

Review article

## Update on the pathophysiology of the epilepsies

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### Abstract

The pathophysiology of convulsive and non-convulsive epilepsies is discussed in its primary generalised forms. Focal, clinical and experimental epilepsies, with emphasis placed on the temporal lobe epilepsies (TLE) and their pathophysiologies are also reviewed. Neurotransmitters and neuromodulators and between them, the second messenger systems are considered in the generation, maintenance or inhibition of the epileptic discharge. Action mechanisms of the more classic antiepileptic drugs are briefly summarized along with the therapeutic strategies that might achieve the final control of abnormal discharges, including genetic control as a promising alternative in the current state of research. We emphasized the study of all type of glutamate and GABA receptors and their relation with mRNA editing in the brain. Some of the genetic studies which have been so fruitful during the last ten years and which have brought new insights regarding the understanding of epileptic syndromes are summarized in this article. © 2000 Elsevier Science B.V. All rights reserved.

**Keywords:** Dentate granule cells; GABA receptors; Glutamate receptors; Sodium channels; Calcium channel; G proteins; mRNA

### 1. Introduction

Epilepsy is a cluster of disorders rather than a single disease. It affects about one in 200 people and may last a person's whole life. The individual, who suffers from epilepsy is vulnerable to economic, social and legal difficulties and might experience deep psychological problems. The goals of epilepsy management are to prevent discharges and to avoid their propagation. The understanding of the pathophysiology of the epilepsies is still incomplete. This paper is a brief review of this subject.

### 2. Basic mechanisms related to generalised tonic-clonic seizures

Generalised seizures are studied by parenteral injection of convulsant drugs, e.g., metrazol in rats. In this animal

model, small bilateral lesions of mammillothalamic tract or anterior thalamus, block the convulsive effect [1] suggesting that the generalised response is unrelated to a diffuse origin of the discharges. Generalised seizures are also evoked by the electrical stimulation of diverse cerebral and brainstem regions. Gale (1990) [2] has divided experimental models of generalised convulsive attacks into three groups, according to motor responses: (a) tonic attacks; (b) clonic attacks; and (c) explosive clonic movements of the four limbs. Tonic attacks (a) would be linked with mesencephalic and hindbrain reticular formation, and fail to disappear with supracollicular resection [3], the response proving to be self-sustained beyond the stimulation. Clonic attacks (b) are elicited by kindling with chronically implanted electrodes in various limbic cerebral areas (septum, amygdala, as well as in the entorhinal cortex, the piriform area and the hippocampus). They may be evoked or suppressed by microinjections in the depth of the prepiriform cortex (area tempestas). Explosive clonic movements (c) are the characteristic response of audiogenic seizures, induced by sound and attributed to inferior colliculus, whereas other models with the same

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response have no clear localization. These experiences underscore the concept of epileptic circuits, perhaps more accurately than that of epileptic neurones or neuronal aggregates.

In adult rats, which were treated with flurotil and muscimol (a GABA<sub>A</sub> agonist) injected bilaterally in substantia nigra (pars reticulata) attacks were suppressed. Bicuculline (a GABA<sub>A</sub> antagonist) facilitated seizures and baclofen (a GABA<sub>B</sub> agonist) had no effect [4]. The red nucleus may share inhibitory properties with the locus niger and the pontine reticular formation may inhibit spinal activity induced by convulsant drugs.

Various generalised tonic–clonic epilepsy models have been developed in animal strains with a genetic charge for this seizure type. The photosensitive baboon, presents a picture remarkably similar to the human “grand mal” type of seizures. Both clinically and electrically, attacks resemble those of human patients, but in contrast, monkeys hardly ever suffer spontaneous attacks. Studies with these models have attempted to find a genetic sequence and a structural enzyme or protein abnormality that may trigger tonic–clonic attacks. Since verapamil is able to depress generalised tonic–clonic activity in rats receiving systemic pentylenetetrazol, the suggestion has been advanced that calcium antagonists may be helpful in treating human epilepsy.

### 3. Basic mechanisms related to generalised absence seizures

The basic mechanism underlying generalised absence seizures seems to involve thalamocortical circuits alternating an inhibitory phase, mediated by GABA<sub>B</sub> and an excitatory phase, mediated by glutamate receptors.

Oscillatory phases result in bilateral spike and wave (SW) discharges that distinguish these attacks. Jasper and Droogleever-Fortuyn [5] showed that electrical stimulation of cat midline and intralaminar nuclei at 3 cycles/s can provoke bilateral synchronous SW as shown in cortical EEG.

In the cat receiving penicillin, the SW pattern seems to be initiated in the cerebral cortex, whereas in rat models, thalamus and cortex are necessary to trigger the SW discharge [6,7]. The thalamic reticular nucleus (TRN) commands oscillatory behaviours in thalamocortical circuits because of its low threshold for Ca<sup>2+</sup>-dependent spikes mediated by the GABA<sub>B</sub> receptors, inhibitory postsynaptic potentials (IPSP), which then generate high rate action potentials [8]. Ethosuximide and trimetadione decrease these Ca<sup>2+</sup> currents and antagonise clinical absence attacks. In contrast, an increase in GABAergic activity potentiates both, clinical and experimental absences.

In experimental epilepsy, during SW discharges recorded both in the cortex and in the thalamus, there is both

GABAergic IPSP preservation and a lack of focal epileptogenic depolarising shifts. GABAergic discharges of TRN and its projections into thalamic specific nuclei would be involved in the initiation and control of absence attacks.

An increase in GABA<sub>A</sub> mediated inhibition, for example by benzodiazepines, may be useful for absence treatment by inhibiting TRN and its GABA<sub>B</sub> discharge. In turn, GABA<sub>B</sub> agonist baclofen exacerbates SW discharges in some experimental models, in a way not achieved using GABA<sub>A</sub> agonist muscimol [9]. In the rat with genetic epileptic absence of Strasbourg (GAERS), gamma-vinyl-GABA and glycine worsen the picture [10].

Long-term activation of NMDA receptors with agonist produces a tonic depolarisation of thalamic neurones, decreases low threshold calcium current occurrence and blocks absence attacks [11]. NMDA antagonists blocks the excitatory postsynaptic potentials (EPSP) necessary to begin the cycle. Therefore, the full sequence would be as follows: (1) EPSP mediated by NMDA; (2) slow IPSP mediated by GABA A and B receptors; (3) low threshold calcium current; and (4) new depolarisation and the onset of a new cycle.

This cycle occurs at the TRN level and its projections. The clinical expression is an absence and the electroencephalographic correlate, the SW complex, reflects an alternate depolarisation and hyperpolarisation.

Hyperpolarisation may be an essential element of the cycle, followed by an increased conductance of voltage-dependent Ca<sup>2+</sup> currents (T channels) with subsequent occurrence of Na<sup>+</sup>-dependent action potentials (AP) and repolarization through voltage-dependent K<sup>+</sup> currents. These findings show that there is a need for more research on drugs that block T channels Ca<sup>2+</sup> currents for absence treatment, allowing selective cycle suppression. Another strategic point is slow IPSP blockade or sustained depolarization on thalamocortical relays to prevent T current activation [12]. Each SW complex appears to begin in the cerebral cortex, but oscillatory pattern requires the integrity of thalamocortical circuits, comprising specific and non-specific thalamic neurones showing that it is better to consider epilepsy as a network dysfunction rather than a single derangement of some nerve cells.

As for L calcium channels, their modulation may be opposite to that of T calcium channels in Wistar rats with spontaneous SW discharges [13]. L channels agonists (such as BAYK 8644) protect against SW discharges in Wistar rats, while antagonist such as nimodipine, facilitate them.

In photosensitive SW activity, Quesney and Reader have postulated a response of the hyperexcitable cerebral cortex to photically evoked thalamocortical discharges that decrease the dopamine cortical release or the net effect of this neurotransmitter which probably has an inhibitory effect on the occipital cortex [14].

There are no animal models that accurately replicate human absence seizures, however there are similar in-

vestigations of animal mechanisms, such as the Wistar or GAERS rats with spontaneous absences, or cats parentally injected with penicillin.

Myoclonic absences have been linked with 12 P trisomy and juvenile myoclonic epilepsy with 6 chromosome short arm. In the last years, 7 chromosomal loci for Mendelian forms of common idiopathic generalized epilepsies have been reported [15].

#### 4. Mechanisms related to partial seizures: temporal lobe epilepsies

The histological pattern of temporal lobe epilepsy has been studied since 1880 by means of light microscopy studies and more recently by immunohistochemistry when the peroxidase–antiperoxidase method became available. Gliosis and neuronal loss are evident to a variable degree, as shown by massive degenerative changes in pyramidal neurones of hippocampus and in granular cells of fascia dentata. Subtler alterations include dendritic spine loss and dendritic bead and spindle deformation [16]. Tissue obtained from patients with a temporal focus provides the usual material for study. The significance of these microscopic changes is uncertain, since they may be due to repeated hypoxic-ischemic insults.

Experimental studies in focal epilepsy [17] have disclosed bioelectrical activation termed paroxysmal depolar-

isation shift (PDS), consisting of: (1) an abrupt 25–40 mV depolarisation for roughly 200 ms recorded with intracellular microelectrodes and underlying spike activity recorded with surface electrodes, e.g., in foci induced in cats by application of topical penicillin, followed by (2) action potential (AP) discharge at the peak of depolarisation (see Fig. 1) and (3) a long-lasting hyperpolarisation. Epileptic focus topography is such that neurones located in its centre exhibit PDS while peripheral ones only record hyperpolarisation [18] which tends to prevent discharge propagation. PDS is the huge EPSP expression probably resulting in synchronous activation of recurrent axonal collaterals [19]. Some of the associated AP may reflect ephaptic effects of the electrical field [20]. In turn, the ability of cortical cell to generate PDS may be linked with a marked decrease or blockade of IPSP [21].

As regards molecular events underlying PDS, depolarisation is the result of cell entry of  $\text{Na}^+$  and  $\text{Ca}^{2+}$  currents.  $\text{Na}^+$  enters with initial EPSP, whereas voltage-sensitive  $\text{Ca}^{2+}$  is activated by the same depolarisation (positive feedback), but also by activation of glutamate NMDA receptors (Fig. 2) that are also voltage-sensitive and capable of maximal effect at  $-30$  mV, being blocked by  $\text{Mg}^{2+}$  presence. During repolarisation,  $\text{K}^+$  and  $\text{Cl}^-$  conductance are raised so that  $\text{K}^+$  ions flow through different channels, voltage-sensitive and calcium-dependent, and precede chloride ion entry, mediated by  $\text{GABA}_A$  receptors, that produce post-hyperpolarisation [22]. Evidence that

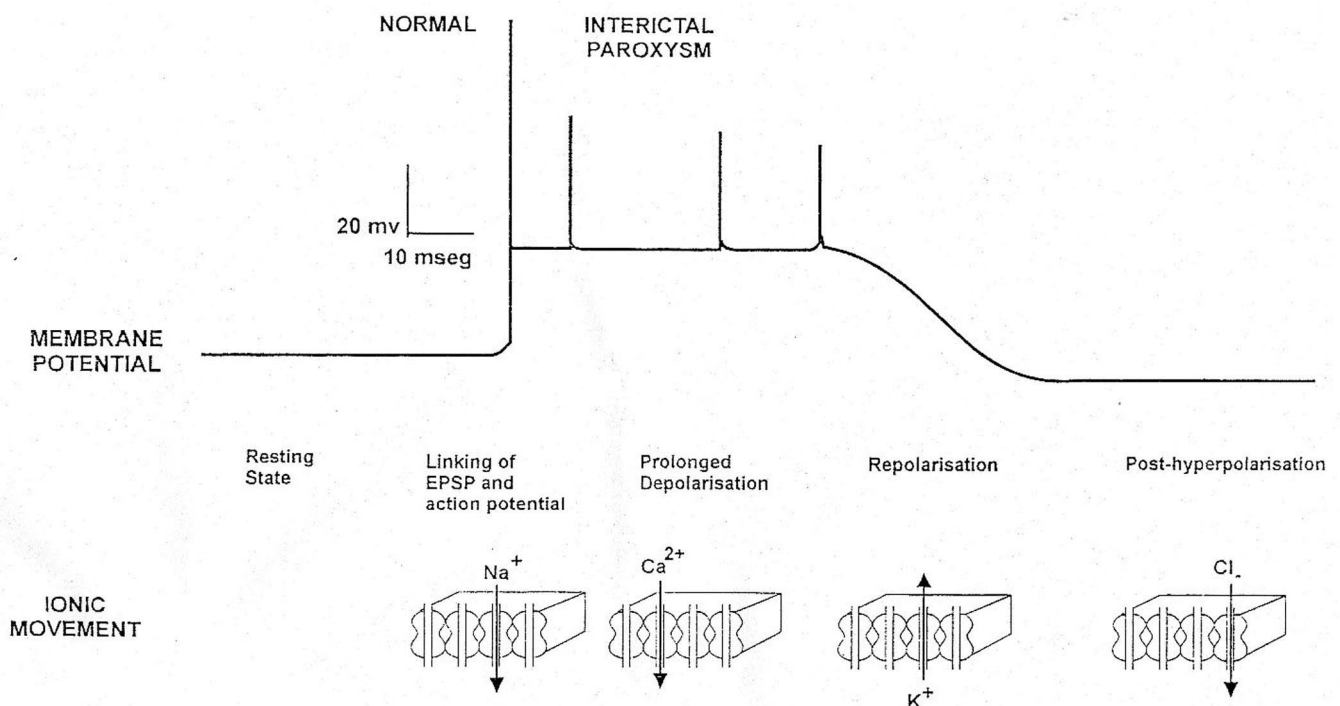


Fig. 1. Paroxysmal depolarisation shift (PDS). An initial excitatory postsynaptic potential (EPSP) and action potential (AP) are linked to  $\text{Na}^+$  entry to the cell, which in turn promotes  $\text{Ca}^{2+}$  entry, thus prolonging depolarisation. Calcium-dependent  $\text{K}^+$  conductance increases, repolarising the cell, an effect which persists (post-hyperpolarisation) due to  $\text{Cl}^-$  entry until metabolic pumps re-establish resting potential. Modified from Lothman EW and Collins RC. Seizures and epilepsy. In: Pearlman AL and Collins RC, editors. Neurobiology of disease. New York: Oxford, 1990: 276–98 [18].

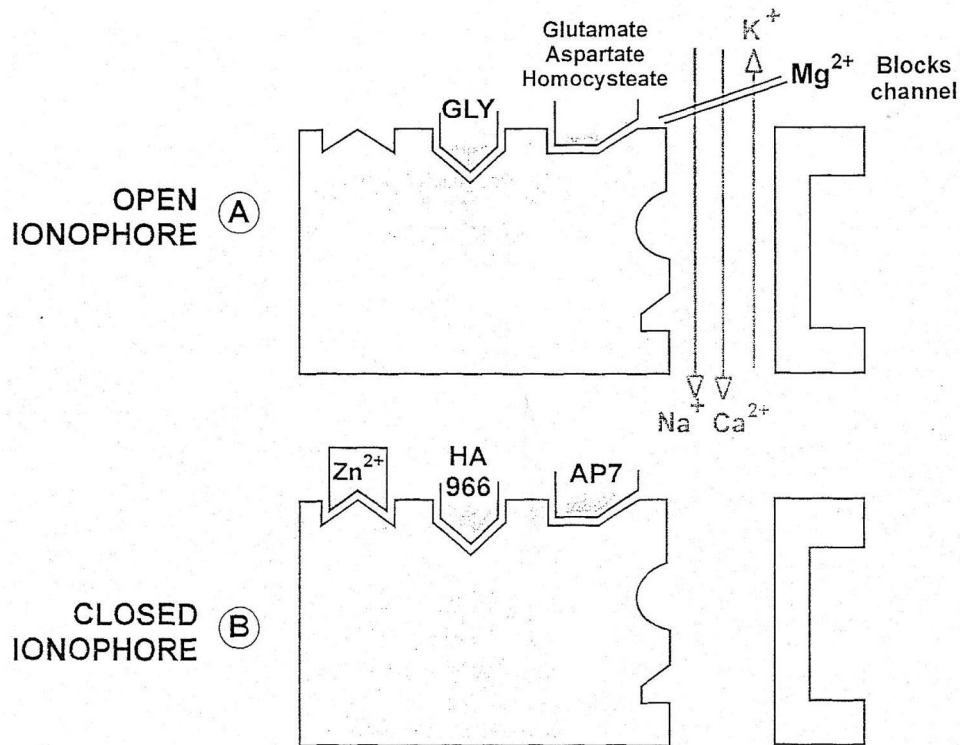


Fig. 2. NMDA receptor: functional states. A: Open ionophore allows  $\text{Na}^+$  and  $\text{Ca}^{2+}$  entry in the presence of glycine and an NMDA receptor agonist; it is blocked by  $\text{Mg}^{2+}$ . B: Closed channel due to action of direct competitive antagonists as AP7, or by agents acting at the glycine site, as HA 966, or at the  $\text{Zn}^{2+}$  site. Modified from Faingold CL and Meldrum BS. Excitant amino acids in Epilepsy [163]. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors. Generalized Epilepsy: Neurobiological Approaches. Birkhäuser: Boston, 1990: 102–117.

$\text{GABA}_B$  receptor activation in the hippocampus induces hyperpolarising currents, by  $\text{K}^+$  efflux, responsible for slow IPSP during post-hyperpolarisation have also been presented [23].

It is generally assumed that temporal lobe attacks may arise in the hippocampus, where patients usually have neuronal loss in CA1 and CA3 and where there is also astrocyte proliferation. In hippocampus slices of adult animals, many proconvulsive treatments lead to recurrent discharges that begin in CA3 and spread to CA1, where ictal events are generated when the slice undergoes an increase in  $\text{K}_e^+$  or a decrease in  $\text{Ca}_e^{2+}$  or  $\text{Mg}_e^{2+}$  [24]. Pyramidal CA3 cells may act as interictal spikes pace-makers, because of a very high  $\text{Ca}^{2+}$  conductance and a profuse disposition of recurrent axonal collaterals, predisposing to PDS. Conversely, CA1 region is able to maintain seizures by itself, without previous interictal spikes [18].

According to Heinemann and co-workers [25], when ictal activity encompasses dentate gyrus, it will increase in CA1, subiculum and entorhinal cortical deep layers, which can initiate the discharges via the perforant pathway, making epilepsy difficult to arrest (Fig. 3). In human temporal epilepsy, mossy fibre budding is seen in dentate gyrus from the granular to the molecular layer, providing the anatomophysiological basis for a recurrent excitation

[26]. There is a good experimental correlation between such budding and hyperexcitability. Electron microscopy shows mossy fibre axonal endings making autosynapses with their own dendrites and with other ones, in both apical and distal sections [27], but also making synapses with inhibitory interneurons, building inhibitory circuits.

According to Lorente de No, mossy axon granular cells at the dentate gyrus project to pyramidal cell dendrites in CA3 [28]. The appearance of spikes in these neurones may depend on: (a) disturbances in their dendrite's physical properties, perhaps linked with the morphological changes mentioned above (spine loss, nodule appearance) and/or (b) increased synaptic input, which may be linked with abnormal proliferation of mossy axons. Another pathological component to be considered in human temporal epilepsy is hilar neuronal loss in dentate gyrus, enhancing granular cell excitability [29]. Other aspects include: (1) synaptic neoformation processes, closer to the concept of neuronal plasticity than one involving cell degeneration, as mentioned with regard to dentate mossy fibres; (2) metabolic depression of interictal glucose, demonstrated by a combination of MRI and PET studies [30]; and (3) osmolality disturbances, that may change a silent area into a consistent epileptiform activity when extracellular osmolality is decreased, shown in experimental conditions in CA1 and in dentate gyrus in the rat [31].

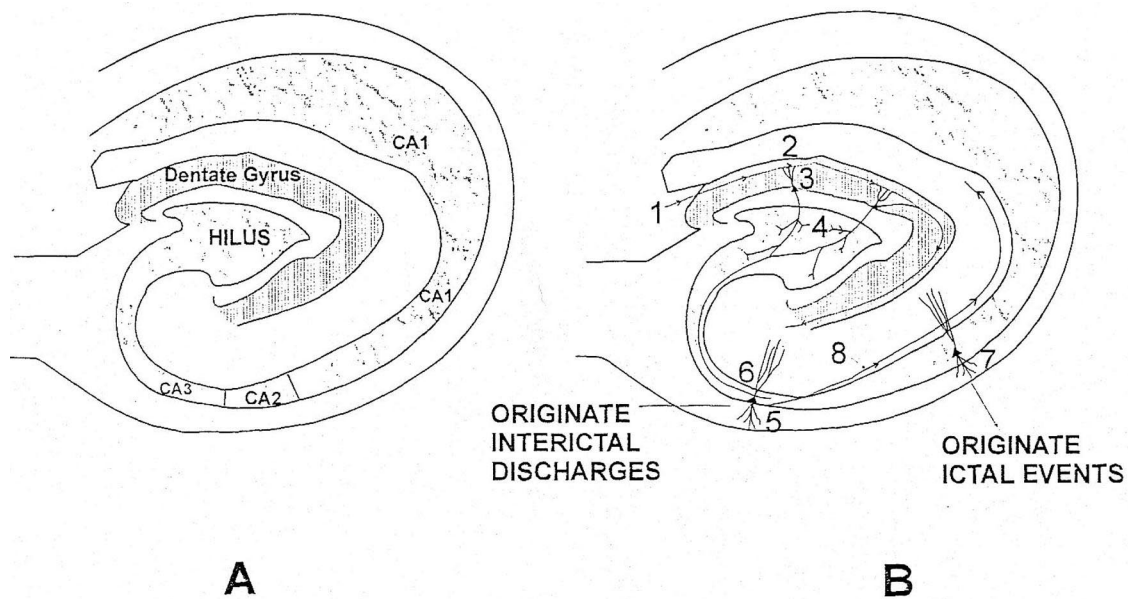


Fig. 3. Normal human hippocampus and its relation with epileptogenesis. Excitatory axons from the entorhinal cortex make up the perforant pathway (1) which ends in the outermost layer of the molecular dentate gyrus (2), exciting the distal portion of dendrites belonging to dentate granule cells (3), which in turn excite the hilar neurones (4) through axonal collaterals, and pyramidal CA3 neurones (5) via mossy fibres (6). CA3 pyramidal neurones excite CA1 pyramidal neurones (7) via Schaffer's collateral bundle (8), subiculum and again entorhinal cortex. The functional state of disinhibition of the circuit led to epileptic discharges. Modified from Sloviter RS. The functional organization of the hippocampal dentate gyrus and its relevance to the pathogenesis of temporal lobe epilepsy. *Ann Neurol* 1994; 35: 640–654 [29].

Studying the transition from interictal to ictal activity, the latter can start without being preceded by interictal spikes. According to Dichter [32], voltage-dependent  $\text{Ca}^{2+}$  currents may contribute to PDS, to interictal–ictal transition and to discharge spread. A  $\text{K}^{+}$  reuptake deficit by astrocytes may operate also as an epileptogenic mechanism. In this regard, various hypotheses have been advanced since  $\text{K}^{+}$  accumulation increases hippocampus excitability [33].

A combination of increased NMDA receptor function and depressed GABAergic transmission due to reduced release, to receptor desensitisation or to genetic abnormality, may well be responsible for the onset of an attack. In fact, NMDA receptors are increased in human temporal lobe epilepsy in the hippocampus [34], while GABAergic transmission appears to be less effective [35].

Decrease in inhibition in dentate granule cells from patients with medial temporal lobe epilepsy have been recently reported [36]. Conversely Babb found GABA hyperinnervation in basket cells in human and rat hippocampal epilepsies between the attacks. It is not sufficient to avoid the attacks [37] as its function appears to be dormant (JC Lacaille, personal communication).

NMDA receptor binding may increase from 20 to 110% in CA1 area and in dentate gyrus of hippocampus in epileptic patients, in surgical specimen studies [38]. The molecular lacunar stratum of CA1 also disclosed a 63% increase in quisqualate-type receptor binding, whereas benzodiazepine (BDZ) and  $\text{GABA}_A$  binding was signifi-

cantly decreased in CA1 and CA4, but not in the dentate gyrus.

Geddes and co-workers [34], documented that the increase in NMDA receptors is located at the parahippocampus, which also shows an increase in binding to kainate-type receptors. Wyler et al. [39] found that there is a decrease in the number of muscarine (cholinergic) receptors and an increase in NMDA receptors in temporal cortex removed from epileptic patients.

When aminoacid levels in tissues removed from patients with temporal epilepsy categorised as: (a) having interictal cortical spikes and (b) lacking cortical spikes — activity confined to amygdala/hippocampus — are compared, there is an increase of glutamate, aspartate and glycine, but not in GABA and the taurine in group (a), which causes a loss of equilibrium between excitatory and inhibitory aminoacids [40].

On studying frontal and temporal foci, Van Gelder and co-workers found a decrease in aspartate and GABA in all the examined areas, but only a decrease in glutamate and taurine and an increase in glycine in the more excitable regions, by comparing aminoacids concentrations in the focus and its surroundings [41]. Conversely, Kish et al. reported a 28% increase in aspartate concentration in surgical specimens from patients with temporal epilepsy [42]. Piredda and Gale [43] have shown aspartate's ability as a convulsant agent, when injected into a rat's prepiriform cortex. Chapman et al. reported a correlation between valproate anticonvulsant strength and its ability to

reduce cerebral aspartate levels [44]. An increase in glutamate and aspartate levels in CSF from epileptic patients has also been documented [45].

Neuronal habituation to subliminal repetitive stimulation (kindling) may produce biochemical changes and facilitate convulsive attacks in animals, when different areas in the limbic system are stimulated. This may provide a model applicable to some issues of human temporal epilepsy. With each stimulation, second messenger systems are activated within cells and proto-oncogens are formed, which in turn activate genes, whose products regulate synaptogenesis and cell survival [46]. In amygdaloid kindling, there is a reduction in acetylcholine (ACh) binding in dentate, hippocampus and amygdala [47], but an increase in BDZ receptor binding in hippocampus [48]. It has been suggested that cholinergic circuits may be directly or indirectly involved in amygdaloid kindling.

Experiments in freezing lesions show that the  $\text{Na}^+ - \text{K}^+$  pump in synapses of primary focus and mirror focus, exhibits changes in its activation that lead to a defect in its net operation [49].  $\text{Na}^+ - \text{K}^+$  pump reduction increases neurone sensitivity to glutamate turning the neurones more excitable [50]. Although the  $\text{Na}^+$  pump protein undergoes up regulation, as illustrated by studies in tissues surgically removed from patients with hippocampal sclerosis, its capacity is limited by low activity levels of cytochrome c oxidase enzyme, which is necessary to provide ATP for pump ( $\text{Na}^+$ ,  $\text{K}^+ - \text{ATPase}$ ) to function. Decreases in cytochrome c oxidase and succine dehydrogenase were found in the periphery of epileptogenic scars in cats [51]. Comparing  $\text{Na}^+$  pump densities in diverse hippocampal areas in human temporal epilepsy, CA1 has the least amount, predisposing its cells to the easiest neuronal death [50].

An inefficient  $\text{Ca}^{2+} - \text{ATPase}$  has been incriminated in audiogenic seizures, a form of epilepsy in which the abnormality seems to be situated in granular cells of dentate fascia [52].

According to Delgado-Escueta and Horan [53] six steps lead from focal epileptogenesis to clinical epilepsy: (1) the generation of enhanced physiological responses; (2) PDS, which in turn lead to interictal spike appearance in EEG; (3) focus spread to perifocal neurones; (4) the utilisation or breakdown of control mechanisms with brain circuits that limit the propagation of seizure discharges via preferred routes of spread; (5) the appearance of secondary foci in regions synaptically linked to the primary focus; and (6) the emergence of clinical seizures.

Experimental models of partial epileptic seizures are obtained through foci generated by topical application of convulsants like penicillin, cobalt or aluminum hydroxide gel, or freeze-focus or by injecting intrahippocampally kainic acid or tetanus toxins. Recent studies have shown that even a single kindled seizure induces apoptotic neuronal death in the dentate gyrus and directly demonstrates that brief seizures provoke neuronal death [54,55].

## 5. Role of neurotransmitters and neuromodulators in relation with neuronal excitability

### 5.1. Aminoacids

(a) Glutamate–Aspartate: These excitatory aminoacids promote  $\text{Na}^+$  and  $\text{Ca}^{2+}$  cell entry through NMDA receptors (Fig. 2) which incorporate 90% of  $\text{Na}^+$  and 10% of  $\text{Ca}^{2+}$  [56]. Their influx is also voltage-dependent, thus increasing during depolarisation, generating a sustained depolarisation, and dependent on channel blockade by  $\text{Mg}^{2+}$  [57], whose depletion in extracellular space increases induced NMDA currents. Both, aspartate and glutamate are able to trigger epileptic seizures and provoke cell damage. According to Curtis and Johnston [58], aspartate and glutamate are the two quantitatively more important neurotransmitters in mammalian CNS.

NMDA receptor may have amplifier properties of excitatory synapses, indistinctly activated by aspartate, glutamate, homocysteate or quinolinic acid. There is evidence that NMDA receptors are more easily activated in the kindling model [59]. According to Krnjevic [60], the main trigger of long-lasting post-tetanic potentiation is the  $\text{Ca}^{2+}$  influx initiated by glutamate and mediated by NMDA receptors. As previously cited, there was an increased binding in the hippocampus [38] and parahippocampus [31] of ligands to NMDA receptors in the cortex removed from patients with temporal epilepsy, as well as an increase in NMDA receptor density [39].

The convulsant ability of aspartate and its possible relation (reverse) with anti-convulsant valproate effect was also cited [44].

While data related to glutamate in epileptic foci are contradictory, indicating indistinctly increased [61] or reduced [41,62] concentration, there is a discernible trend towards a “glutamate leakage” in experimental conditions, both in animals [63] and in humans [64]. Mutani and co-workers reported a decrease in CSF glutamate from epileptic patients with normal values of aspartate and other aminoacids [65].

The study of glutamate receptors has resulted in new classifications. AMPA has replaced quisqualate and currently there are: (1) ionotropic receptors; (a) Glu-R, subdivided into seven subunits, 1–4 sensitive to AMPA and 5–7 sensitive to kainate; (b) Ka (kainate) receptors 1 and 2; (c) NMDAR, 1 and 2 a-b-c-d; and (2) metabotropic receptors mGlu-R; coupled at least to seven different G proteins (1–7).

Merlin and Wong suggest that mGlu-R also participate in epileptic activity [66]. Benaise et al. show that increased AMPA activity in rat area tempestas changes occasional attacks into sustained epileptic activity [67]. According to Comair et al., raised NMDA receptor activation may contribute to hippocampal hyperexcitability in epileptic patients [68]. Hyperexpression of Glu6 (kainate) produces permanent changes in hippocampal excitability [69] which in turn may provoke epileptogenesis.

The role of glutamate as a convulsant and excitotoxic agent is well documented. It is also a precursor of the powerful inhibitor GABA and glutamate/GABA ratio which might exert more influence than any component per se in the generation of paroxysmal neuronal discharges.

Quinolinic acid is an endogenous ligand of the NMDA receptor which can be raised in the brain of some epileptic patients. Feldblum and co-workers describe a decrease in quinolinphosphoribosyltransferase in the frontal and temporal cortex in epileptic human tissue, that could lead to quinolinic acid accumulation with the corresponding amplification of some excitatory synapses, thus predisposing to epileptogenesis [70].

(b) Glycine and taurine: Glycine is a short chain amino acid which shows powerful inhibitory actions, long known in spinal cord, medulla and brain stem. Yet at the NMDA receptor level, glycine increases the affinity of this receptor for glutamate, thus proving excitatory for these synapses [71].

Conversely,  $Zn^{2+}$  modulates the receptor in the opposite direction [72]. Nonetheless, intracellular injection of  $Zn^{2+}$  is epileptogenic, possibly due to its inhibitory effect on  $Na^+$ ,  $K^+$ -ATPase [73] and/or its stimulatory action on urinary excretion of taurine [74].

Taurine appears to be a weak inhibitor in the cerebral cortex, by decreasing the number of cobalt-induced epileptic seizures in mice and cats, as well as delaying the occurrence of seizures induced by ouabaine in rats, so that it could exert an anti-epileptic action [73]. Taurine may inhibit  $Ca^{2+}$  entry to synaptosomes, it increases spontaneous efflux of GABA and inhibit its release by stimulation [75]. Taurine reduction in human epileptic foci has been previously reported [41].

(c) GABA: GABA is the main inhibitory neurotransmitter in vertebrates as roughly 40% of all cerebral synapses are GABAergic. However, its powerful anticonvulsant action is site-dependent; thus, at the nigrocollicular projection, GABA action appears to be pro-convulsant [76]. Locus niger infusion of GABA<sub>A</sub> agonist muscimol is also pro-convulsant in rats treated with fluorotil [77].

A schematic representation of GABA<sub>A</sub> receptor (Fig. 4) includes four subunit families: alpha (1–6), beta (1–3), gamma (1–3) and delta. Selection of one of the subunits from the above 13 types allows for the possible existence of more than 10 000 pentameric subunit combination, although it has been suggested that less than ten major subtypes of GABA<sub>A</sub> receptors actually exist in the adult mammalian brain [78]. This receptor promotes the  $Cl^-$  influx thus provoking hyperpolarisation with a decrease in cellular excitation state, the inhibition being postsynaptic. BDZ and barbiturates increase the strength of GABA<sub>A</sub>-mediated inhibitions [79]. BDZ increases the likelihood of opening the  $Cl^-$  ionophore [80], while barbiturates prolong opening time [81]. Picrotoxin and pentylenetetrazol may block the ionophore [82], while bicuculline is a GABA<sub>A</sub> competitive antagonist [83]. GABA<sub>B</sub> receptors mediate pre- and post-synaptic inhibitions, by efflux  $K^+$  currents or

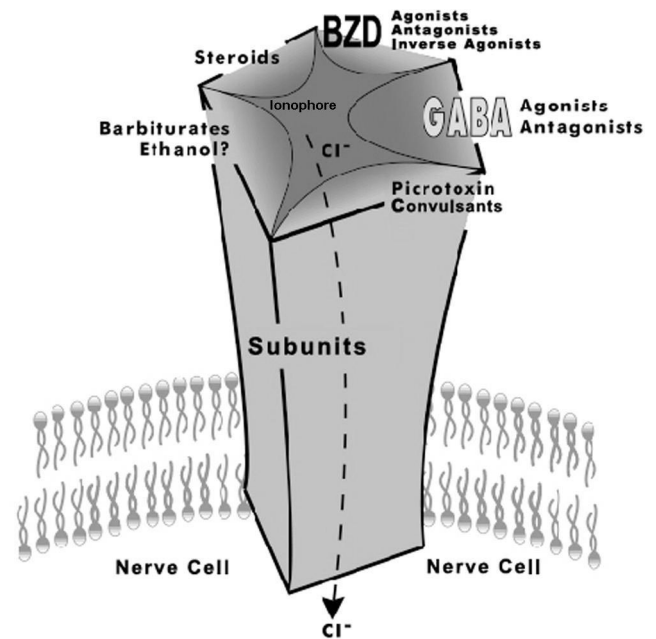


Fig. 4. A schematic representation of the GABA<sub>A</sub> receptor complex. Polymers made up by combinations of multiple subunit subtypes (alpha, beta, gamma and delta). The exact emplacement of each subunit is unknown. The schematic representation is hazardous. Binding site of benzodiazepines,  $Cl^-$  ionophore and GABA are also represented. Other domains allow binding of picrotoxin and barbiturates. The ligand-gated chloride channel proposed to be a pentamer composed by different subunits. Modified from Upton N and Blackburn T. Pharmacology of mammalian GABA<sub>A</sub> receptors. In: Enna SJ and Bowery NG, editors. The GABA Receptors, 2nd Edition. Totowa New Jersey: Humana Press Inc., 1997: 83–120 [164].

a decrease in  $Ca^{2+}$  influx. The role of these receptors in the production of slow post-hyperpolarisation IPSP is discussed in another section of this article. According to Ribak and co-workers [84], in monkeys subjected to aluminium gel, there is a significant reduction of GABAergic impulses in the affected cortices, with a decrease in the number of GABAergic nervous endings. Reduction in BDZ receptor density in epileptogenic foci of patients with partial epilepsy has been reported [85]. In tonic-clonic generalised seizures, enhanced binding to BDZ receptors in cerebellar nuclei and a decrease in thalamus has been described, due to the corresponding modifications in receptor density [86].

Although the data in the literature is contradictory, there is a consensus in stressing that GABAergic function may be depressed in focal epilepsies and in some models of generalised tonic-clonic epilepsy, whereas it is increased in generalised absences.

## 5.2. Biogenic amines

(a) Acetylcholine (ACh): ACh excites cortical neurones by the activation of a depolarising muscarinic receptor acting by blockade of sustained  $K^+$  currents operating during the resting potential. It also blocks  $Ca^{2+}$  dependent

slow  $K^+$  currents occurring during hyperpolarisation (after hyperpolarisation, AHP). AHP provides a useful tool to stop discharges in fast firing systems, as for example pyramidal neurones of the hippocampus.

Although little is known about cerebral nicotine receptors, topical cerebral application of nicotine exerts convulsant effects, as also does its systemic administration [87].

In spite of the conflicting data in the literature, there may be only an epileptogenic action for nicotine receptors, while muscarine receptors may play an excitatory or suppressor role, depending both on the circuit involved and on the neurotransmitter concentration [88]. An increase in ACh synthesising and degrading enzymes in the epileptic cortex of patients undergoing anterior temporal lobectomy has been reported [89], but discussion persists as regards the exact significance of such changes.

The presence of down-regulation of the cholinergic receptor in the temporal lobe of epileptic patients, has also been described [39], which may be a homeostatic mechanism for decreasing excitatory input.

(b) Serotonine (5HT): This neurotransmitter has been linked with regulation of impulses related to hormonal secretion, sleeping, mood, eating, drinking, and sexual drive. It causes the activation of hyperpolarising currents capable of stopping spontaneous neuronal discharges [90]. It may also produce: (1) depolarisation by blocking sustained  $K^+$  currents, as in muscarine receptor activation [91]; (2) blockade of post-discharge hyperpolarisation (post-hyperpolarisation), as muscarine or beta-adrenergic receptors [92]; (3) blockade of slow IPSP obtained in CA1; and (4) blockade of fast IPSP and increased spontaneous and evoked IPSP in CA1 [93].

It is accepted that 5HT delays seizure generation, while a reduction in 5HT promotes their appearance [94]. The 5HT depletion with *p*-chlorophenylalanine decreases the discharge threshold in rats pre-treated with pentylentetrazol [95], while administration of precursor 5HTP increases it [96]. Browning suggests that 5HT attenuates the tonic component of maximal electroshock seizures [97]. 5HT and ACh effects as neurotransmitters and neuro-modulators are complex, assuming at a postsynaptic level at least five different effects for ACh and four for 5HT, as well as additional actions mediated at presynaptic sites. A preponderance of inhibitory actions for 5HT and excitatory ones for ACh is assumed, although receptor 5HT<sub>2</sub> would play a major role in amygdaloid kindling acquisition.

(c) Noradrenaline (NA): A large body of work has attempted to correlate NA levels with epileptic seizure susceptibility. According to Brière et al., a reduction in  $\alpha_1$  receptors in tissues removed from temporal epileptic patients may be the background of a noradrenergic hypofunction, which in turn may reduce inhibition of epileptic foci [98]. Adrenaline, noradrenaline and homovanillic acid concentrations in CSF increase after a generalised tonic-clonic seizure [99], perhaps as an inhib-

itory compensatory reaction, or simply due to higher overall activity.

On studying amygdaloid kindling in rats injected with 6-hydroxydopamine in ascending noradrenergic fibres, Corcoran and Masson [100] found that animals with NA depletion, rapidly developed generalised seizures. In rats with audiogenic attacks, a reduction was found in  $\alpha_1$  and  $\alpha_2$  receptors concomitantly with seizure susceptibility [101]. In another rat model, there is an increase in receptors,  $\alpha_2$  agonists proving proconvulsant and  $\alpha_2$  antagonists, anti-convulsant [102]. Beta<sub>2</sub> receptor activation may be anticonvulsant, acting through cAMP.

In rats, locus coeruleus stimulation may delay amygdaloid kindling development [103]. It is assumed that NA acts as an inhibitory neurotransmitter within the brain, partly facilitating GABAergic transmission. In animals the higher monoamine level produced by monoamine oxidase inhibitors, protects them against seizure susceptibility.

(d) Dopamine (DA): DA applied by iontophoresis in hippocampal slices can provoke and/or prolong hyperpolarisation, which could be associated with an anti-epileptic action [104]. Quesney and Reader have suggested that photosensitive epilepsy could be related to a cortical deficit in DA transmission, a condition which could be counteracted by apomorphine administration [14]. Johnson et al. [105] indicate that cortical DA contents are reduced in photosensitive recessive autosomal homozygote chickens. DA administration by microiontophoresis on the frontoparietal or the occipital cortex in rats or cats usually decreases evoked discharges [106]. Opposite actions are attributable to D<sub>1</sub> and D<sub>2</sub> receptors. The former may be proconvulsant and the latter may be found to be anticonvulsant.

### 5.3. Second messengers

(a) Cyclic nucleotides (cN): cAMP elevation commonly develops subsequent to epileptic seizures, while cGMP elevation could be related to seizure triggering mechanisms [107].

Aminophylline inhibits cAMP accumulation in different cerebral regions and promotes a faster development of tonic attacks induced by pentylentetrazol [107].

The reason of cAMP increment at seizure onset would be to avoid discharge propagation and may respond to raised adenylate cyclase mediated by adenosine and/or biogenic amines. In turn, depolarisation and  $Ca^{2+}$  entry into cells, may activate guanylate cyclase that regulates cGMP levels [107].

Hoffer et al. demonstrated that cGMP applied by iontophoresis excites and induces epilepsy in pyramidal neurones of grafted hippocampus [108]. In contrast, cAMP invariably diminishes spontaneous and evoked activity in guinea pig hippocampal slices in vitro [107].

Isobutylmethylxantine, which raises cGMP and reduces



cAMP, increases spontaneous and evoked potential duration [107].

cAMP may be the second messenger for biogenic amines and adenosine, and may prove to be responsible for the anticonvulsant action of these compounds. In contrast, cGMP could be responsible for the initiation and maintenance of epileptic discharges. Regulatory G proteins, binding to guanine nucleotides, are involved in kindling epileptogenesis. Iwasa et al., reported an increase in  $G_i$  2  $\alpha$  mRNA on the side of stimulated cortex, 24 h after the last generalised seizure, persisting 3 or 4 weeks, in rats undergoing amygdaloid kindling [109].

(b) Phosphoinositides (PI): The phosphoinositides (PI) makes up another system which, as cyclic nucleotides, may act as the second messenger in cerebral neurones (Fig. 5).

According to Dubeau and co-workers, PI turnover is significantly reduced in active epileptic cortex vs. inactive cortex, as defined by electrocorticography, suggesting a PI cycle uncoupling that would prevent modulation of neuronal activity [110]. The main function of this system is to prevent excessive  $Ca^{2+}$  accumulation in endoplasmic reticulum. Cholinergically induced PI formation is reduced by epileptic activity, as also seen in the kindling model by noradrenergic and cholinergic deactivation [111]. Likewise, stimulation by excitatory aminoacids tends to reduce the efficiency of this cycle in the epileptic cortex [110].

The importance of this second messenger system is illustrated by the fact that it appears to be related at least with 17 different receptor subtypes. Among these, the muscarine receptors ( $M_1$  and  $M_2$ ),  $\alpha_1$  (adrenergic),  $H_1$  (histaminergic),  $5HT_2$  and AMPA stimulate the cycle, while  $D_2$  receptors as well as adenylate cyclase prove to be inhibitory [112].

Epileptic seizures increase diacylglycerol (DAG) by IP<sub>2</sub> [113]. DAG accumulation activates PKC, which phosphorylates  $\alpha_1$  receptors, inhibiting its response, thus disrupting the cycle. Bradichinine, a strong activator of the PI cycle, produces a biphasic response comprising transient initial hyperpolarisation followed by a depolarisation. Intracellular application of IP<sub>3</sub> mimics the initial phase through a calcium-sensitive  $K^+$  current, and may be subsequent to calcium mobilisation from its intracellular stores [114].

(c) Calcium ( $Ca^{2+}$ ): Calcium provokes repetitive sustained discharges, through its entry to the neurones, by voltage-dependent currents and/or by NMDA receptor channel (see Fig. 2). There are various types of voltage-dependent  $Ca^{2+}$  channels: the presynaptic N channel, related to receptors and G proteins, the L channel in neuronal soma and the T channel, whose dysfunction was mentioned in relation with epileptogenic mechanisms of generalised absences, among other channels (P, Q, R).

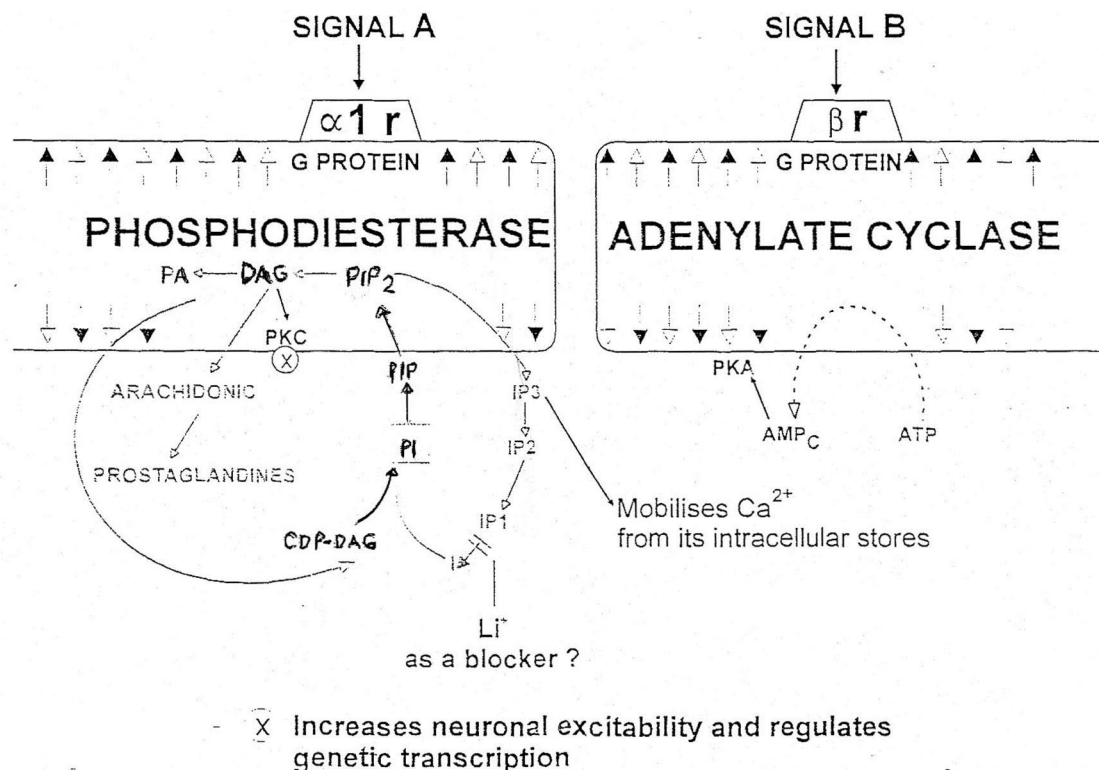


Fig. 5. Transmembrane signalling systems: phosphoinositide (PI) cycle. PI, phosphatidylinositol; PIP, phosphatidylinositol 4-P; PIP<sub>2</sub>, phosphatidylinositol 4,5-bis-P; DAG, 1,2-diacylglycerol; PA, phosphatidic acid; PKC, protein kinase C; IP<sub>3</sub>, D-myo-inositol 1,4,5-tris-P; IP<sub>2</sub>, D-myo-inositol 1,4-bis-P; IP<sub>1</sub>, D-myo-inositol 1-P; I, D-myo-inositol; CDP-DAG, citidine diphospho-diacylglycerol; PKA, protein kinase A. Modified from Hirasawa K and Nishizuka Y. Phosphatidylinositol turnover in receptor mechanism and signal transduction. *Ann Rev Pharmacol Toxicol* 1985; 25: 147–170 [165].

Channel N is important for neurotransmitter release and is exclusive of neurones. Little is known about the L and T channels, that appear to be related to control of neuronal excitability. It should be recalled that, as in the case of cyclic nucleotides and PI, calcium is an intracellular transducer of signals coded by membrane receptors, that function with such receptors and with their configuration changes (second messengers). Ferrendelli et al. demonstrated the ability of phenobarbital, phenytoin and carbamazepine to block voltage-dependent  $\text{Ca}^{2+}$  entry to isolated nerve ending preparations, which is also valid for BDZ [115].

Calcium activation by NMDA receptors generates fast and long-lasting currents, leading in some instances to fast depolarisation and persistent neuronal changes [116].

The discovery of a binding calcium protein calmoduline (CaM) was the first step in understanding the mechanism postulating  $\text{Ca}^{2+}$  as a second messenger. CaM binds  $\text{Ca}^{2+}$  and is, in turn, attacked by CaMK2. Anti-convulsant drugs antagonise the system by activating the enzyme CaMK2, which inhibits CaM [117]. CaMK2 levels are impaired in hippocampal neurones in animals undergoing kindling [118]. PKC is another protein regulating neuronal excitability and influenced by  $\text{Ca}^{2+}$ . Calcium, by itself, is a powerful modulator of the PI system [114].

PKC is the convergence site of second messengers. The enzyme can interact with both pre-synaptic and post-synaptic membranes, regulating neurotransmission and genetic transcription. Multiple subtypes have been characterised. PKC activation inhibits several types of  $\text{K}^+$  channels, thus increasing neuronal excitability, perhaps through phosphorylation of  $\text{G}_i$ , an inhibitory G protein, that depresses adenylate cyclase and therefore, decreases bi-directional modulation exerted by cAMP on neurotransmission. It also can reduce sodium currents by phosphorylation of the alpha subunit of the sodium channel protein itself [119].

In experimental epilepsy, paroxysmal discharges are accompanied by a reduction in  $\text{Ca}_e^{2+}$  level [120], which occurs in human epileptic tissues. Such  $\text{Ca}^{2+}$  mobilisation may operate as a common path for the execution of epileptic discharges.

#### 5.4. Other neuromodulator substances

(a) Purines: During epileptic seizures, adenosine is released, acting as a powerful endogenous anti-convulsant [121]. In vitro studies have shown the anti-convulsant effect of adenosine in audiogenic attacks [122], seizures induced by maximal electroshock [107] and biochemically induced seizures [123]. Adenosine uptake inhibitors reduce severity and duration of seizures induced by amygdaloid kindling [124].

In hippocampus preparations, adenosine reduces the synaptic transmission, acting on pre- and post-synaptic receptors [125] and decreasing the release of excitatory

neurotransmitters [126]. In slices, adenosine suppresses the synaptic transmission of CA3 to CA1 by Schaffer collaterals and the stimulation of dentate granular cells by the perforant tract [114], an effect perhaps mediated by  $\text{A}_1$  receptors. It may be concluded that adenosine precludes the initiation and maintenance of epileptic discharges [127].

(b) Opioid peptides: A variety of experiments support the role of opioid peptides as proconvulsant or anticonvulsant. In favour of their epileptogenic actions: (1) beta-endorphin and leu-enkephalin administered IV induced ictal EEG activity in the rat [128,129]; (2) mu and delta receptors are likewise pro-convulsant [130]; and (3) morphine causes generalised seizures [131] and can antagonise this effect [132]. Conversely: (1) naloxone shortens the postictal state that follows kindling or electroshock [133,134]; and (2) various kappa agonists show anti-convulsant activity against seizures induced by ECT or fluoritil, while dinorphine shares the same effect.

Lanerolle et al. reported increased immunoreactivity for dinorphine A in granular cells of human hippocampal epileptic foci [135]. However, the interpretation of this finding and of other neuropeptide alterations remains uncertain. Mayberg et al. documented a significant increase in mu receptor binding in epileptic temporal cortex [136]. The anti-convulsant effect of the peptide ACTH should be mentioned, but the explanation of its anti-epileptic action remains controversial.

The role of opioid systems in the suppression of seizures in a variety of experimental models of epilepsy is better documented than is the role of opioid systems in the initiation of the attacks. Current evidence suggests that the anti-convulsant effects of peptide and non-peptide opioids can be mediated through mu, delta or kappa opiate receptors. Frost has suggested that non-mu opiate receptors are elevated in temporal lobe epilepsy [137].

## 6. Anti-epileptic therapy

### 6.1. Drugs in use

Better known anticonvulsant drugs generally act: (1) by increasing GABAergic action mechanisms, that are predominantly inhibitory; (2) by depressing excitatory glutamate neurotransmission; (3) by depressing voltage-dependent  $\text{Ca}^{2+}$  mobilising mechanisms; or (4) by depressing  $\text{Na}^+$  conductance, which is also voltage-dependent.

According to Macdonald, the  $\text{Na}^+$  channel blockade is an important action in phenytoin, CBZ and valproate, less important in phenobarbital and clonazepam and null in ethosuximide. The blockade of T calcium channels is significant for valproate and ethosuximide, doubtful for clonazepam and CBZ, and null for phenobarbital and phenytoin. Finally, the GABA channel activation is important in the clonazepam action, less but still significant

for phenobarbital, debatable in the case of valproate and null for phenytoin, CBZ and ethosuximide [138].

Drugs acting against  $\text{Na}^+$  channels provide the treatment of choice for generalised and partial tonic-clonic seizures; those acting at the T calcium channel level afford satisfactory therapy for generalised absences, and those acting at GABA receptors are useful to treat myoclonic seizures.

This brief discussion has not reviewed the action of more recently available drugs, such as oxcarbazepine, vigabatrin, lamotrigine, phelbamate, gabapentin, topiramate and zonisamide, among others. Their use may benefit patients otherwise categorised as refractory. However, their mechanisms of action remain controversial.

## 6.2. New therapeutical approaches

(a) Genetic studies: There is a bulk of recent research about genetic linkage studies in familial epilepsies with special reference to the following clinical forms: autosomal dominant nocturnal frontal lobe epilepsy (ADNFLE), autosomal dominant lateral temporal lobe epilepsy (ADLTE), familial partial epilepsy with variable foci (FPVEF), benign familial neonatal convulsions (BFNC), familial febrile convulsions (FFC), benign epilepsy of childhood with centrotemporal spikes (BECTS) and juvenile myoclonic epilepsy (JME).

ADNFLE: All families reported show that there is an autosomal dominant inheritance with incomplete penetrance. Affected individuals develop predominantly nocturnal seizures with frontal lobe semiology. Phillips et al. [139] pointed out that the gene for ADNFLE maps to chromosome 20q13.2–q13.3 and found a second locus at 15q24 [140].

A missense mutation in the gene for the alpha-4 subunit (CHRNA4) of the neuronal nicotinic acetylcholine receptor (nAChR) was described in an Australian family [139] and another mutation in the same gene in a Norwegian family [141]. This gene maps in the same region of 20q. The missense mutation replaces serine with phenylalanine at codon 248, in the second transmembrane domain. The gene is expressed in all layers of the frontal cortex [142].

ADLTE: This syndrome is inherited in an autosomal dominant mode, with a penetrance of about 80%. It is characterized by rare partial seizures, usually secondary generalized, arising mostly during sleep. The partial component, the interictal EEG abnormalities and the results of SPECT studies suggest a lateral temporal lobe origin. The genetical analysis found linkage to chromosome 10q [143].

FPVEF: This group differs from the other inherited partial epilepsies because the clinical features of the seizures and interictal EEG foci differed among family members and include frontal, temporal, occipital and centroparietal seizures. A suggesting of linkage was found on chromosome 2q [144].

BFNC: It is a rare autosomal dominant form of epilepsy.

Genetic heterogeneity is reported with a first suggested locus on chromosome 20q and a second one on chromosome 8q [145].

FFC: Wallace et al. [146] reported linkage of a putative autosomal dominant febrile convulsion gene to chromosome 8q13–21, and Johnson et al. [147] reported another autosomal dominant febrile convulsion gene on chromosome 19p13.3 between loci D19S591 and 19S395.

BECTS: Neubauer et al. [148] found evidence for linkage of BECTS to a region on chromosome 15q14. This disorder is genetically heterogeneous.

JME: Weissbecker et al. [149] reported confirmation of a JME locus to the HLA region of chromosome 6p. More recently, chromosomal regions harbouring genes for nAChR subunits were tested for linkage to the JME trait in 34 pedigrees. Significant evidence for linkage with heterogeneity was found to polymorphic loci encompassing the region in which the gene encoding the alpha 7 subunit of nAChR maps to chromosome 15q14 [150].

(b) Regulatory events at the genetic level: Second messenger systems interact with proteins controlling genetic transcription. Thus, a “third messenger” has been postulated acting in the cell nucleus in order to control genetic expression. These are proteins translated from protooncogenes. Phorbol esters, by activating PKC, induce protooncogene transcription as Fos and Jun. These genes exert regulatory functions interacting in adaptative responses to environmental stimuli or biologic stress, e.g., epileptic seizures. Within this regulation, it is possible to include neuronal metabolism, as well as neurotransmitter synthesis and degradation. A second cascade of regulatory events may involve not only the number of receptors, but also their subtypes as well as the coupling to the receptor-G protein second messenger.

Thus, mRNA of various genes with early immediate response, initially identified in fibroblasts, are rapidly increased within the brain after convulsive seizures [151] (c-fos, c-jun, jun-b y zif/268). PKC could raise mRNA levels for these transcriptional factors [152]. This mRNA increase occurs mainly in granule cells of the dentate gyrus [114].

According to Sprengel et al., the genetic mechanism of RNA editing appears to be involved in the control of rapid glutamate-elicited postsynaptic  $\text{Ca}^{2+}$  transients. Analysis of the GluR-B gene subunits transcribed in mice which were genetically engineered to harbor one allele incapacitated for editing, revealed that 30% of the GluR-BmRNA occurred in its unedited form, which was predictive of an increase in the  $\text{Ca}^{2+}$  permeability of AMPA receptors, and may lead the mice to have seizures [153].

New pharmacological designs must consider “second and third messenger” processes, in order to control epileptic seizure physiopathology.

(c) The control of neurotransmitters, their receptors an ionic flux: In generalised absences, pharmacological approaches attempt to achieve: (1) NMDA receptor inactiva-

tion; (2) GABAB receptor inactivation; and (3) inactivation of low threshold T calcium currents.

In focal epilepsy, with or without secondary generalisation, and in primary generalised epilepsy, the pharmacological approaches attempt to achieve: (1) blockade of  $\text{Na}^+$  conductance that promotes repetitive voltage-dependent discharges; (2) blockade of voltage-dependent  $\text{Ca}^{2+}$  currents; (3) inactivation of NMDA receptors and other types of the GluR superfamily; (4) an increase in  $\text{K}^+$  conductance; (5) an increase in  $\text{Cl}^-$  entry; (6) activation of GABAergic receptors; (7) interactions with cyclic nucleotide and PI systems (second messengers); and (8)  $\text{Na}^+$ ,  $\text{K}^+$ -ATPase stimulation.

(d) Surgical control of epileptic discharges: Surgical procedures are in vogue for treatment of partial epilepsies and temporal lobe epilepsies (e.g., focus surgery, amygdalohippocampectomy, anterior temporal lobectomy or total lobe resection), unilateral hemispherical lesions e.g., Rasmussen syndrome or hemimegalencephaly (hemispherectomy or hemispherotomy) and uncontrollable Lennox syndrome (corpus callosum section) [154]. Subpial transections are performed when other techniques are not recommended in order to spare the eloquent areas (e.g., Landau-Kleffner syndrome). These procedures, as well as vagal or thalamic stimulation, have succeeded where medical treatment has failed.

A global survey on epilepsy surgery which covers a period of ten years, reported by the International League Against Epilepsy (ILAE) provided the following data: In adults: (1) Anterior temporal lobectomies were performed in 71% of the surgical patients and there was a reduction or suppression of seizures in almost 90% of them; (2) selective amygdalo hippocampectomies were done in 10% of the intervened patients with marked post-operative improvements in almost 80% of them; (3) frontal lobe resections in another 10% were less effective to render the patients seizure-free; and (4) parietal and occipital resections were done infrequently and their results were a little better than the frontal lobe resections but no so effective as temporal lobe resections. In children: (1) anterior or complete callosotomies were performed, leading to a marked reduction of seizures, although the children were not completely seizure-free in more than 90% of the cases; and (2) hemispherectomies were performed in almost 24% of the patients and their results were much better than the callosotomies, which were performed in order to render the children seizure-free [155].

## 7. Conclusions

The basic mechanisms of the epilepsies were considered separately in: (a) convulsive primary generalised epilepsy; (b) non-convulsive primary generalised epilepsy; and (c) partial epilepsies.

(a) The discharge synchronisation mechanisms generating this seizure type are poorly understood, but correlate with epileptogenic circuits in the brain stem (tonic phase?) and/or the limbic system (clonic phase?).

(b) The model of SW complex at 3–6 cycles/s, generated in the cat after parenteral injection of penicillin, shows an alternate sequence of depolarisation–hyperpolarisation, in which the spike (S) corresponds to an EPSP and the slow wave (W) to an IPSP [156]. There are cycles in the thalamocortical complexes that include: (1) NMDA activation; (2) activation of GABA receptors; (3) activation of T calcium channels; and (4) restarting of the cycle. Perhaps these concepts may be extrapolated to generalised absences suffered by human epileptic patients.

(c) Partial transition to seizures is a dynamic intracellular and network event which closely correlates with the paroxysmal depolarisation shifts (PDS), considered a giant EPSP triggered by increased excitatory input in neurones that undergo one or more abnormalities. These are as follows: (1) proliferation of recurrent axonal collaterals; (2) excess of  $\text{Ca}^{2+}$  conductance; (3) GluR hyperactivity, in one or various subtypes (NMDA, Ka, AMPA, mGluR, among others); (4) impaired GABA<sub>A</sub> receptor activity; (5) alterations in the extracellular compartment with  $\text{K}^+$  accumulation or  $\text{Ca}^{2+}$  and/or  $\text{Mg}^{2+}$  depletion; (6) functional deficits in ATPases involved with cationic transport against chemical gradients; (7) deficient aerobic metabolism of the neurone itself; (8) sudden changes in pH and/or osmolality (alkalosis and hyponatremia favour the appearance of seizures); and/or (9) genetic abnormalities, etc. An example of GluR hyperactivity is the region CA3 of the hippocampus, which has the highest density of Ka receptors [157] and is a pacemaker of interictal spikes in chronic epileptic foci. It has been claimed that circulating autoantibodies react with GluR3 in Rasmussen encephalitis [158].

Regarding the influence of neurotransmitters and neuromodulators, other than glutamate and GABA, in relation to neuronal excitability, let us speculate as follows: (a) glycine is mainly an excitatory agent and taurine is an inhibitory agent; (b) Ach is predominantly excitatory; 5HT is predominantly inhibitory; (c) the adrenergic receptors are  $\alpha_2$  excitatory;  $\alpha_1$  and  $\beta$  inhibitory; (d) the dopaminergic receptors  $\text{D}_1$  agonists are pro-convulsant and the  $\text{D}_2$  agonists are anti-convulsant; (e) adenosine with inhibitory effects as cAMP, while cGMP has opposite effects; (f) opioid peptides with contrasting effects:  $\beta$ -endorphin and leuкоencephaline excitatory, naloxone and dinorphine inhibitory, while morphine presents dual effects; and (g)  $\text{Zn}^{2+}$  exerts dual effects.

Besides regulating ionic channels by the action of voltage and by neurotransmitter-receptor coupling, a transmembrane signalling system (Fig. 5) helps the neurones to regulate their excitability by binding the receptor inside the membrane to G protein which in turn binds cyclic nucleotides and PI. These substances and calcium ( $\text{Ca}^{2+}$ ) make

up three systems of second messengers that have been thoroughly studied.

Several receptors operate with these systems, including GABA<sub>B</sub> and mGluR. GABA<sub>B</sub> receptor exerts antagonistic effects; it inhibits presynaptic release of GABA, thus proving excitatory, but it also inhibits presynaptic release of glutamate, thus proving inhibitory. Besides, it also prolongs postsynaptic hyperpolarisation by K<sup>+</sup> efflux and the appearance of Ca<sup>2+</sup>-dependent spikes in experimental models of absence seizures. According to Mody et al. new evidence demonstrates that mGluR are involved in the regulation of cortical and hippocampal excitability, long time potentiation and epileptogenesis [159].

According to Vornov and Coyle [157] “multiple neurotransmitters may modulate the same mechanism of second messengers, but an isolated neurotransmitter may activate different second messengers”. Second messenger systems converge in PKC, membrane enzyme that regulates cell excitability but also promotes transcriptional changes which can operate in the regulation of neuronal metabolism, neurotransmitter synthesis and transport, receptor density and their eventual coupling to G protein. A “third messenger system”, is currently envisaged, having mRNA as subject [160], whose level increases after electrical stimulation or convulsive seizure.

Factors such as morphological changes, electrophysiological abnormal responses, failure in neurotransmitter functioning or in metabotropic receptors coupled with second messengers systems or genetic mutations could be combined [161] in relation with a genetic charge favouring epileptogenesis [153,162] and acquired lesion [16,26] or a combination of both, decreasing the discharge threshold which in turn will drive Na<sup>+</sup> and Ca<sup>2+</sup> into the nerve cells, a common path for the execution of epileptic discharges.

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## References

- [1] Ferrendelli JA. Some subcortical mechanisms involved in experimental generalized seizures. In: Dichter MA, editor, Mechanisms of epileptogenesis — the transition to seizure, New York: Plenum Press, 1988, pp. 101–10.
- [2] Gale K. Animal models of generalized convulsive seizures: some neuroanatomical differentiation of seizure types. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, Generalized epilepsy: neurobiological approaches, Boston: Birkhäuser, 1990, pp. 329–44.
- [3] Bergman F, Costin A, Gutman J. A low threshold convulsive area in the rabbit mesencephalon. *Electroencephalogr Clin Neurophysiol* 1963;15:683–90.
- [4] Moshe S, Sperber EF. Substantia nigra mediated control of generalized seizures. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, Generalized epilepsy: neurobiological approaches, Boston: Birkhäuser, 1990, pp. 355–67.
- [5] Jasper HH, Droogleever-Fortuyn J. Experimental studies on the functional anatomy of petit mal epilepsy. *Res Publ Assoc Nerv Ment Dis* 1947;26:272–98.
- [6] Avoli M, Gloor P, Kostopoulos G, Gotman J. An analysis of penicillin-induced generalized spike and wave discharges using simultaneous recordings of cortical and thalamic single neurons. *J Neurophysiol* 1983;50:819–37.
- [7] Vergnes M, Marescaux C, Depaulis A. Mapping of spontaneous spike and wave discharges in Wistar rats with genetic generalized non-convulsive epilepsy. *Brain Res* 1990;523:87–91.
- [8] Von Krosigk M, Bal T, McCormick DA. Cellular mechanisms of a synchronized oscillation in the thalamus. *Science* 1993;261:361–4.
- [9] Snead OC. Evidence for GABAB mediated mechanisms in experimental absence seizures. *Eur J Pharmacol* 1992;213:343–9.
- [10] Liu Z, Seiler N, Marescaux C, Depaulis A, Vergnes M. Potentiation of gamma-vinyl GABA (vigabatrin) effects by glycine. *Eur J Pharmacol* 1990;182:109–15.
- [11] Snead OC. Basic mechanisms of generalized absence seizures. *Ann Neurol* 1995;37:146–57.
- [12] Snead OC, Hechler V, Vergnes M, Marescaux C, Maitre M. Increased gamma-hydroxybutyric acid receptors in thalamus of a genetic animal model of petit mal epilepsy. *Epilepsy Res* 1990;7:121–8.
- [13] Van Luijcklaar E, Ates N, Coenen A. Role of L-type calcium channel modulation in nonconvulsive epilepsy in rats. *Epilepsia* 1995;36(1):86–92.
- [14] Quesney LF, Reader TA. Role of dopamine in generalized photosensitive epilepsy: electroencephalographic and biochemical aspects. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, Generalized epilepsy: neurobiological approaches, Boston: Birkhäuser, 1990, pp. 298–313.
- [15] Delgado-Escueta AV, Gee MN, Medina MT, Alonso ME, Cordova F, Rubio-Donnadieu S et al. Genetics of common idiopathic generalized epilepsies. *Epilepsia* 1997;38(Suppl 7):S6–7.
- [16] Scheibel ME, Crandall PH, Scheibel AB. Hippocampal-dentate complex in temporal lobe epilepsy. A Golgi study. *Epilepsia* 1974;15:55–80.
- [17] Prince DA. The depolarization shift in epileptic neurons. *Exp Neurol* 1968;21:467–85.
- [18] Lothman EW, Collins RC. Seizures and epilepsy. In: Pearlman AL, Collins RC, editors, Neurobiology of disease, New York: Oxford, 1990, pp. 276–98.
- [19] Ayala GF, Dichter M, Gumnit RJ, Matsumoto H, Spencer W. Genesis of epileptic interictal spikes. New knowledge of cortical feedback systems suggests a neurophysiological explanation of brief paroxysms. *Brain Res* 1973;52:1–17.
- [20] Tancredi V, Avoli M. Control of spontaneous epileptiform discharges by extracellular potassium: an “in vitro” study in the CA1 subfield of the hippocampal slice. *Exp Brain Res* 1987;67:363–72.
- [21] Dingledine R, Gjerstad L. Reduced inhibition during epileptiform activity in the vitro hippocampal slice. *J Physiol (Lond)* 1980;305:297–313.
- [22] Avoli M. GABAergic mechanisms and epileptic discharges. In: Avoli M, Reader TA, Dykes RWY, Gloor P, editors, Neurotransmitter and cortical function: from molecules to mind, New York: Plenum, 1988, pp. 187–205.

- [23] Dutar P, Nicoll RA. A physiological role for GABAB receptors in the central nervous system. *Nature* 1988;332:156–8.
- [24] Traynelis SF, Dingledine R. Potassium-induced spontaneous electrophysiological seizures in the rat hippocampal slice. *J Neurophysiol* 1988;59:259–76.
- [25] Heinemann U, Draguhn A, Ficker E, Stabel J, Zhang C. Strategies for the development of drugs for pharmacoresistant epilepsies. *Epilepsia* 1994;35(Suppl 5):S10–21.
- [26] Babb TL, Kupfer WR, Pretorius JK, Crandall PH, Levesque MF. Synaptic reorganization by mossy fibers in human epileptic fascia dentata. *Neuroscience* 1991;42:351–63.
- [27] Franck JA, Pokorny J, Kunkel D, Schwartzkroin P. Physiologic and morphologic characteristics of granule cell circuitry in human epileptic hippocampus. *Epilepsia* 1995;36(6):543–58.
- [28] Lorente de No R. Studies on the structure of the cerebral cortex II. Continuation of the study of the ammonic system. *J Psychol Neurol* 1934;46:113–77.
- [29] Sloviter RS. The functional organization of the hippocampal dentate gyrus and its relevance to the pathogenesis of temporal lobe epilepsy. *Ann Neurol* 1994;35:640–54.
- [30] Semah F, Baulac M, Hasboun D, Frouin V, Mangin J, Papageorgiou S et al. Is interictal hypometabolism related to mesial temporal sclerosis? A positron emission tomography/magnetic resonance imaging confrontation. *Epilepsia* 1995;36(5):447–56.
- [31] Roper SN, Obenaus A, Dudek FE. Osmolality and nonsynaptic epileptiform bursts in rat CA1 and dentate gyrus. *Ann Neurol* 1992;31:81–5.
- [32] Dichter MA. Emerging insights into mechanisms of Epilepsy: implications for new antiepileptic drugs development. *Epilepsia* 1994;35(Suppl 4):S51–7.
- [33] Izquierdo I, Nasello AG, Marichich ES. The dependence of hippocampal function on extracellular potassium levels. In: *Currents in modern biology*, Amsterdam: North-Holland Publishing Co, 1971, pp. 35–46.
- [34] Geddes JW, Cahan LD, Cooper SM, Kim RC, Choi BH, Wotman CW. Altered distribution of excitatory amino acid receptors in temporal lobe epilepsy. *Exp Neurol* 1990;108:214–20.
- [35] Ribak CE, Harris AB, Vaughn JE, Roberts E. Inhibitory GABAergic nerve terminals decrease at sites of focal epilepsy. *Science* 1979;205:211–4.
- [36] Williamson A, Patrylo P, Spencer DD. Decrease in inhibition in dentate granule cells from patients with medial temporal lobe epilepsy. *Ann Neurol* 1999;45:92–9.
- [37] Babb TH. Synaptic reorganizations in human and rat hippocampal epilepsy. In: Delgado Escueta AV, Wilson WA, Olsen RW, Porter RJ, editors, *Jasper's basic mechanisms of the epilepsies*, 3rd ed, *Advances in Neurology*, vol. 79, Philadelphia: Lippincott Williams and Wilkins, 1999, pp. 763–79.
- [38] McDonald JW, Garofalo EA, Hood T, Sackellares JC, Gilman S, McKeever PE et al. Altered excitatory and inhibitory amino acid receptor binding in hippocampus of patients with temporal lobe epilepsy. *Ann Neurol* 1991;29:529–41.
- [39] Wyler AR, Nadi S, Porter RJ. Acetylcholine, GABA, benzodiazepine and glutamate receptors in the temporal lobe of epileptic patients. *Neurology* 1987;37(Suppl 1):103.
- [40] Sherwin A, Robitaille Y, Quesney F, Olivier A, Villemure J, Leblanc R et al. Excitatory amino acids are elevated in human epileptic cerebral cortex. *Neurology* 1988;38:920–3.
- [41] Van Gelder NM, Sherwin AL, Rasmussen T. Amino acid content of epileptogenic human brain: focal vs. surrounding regions. *Brain Res* 1972;40:385–93.
- [42] Kish SJ, Dixon LM, Sherwin AL. Aspartic acid aminotransferase activity is increased in actively spiking compared with non-spiking human epileptic cortex. *J Neurol Neurosurg Psychiatr* 1988;51:552–6.
- [43] Piredda S, Gale K. Role of excitatory amino acid transmission in the genesis of seizures elicited from the deep prepiriform cortex. *Brain Res* 1986;377:205–10.
- [44] Chapman AG, Meldrum BS, Mendes E. Acute anticonvulsant activity of structural analogues of valproic acid and changes in brain GABA and aspartate content. *Life Sci* 1983;32:2023–7.
- [45] Logothetis J, Bovis M. Free amino acid changes in the spinal fluid of patients with central nervous system degeneration and chronic epileptic disorders. *World Neurol* 1962;3:466–74.
- [46] Isakson PJ, Huntsman MM, Murray KD, Gall CM. BDNF mRNA expression is increased in adult rat forebrain after limbic seizures: temporal patterns of induction distinct from NFG. *Neuron* 1991;6:937–48.
- [47] Dashieff RM, Byrne MC, Patrone V, McNamara JO. Biochemical evidence of decreased muscarinic cholinergic neuronal communication following amygdala-kindled seizures. *Brain Res* 1981;206:233–8.
- [48] McNamara JO, Peper AM, Patrone V. Repeated seizures induce long-term increase in hippocampal benzodiazepine receptors. *Proc Natl Acad Sci USA* 1980;77(5):3029–32.
- [49] Delgado-Escueta AV, Davidson D, Reilly EL. Potassium transport within synaptosomes isolated from epileptogenic foci. *Brain Res* 1974;78:223–37.
- [50] Brines M, Tabuteau H, Sundaresan S, Kim J, Spencer D, de Lanerolle N. Regional distributions of hippocampal  $\text{Na}^{++}$ ,  $\text{K}^{+}$ –ATPase, cytochrome oxidase, and total protein in temporal lobe epilepsy. *Epilepsia* 1995;36(4):371–83.
- [51] Mison-Crighel N, Constantinescu E, Costa-Foru D, Crighel E. Changes in the activity of some enzyme systems determined in the cortical scar and at the level of secondary degenerative lesions. *Acta Neurol Scand* 1962;38(Suppl 1):S81.
- [52] Rosenblatt DE, Lauter CJ, Trams EG. Deficiency of a  $\text{Ca}^{++}$ –ATPase in brains of seizure prone mice. *J Neurochem* 1976;29:1299–304.
- [53] Delgado-Escueta AV, Horan MP. Neurobiology. General principles related to epilepsy. In: Glaser GH, Penry JK, Woodbury DM, editors, *Antiepileptic drugs. Mechanisms of action*, New York: Raven Press, 1980, pp. 85–126.
- [54] Bengzon J, Kokaia Z, Elmer E et al. Apoptosis and proliferation of dentate gyrus neurons after single and intermittent limbic seizures. *Proc Natl Acad Sci USA* 1997;94:10432–7.
- [55] Zhang LX, Smith MA, Li X et al. Apoptosis of hippocampal neurons after amygdala kindled seizures. *Mol Brain Res* 1998;55:198–208.
- [56] Pumain R, Kurcewicz Y, Louvel J. Ionic changes induced by excitatory amino acids in the rat cerebral cortex. *Can J Physiol Pharmacol* 1987;65:1067–77.
- [57] Nowak L, Bregestovski P, Ascher P, Herbet A, Prochiantz A. Magnesium gates glutamate-activated channels in mouse central neurones. *Nature* 1984;307:462–5.
- [58] Curtis DR, Johnston GAR. Amino acid transmitter in the mammalian central nervous system. *Ergeb Physiol* 1974;69:97–188.
- [59] Mody I, Stanton PK, Heinemann U. Activation of *N*-methyl-D-aspartate receptors parallels changes in cellular and synaptic properties of dentate granule cells after kindling. *J Neurophysiol* 1988;59:1033–54.
- [60] Krnjevic K. Role of neurotransmitters in the genesis of epileptiform discharges. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, *Generalized epilepsy: neurobiological approaches*, Boston: Birkhäuser, 1990, pp. 86–101.
- [61] Perry TL, Hansen S. Amino acid abnormalities in epileptogenic foci. *Neurology* 1981;31:872–6.
- [62] Tower DB. Nature and extent of the biochemical lesion in human epileptogenic cerebral cortex. *Neurology* 1955;5:113–30.
- [63] Berl S, Purpura DP, Girado M, Waelsh H. Amino acid metabolism in epileptogenic and nonepileptogenic lesions of the neocortex (cat). *J Neurochem* 1959;4:311–7.
- [64] Tower DB. The neurochemistry of convulsive states and allied disorders. In: Folch Pi J, editor, *Chemical pathology of the nervous system*, Oxford: Pergamon, 1961, pp. 307–44.

- [65] Mutani R, Monaco F, Durelli L, Delsedime M. Free aminoacids in the cerebrospinal fluid of epileptic subjects. *Epilepsia* 1974;15:593–7.
- [66] Merlin LR, Wong RKS. Effects of mGluR antagonists on epileptiform activities in hippocampal slices. *Soc Neurosci Abstr* 1994;20:398.
- [67] Benaïse L, Fornai F, Dybdal D, Gale K. Critical role of AMPA receptors in area tempestas in generating status epilepticus in rats. *Soc Neurosci Abstr* 1994;20:403.
- [68] Comair Y, Tocco G, Najm I, Kaakaji R, Luders H, Baudry M. Changes in hippocampal glutamate/AMPA receptors in epileptic human hippocampus. *Soc Neurosci Abstr* 1994;20:1452.
- [69] Telfeian AE, During M, Federoff HJ, Mirchandani G, Williamson A. Long term changes in hippocampal excitability following over-expression of GluR6. *Soc Neurosci Abstr* 1994;20:1667.
- [70] Feldblum S, Rougier A, Loiseau H, Cohadon F, Morselli PL, Lloyd KG. Quinolinic-phosphoribosyl transferase activity is decreased in epileptic human tissue. *Epilepsia* 1988;29:523–9.
- [71] Thomson AM, Walker VE, Flynn DM. Glycine enhances NMDA-receptor mediated synaptic potentials in neocortical slices. *Nature* 1989;338:422–4.
- [72] Peters S, Koh J, Choi DW. Zinc selectively blocks the action of *N*-methyl-D-aspartate on cortical neurons. *Science* 1987;236:589–93.
- [73] Barbeau A, Donaldson J. Zinc, taurine and epilepsy. *Arch Neurol* 1974;30:52–8.
- [74] Anthony WL, Woosley RL, Hsu JM. Urinary excretion of radiosulfur following taurine-35 S injection in zinc deficient rats. *Proc Soc Exp Biol Med* 1971;138:989–92.
- [75] Pasantes-Morales H, Arzate NE, Cruz C. The role of taurine in nervous tissue: its effects on ionic fluxes. *Adv Exp Med Biol* 1981;139:273–92.
- [76] Gale K. GABA in epilepsy: the pharmacologic basis. *Epilepsia* 1989;30(Suppl 3):S1–S11.
- [77] Sperber EF, Wong BY, Wurlpel JN, Moshe SL. Nigral infusions of muscimol or bicuculline facilitate seizures in developing rats. *Brain Res* 1987;465:243–50.
- [78] McKernan RM, Whiting PJ. Which GABAA receptor-subtypes really occur in the brain. *Trends Neurosci* 1996;19:139–43.
- [79] Nicoll RA, Eccles JC, Oshima T, Rubia R. Prolongation of hippocampal inhibitory postsynaptic potentials by barbiturates. *Nature* 1985;258:625–7.
- [80] Study RE, Barker JL. Diazepam and pentobarbital: Fluctuation analysis reveals different mechanisms for potentiation of gamma-amino butyric acid responses in cultured spinal cord neurons. *Proc Natl Acad Sci USA* 1981;76:7180–4.
- [81] Mathers DA, Barker JL. Pentobarbital opens ion channels of long duration in cultured mouse spinal neurons. *Science* 1980;209:507–9.
- [82] Macdonald RL, Barker JL. Pentylenetetrazol and penicillin are selective antagonists of GABA-mediated post-synaptic inhibition in cultured mammalian neurons. *Nature* 1977;267:720–1.
- [83] Enna SJ, Gallagher JP. Biochemical and electrophysiological characteristics of mammalian GABA receptors. *Int Rev Neurobiol* 1983;24:181–212.
- [84] Ribak CE, Harris AB, Vaughn JE, Roberts E. Immunocytochemical changes in cortical neurons in a monkey model of epilepsy. In: Marselli PL, Lloyd KG, Loscher H et al., editors. *Neurotransmitters, seizures and epilepsy*, New York: Raven Press, 1981, pp. 11–22.
- [85] Savic I, Persson A, Roland P, Pauli S, Sedvall G, Widen L. In-vivo demonstration of reduced benzodiazepine receptor binding in human epileptic foci. *The Lancet* 1988;15:863–6.
- [86] Savic I, Pauli S, Thorell JO, Blomqvist G. In-vivo demonstration of altered benzodiazepine receptor density in patients with generalized epilepsy. *J Neurol Neurosurg Psychiatr* 1994;57:797–804.
- [87] Silvette H. The actions of nicotine on central nervous system functions. *Pharmacol Rev* 1962;14:137–73.
- [88] Segal M. Acetylcholine and epilepsy. In: Fisher RS, Coyle JT, editors. *Neurotransmitters and epilepsy*, New York: Wiley-Liss, 1991, pp. 95–101.
- [89] Kish SJ, Olivier A, Dubeau F, Robitaille Y, Sherwin AL. Increased activity of choline acetyltransferase and acetylcholinesterase in actively epileptic human cerebral cortex. *Epilepsy Res* 1988;2:227–31.
- [90] Segal M. Serotonin and epilepsy. In: Fisher RS, Coyle JT, editors. *Neurotransmitters and epilepsy*, New York: Wiley-Liss, 1991, pp. 103–8.
- [91] Colino A, Halliwell JV. Differential modulation of three separate K conductances in hippocampal CA1 neurons by serotonin. *Nature* 1987;328:73–6.
- [92] Andrade R, Nicoll RA. Pharmacologically distinct actions of serotonin on single pyramidal neurons of the rat hippocampus recorded in vitro. *J Physiol* 1987;394:99–124.
- [93] Segal M. Developmental changes in serotonin actions in rat hippocampus. *Dev Brain Res* 1990;52:247–52.
- [94] Waterhouse BD. Electrophysiological assessment of monoamine synaptic function in neuronal circuits of seizure susceptible brains. *Life Sci* 1986;39:807–18.
- [95] Alexander GJ, Kopeloff LM. Metrazol seizures in rats: Effect of *p*-chlorophenylalanine. *Brain Res* 1970;22:231–5.
- [96] Killian M, Frey HH. Central monoamines and convulsive thresholds in mice and rats. *Neuropharmacol* 1973;12:681–92.
- [97] Browning RA. Role of the brain stem reticular formation in tonic-clonic seizures: lesion and pharmacological studies. *Fed Proc* 1985;44(8):2425–31.
- [98] Brière R, Sherwin AL, Robitaille Y, Olivier A, Quesney LF, Reader TA. Alpha 1 adrenoceptors are decreased in human epileptic foci. *Ann Neurol* 1986;19:26–30.
- [99] Devinsky O, Emoto S, Goldstein D, Estull R, Porter R, Theodore W et al. Cerebrospinal fluid and serum levels of DOPA, catechols and monoamine metabolites in patients with epilepsy. *Epilepsia* 1992;33:263–70.
- [100] Corcoran ME, Mason ST. Role of forebrain catecholamines in amygdaloid kindling. *Brain Res* 1980;190:473–84.
- [101] Grailliot C, Baumann N, Maurin Y. Modulation of alpha 1 and alpha 2-adrenoceptor binding sites in the brain of audiogenic seizure susceptible mice (DBA/2J). *Eur J Pharmacol* 1985;118:231–7.
- [102] Chermat R, Lachapelle F, Baumann N, Simon P. Anticonvulsant effect of yohimbine in quaking mice: antagonism by clonidine and prazosine. *Life Sci* 1979;125:1471–6.
- [103] Jimenez-Rivera C, Voltura A, Weiss GK. Effects of locus coeruleus stimulation on the development of kindled seizures. *Exp Neurol* 1987;95:13–20.
- [104] Benardo LS, Prince DA. Dopamine action on hippocampal pyramidal cells. *J Neurosci* 1982;2(4):415–23.
- [105] Johnson DD, Jaju AT, Ness L, Richardson JS, Crawford RD. Brain norepinephrine, dopamine and 5-hydroxytryptamine concentration abnormalities and their role in the high seizure susceptibility of epileptic chickens. *Can J Physiol Pharmacol* 1981;59:144–9.
- [106] Reader TA. The effects of dopamine, noradrenaline and serotonin in the visual cortex of the cat. *Experientia* 1978;34:1586–8.
- [107] Ferrendelli JA. Roles of biogenic amines and cyclic nucleotides in seizure mechanisms. *Ann Neurol* 1984;16(Suppl):S98–S103.
- [108] Hoffer B, Seiger A, Freedman R, Olson L, Taylor D. Electrophysiology and cytology of hippocampal formation transplants in the anterior chamber of the eye II. Cholinergic mechanisms. *Brain Res* 1977;119:108–32.
- [109] Iwasa H, Kikuchi S, Suzuki K, Sato T, Hasegawa S. Persistent increase in G protein subclass mRNAs in kindling-elicited epileptogenesis. *Epilepsia* 1995;36(Suppl 3):S49.
- [110] Dubeau F, Sherwin A, Olivier A, Villemure R, Leblanc L, Quesney LF et al. Excitatory amino acids modulate phosphoinositide signal transduction in human epileptic neocortex. *Epilepsia* 1992;33(2):255–62.

- [111] Stelzer A, Feasey KJ, Moneta ME, Sincini E, Bruggencate GT, Noble EP. Inositol 1-phosphate formation in long term potentiation and kindling. *Brain Res* 1989;490:41–7.
- [112] Simmonds SH, Strange PG. Inhibition of inositol phospholipid breakdown by D2 dopamine receptors in dissociated bovine anterior pituitary cells. *Neurosci Lett* 1985;60:267–72.
- [113] Bazán NG, Birkle DL, Tang W, Reddy TS. The accumulation of free arachidonic acid and the formation of prostaglandins and lipoxygenase reaction products in the brain during experimental epilepsy. In: Delgado Escueta AV, Ward Jr. AA, Woodbury DM, editors, *Advances in neurology, Basic mechanisms of the epilepsies. Molecular cellular approaches*, vol. 44, New York: Raven Press, 1986, pp. 879–902.
- [114] Baraban JM, Cole AJ, Stratton KR, Pritchett J, Alvaro J, Abu-Shakra S et al. Neuronal excitability: focus on second messengers system. In: Fisher RS, Coyle JT, editors, *Neurotransmitters and epilepsy*, New York: Wiley-Liss, 1991, pp. 33–45.
- [115] Ferrendelli JA, Daniels MC, Queen S. Comparative actions of phenytoin and other anticonvulsant drugs on potassium - and veratridine-stimulated calcium uptake in synaptosomes. *J Pharmacol Exp Ther* 1982;220:29–34.
- [116] De Lorenzo RJ. Ionic channels, membranes and molecules understanding epilepsy and neuronal excitability. In: Dodson WE, Pellock JM, editors, *Pediatric epilepsy. Diagnosis and therapy*, New York: Demos Publications Inc, 1993, pp. 17–25.
- [117] De Lorenzo RJ. A molecular approach to the calcium signal in brain: relationship to synaptic modulation and seizure discharge. *Adv Neurol* 1986;44:435–64.
- [118] Goldenring JR, Wasterlain CG, Oestreicher A, de Graan P, Farber D, Glaser G et al. Kindling induces a long-lasting change in the activity of a hippocampal membrane calmodulin-dependent protein kinase system. *Brain Res* 1986;377:47–53.
- [119] Costa MR, Caterall WA. Phosphorylation of the alpha subunit of the sodium channel by protein kinase C. *Cell Mol Neurobiol* 1984;4:291–7.
- [120] Pumain R, Heinemann U. Extracellular ions in epileptiform discharge. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, *Generalized epilepsy: neurobiological approaches*, Boston: Birkhäuser, 1990, pp. 61–85.
- [121] Chin JH. Adenosine receptors in brain: neuromodulation and role in epilepsy. *Ann Neurol* 1989;26:695–8.
- [122] Maitre M, Ciesielski L, Lehmann A, Kempf E, Mandel P. Protective effect of adenosine and nicotinamide against audiogenic seizures. *Biochem Pharmacol* 1974;23:2807–16.
- [123] Dunwiddie TV, Worth T. Sedative and anti-convulsant effects of adenosine analogs in mouse and rat. *J Pharmacol Exp Ther* 1982;220:70–6.
- [124] Dragunow C, Goddard GV, Lavery R. Is adenosine an endogenous anticonvulsant? *Epilepsia* 1985;26(5):480–7.
- [125] Haas HL, Greene RW. Endogenous adenosine inhibits hippocampal CA1 neurons: further evidence from extra- and intracellular recording. *Naunyn Schmiedeberg Arch Pharmacol* 1988;337:561–5.
- [126] Dunwiddie TV, Haas HL. Adenosine increases synaptic facilitation in the in-vitro rat hippocampus: evidence for a presynaptic site of action. *J Physiol (Lond)* 1989;369:365–77.
- [127] During MJ, Spencer DD. Adenosine: a potential mediator of seizure arrest and postictal refractoriness. *Ann Neurol* 1992;32:618–24.
- [128] Henriksen SJ, Bloom FE, McCoy F, Ling N, Guillemin R.  $\beta$ -endorphin induces non-convulsive limbic seizures. *Proc Natl Acad Sci USA* 1978;75:5221–5.
- [129] Frenk H, Urca G, Liebeskind JC. Epileptic properties of leucine- and methionine-enkephalin: comparison with morphine and reversibility naloxone. *Brain Res* 1978;147:327–37.
- [130] Snead OC. Opiate-induced seizures: a study of mu and delta specific mechanisms. *Exp Neurol* 1986;93:348–58.
- [131] Urca G, Frenk H, Liebeskind JC, Taylor AN. Morphine and enkephalin: analgesic and epileptic properties. *Science* 1977;197:83–6.
- [132] Urca G, Frenk H. Systemic morphine blocks the seizure induced by intracerebroventricular (i.c.v.) injections of opiates and opiate peptides. *Brain Res* 1982;246:121–6.
- [133] Frenk H, Engel J, Ackermann RF, Sharit Y, Liebeskind JC. Endogenous opioids may mediate postictal behavior depression in amygdaloid kindled rats. *Brain Res* 1979;167:435–40.
- [134] Holaday JW, Belenky G. Opiate-like effects of electroconvulsive shock in rats: a differential effect of naloxone on nociceptive measures. *Life Sci* 1980;27:1929–34.
- [135] de Lanerolle NC, Sundaresan S, Brines ML, Spencer DD. Phenotypic changes of hippocampal neurons and dynorphin in human epilepsy. *Soc Neurosci Abstr* 1989;15:340.
- [136] Mayberg HS, Sadzot B, Meltzer CC, Fisher RS, Lesser RP, Dannals RF et al. Quantification of Mu and Non-Mu opiate receptors in temporal lobe epilepsy using positron emission tomography. *Ann Neurol* 1991;30:3–11.
- [137] Frost JJ. Imaging mu-opiate receptors in epilepsy by positron emission tomography. *Semin Neurol* 1989;9:317–22.
- [138] Macdonald RL. Antiepileptic drug actions on neurotransmitters receptors and ion channels. In: Fisher RS, Coyle JT, editors, *Neurotransmitters and epilepsy*, New York: Wiley-Liss, 1991, pp. 231–45.
- [139] Phillips HA, Scheffer IE, Berkovic SF, Hollway GE, Sutherland GR, Mulley JC. Localization of a gene for autosomal dominant nocturnal frontal lobe epilepsy to chromosome 20q13.2 (letter). *Nat Genet* 1995;10(1):117–8.
- [140] Phillips HA, Scheffer IE, Crossland KM, Bhatia KP, Fish DR, Marsden CD et al. Autosomal dominant nocturnal frontal lobe epilepsy: genetic heterogeneity and evidence for a second locus at 15q24. *Am J Hum Genet* 1998;63(4):1108–16.
- [141] Nakken KO, Magnusson A, Steinlein OK. Autosomal dominant nocturnal frontal lobe epilepsy. An electroclinical and genetic description of a Norwegian family with ten affected members. *Tidsskr Nor Laegeforen* 1998;118(5):716–8.
- [142] Steinlein OK, Mulley JC, Propping P, Wallace RH, Phillips HA, Sutherland GR, Scheffer IE, Berkovic SF. A missense mutation in the neuronal nicotinic acetylcholine receptor alpha 4 subunit is associated with autosomal dominant nocturnal frontal lobe epilepsy. *Nat Genet* 1995;11(2):201–3.
- [143] Poza JJ, Saenz A, Martínez Gil A, Cheron N, Cobo AM, Urtasun M et al. Autosomal dominant lateral temporal epilepsy: clinical and genetic study of a large Basque pedigree linked to chromosome 10q. *Ann Neurol* 1999;45(2):182–8.
- [144] Scheffer IE, Phillips HA, O'Brien CE, Sailing MM, Wrennall JA, Wallace RH et al. Familial partial epilepsy with variable foci: a new partial epilepsy syndrome with suggestion of linkage to chromosome 2. *Ann Neurol* 1998;44(6):890–9.
- [145] Steinlein O, Schuster V, Fischer C, Haussler M. Benign familial neonatal convulsions: confirmation of genetic heterogeneity and further evidence for a second locus on chromosome 8q. *Hum Genet* 1995;95(4):411–5.
- [146] Wallace RH, Berkovic SF, Howell RA, Sutherland GR, Mulley JC. Suggestion of a major gene for familial febrile convulsions mapping to 8q13-21. *J Med Genet* 1996;33(4):308–12.
- [147] Johnson EW, Dubovsky J, Rich SS, O'Donovan CA, Orr HT, Anderson VE et al. Evidence for a novel gene for familial febrile convulsions, FEB2 linked to chromosome 19p in an extended family from the Midwest. *Hum Mol Genet* 1998;7(1):63–7.
- [148] Neubauer BA, Fiedler B, Himmelein B, Kampfer F, Lassar U, Schwabe G et al. Centrottemporal spikes in families with rolandic epilepsy. Linkage to chromosome 15q14. *Neurology* 1998;51(6):1608–12.
- [149] Weissbecker KA, Durner M, Janz D, Scaramelli A, Sparkes RS, Spence MA. Confirmation of linkage between juvenile myoclonic epilepsy locus and the HLA region of chromosome 6. *Am J Med Genet* 1991;38(1):32–6.
- [150] Elmslie FV, Rees M, Williamson MP, Kerr M, Kjeldsen MJ, Pang



- KA et al. Genetic mapping of a major susceptibility locus for juvenile myoclonic epilepsy on chromosome 15q. *Hum Mol Genet* 1997;6(8):1329–34.
- [151] Morgan JI, Cohen DR, Hempstead JL, Curran T. Mapping patterns of c-fos expression in the central nervous system after seizure. *Science* 1987;237:192–7.
- [152] Lamph WW, Wamsley P, Sassone-Corsi P, Verma IM. Induction of proto-oncogene JUN/AP-1 by serum and TPA. *Nature* 1988;334:629–31.
- [153] Sprengel R, Higuchi M, Monyer H, Seeburg PH. Glutamate receptor channels: a possible link between RNA editing in the brain and epilepsy. In: Delgado Escueta AV, Wilson WA, Olsen RW, Porter RJ, editors, *Jasper's basic mechanisms of the epilepsies*, 3rd ed, *Advances in neurology*, vol. 79, Philadelphia: Lippincott Williams and Wilkins, 1999, pp. 525–34.
- [154] Engel Jr. J. Update on surgical treatment of the epilepsies. *Neurology* 1993;43:1612–7.
- [155] ILAE Commission Report. A global survey on epilepsy surgery, 1980–1990: a report by the Commission on Neurosurgery of Epilepsy. *Epilepsia* 1997;38(2):249–255.
- [156] Avoli M, Gloor P, Kostopoulos G. Focal and generalized epileptiform activity in the cortex. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, *Generalized epilepsies: neurobiological approaches*, Boston: Birkhäuser, 1990, pp. 213–31.
- [157] Vornov JJ, Coyle JT. Mechanisms of neurotransmitters receptor actions. In: Fisher RS, Coyle JT, editors, *Neurotransmitters and epilepsy*, New York: Wiley-Liss, 1991, pp. 17–31.
- [158] Rogers SW, Andrews PI, Gahring LC, Whisenand T, Cauley K, Crain B et al. Autoantibodies to glutamate receptor 3 in Rasmussen's encephalitis. *Science* 1994;265:648–51.
- [159] Mody I, Hablitz J, Malenka R, Schoepp D. Excitation and inhibition through GABAB and metabotropic glutamate receptors. *Epilepsia* 1993;34(Suppl.6):S1.
- [160] Gloor P. Epilepsy: relationship between electrophysiology and intracellular mechanisms involving second messengers and gene expression. *Can J Neurol Sci* 1989;16:8–21.
- [161] Ure JA. Biochemical aspects of the epilepsies. *Neuropsychiatria* 1976;2:17–25.
- [162] Brusa R, Zimmermann F, Koh DS et al. Early-onset epilepsy and postnatal lethality associated with an editing-deficient GluR-B allele in mice. *Science* 1995;270:1677–80.
- [163] Faingold CL, Meldrum BS. Excitant amino acids in epilepsy. In: Avoli M, Gloor P, Kostopoulos G, Naquet R, editors, *Generalized epilepsy: neurobiological approaches*, Boston: Birkhäuser, 1990, pp. 102–17.
- [164] Upton N, Blackburn T. Pharmacology of mammalian GABA<sub>A</sub> receptors. In: Enna SJ, Bowery NG, editors, *The GABA receptors*, 2nd ed, Totowa, New Jersey: Humana Press Inc, 1997, pp. 83–120.
- [165] Hirasawa K, Nishizuka Y. Phosphatidylinositol turnover in receptor mechanism and signal transduction. *Ann Rev Pharmacol Toxicol* 1985;25:147–70.