

#### Eric Marcon

Motivation

Neutral diversity

Estimation

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Partitioning Diversity

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Phylogenetic

diversity

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. . . . . . . . .

Functional Diversity

Conclusion

# Diversity Metrics

Eric Marcon

October 8, 2018



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### Motivation



## The Questions

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Which diversity metrics make more sense than mere indices?

Which of them may be less sensitive to metabarcoding issues?



## Diversity of what?

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Usually species (definition of E.O. Wilson).

May be any partitioned set.

Usually species in a clade (or a group), considered as a community: e.g. trees in a forest habitat.

Usually asymptotic diversity of a community that does not exist physically.



### Course material

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- Book: Marcon, E. (2017). Mesures de la Biodiversité.
   Kourou, France: UMR EcoFoG. https: //hal-agroparistech.archives-ouvertes.fr/cel-01205813
- English version: https://github.com/EricMarcon/BDmeasurement
- R package entropart (Marcon and Hérault 2015b)



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# Neutral diversity



### Richness and Evenness



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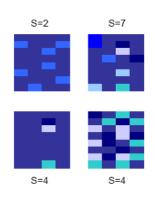
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Which is the most diverse community?



# Measuring Complexity

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### Define a character string:

- Length n
- Each letter has a probability

### Example:

- 3 letters, {a, b, c}, probabilities (1/2, 1/3, 1/6)
- How many 60-character strings?
- ullet The logarithm of the number of strings is n times entropy: 61

Shannon's entropy measures the *complexity* of the distribution of  $\{a, b, c\}$ , independently of n: 1.01



## Entropy in Information Theory

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An experiment with several outcomes:

• The probability to obtain  $r_s$  is  $p_s$ .

Information function:  $I(p_s)$ , between  $I(0) = +\infty$  and I(1) = 0.

- Definition: rarity is  $1/p_s$ .
- The logarithm of rarity is Shannon's information function.

The expectation of the information carried by an individual is Shannon's entropy:

$$\sum_{s} p_s \ln \frac{1}{p_s}$$

The average information is equivalent to complexity.



## Generalized Entropy

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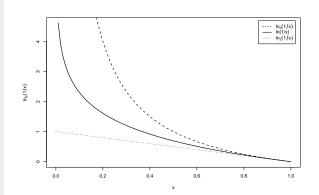
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Other entropies: Rényi, Shorrocks... Tsallis (1988)

Parametric, to focus on rare or abundant species.

Deformed logarithm:  $\ln_q x = \frac{x^{1-q}-1}{1-q}$ 





# Tsallis (HCDT) Entropy 1/2

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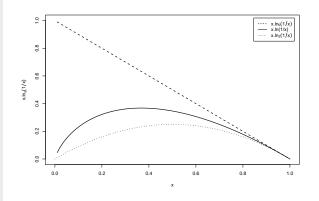
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Contribution of a species to entropy of order q = 0, 1, 2.



# Tsallis (HCDT) Entropy 2/2

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Tsallis entropy is the average (deformed, of order q) logarithm of rarity (Tsallis 1994)

The order q stresses small or high probabilities.

- Entropy of order 0: the number of species -1.
- Entropy of order 1: Shannon.
- Entropy of order 2: Simpson.

## Hill Numbers

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The number of equiprobable outcomes that have the same entropy as the observed system (Hill 1973): *effective number of species*.

They are the (deformed, of order q) exponential of entropy (Marcon et al. 2014).

$$e_q^x = [1 + (1 - q)x]^{1/(1-q)}.$$

Diversity is noted  ${}^qD(\mathbf{p_s})$ .



## **Diversity Profiles**

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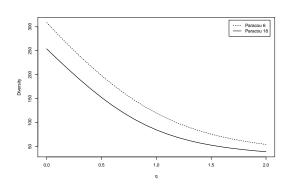
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A diversity profile is  ${}^qD \sim q$ .

Compare two communities:



# Summary

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Entropy is the average log of rarity:

$${}^{q}H(\mathbf{p_s}) = \sum_{s} p_s \ln_q (1/p_s)$$

Diversity is its exponential:

$${}^{q}D(\mathbf{p_s}) = e_q^{{}^{q}H(\mathbf{p_s})}$$



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### Estimation bias

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Rare species are difficult to sample

ightarrow diversity is generally underestimated.

Sampling effort is measured by n, the sample size.

The estimation bias decreases with n and q.

Simpson diversity is almost unbiased if  $n/(n-1) \approx 1$ .



### Estimation of richness

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High bias but easiest correction methods.

Jackknife 1 estimator: just add the number of singletons  $S_1$ .

Correct if sample completeness > 3/4, i.e. singletons are less than 1 species out of 4.

Example: 225 species including 19 singletons  $\rightarrow$  244 species.



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Conclusion

Many estimators of entropy, most based on sample coverage. See section 4.6 of the book.

Entropy is estimated, then transformed to diversity.

Sample coverage is the probability of an individual of the community to belong to a sample species.

Far more than sample completeness.

Estimated by

$$\hat{C} = 1 - \frac{S_1}{n}$$
.



### Practical estimation

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All estimators available in *entropart::Diversity*.

### Rule of thumb:

- Chao-Wang-Jost estimator is the best if singletons are less than 1 species out of 4.
- Else, the unveiled estimator chooses the appropriate
   Jackknife estimator for richness and is less biased but has higher variance.



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## Practical 1



# Data

Use Paracou forest tree inventory (2016).

Eric Marcon library("secret") library("openssl") Motivation # Requires a private key to read the data

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unite(col = spName, Genus, Species, remove = FALSE) %>% group\_by(Plot, spName) %>% summarize(Abundance = length(Species)) ->

species name field.

Abundances

Paracou %>%

Prepare a named vector for plot 6 data.

Abundances %>% filter(Plot==6) -> AbundancesP6 AbdP6 <- AbundancesP6\$Abundance names (AbdP6) <- AbundancesP6\$snName

Paracou <- get\_secret("Paracou", key = read\_key("Cours-Biodiversite\_rsa"

Summarize the list of trees into abundance table. First prepare a



## Richness

## None

334

sum(AbdP6 == 1)

471

## [1] 98

Number of species in plot 6.

AbdP6 %>% Richness(Correction="None")

Number of singletons in plot 6.

AbdP6 %>% Richness(Correction="Jackknife")

library("entropart")

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Jackknife 2



## Sample coverage

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### Coverage (AbdP6)

## ZhangHuang ## 0.9723336

More than 1/3 of species are not observed but they contain less than 3% of the number of trees.



## Diversity Profile

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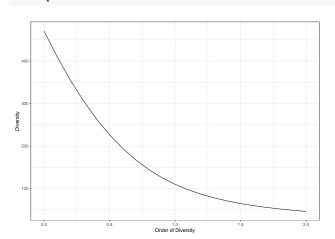
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### Unveiled-Jackknife estimator.

CommunityProfile(Diversity, AbdP6, Correction="UnveilJ") %>% autoplot





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# Partitioning Diversity



## $\alpha$ , $\beta$ and $\gamma$ diversity

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### Defined by Whittaker (1972):

- $\alpha$ : Average richness of a set of habitats, i.e.  $\bar{S}$  species per habitat.
- ullet  $\gamma$ : Richness of the assemblage, i.e. S species
- $\beta$ : The ratio  $\gamma/\alpha$ , i.e. a number of habitats.

### Extensions to:

- All orders of diversity.
- Any embedded spatial scales.



## The controversy

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After Lande (1996): additive partitioning of diversity.

### Resolution:

- $\beta$  diversity = the ratio  $\gamma/\alpha$ .
- $\beta$  entropy = the difference  $\gamma \alpha$ .

Exponentials multiply, logs sum...



### Effective number of communities

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 $\alpha$  diversity at the community level is an effective number of species / community.

 $\gamma$  diversity at the assemblage ("meta-community") level is an effective number of species.

ightarrow eta diversity is an effective number of communities.



### Effective number of communities

```
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```

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```
## HCDT diversity partitioning of order 1
## of metaCommunity Paracou618.MC
## with correction: UnveilJ
## Alpha diversity of communities:
##
        P006
                  P018
   84.52352 119.50807
## Total alpha diversity of the communities:
## [1] 98.02742
## Beta diversity of the communities:
   UnveilJ
## 1.416315
## Gamma diversity of the metacommunity:
## UnveilJ
## 138.8377
```



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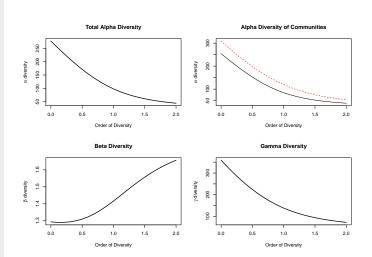
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# Relative Entropy

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Conclusion

Departure of the observed distribution from the expected distribution (Kullback and Leibler 1951's divergence).

Generalization to order q (Marcon et al. 2014).

If the expected distribution is the average distribution (that of the assemblage) then the relative entropy is the difference between the entropy of the assemblage  $(\gamma)$  and that of each community  $(\alpha)$ : it is  $\beta$  entropy.

Useless but important.



## Differentiation vs Proportional Diversity

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Conclusion

This definition of  $\beta$  diversity measures the average departure of a community from the meta-community.

 $\rightarrow$  Proportional Diversity.

Other measures exist to define how different two communities are from each other.

ightarrow Differentiation Diversity.



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## Practical 2



## Data

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Formatting the data as a dataframe, species in lines, plots in columns.

```
Abundances %>%

spread(key = Plot, value = Abundance, fill=0) %>%

as.data.frame -> df

# Name rows and columns

rownames(df) <- df$spName

df <- df[, -1]

colnames(df) <- paste("P", colnames(df), sep="")

# Create a MetaCommunity object

ParacouMC <- MetaCommunity(df, Weights = colSums(df))
```



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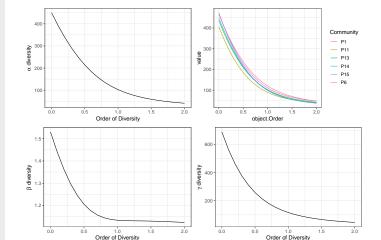
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## Ultrametric phylogeny

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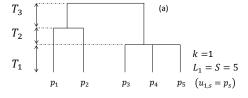
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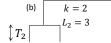
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$$u_{2,1}$$
  $u_{2,2}$   $u_{2,3}$   
=  $p_1$  =  $p_2$  =  $p$ 

$$= p_3 + p_4 + p_5$$

c) 
$$\int_{3} T_3 \qquad k = K = 3$$
$$L_3 = 2$$

$$u_{3,1}$$
  $u_{3,2}$   
=  $p_1 + p_2$   $u_{3,2}$   
=  $p_3 + p_4 + p_5$ 



## Phylogenetic Entropy and Diversity

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Entropy sums along the tree.

Diversity is its exponential (Marcon and Hérault 2015a). It is the number of equiprobable species in a star phylogeny of height 1.

ightarrow Estimate entropy along the tree, average it, transform the phyloentropy into phylodiversity.



## Phylogenetic Diversity

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Phylogenetic diversity of order 0 is called PD. It reduces to richness in a star tree of height 1.

Phylogenetic entropy of order 2 is Rao's quadratic entropy. It reduces to Simpson's entropy in a star tree of height 1.



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### Make a taxonomic tree.

```
library("ape")
library("magrittr")
Paracou %>%
  filter(Plot == 6) %>%
  select(Family:Species) %>%
  unite(col=spName, Genus, Species, remove=FALSE) %>%
  mutate_if(is.character, as.factor) %>%
  {as.phylo(-Family/Genus/spName, data=., collapse=FALSE)} %>%
  compute.brlen(method=1) %>%
  collapse.singles %>%
  multi2di %T>%
  plot(show.tip.label = FALSE) -> p6Phylo
```





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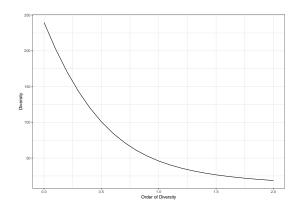
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### Same estimator as that of neutral diversity.

```
dp <- CommunityProfile(function(Abd, q, CheckArguments)
   PhyloDiversity(Abd, q, Correction="UnveilJ", Tree=p6Phylo)$Total,
   AbdP6)
autoplot(dp)</pre>
```





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# Functional Diversity



## Functional space

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Distance between species are not ultrametric.

Metrics based on the distance matrix.

No time here: see part 3 of the book.



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## Entropy to unify diversity measures



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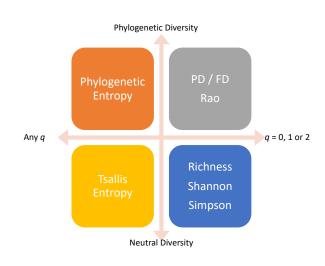
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# Metabarcoding

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Satisfactory results when supervised clustering is possible (but no estimation bias correction available)

Risky results with unsupervised clustering. Seems to work quite well around Shannon's diversity.



### References

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