

Class10

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Introduction to the RCSB Protein Data Bank (PDB)

The Protein Data Bank(PDB) is the main repository of biomolecular structure. ### PDB statistics

```
PDB.data <- read.csv('Data Export Summary.csv',row.names = 1)
PDB <- data.frame(PDB.data)
PDB
```

	X.ray	EM	NMR	Integrative	Multiple.methods
Protein (only)	178,795	21,825	12,773	343	226
Protein/Oligosaccharide	10,363	3,564	34	8	11
Protein/NA	9,106	6,335	287	24	7
Nucleic acid (only)	3,132	221	1,566	3	15
Other	175	25	33	4	0
Oligosaccharide (only)	11	0	6	0	1
	Neutron	Other	Total		
Protein (only)	84	32	214,078		
Protein/Oligosaccharide	1	0	13,981		
Protein/NA	0	0	15,759		
Nucleic acid (only)	3	1	4,941		
Other	0	0	237		
Oligosaccharide (only)	0	4	22		

Q1: What percentage of structures in the PDB are solved by X-Ray and Electron Microscopy.

The comma in these numbers leads to the numbers here being read as character.

```
library(readr)
stats <- read_csv('Data Export Summary.csv')
stats

# A tibble: 6 x 9
  `Molecular Type` `X-ray`    EM    NMR Integrative `Multiple methods` Neutron
  <chr>           <dbl>    <dbl> <dbl>        <dbl>           <dbl>    <dbl>
1 Protein (only)   178795  21825 12773        343          226     84
2 Protein/Oligosacch~ 10363   3564   34          8          11      1
3 Protein/NA       9106    6335   287         24          7      0
4 Nucleic acid (only) 3132    221   1566         3         15      3
5 Other            175     25    33          4          0      0
6 Oligosaccharide (~ 11      0     6          0          1      0
# i 2 more variables: Other <dbl>, Total <dbl>

n.Xray <- sum(stats$`X-ray`)
n.Total <- sum(stats$`Total`)
per.Xray <- n.Xray/n.Total
per.Xray
```

```
[1] 0.8095077
```

80.95% of structures in the PDB are solved by X-Ray and Electron Microscopy.

Q2: What proportion of structures in the PDB are protein?

```
per.protein <- stats[1,'Total']/n.Total
per.protein
```

```
      Total
1 0.8596889
```

85.97% of structures in the PDB are protein (only).

Q3: Type HIV in the PDB website search box on the home page and determine how many HIV-1 protease structures are in the current PDB?

644 HIV-1 protease structures are in the current PDB.

Visualizing the HIV-1 protease structure

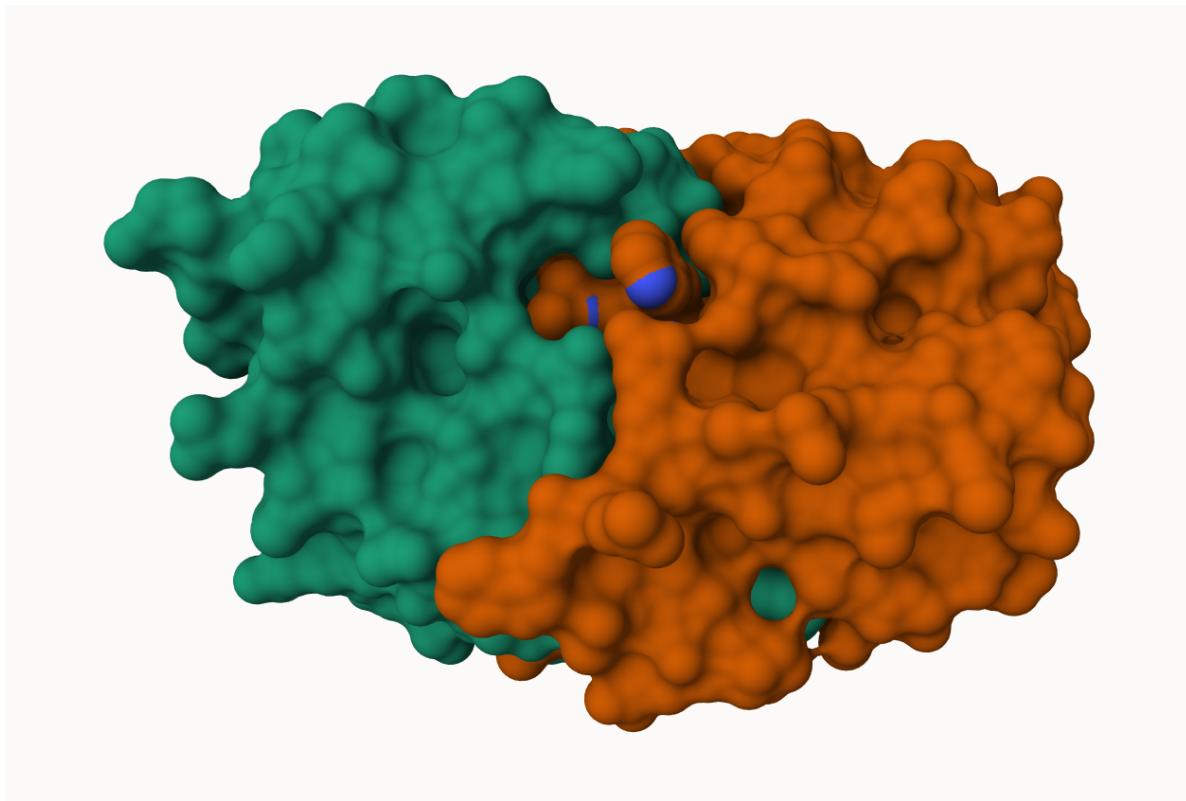


Figure 1: HIV-pr 1HSG with surface display showing ligand binding

Q4: Water molecules normally have 3 atoms. Why do we see just one atom per water molecule in this structure?

The structure was constructed using X-RAY DIFFRACTION according to the PDB website. As the electron density of hydrogen atoms was too weak to be detected, the X-RAY DIFFRACTION only showed the oxygen atoms as they have a much stronger electron density.

Q5: There is a critical “conserved” water molecule in the binding site. Can you identify this water molecule? What residue number does this water molecule have

HOH308

Q6: Generate and save a figure clearly showing the two distinct chains of HIV-protease along with the ligand. You might also consider showing the catalytic residues ASP 25 in each chain and the critical water (we recommend “Ball & Stick” for these side-chains). Add this figure to your Quarto document.

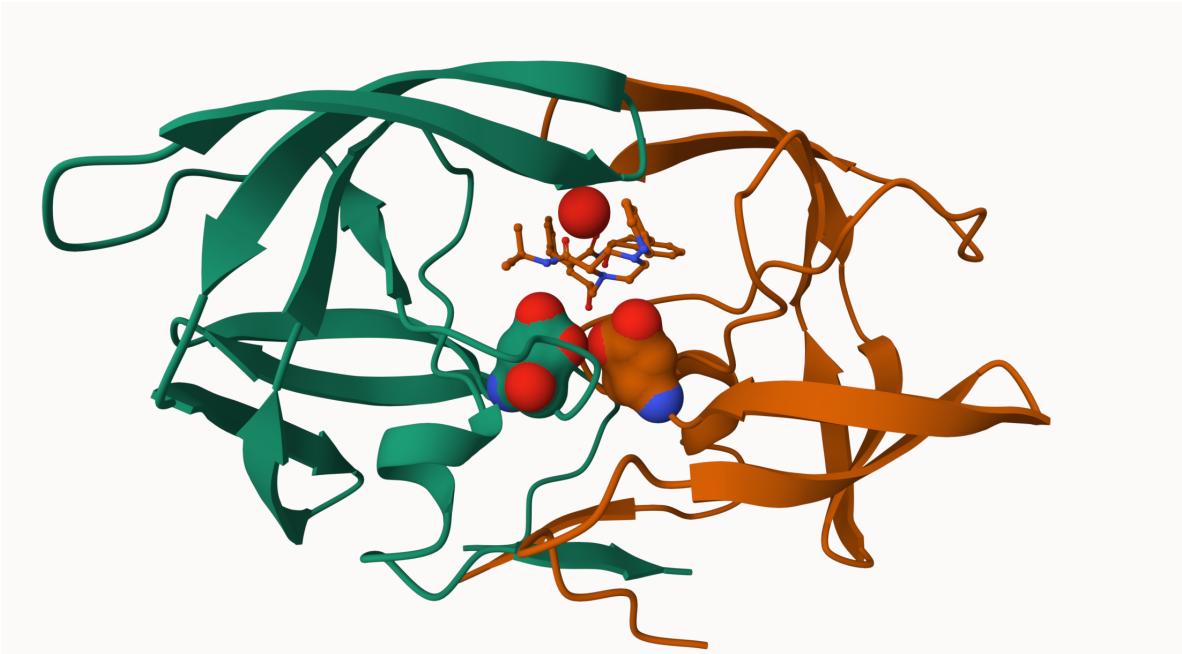


Figure 2: HIV-pr 1HSG highlighting ASP25 amino acids in both chains of the HIV-PR dimer along with the inhibitor and the all important active site water

Bio3D package for structural bioinformatics

```
library(bio3d)
pdb <- read.pdb("1hsg")
```

Note: Accessing on-line PDB file

```
 pdb
```

```
Call: read.pdb(file = "1hsg")
```

```
Total Models#: 1
Total Atoms#: 1686, XYZs#: 5058 Chains#: 2 (values: A B)
```

```
Protein Atoms#: 1514 (residues/Calpha atoms#: 198)
Nucleic acid Atoms#: 0 (residues/phosphate atoms#: 0)
```

```
Non-protein/nucleic Atoms#: 172 (residues: 128)
Non-protein/nucleic resid values: [ HOH (127), MK1 (1) ]
```

Protein sequence:

```
PQITLWQRPLVTIKIGGQLKEALLDTGADDTVLEEMSLPGRWPQMIGGIGGGFIKVRQYD
QILIEICGHKAIGTVLVGPTPVNIIGRNLLTQIGCTLNFPQITLWQRPLVTIKIGGQLKE
ALLDTGADDTVLEEMSLPGRWPQMIGGIGGGFIKVRQYDQILIEICGHKAIGTVLVGPTP
VNIIGRNLLTQIGCTLNF
```

```
+ attr: atom, xyz, seqres, helix, sheet,
calpha, remark, call
```

Q7: How many amino acid residues are there in this pdb object?

198

Q8: Name one of the two non-protein residues?

HOH (127)

Q9: How many protein chains are in this structure?

2 (chain A &B)

```
head(pdb$atom)
```

	type	eleno	elety	alt	resid	chain	resno	insert	x	y	z	o	b
1	ATOM	1	N	<NA>	PRO	A	1	<NA>	29.361	39.686	5.862	1	38.10
2	ATOM	2	CA	<NA>	PRO	A	1	<NA>	30.307	38.663	5.319	1	40.62
3	ATOM	3	C	<NA>	PRO	A	1	<NA>	29.760	38.071	4.022	1	42.64
4	ATOM	4	O	<NA>	PRO	A	1	<NA>	28.600	38.302	3.676	1	43.40
5	ATOM	5	CB	<NA>	PRO	A	1	<NA>	30.508	37.541	6.342	1	37.87
6	ATOM	6	CG	<NA>	PRO	A	1	<NA>	29.296	37.591	7.162	1	38.40
	segid	elesy	charge										
1	<NA>	N	<NA>										
2	<NA>	C	<NA>										
3	<NA>	C	<NA>										
4	<NA>	O	<NA>										
5	<NA>	C	<NA>										
6	<NA>	C	<NA>										

```
#library(bio3dview)
#library(NGLviewR)
#view.pdb(pdb) |>
#  setSpin()

# Select the important ASP 25 residue
#sele <- atom.select(pdb, resno=25)

# and highlight them in spacefill representation
#view.pdb(pdb, cols=c("navy", "teal"),
#          highlight = sele,
#          highlight.style = "spacefill") |>
#  setRock()
```

Predicting functional motions of a single structure

Read an ADK structure from the PDB database:

```
adk <- read.pdb("6s36")
```

Note: Accessing on-line PDB file
 PDB has ALT records, taking A only, rm.alt=TRUE

```
adk
```

Call: read.pdb(file = "6s36")

```
Total Models#: 1
Total Atoms#: 1898,  XYZs#: 5694  Chains#: 1  (values: A)

Protein Atoms#: 1654  (residues/Calpha atoms#: 214)
Nucleic acid Atoms#: 0  (residues/phosphate atoms#: 0)

Non-protein/nucleic Atoms#: 244  (residues: 244)
Non-protein/nucleic resid values: [ CL (3), HOH (238), MG (2), NA (1) ]

Protein sequence:
MRIILLGAPGAGKGTQAQFIMEKYGIPQISTGMLRAAVKSGSELGKQAKDIDMAGKLVT
DELVIALVKERIAQEDCRNGFLLDGFPRTIPQADAMKEAGINVVDYVLEFDVPDELVDKI
```

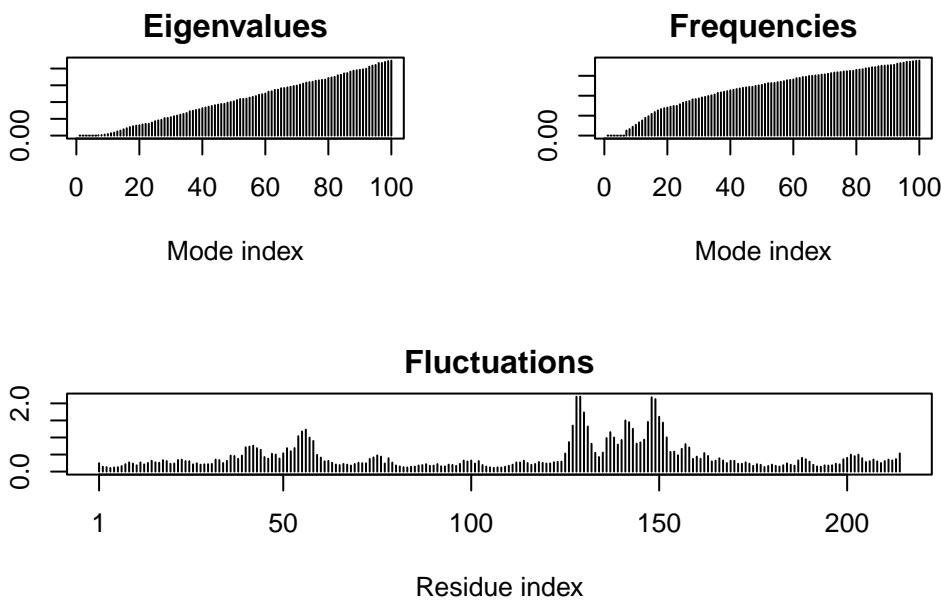
```
VGRRVHAPSGRVYHVKFNPPKVEGKDDVTGEELTTRKDDQEETVRKRLVEYHQMTAPLIG  
YYSKEAEAGNTKYAKVDGTPVAEVRADLEKILG
```

```
+ attr: atom, xyz, seqres, helix, sheet,  
       calpha, remark, call
```

```
m <- nma(adk)
```

```
Building Hessian...           Done in 0.012 seconds.  
Diagonalizing Hessian...     Done in 0.052 seconds.
```

```
plot(m)
```



Write out our results as we trajectory/movie of predicted motions:

```
mktrj(m, file="adk_m7.pdb")
```