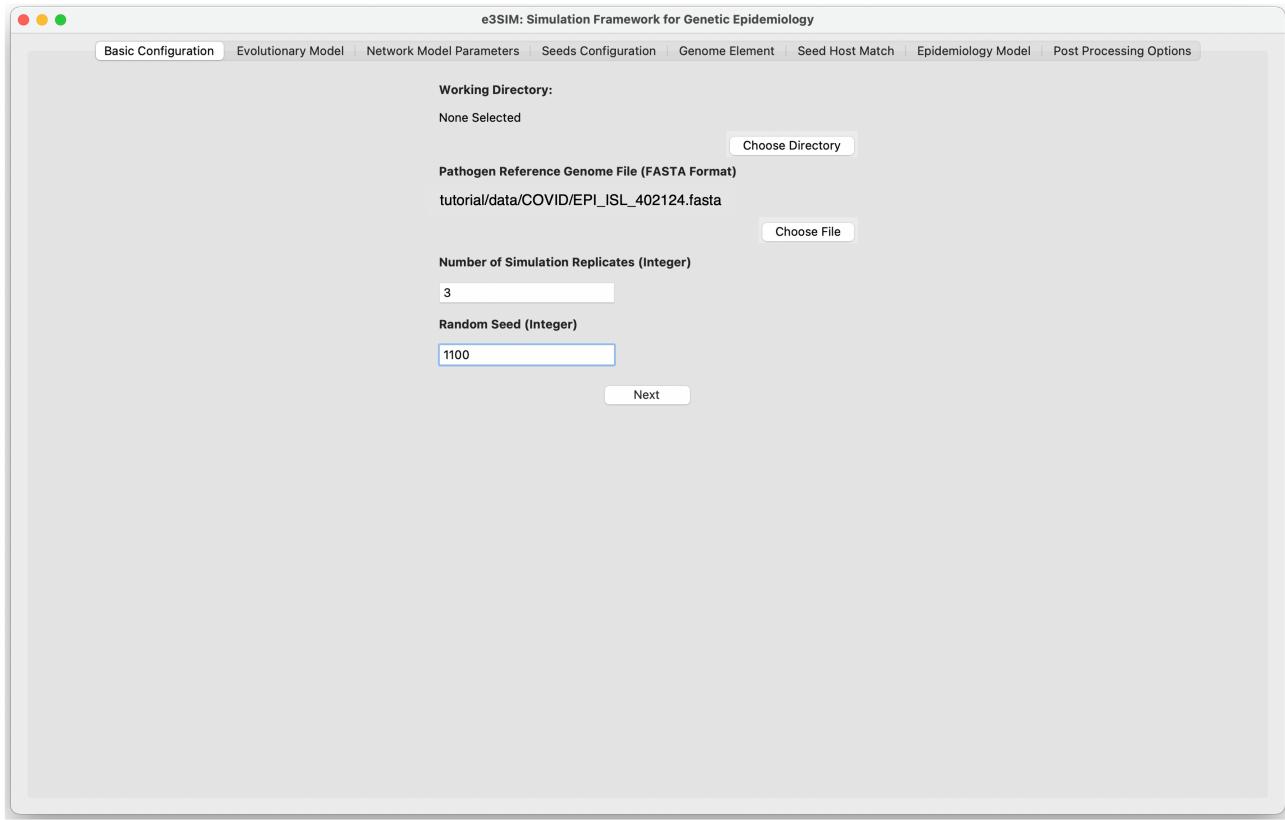


Figure 7.1: The first tab of the GUI: Basic configuration.

In the “Basic Configuration” tab (Figure 7.1), some basic parameters are set, including the working directory and the random number generator, which are used in subsequent tabs where pre-simulation modules are executed. By clicking the “Choose Directory” button after the “Working Directory” item, you can navigate to the path you would like to choose as your working directory, and all subsequent pre-simulation modules will generate files in this path you chose. Note that when clicking on the button, the default path popping out will be the current path where you launched the GUI.

Then, you can navigate to choose the reference genome in FASTA format by clicking on the “Choose File” button. If the reference genome has an incorrect format, an error message will pop out. The “Number of simulation replicates” item lets you choose how many simulations with the same parameter set but different random seeds you want to run; the integer you entered here will be logged under the “n_replicates” key in the “BasicRunConfiguration” section of the configuration file (Section 3.2.1).

- Note: The SARS-CoV-2 reference genome FASTA file needed to reproduce the example in Section 5.2 (as shown in the GUI screenshots in this section) is located at `tutorial/data/COVID/EPI_ISL_402124.fasta`.

Lastly, you could optionally specify a random seed, which will be used for all pre-simulation modules in the GUI, and will be logged under the “random_number_seed” key in the “BasicRunConfiguration” section of the configuration file (Section 3.2.1). Note that the random seed improves run-to-run reproducibility on a given setup, but your output may differ slightly from the output in Section 5.2 across operating systems, hardware, or software/library versions.

After configuring everything, make sure to click on the “Next” button to store all your choices in the configuration file. If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

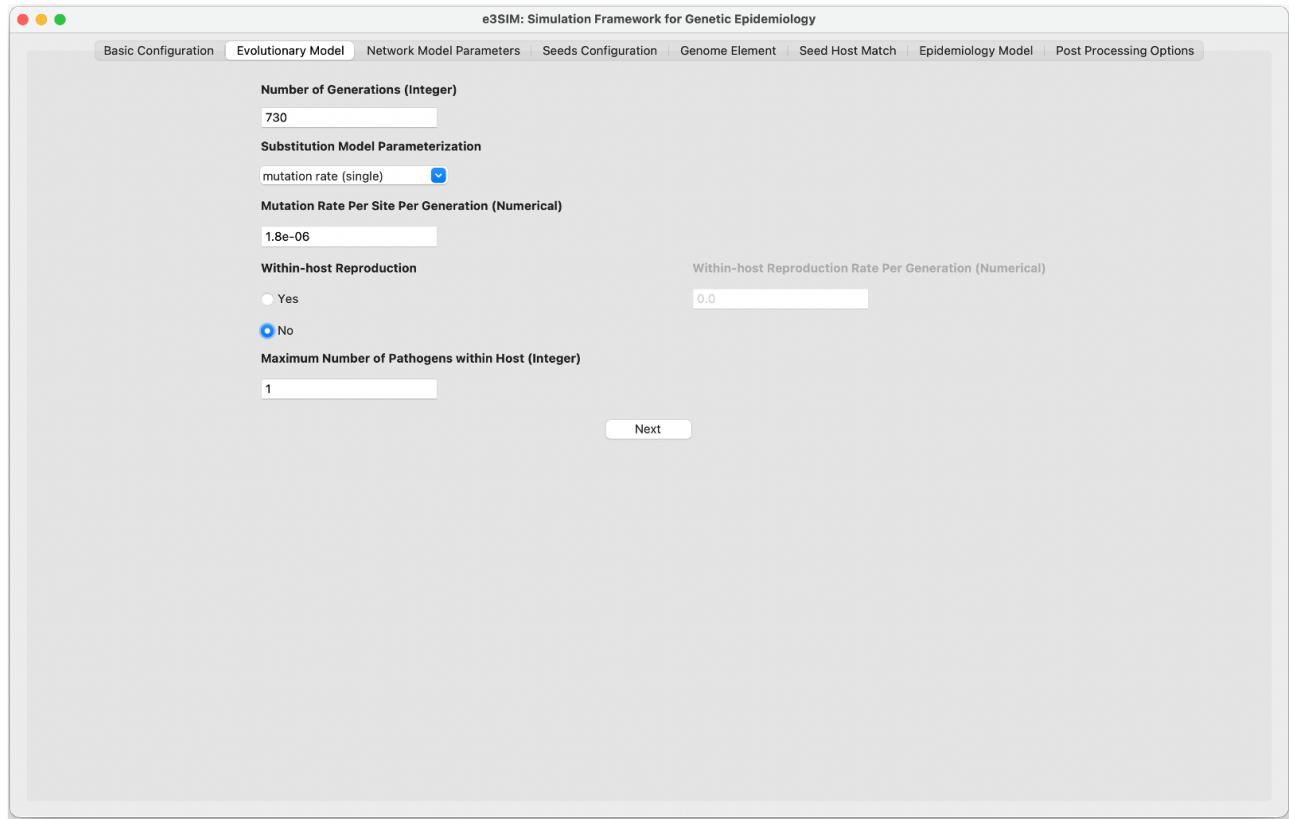


Figure 7.2: The second tab of the GUI: Evolutionary model.

In the “Evolutionary Model” tab (Figure 7.2), configurations for the evolutionary model and the within-host reproduction mode are set. This evolutionary model will be used in the main simulator module, and can also be directly loaded in the *SeedGenerator* module. Note that all the settings in this tab configure the *OutbreakSimulator* model, rather than the seed sequence generation process, which will be specified separately and executed in the fourth tab. The “Number of Generations” item lets you specify the number of ticks you will run for the simulation (see Section 4.1 for the definition of ticks).

You can use the drop-down menu to choose a substitution model: if you choose “mutation rate (single)”, an item will pop up to let you define a mutation rate per site per generation (tick) for the pathogen genome, and the substitution model will be default to Jukes-Cantor (see Section 4.1 for details); if you choose “mutation rate matrix” in the drop-down menu, a substitution probability matrix will pop up, as shown in Figure 7.2. The rows show the current allele in the order of ACGT, and the columns show the target mutated allele in the order of ACGT. For example, the number in the first row and third column specifies the probability of transition from an A allele to a G allele per tick. These settings will be logged in the “EvolutionModel” section of the configuration file.

You can choose whether to enable within-host reproduction. The “within-host reproduction rate” field becomes available only when you choose “Yes” (stored under the “within_host_reproduction” key of the configuration file), and will be stored under the “within_host_reproduction_rate” key of the configuration file. The maximum number of pathogens within a host can also be specified (stored under the “cap_withinhost” key of the configuration file).

After configuring everything, make sure to click on the “Next” button to store all your choices in the configuration file. If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

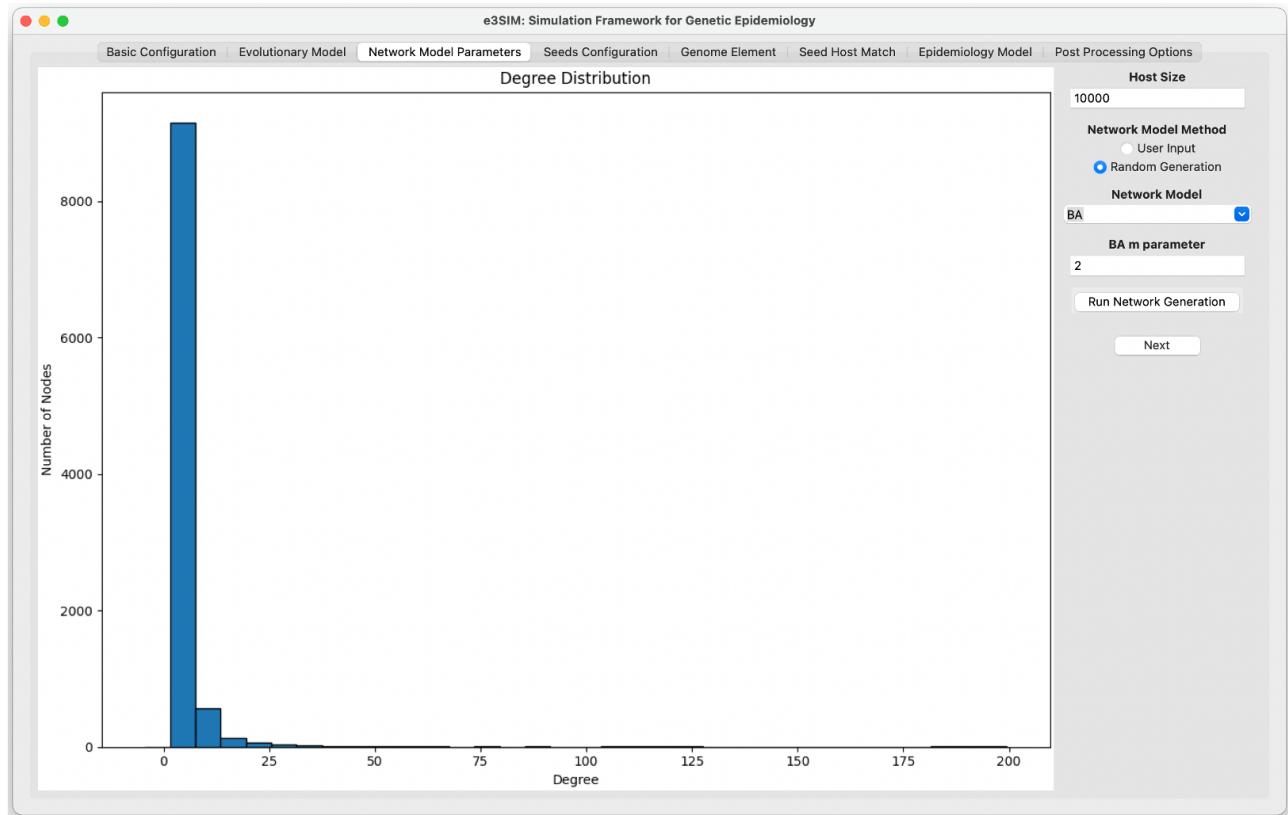


Figure 7.3: The third tab of the GUI: Network Model Parameters.

In the “Network Model Parameters” tab (Figure 7.3), configurations for the host population are set, and *NetworkGenerator* is run. The generated contact network will be used in the main simulator and the *Host-SeedMatcher* module. After setting all the parameters, clicking on the “Run Network Generation” button will generate a random network and visualize the network in the left panel. Please see Section 2.1 for all the modes of the host contact network generation.

Once a network model method is selected, corresponding variables required for the selected method will pop up. For example, in Figure 7.3, the user specifies a random generation method for the network and specifies a Barabási–Albert graph, so that the “BA probability parameter” is enabled. After running the network generation and viewing the degree distribution to make sure this is the contact network desired, click on the “Next” button to proceed and store all your choices in the “NetworkModelParameters” section of the configuration file.

If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

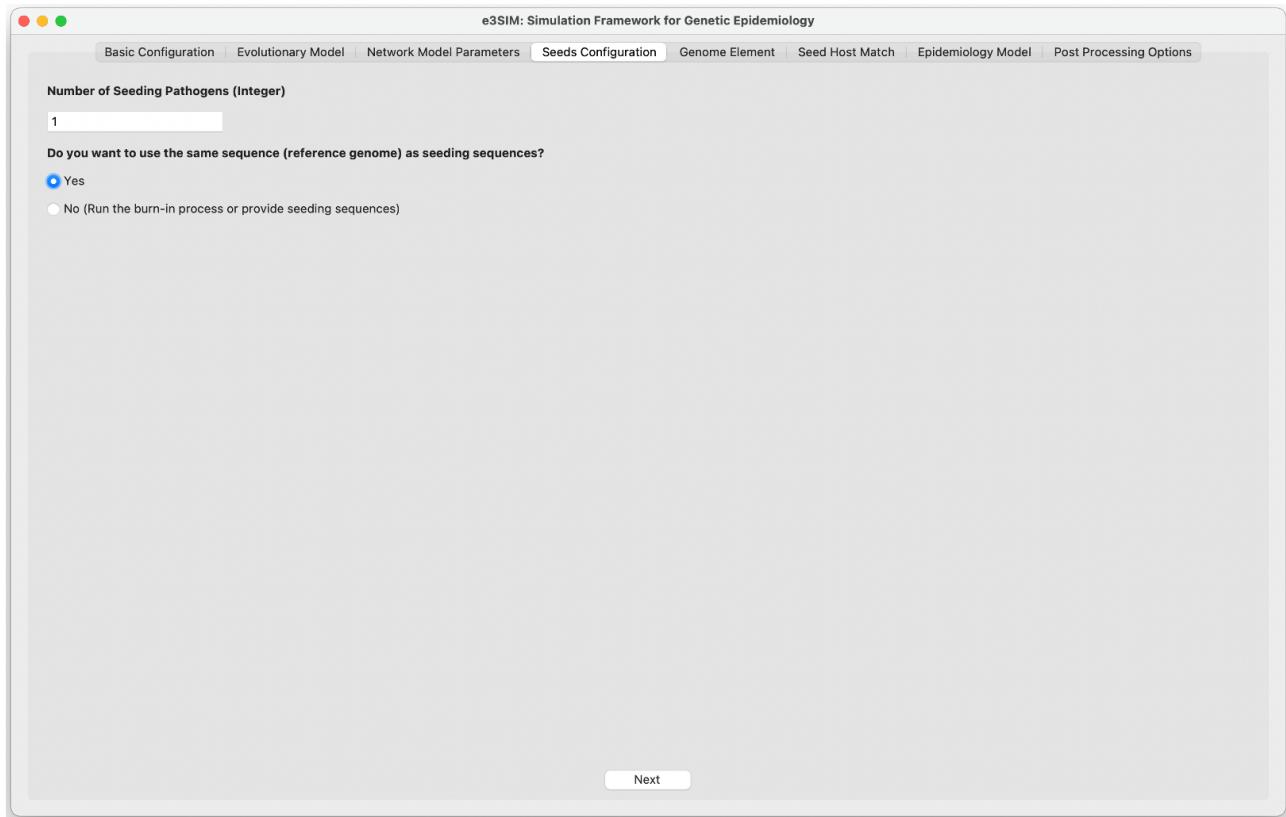


Figure 7.4: The fourth tab of the GUI: Seeds Configuration.

In the “Seeds Configuration” tab (Figure 7.4), configurations for the seeding sequences are set, and *SeedGenerator* is run. The generated seeding sequences will be used in the main simulator, the *GeneticEffectGenerator* module and the *HostSeedMatcher* module. After setting all the parameters, clicking on the “Run Seed Generation” button will generate seeding sequences and their genealogy to store in the working directory. Please refer to Section 2.2 for how and why these parameters should be specified.

The mutation specifications could be different from the main simulator specifications specified in Tab 2, but can also be directly imported from Tab by clicking on “Load mut.rate(s) from Evolutionary Model”. Since this step includes running a SLiM simulation, it might take some time to finish, during which the GUI will be frozen. A pop-up window indicating the completion of the seed generation will appear once finished.

After configuring everything, make sure to click on the “Next” button to store all your choices in the configuration file. Settings here will be stored in the “SeedsConfiguration” section of the configuration file, and only the “Number of seeding pathogens” will be used to run *OutbreakSimulator*, while all other settings are just for generating the seeding sequences and are irrelevant to the actual outbreak simulation.

If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

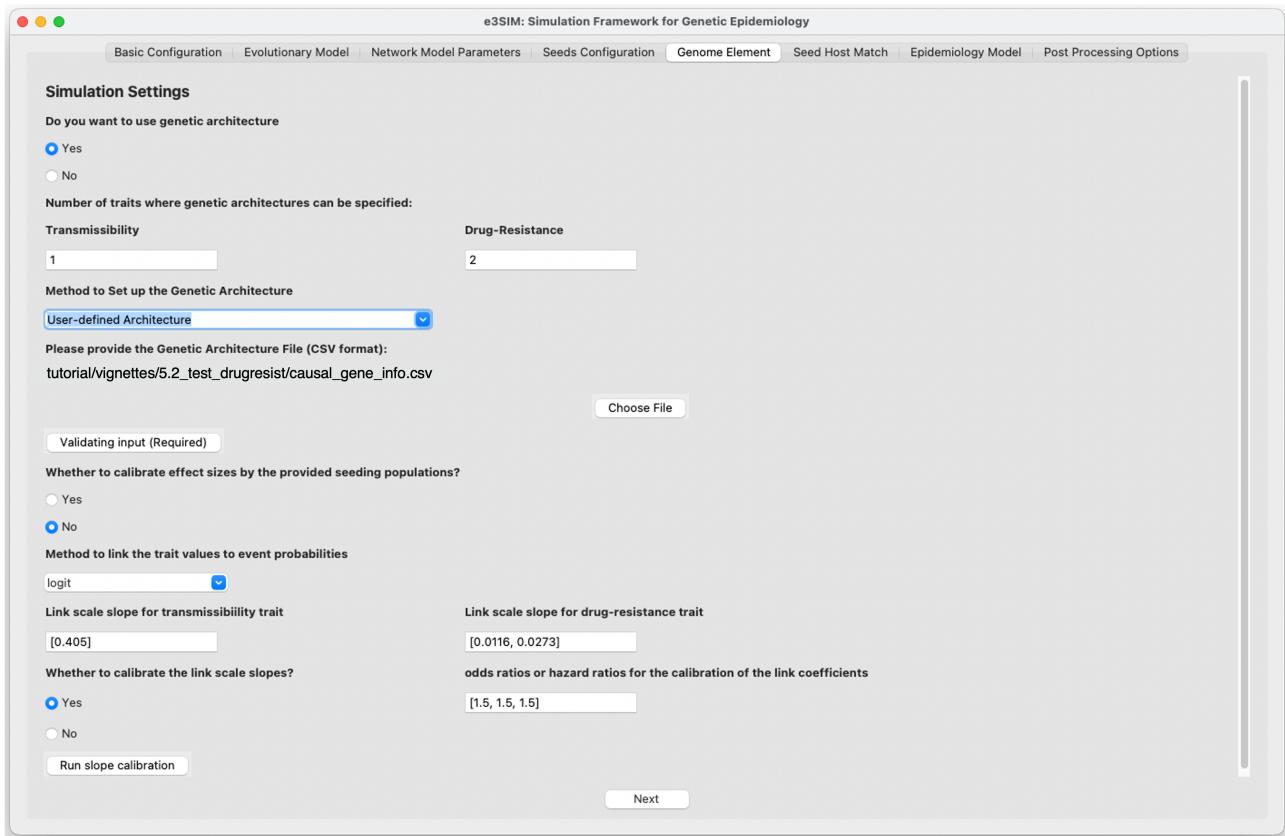


Figure 7.5: The fifth tab of the GUI: Genome Element.

In the “Genome Element” tab (Figure 7.5), the genetic architecture is set, and *GeneticEffectGenerator* is run to calculate and store the trait values of the seeding sequences. The genetic architecture is stored in the working directory. Please refer to Section 2.3 for how and why these parameters should be specified.

If you choose “No” for the question “Do you want to use genetic architecture”, all the subsequent options will disappear from the GUI, and your simulation will be a neutral simulation where mutations do not affect the epidemiological process.

If you choose “Yes”, you will be prompted to set up the genetic architecture for transmissibility and drug-resistance. Users can choose the method to set up the effect sizes, and provide either the candidate regions or a user-provided genetic architecture file. For the random generation mode, expected fractions and dispersion of the fraction need to be provided.

- Note: The genetic architecture file `causal_gene_info.csv` needed to reproduce the example in Section 5.2 (as shown in the current GUI screenshots) using the “User-defined Architecture” mode is located in `tutorial/vignettes/5.2_test_drugresist`.

If users want to calibrate the effect sizes, they should select “Yes” for the “Whether to calibrate effect sizes by the provided seeding populations”, and click on “Run effect size generation” again to calibrate the effect sizes. The link-scale slope has to be set for all traits. Users can click on the “Run slope calibration” button at the bottom of the page, and the calibrated slope alpha will be shown in the terminal. Users will have to enter the alpha(s) into the GUI manually.

If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

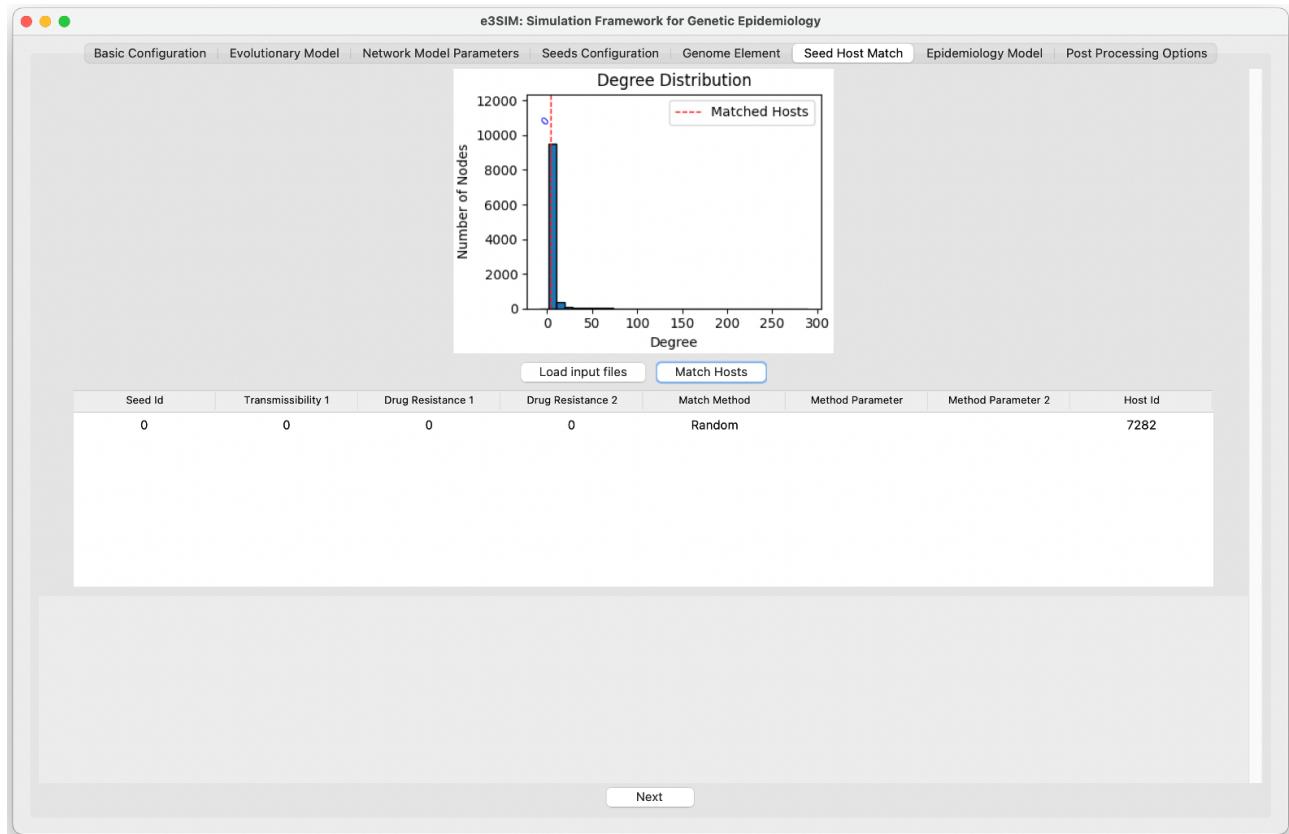


Figure 7.6: The sixth tab of the GUI: Seed Host Match.

In the “Seed Host Match” tab (Figure 7.6), the generated or input network is loaded (by clicking “Load input files”), and HostSeedMatcher is run to match the seeding sequences to the hosts’ IDs. The matching file is stored in the working directory. The table in the panel below the degree distribution plot shows the trait values of each seeding sequence, using the sequence generated in the “Seeds Configuration” tab and the genetic architecture defined in the “Genome Element” tab. For each seed, one matching method can be chosen by double-clicking on the cell in the “Match Method” column.

All the seeds default to random matching, which matches seeds to a random host from the host contact network. However, you can change the method to “ranking” or “percentile” by double-clicking on the drop-down menu for each seed in the “Match Method” column. By choosing methods different from random, parameters are required for each method. Please refer to Section 2.4 for details on how to set these parameters. They should be typed in the “method parameter” columns.

After clicking on the “Match Hosts” button, the “Host id” column will be filled with the matched hosts’ IDs according to the method you chose, and the contact degree of the matched hosts will be plotted on the above panel as vertical lines in the degree distribution plot. You can adjust the methods and parameters to re-match until you are satisfied with the matching.

After configuring everything, make sure to click on the “Next” button to store all your choices in the configuration file. If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

Basic Configuration Evolutionary Model Network Model Parameters Seeds Configuration Genome Element Seed Host Match Epidemiology Model Post Processing Options

Compartmental Model

SEIR None selected Choose File

Number of Epochs (Integer) 3 [250, 500]

Evolutionary Components Setting

Which Transmissibility Trait to Use for Each Epoch (List integer) [1,1,1]

Which Drug-resistance Trait to Use for Each Epoch (List Integer) [0,1,2]

Transition Probabilities between Compartments

Transmission Prob. β (List Numerical) [0.04, 0.02, 0.02] Latency Probability ζ (List Numerical) [1, 1, 1]

Activation Prob. ν (List Numerical) [0.3, 0.3, 0.3] Latent Recovery Prob. τ (List Numerical) [0, 0, 0]

De-activation Prob. ϕ (List Numerical) [0, 0, 0]

Active Recovery Prob. γ (List Numerical) [0.03, 0.06, 0.05]

Immunity Loss Prob. ω (List Numerical) [0.05, 0.1, 0.1]

Base Survival under Treatment Prob. s (List Numerical) [0.01, 0.96, 0.96]

Sample Prob. e (List Numerical) [0.0002, 0.0003, 0.0003]

Post-sampling Recovery Prob. δ (List Numerical) [0, 0, 0]

Massive Sampling Events

How Many Massive Sampling Events Will Be Simulated (Integer) 0

In Which Generation (s) Will the Massive Sampling Events be Simulated (List Integer) []

Probability of Being Sampled for Every Pathogen Genome in Each Massive Sample Event (List Numerical) []

Probability of Recovering Once Sampled in Each Massive Sampling Event (List Numerical) []

Enable Super-infection Events? Yes No

Next

Figure 7.7: The seventh tab of the GUI: Epidemiology Model, with a valid example of values entered.

In the “Epidemiology Model” tab (Figure 7.7), the configuration of the epidemiological model of the simulation is set. The epoch structure is also specified. Please refer to Section 4.4 for the correct format of these parameters.

After configuring everything, make sure to click on the “Next” button to store all your choices in the configuration file. If the “Next” button does not appear in the interface, please switch to full-screen mode.

Chapter 7 Introduction to the GUI

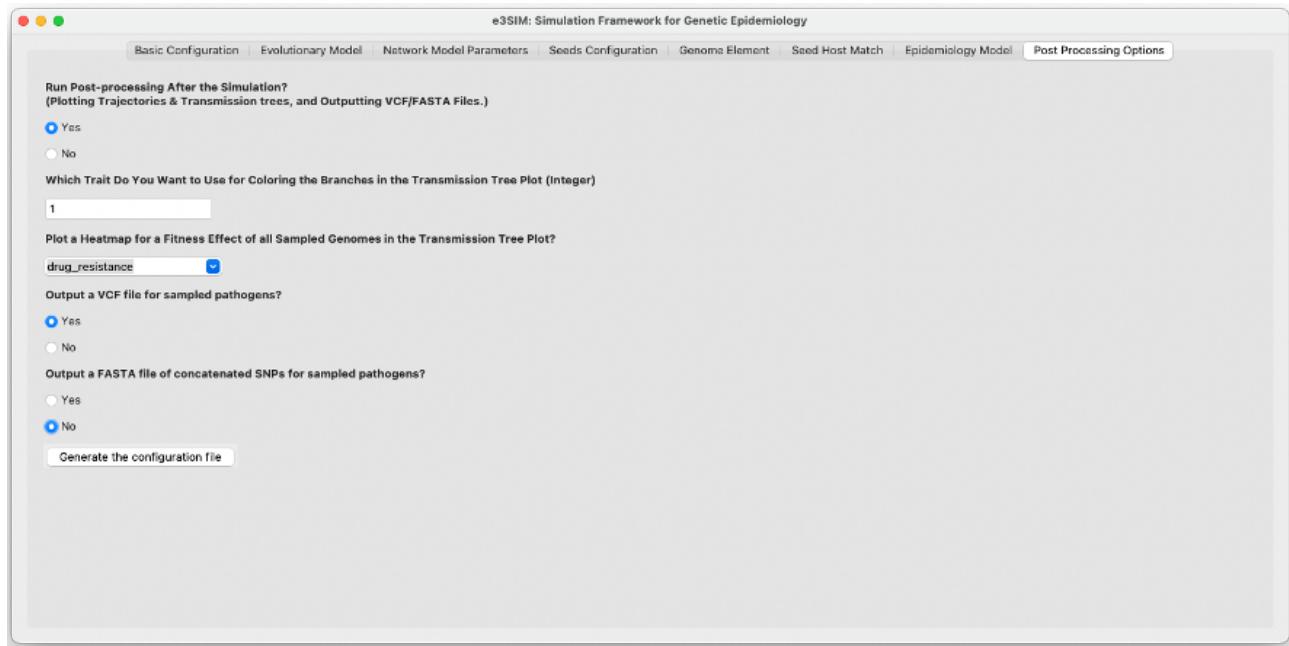


Figure 7.8: The eighth tab of the GUI: Post Processing Options.

In the “Post Processing Options” tab (Figure 7.8), the post-processing steps after the simulation are configured. For the question “Which trait do you want to use for coloring the branches in the transmission tree plot”, if “0” is specified, the branches will be colored by seed ID. If “1” is specified, the first trait value will be used. If three traits are specified, where two are transmissibility and one is drug resistance, then “1” refers to the first transmissibility trait, and “3” refers to the drug-resistance trait. Please see Part III and Section 3.2.7 for more details on the post-processing options.

Please make sure to click on the “Generate the configuration file” button to generate the full configuration file in the working directory before you exit the GUI.

It is recommended that first-time users do not close the GUI before making sure every entry is correct, since the GUI does not perform validity checks for all entries. The complete validity check will be performed when you run OutbreakSimulator. You can go back to previous tabs to make edits and regenerate the configuration file as many times as you wish.

Part V

Advanced usage

Chapter 8

SLiM code of OutbreakSimulator

The main simulator module of e3SIM uses SLiM 4 in the backend. In `OutbreakSimulator`, e3SIM assembles Eidos code blocks to generate a customized script for the simulation configuration specified by the user and executes it. Advanced users can modify these code blocks to implement their own model. All the Eidos code blocks are located in the `slim_scripts` directory. For instructions on how to write and modify Eidos code and for the logic of SLiM, please refer to the [SLiM tutorial](#). We describe how e3SIM assembles the code blocks below, so that experienced users can modify the code accordingly.

Note that the random seed for the SLiM simulations can be found in SLiM's log file, `slim.stdout`, in each replicate's directory after the simulation finishes.

Chapter 8 SLiM code of OutbreakSimulator

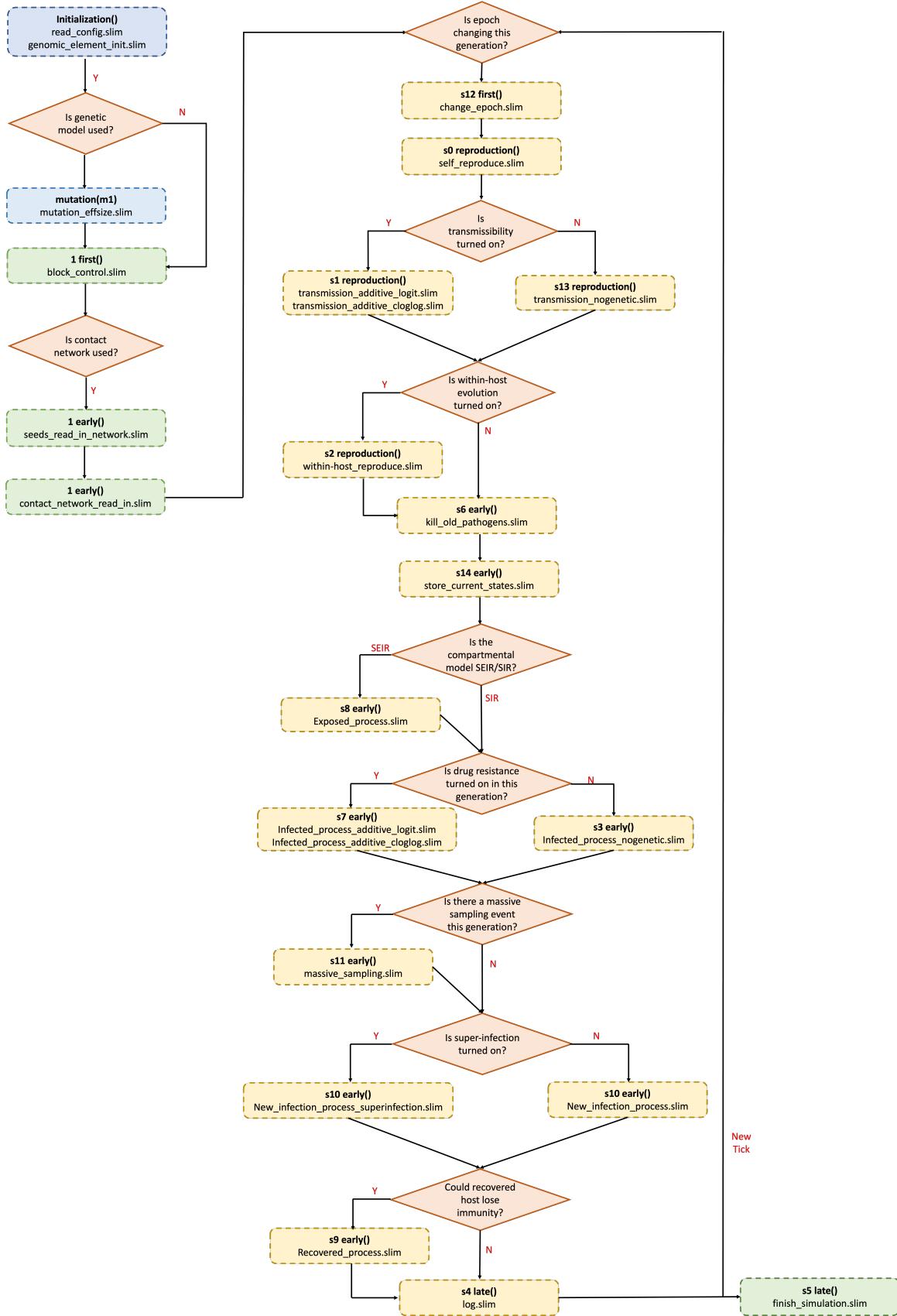


Figure 8.1: Flowchart showing the logic of code-block assembly for **OutbreakSimulator**