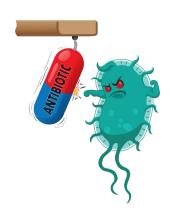
Predicting Antibiotic Resistance: Challenges, Findings, and Lessons Learned



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The Prescription Challenge

 When a patient presents with a bacterial infection, clinicians must choose among several possible antibiotics, often before lab results are available

Resistance testing takes 24–72
 hours and may not be routinely available in all hospitals.

 Incorrect or delayed antibiotic choices can worsen patient outcomes and contribute to rising resistance.



Aims & Objectives

• <u>Objective:</u> Explore whether patient and microbiological data can **predict antibiotic resistance**.

• Impact:

- Support clinicians in making data-informed antibiotic prescriptions while awaiting lab results.
- Identify key resistance risk factors to guide more effective treatment decisions.

Approach:

- Train machine learning models that classify each antibiotic as susceptible or resistant.
- Examine which patient and clinical features (e.g., age, ward type, prior antibiotic exposure)
 most influence resistance.



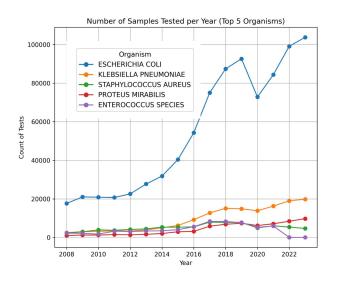
Dataset & Preliminary Preprocessing

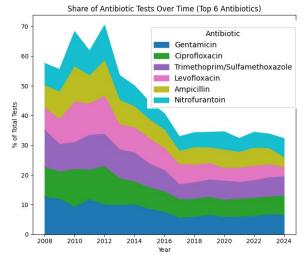
- We use the <u>Dryad database</u> (Fateme et. al, 2025):
 ~750k microbiological culture records from ~300k unique patients (1999-2024).
- Features: age, prior infections, prior antibiotics, adi score, nursing home visits, ward
- Preliminary Preprocessing:
 - Removed duplicates
 - Removed repeated visits → first visit only
 - One-hot encoded categorical features (e.g. ward info)

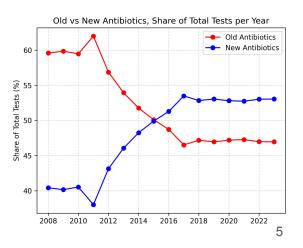
Anon ID	Prior Organism Exposure	Prior Antibiotics Used	Age of Subject	Gender	Area Deprivation Index (ADI Score)	ADI State Rank	Days between Nursing Home Visits & Blood Culture	Bacterial Culture from Inpatients	Bacterial Culture from Outpatients	Bacterial Culture from Emergency Room	Bacterial Culture from ICU
1164374	2	1	70	0	6	3	0	0	1	0	0
1064815	0	1	50	0	22	7	0	0	1	0	0

Additional Preprocessing Based on EDA

- Focused on *E. coli* isolates only (most frequently tested organism)
- Selected data from 2016 onward to reflect current resistance patterns and antibiotic guidelines
- Kept only the 9 antibiotics tested for ≥75% of *E. coli* patients







Modeling Approach

Goal: Predict if a patient's *E. coli* isolate is resistant (1) or susceptible (0).

Target: Each antibiotic susceptibility/resistance

Models Tested:

- Dummy Classifier baseline
- Logistic Regression
- Random Forest
- XGBoost
- Support Vector Machine (SVM)
- K-Nearest Neighbors (KNN)

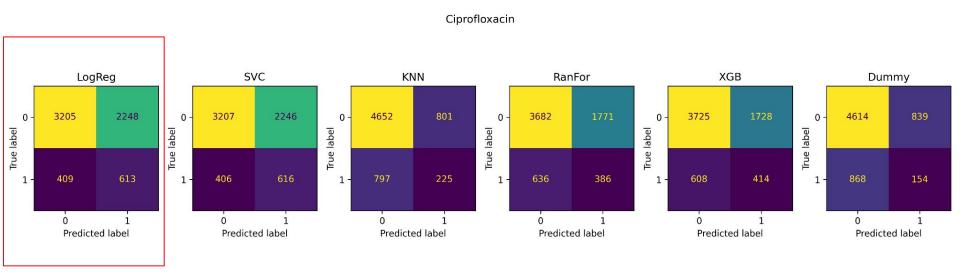
Pipeline Highlights:

- Data split 80% train / 20% test
- Outer 5-fold CV on training set for model selection
- Inner 3-fold CV for hyper-parameter tuning with Randomized Search CV
- StandardScaler applied for distance-based models (SVM, KNN, LogReg)
- SMOTE enhance learning performance over highly imbalance data
- Downsample on larger class to balance the dataset 50/50

Evaluation Metrics

- **F1 Weighted Score:** Harmonic mean of precision & recall
- False Negative Rate (FNR): Critical metric representing the % of resistant cases misclassified as susceptible. (Should be minimized, since missing a resistant infection can lead to ineffective treatment.)
- **Precision:** Correctly predicted resistant out of all predicted resistant
- Recall / Sensitivity: Correctly identified resistant cases
- Accuracy: Overall correctness
- RR AUC: Correctly distinguishing between positive/negative cases

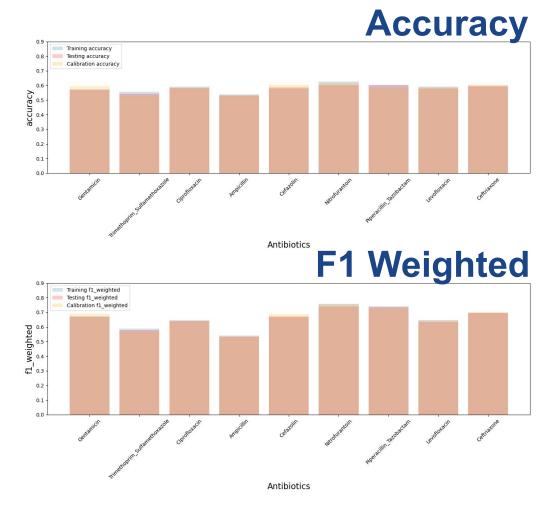
Confusion Matrices



Chosen model: logistic regression

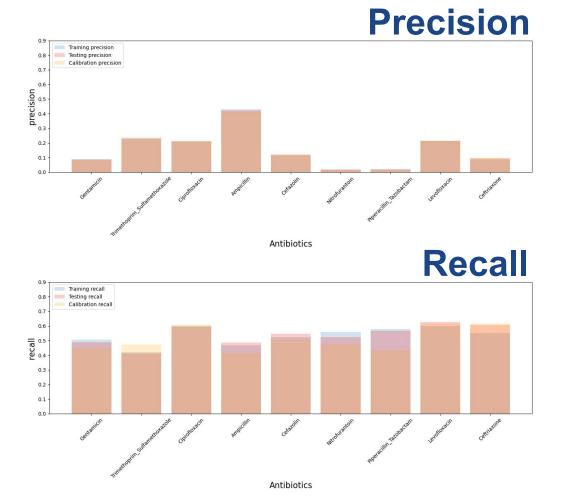
Results

- Looked at evaluation metrics between training, testing, and calibration dataset for any sign of over or under training.
- Stable accuracy and small variability in F1 weighted across antibiotics.



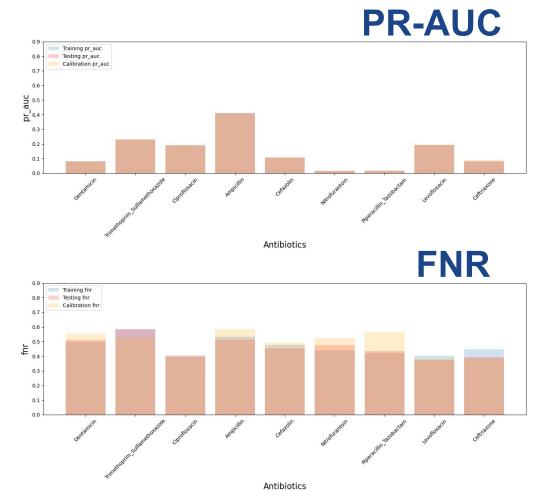
Results

 Large variability in Precision and small variability in recall across antibiotics



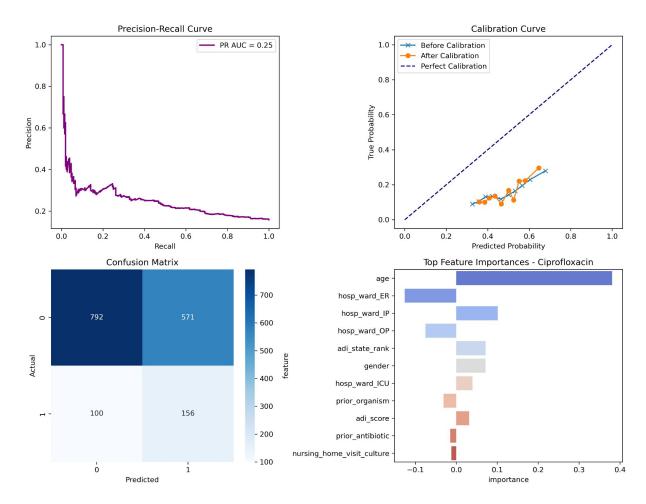
Results

 Large variability in PR-AUC and small variability in recall across antibiotics



Logistic Regression Model Evaluation for Ciprofloxacin

Results



Conclusions

- Completed rigorous data cleaning, preprocessing, and model calibration.
- Achieved decent accuracy and F1 weighted score across most antibiotics.
- False-negative rates remain high, signaling limited sensitivity to predicting resistance.

1. Feature Limitations

Datasets lack detailed biological or treatment features due to privacy, limiting model depth.

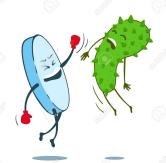
2. Data Imbalance

Resistant cases are rare (e.g., Ceftriaxone: 7%). Even with SMOTE, models biased toward accuracy inflate false negatives.

3. Biological Complexity

 Resistance stems from diverse, nonlinear mechanisms—mutations, plasmids, efflux pumps—hard to capture computationally.





Future Directions

1. Time-Series Modeling

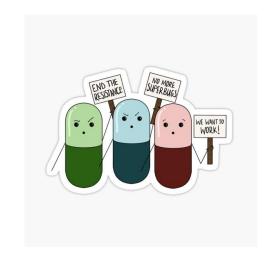
Track monthly resistance trends and forecast future shifts.

2. Extended Feature Importance Exploration

Use richer data to pinpoint key biological, clinical, and socioeconomic factors.

3. Cross-Antibiotic Correlation

Analyze co-resistance patterns to reveal links across different drug classes.



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