

Synapses, neurons and brains

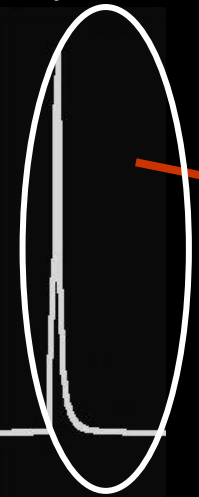
Idan Segev



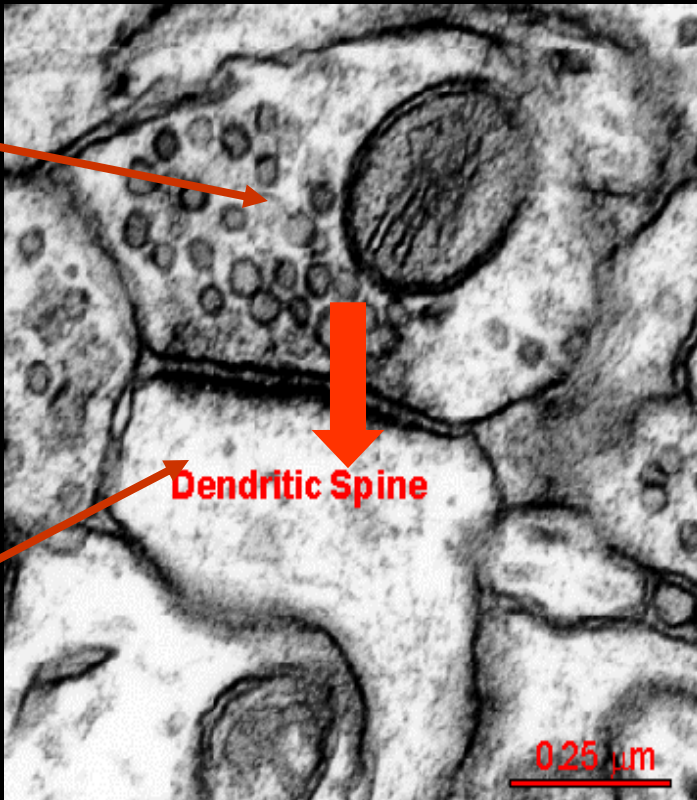
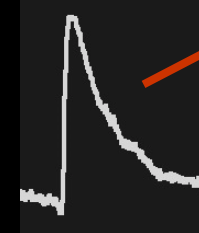
Lesson #4

Electrifying brains – active electrical signals *the spike*

Spike in axon



Syn. Poten.
in dendrite



The excitable (“spiking”) axon .1

The Hodgkin & Huxley experiments .2

Space clamp and voltage clamp .3

Membrane conductances/currents .4
underlying the spike

The H&H model for spike initiation .5

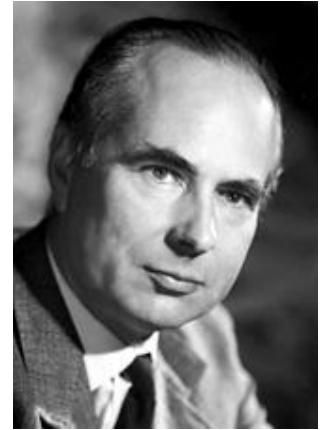
Spike propagation in axons .6

From synapses to spikes .7

THE SPIKE - Hodgkin and Huxley (H&H) – NOBEL 1963)

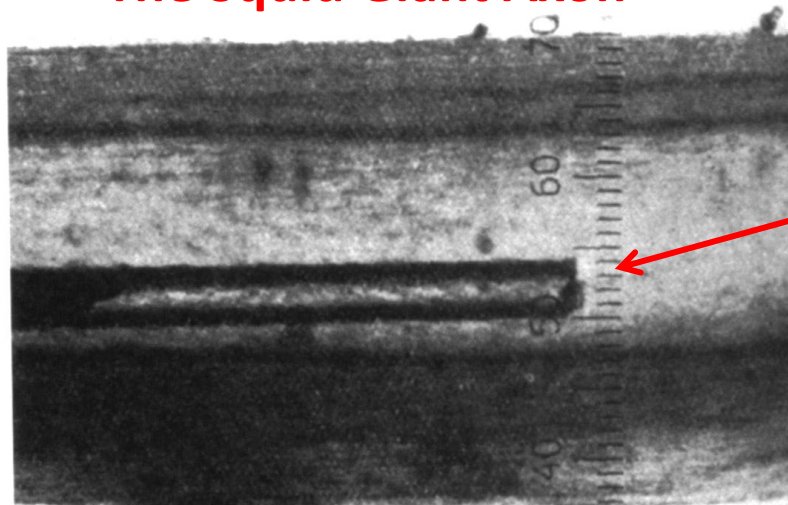


**Sir Alan Lloyd
Hodgkin**



**Sir Andrew Fielding
Huxley**

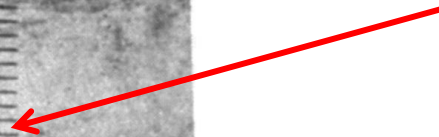
The Squid Giant Axon



~ 0.5 mm



Axial electrode

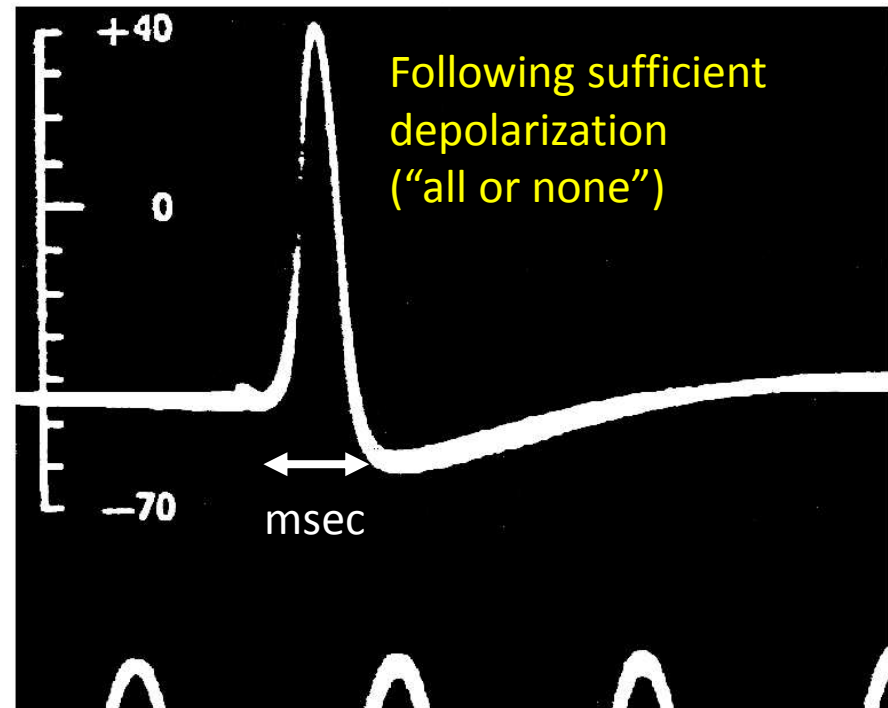


First direct (intracellular) recorded **action-potential (spike)**
in the **squid giant axon** (*Hodgkin and Huxley, 1939*)



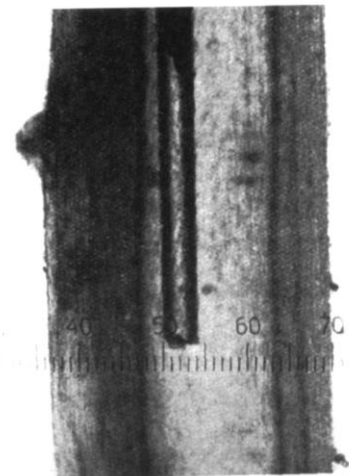
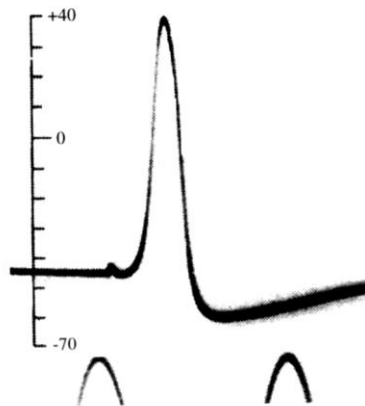
(B)

100 mV



A

B

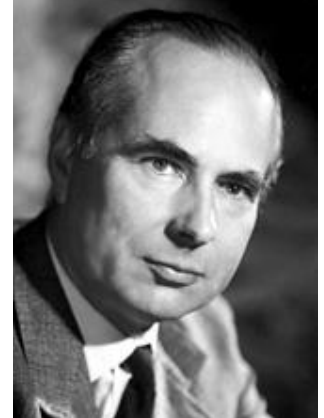
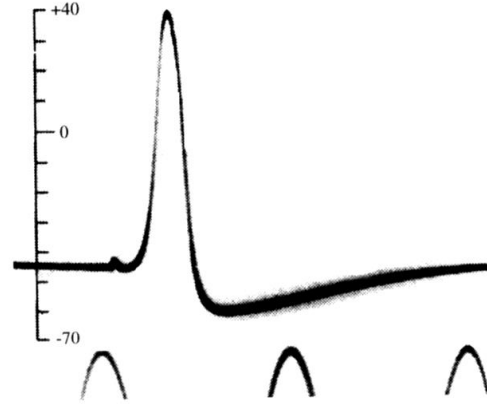
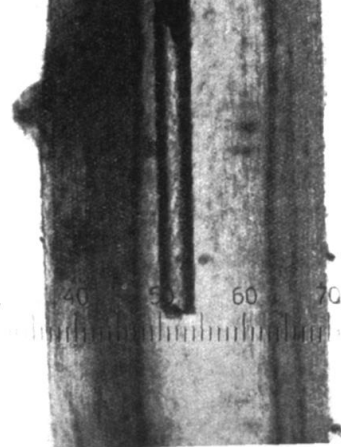


The H&H equations for spike initiation

The triumph of theory



Sir Alan Lloyd
Hodgkin



Sir Andrew Fielding
Huxley

$$I = C_m \dot{V} + g_{Na} h m^3 (V - V_{Na}) + g_K n^4 (V - V_K) + G_L (V - V_L) \quad (1)$$

$$\frac{d}{dt} m = \alpha_m (V) (1 - m) - \beta_m (V) m \quad (2)$$

$$\frac{d}{dt} n = \alpha_n (V) (1 - n) - \beta_n (V) n \quad (3)$$

$$\frac{d}{dt} h = \alpha_h (V) (1 - h) - \beta_h (V) h \quad (4)$$

The “all or none” nature of the spike

Magnitude of depolarizing current stimuli

CURRENT-VOLTAGE RELATION IN NERVE

Supra-threshold

RESULTS

Stimulation with brief currents

investigating the effect of a constant voltage it was important that the membranes studied were capable of giving normal responses. This was done by applying a brief shock to one internal electrode. The recording

Sub-threshold

38



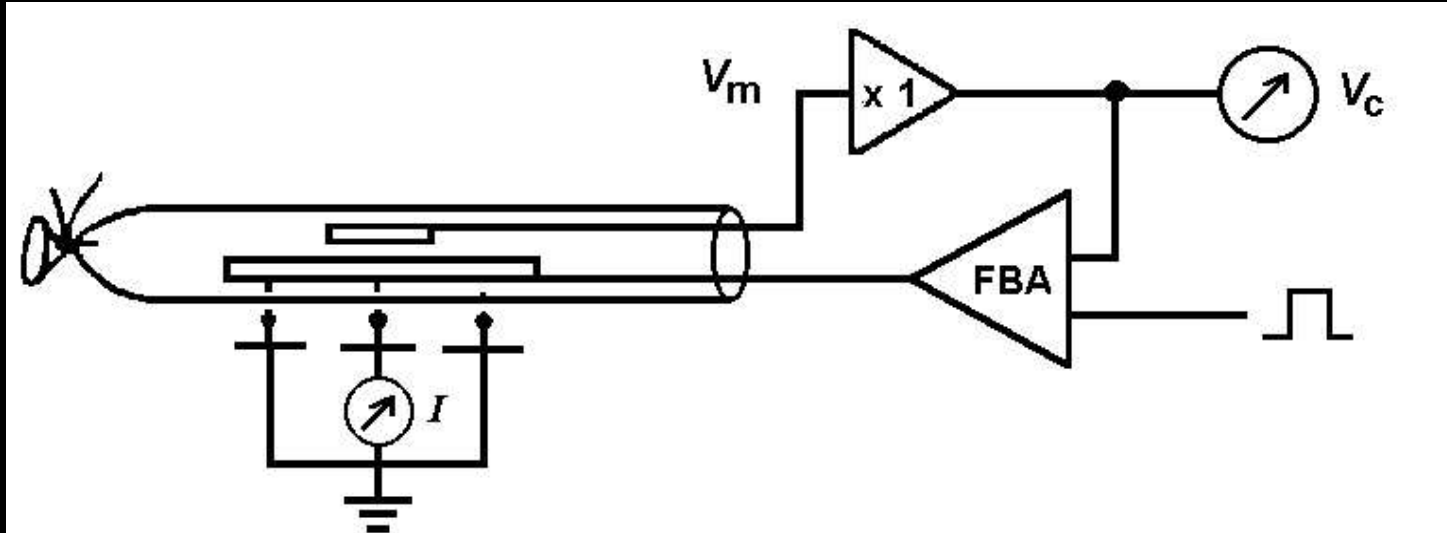
are given on p. 431; typical results are shown in Fig. 8 (23°

Hodgkin, Huxley and Katz, 1952

What are the membrane currents
underlying the spike?

The voltage clamp (+ space clamp)

The technique that made the whole difference



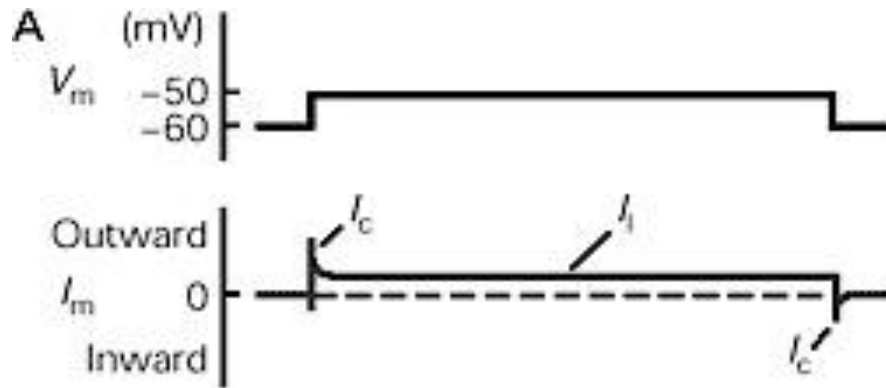
Space clamp - making the (long) axon effectively **isopotential** via the insertion of an axial conductive wire.

Voltage clamp – enables the experimenter to dictate the desired voltage difference between the inside and the outside of the membrane.

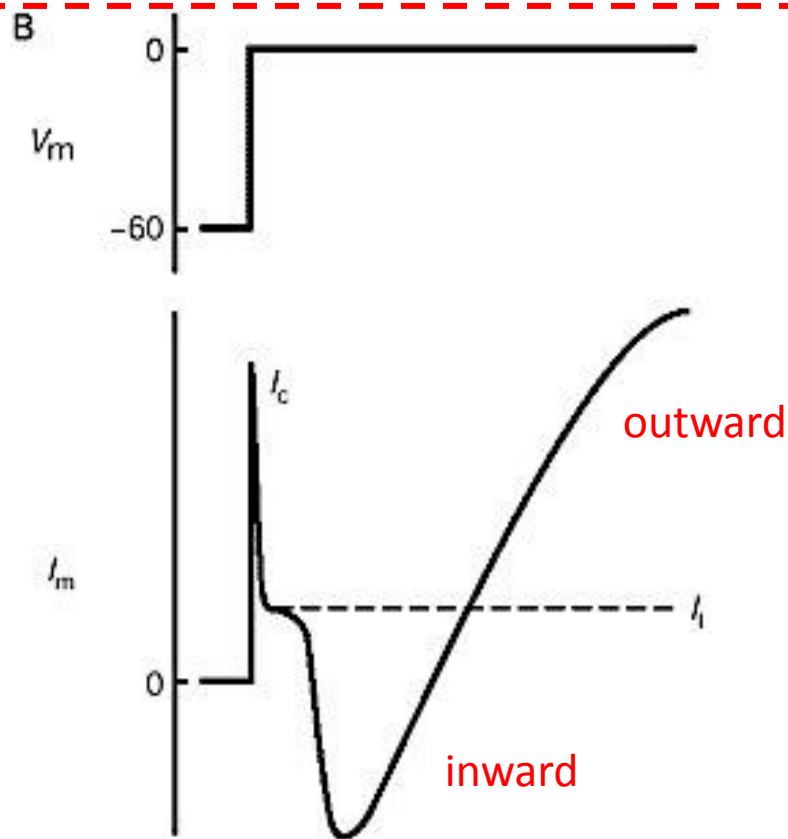
The electronic feedback system injects current to exactly counterbalance the (**excitable – voltage-dependent**) membrane current.

Developed by Kenneth Cole and George Marmont

Membrane current in response to voltage clamp (VC)



For **subthreshold** depolarizing voltage clamp, the recorded membrane current is the current that flows via the leak (passive) conductance + a small capacitive current (at start and end of the VC)

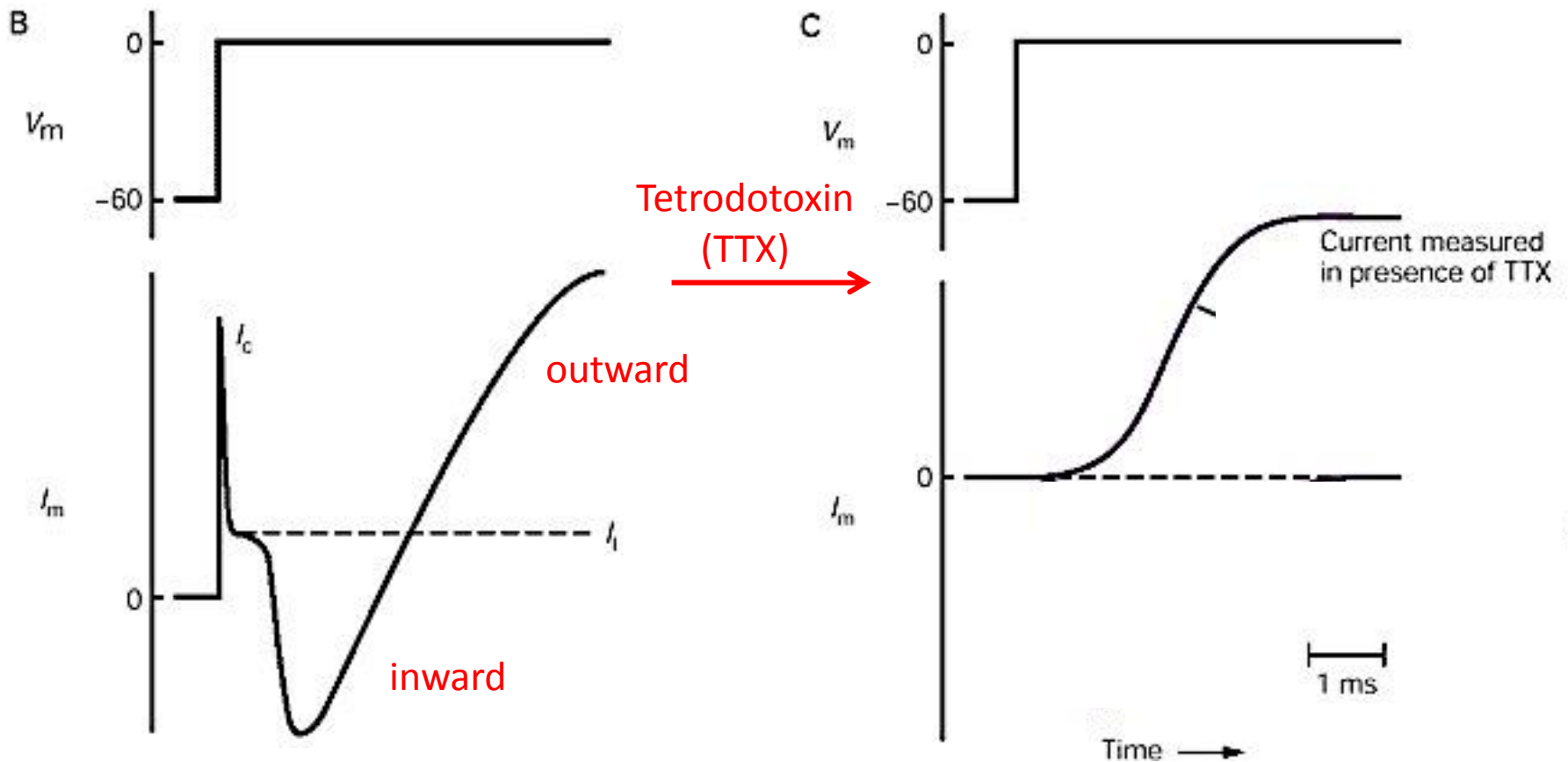


For **suprathreshold** depolarizing voltage clamp, the recorded membrane current (after the fast capacitive current) flows **first inwards** (into the axon) and later **outward** (from inside to the outside)

Separating voltage-dependent active (excitable) currents Using pharmacological agents

2 different currents flow via the membrane during the spike

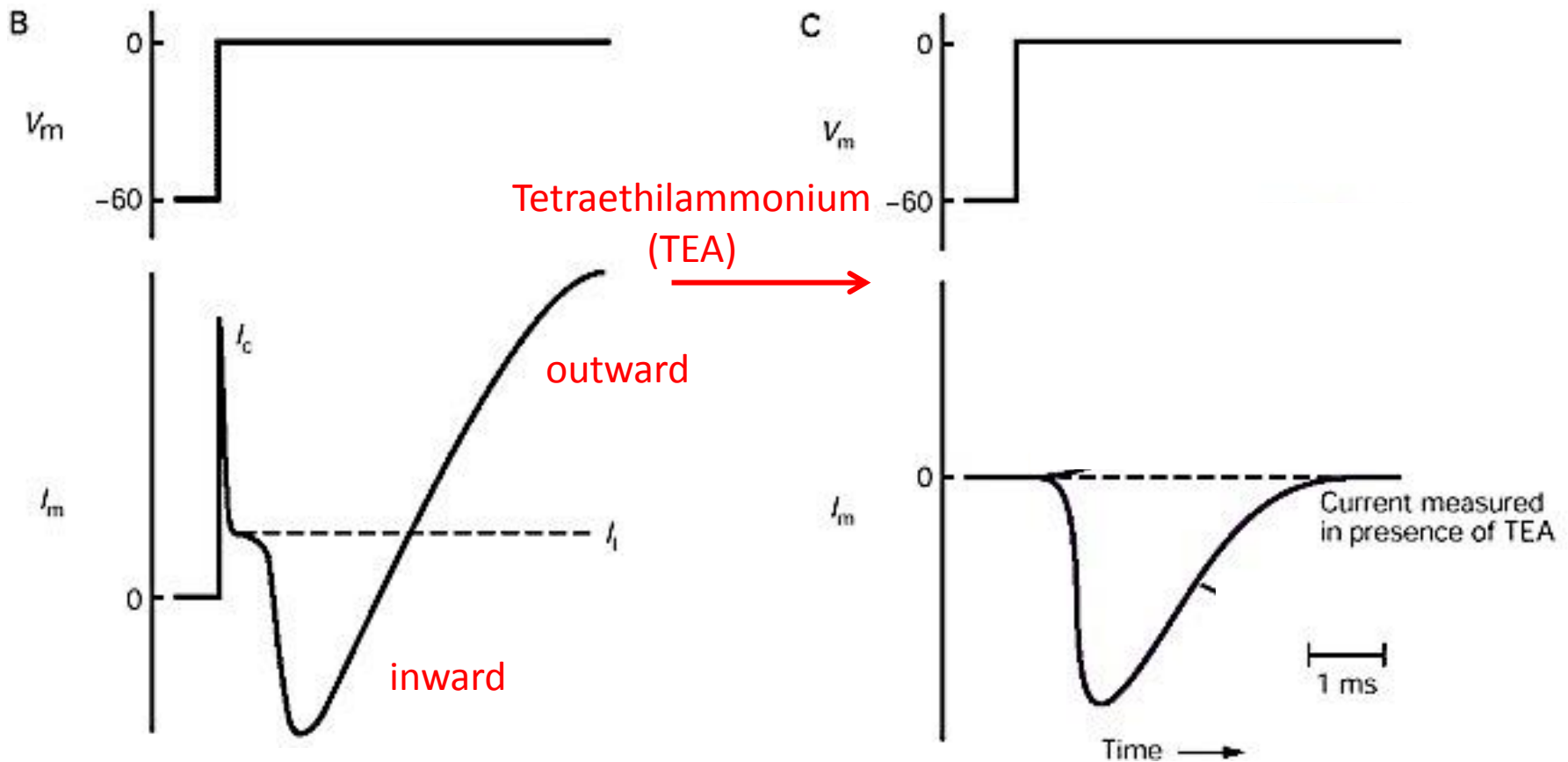
TTX blocks the early – fast inactivating current



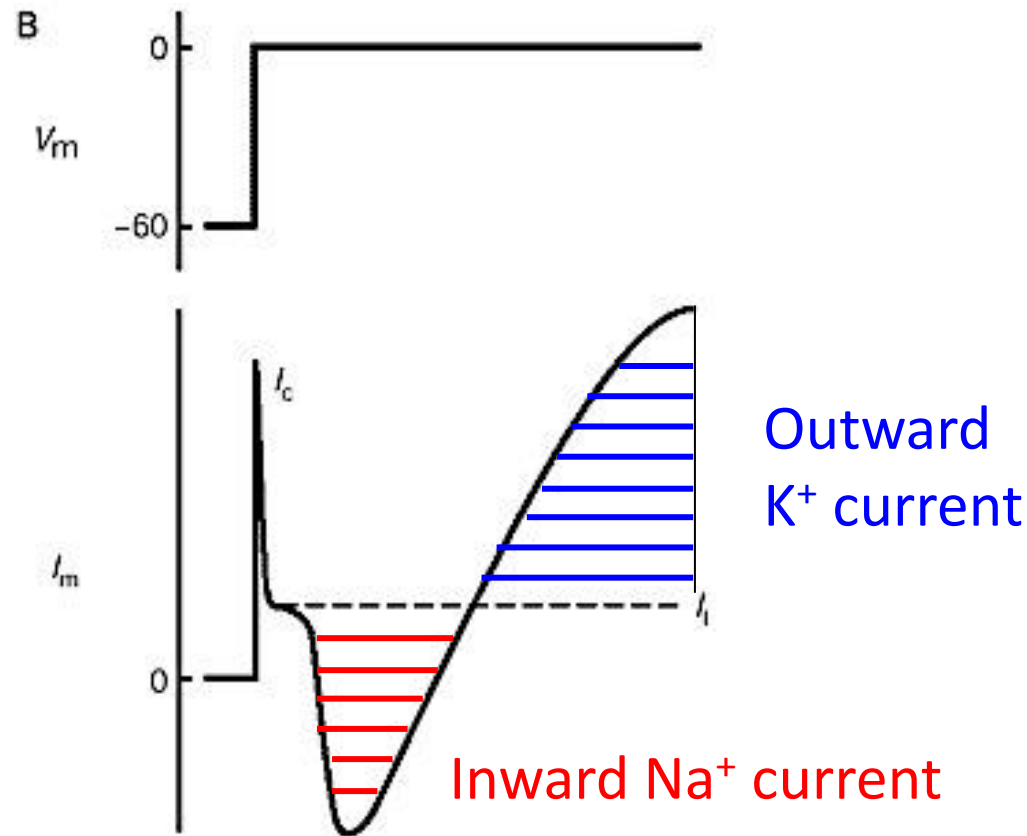
Separating voltage-dependent active (excitable) currents Using pharmacological agents

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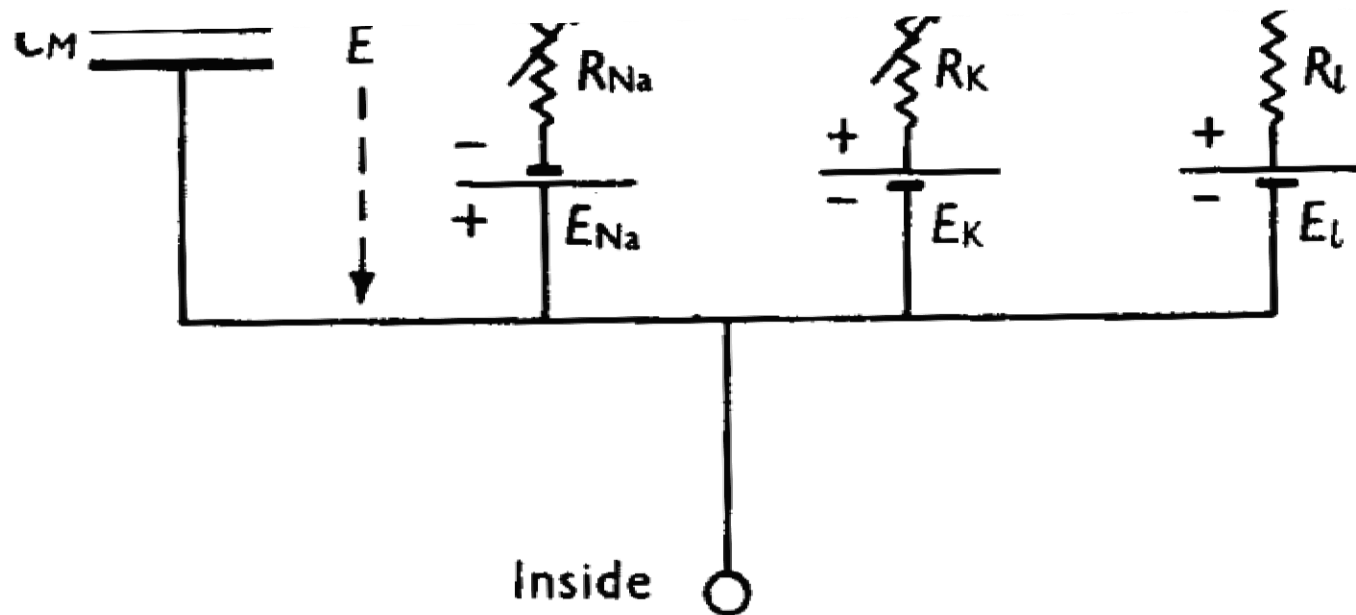
TEA blocks the late – slow non-inactivating current



Changing ion concentration at bath with giant axon showed that early current is carried by Na^+ ions and late one by K^+ ions



The electrical circuit for the squid axon (excitable) membrane



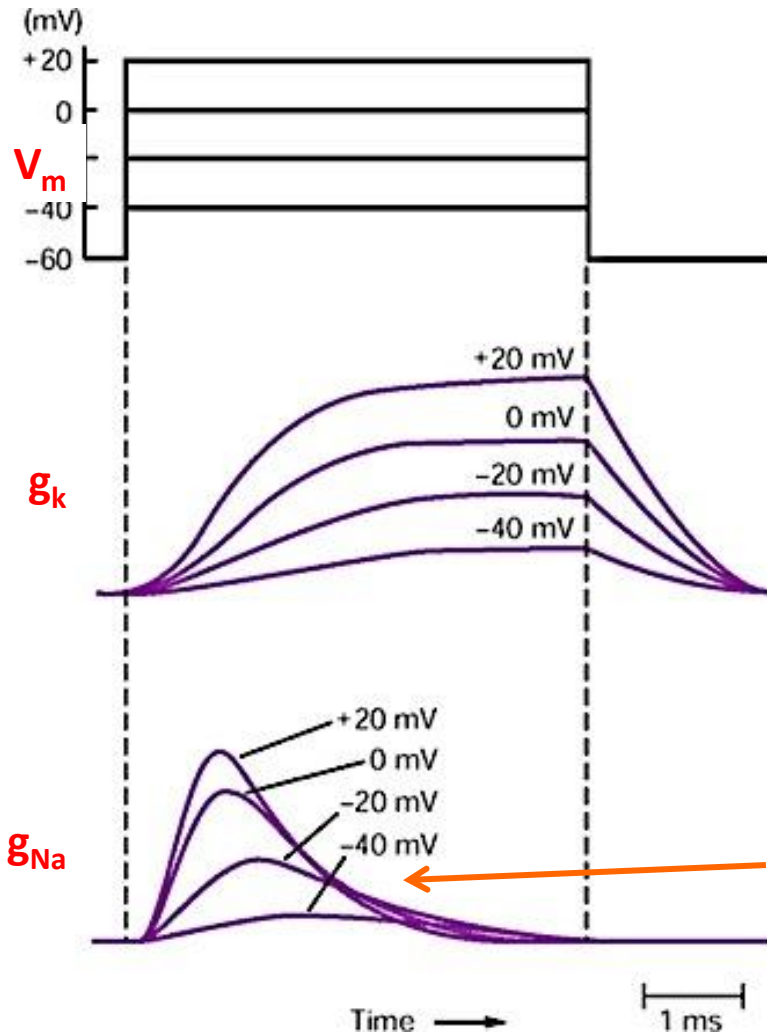
Electrical circuit representing membrane. $R_{Na} = 1/g_{Na}$; $R_K = 1/g_K$; $R_l = 1/\bar{g}_l$. R_{Na} and R_K vary with time and membrane potential; the other components are constant.

The nature of the permeability changes

represent the thickness and composition of the excitable membrane are not known. Our experiments are therefore unlikely to give any certain information about the nature of the molecular events underlying changes in permeability. The object of this section is to show that certain types of theory are

*Ion currents (K^+ and Na^+) for various depolarizing voltage clamp
(and extracting respective ion conductances)*

$$I_K = g_K (V_m - E_K); \quad I_{Na} = g_{Na} (V_m - E_{Na})$$

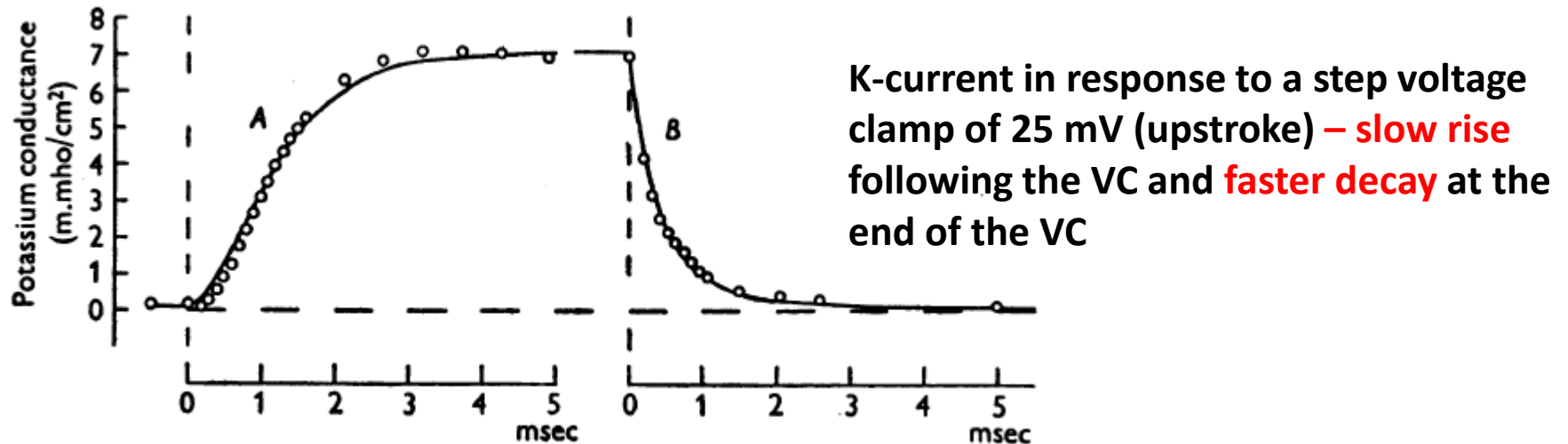


1. The slow (K) current (conductance) does not inactivate during VC

2. The K conductance rises slower than it decays at end of VC

3. The fast (early) Na conductance inactivates during VC

Fitting an equation for the K current (K-conductance) during/following VC



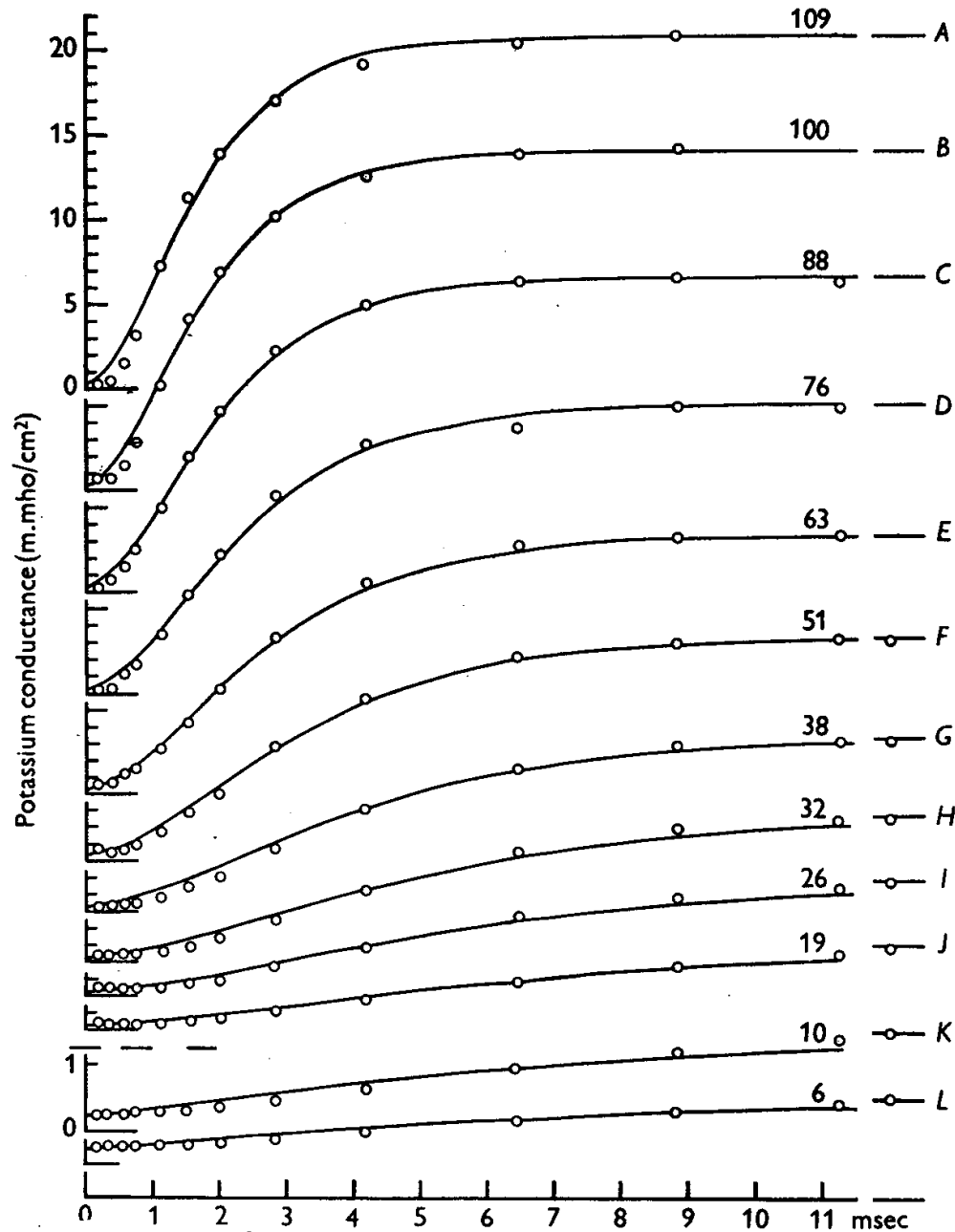
Mathematically – the rising phase of K-current can be described as a power of 4 (namely as $(1 - \exp(-t))^4$ and the decay as $\exp(-4t)$)

$$g_K = \bar{g}_K n^4$$

n represents the proportion of K-ion channels in the open state

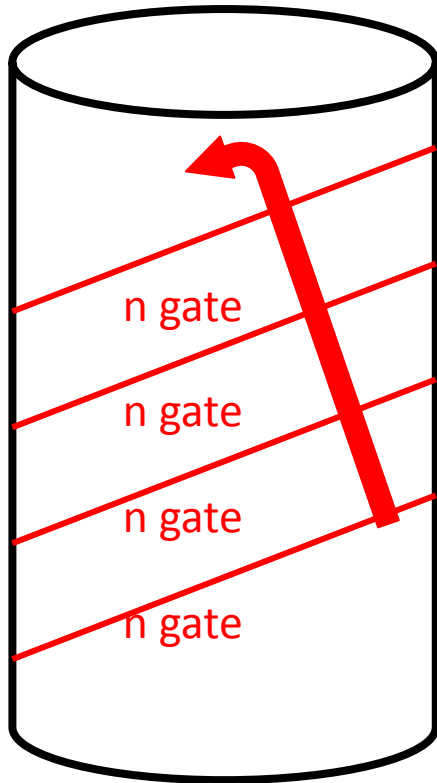
"These equations may be given a physical basis if we assume that potassium ions can only cross the membrane when four similar particles occupy a certain region of the membrane..." Hodgkin AL, Huxley AF. 1952 J Physiol (Lond) 117:500–544

Fitting K current for different VC depolarizing values



Graphical interpretation of H&H model for the K channel

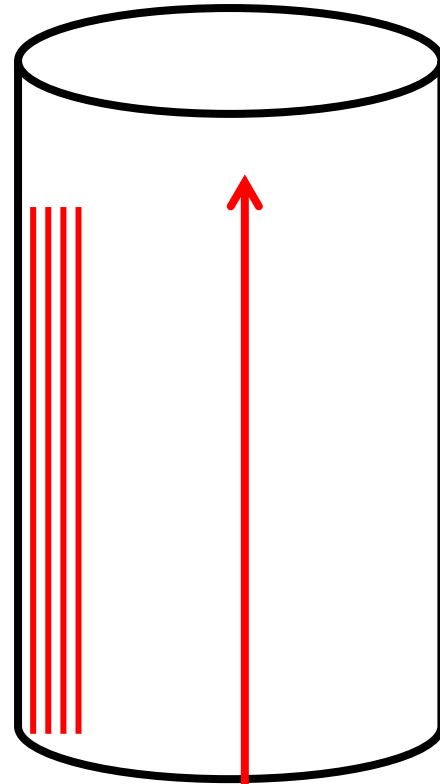
Closed K channel (by 4 n gates)



K^+
INSIDE

4 n gates open with
depolarization

Open K channel (by 4 n gates)



K^+
INSIDE

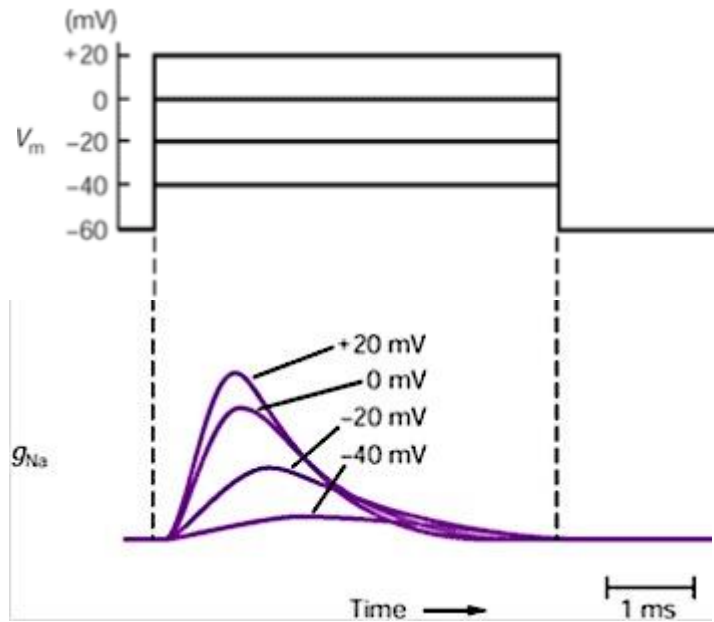
The activation function, n , and the rate functions α_n and β_n

fer in the opposite direction
| increase and β_n should decrease

equations will be discussed in

can only cross the membrane when some particles occupy a certain region of the membrane. n represents the proportion of the particles in a certain position (for example at the inside of the membrane) and $1-n$ represents the proportion that are somewhere else (for example at the outside of the membrane). α determines the rate of transfer from outside to inside, while

Similar procedure is used to extract the activation (m) and inactivation (h) parameters for the Na current

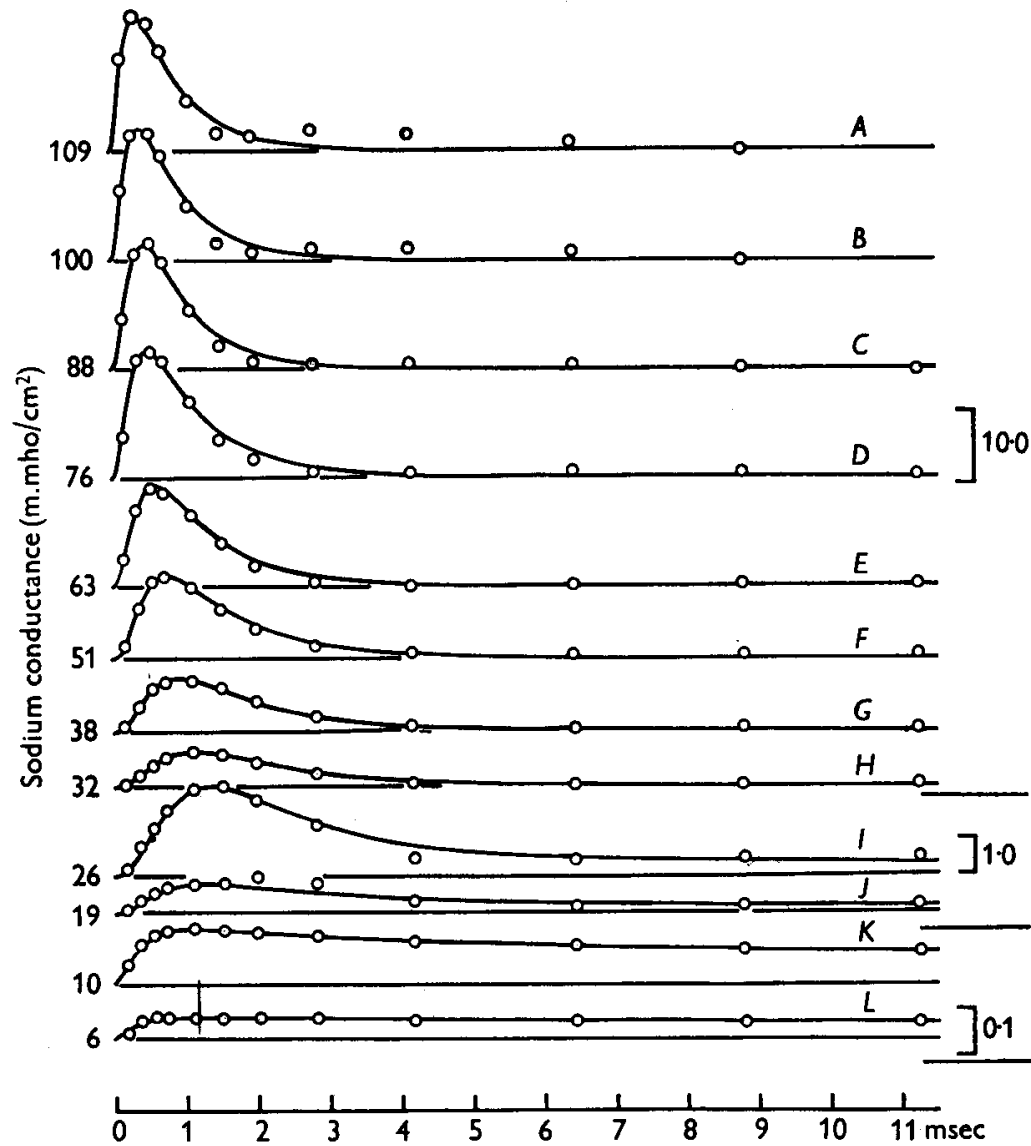


$$g_{Na} = m^3 h \bar{g}_{Na},$$

$$\frac{dm}{dt} = \alpha_m (1 - m) - \beta_m m,$$

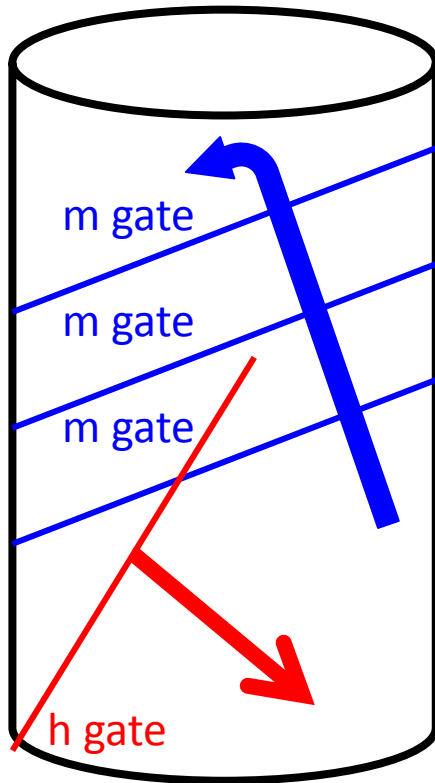
$$\frac{dh}{dt} = \alpha_h (1 - h) - \beta_h h,$$

Fitting Na current for different VC depolarizing values



Graphical interpretation of H&H model for the Na channel

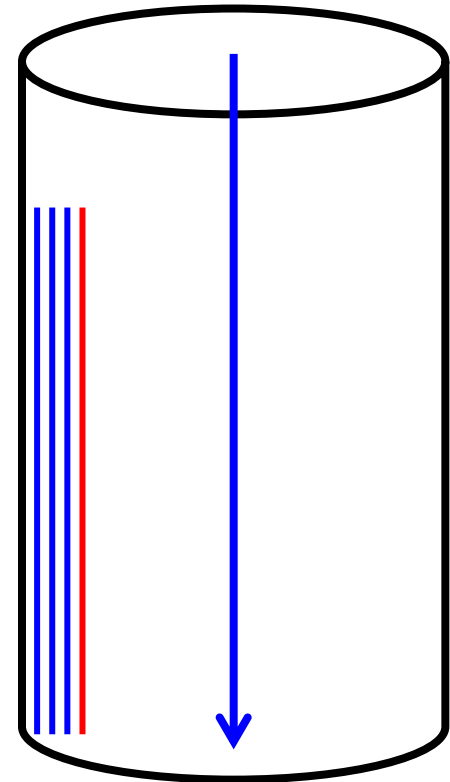
Na channel (by 3 activated m gates and 1 inactivated h gate)



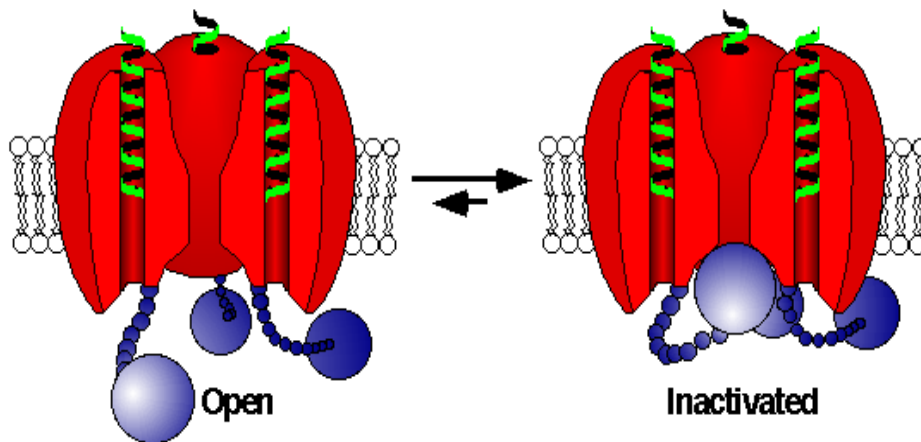
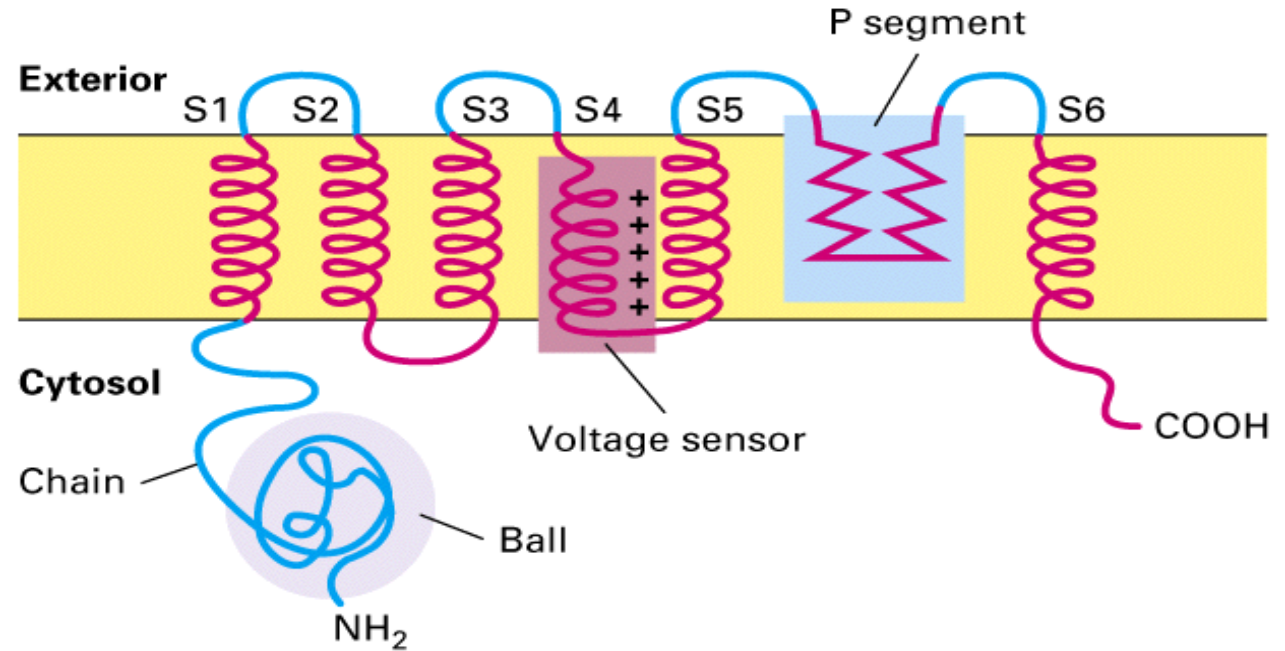
3 (fast) m (activated) gates open with depolarization

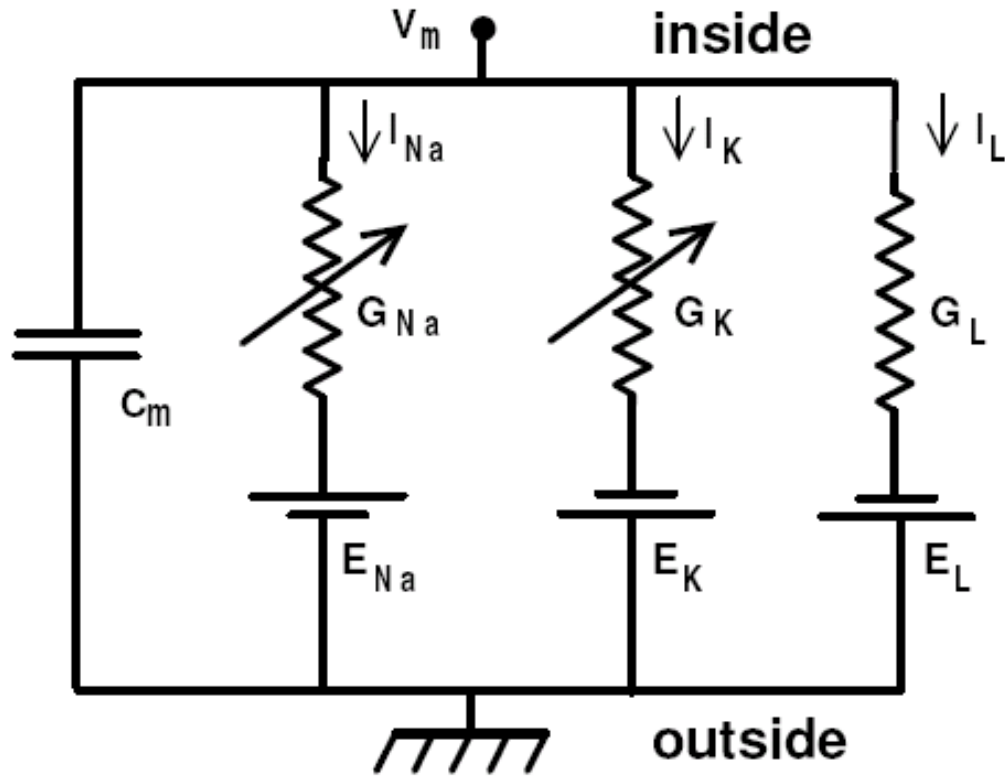
1 (slow) h (inactivated) gate closes with depolarization

Open Na channel
Na outside



Voltage-dependent Ion channels underlying the AP (later than H&H)





$$I = C_m \dot{V} + g_{Na} h m^3 (V - V_{Na}) + g_K n^4 (V - V_K) + G_L (V - V_L) \quad (1)$$

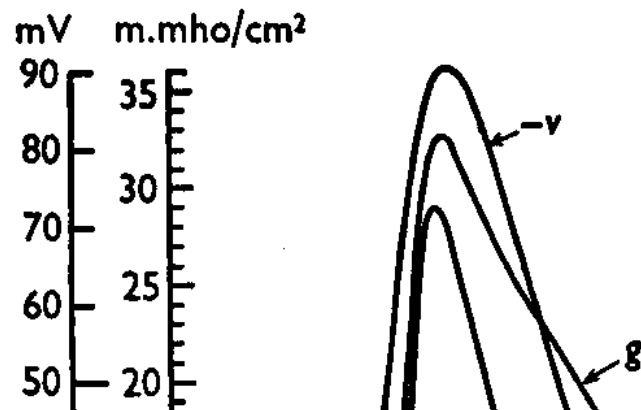
$$\frac{d}{dt} m = \alpha_m (V) (1 - m) - \beta_m (V) m \quad (2)$$

$$\frac{d}{dt} n = \alpha_n (V) (1 - n) - \beta_n (V) n \quad (3)$$

$$\frac{d}{dt} h = \alpha_h (V) (1 - h) - \beta_h (V) h \quad (4)$$

Overlay of the action potential (voltage) and underlying Na and K conductances

conductance peak in Fig. 16*A* and the half-amplitude of the bridge output in Fig. 16*B* is due simply to the choice of scale. Nevertheless, our calculated action potentials agree well with Cole & Curtis's results in this respect. These authors found that the average membrane resistance at the peak of the impedance change was $25 \Omega \cdot \text{cm}^2$, corresponding to a conductance of 40 m.mho/cm^2 . The peak conductances in our calculated action potentials ranged from 31 to 53 m.mho/cm^2 according to the conditions, as shown in Table 4.



Model verification

MEMBRANE CURRENT IN NERVE

525

RESULTS

Membrane action potentials

Model

Form of action potential at 6° C. Three calculated membrane action potentials, with different strengths of stimulus, are shown in the upper part of Fig. 12. Only one, in which the initial displacement of membrane potential was 15 mV, is complete; in the other two the calculation was not carried beyond the middle of the falling phase because of the labour involved and because the solution



Experiment

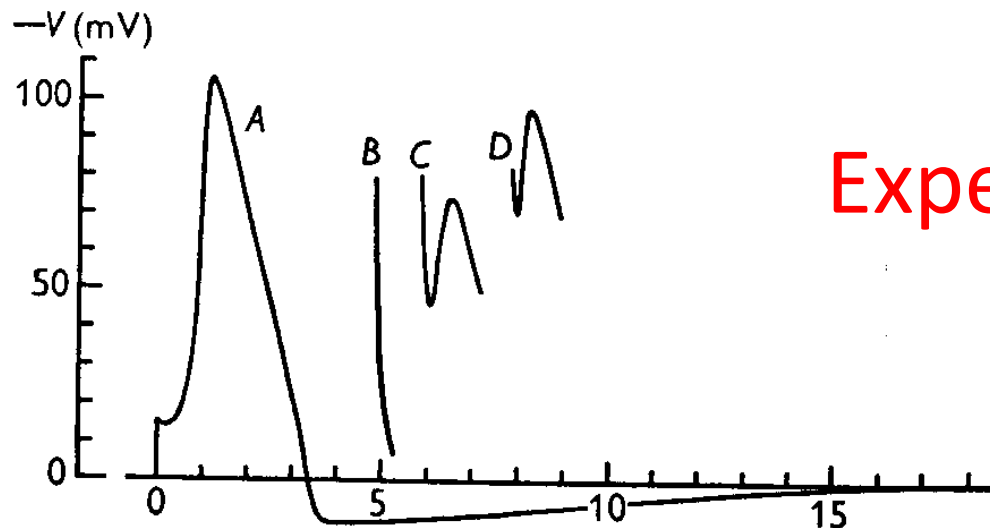
The refractory period

34

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settles down after a heavily damped oscillation of small amplitude as seen in the figure.

Responses to stimuli during positive phase. We call this the membrane when it was suddenly depolarized by a stimulus during the positive phase of the membrane action potential shown by the upper curves in Fig. 20. After the



Model

Experiment

H&H explanation for the refractory period

Refractory period



532

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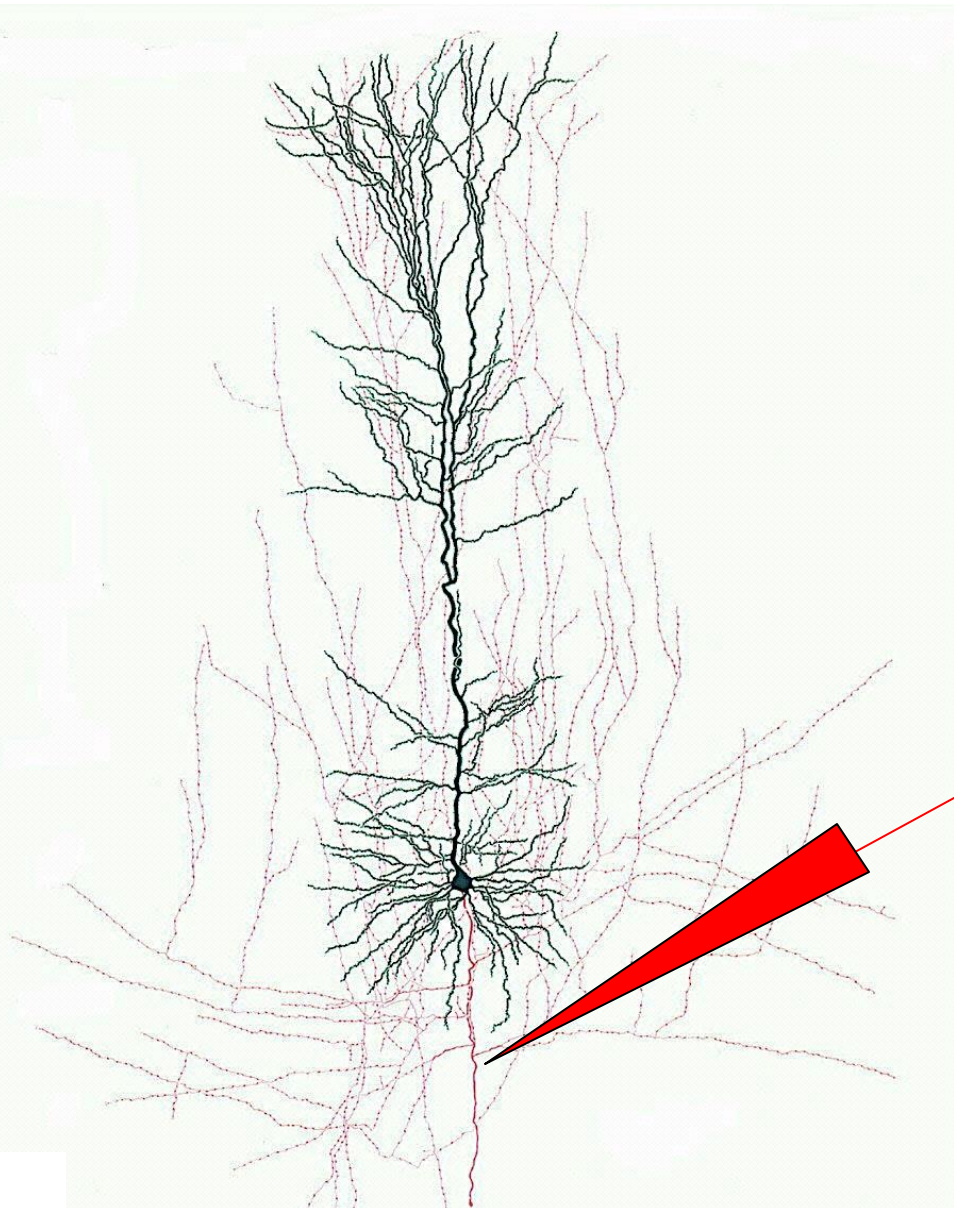
impulse. This has been done for the four complete action potentials that have been calculated, and the results are given in Table 5. It will be seen that the values at 18.5° C are in good agreement with the values found experimentally by Keynes (1951) and Keynes & Lewis (1951), which were obtained at comparable temperatures.

Ionic fluxes. The flux in either direction of an ion can be obtained from the current and the equilibrium potential for that ion if the ionic

Slow recovery from **Na inactivation (h)** and slow kinetics of K conductance, both make the following spike impossible (**absolute refractory**) or hard (**relative refractory**) to be initiated.

The universal nature of the spike

(from squid to cortical axons - carrying the **brain code**; effecting plasticity)



Response of
Cortical
Pyramidal Cell
to Current
Injection

Courtesy of D. McCormick (Yale Univ.)