

On evaluating the accuracy and biological plausibility of diffusion MRI tractograms

David Romascano¹, Alessandro Dal Palù², Jean-Philippe Thiran^{1,3}, and Alessandro Daducci^{1,4}

¹Signal Processing Laboratory (LTSS), École Polytechnique Fédérale de Lausanne, Lausanne, Vaud, Switzerland, ²Department of Mathematics and Computer Science, University of Parma, Parma, Italy, ³Department of Radiology, University Hospital Center and University of Lausanne, Lausanne, Vaud, Switzerland, ⁴Center for Biomedical Imaging, Signal Processing Core, Lausanne, Vaud, Switzerland

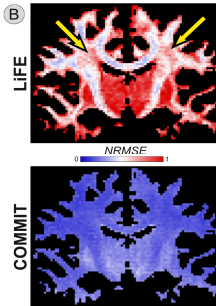
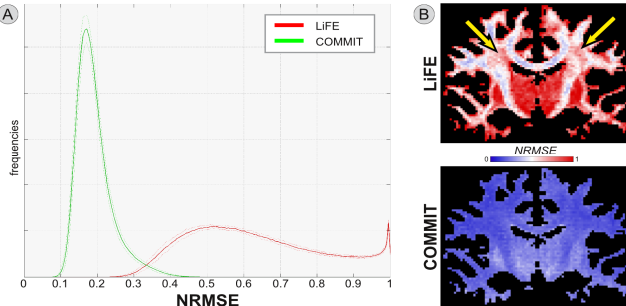
INTRODUCTION. One of the major limitations of diffusion MRI tractography is that the tractograms, i.e. set of fiber tracts, recovered by existing algorithms are not truly quantitative. Several orders of magnitude separate in fact the resolution achievable with MRI from the actual size of the axons and, consequently, each reconstructed trajectory has to be considered as representative of a coherent set of real anatomical fibers, the amount of which is not easy to assess. Hence, the structural connectivity between different brain regions, a.k.a. connectomics, is nowadays quantified by counting the number of recovered pathways or averaging some scalar maps along them; in both cases, these estimates provide only indirect measures of the true underlying neuronal connectivity¹. A number of methods have recently started to appear to address this limitation²⁻⁶; in particular, COMMIT² and LiFE³ have been developed upon the recently proposed framework that showed how to formulate tractography as an efficient system of linear equations⁴, opening de facto the door for the practical possibility to evaluate and compare the accuracy of the tractograms.

PURPOSE. These two models follow different strategies to describe the signal in each voxel. On one hand, COMMIT² uses a forward-model that takes into account that the diffusion MR signal can originate from distinct water pools⁷, e.g. intra- and extra-cellular. On the other hand, LiFE³ models the signal as consisting only of contributions arising from the tracts passing through each voxel (i.e. restricted diffusion). The extra-cellular space around the axons (i.e. hindered diffusion) and any partial volume that can occur with non white-matter (WM) tissue (i.e. isotropic diffusion) are not directly considered, but are “removed” with a de-meaning procedure. However, as shown by several independent studies⁸⁻¹¹, the relative contribution of these compartments is not homogeneous in the WM and can change considerably. The schematic representation in the top-right figure depicts such a situation: the callosal fibers projecting from the corpus callosum (CC) and the corticospinal tract (CST) consist both of tightly-packed axons (yellow circles) that progressively fan-out and eventually cross. Differences in the axonal packing density are compensated by variations in the spacing surrounding the axons themselves, i.e. extra-cellular space. This consideration is implicitly or explicitly assumed in most state-of-the-art techniques for voxelwise microstructure imaging⁷⁻¹⁰ and independent histological studies also corroborate this hypothesis¹¹. Furthermore, to be sensitive with diffusion MRI to the tissue microstructure, multiple b-values have been proven necessary⁹. In this abstract we investigated the importance of using (i) proper multi-compartment models and (ii) adequate multiple b-value acquisitions in order to be able to evaluate the accuracy and the biological plausibility of the tractograms using these global approaches.

METHODS. COMMIT² implements a very general and flexible framework, and LiFE³ can be considered as a special case of it where only the contributions of the tracts are considered in the signal forward-model. Thus, we evaluated both models using the code available at <https://github.com/daducci/COMMIT>. For a first assessment of the goodness of the fit of each model, we downloaded 10 subjects from the Human Connectome Project¹² (90 images with spatial resolution 1.25 mm³ at b=2000 s/mm²) and used the same experimental settings as in the LiFE manuscript³, i.e. CSD-based probabilistic tractography with 500K tracts and NRMSE (reported as percentages) between the measured and modeled signal as quality metric. In addition, to be able to extract the relative fractions of the intra- (icvf) and extra-cellular (ecvf) compartments in each voxel, and thus evaluate the biological plausibility of the tractograms, we used a 2-shell acquisition (81 images with spatial resolution 1.8x1.8x2.5 mm³ at b=700 s/mm² and b=2000 s/mm²) and the same experimental setup as in the COMMIT paper², i.e. using the state-of-the-art global tracking algorithm¹³ with 100K candidate tracts.

RESULTS AND DISCUSSION. The left figure compares the ability of the two models to adequately describe the signal in each voxel. Panel (A) shows the histograms of the fitting errors (NRMSE, mean±std) across the 10 datasets. The plot clearly highlights significantly higher fitting errors with LiFE (68.7%±17.9%) as compared to COMMIT (19.5%±4.6%). Visual inspection in panel (B) shows that LiFE presents the highest errors in areas with crossing fibers (yellow arrows) and with partial volume with gray-matter, where the lack of the extra-cellular compartment in the model prevents an accurate fit; conversely, the smallest errors are observed in regions with densely packed axons, where indeed the extra-cellular space is greatly reduced. On the other hand, COMMIT exhibits an homogeneously low spatial distribution of the errors and definitely appears to fit the data more accurately.

In the bottom-right figure we tested the two models with multi-shell data. The NRMSE map confirms the same observations previously drawn, with LiFE exhibiting much



higher fitting errors (64.8%±18.1%) than COMMIT (10.1%±2.9%) also in this case. The icvf map estimated with LiFE shows a spatial distribution that does not follow the expected pattern of neuronal density as found in previous studies⁸⁻¹¹; this is a clear sign of an incorrect assessment of the tract contributions. On the contrary, the icvf and ecvf maps estimated with COMMIT show a spatial distribution which is in agreement with the known brain anatomy and previous independent studies⁸⁻¹¹. The highest icvf values are in fact found in the major WM bundles, e.g. CC and CST, an homogeneous distribution is observed in crossing regions and a decreased fraction close to gray-matter. The ecvf map shows the opposite behavior, as expected. It is worth noting that this map (not considered in the LiFE model) follows the same spatial pattern as observed in the map of the fitting errors with LiFE. We speculate that this observation reflects the need to consider in the model the non-homogeneous extra-cellular contributions in order to adequately describe the signal in each voxel and, consequently, to be able to estimate the actual contributions of the tracts.

CONCLUSION. Our results indicate that, to be able to evaluate the accuracy and biological plausibility of tractograms with these novel global approaches²⁻⁶, both (i) proper models which account for all possible water pools that can contribute to the diffusion MR signal and (ii) adequate multi-shell acquisitions appear to be mandatory. The lack of either of these two conditions leads to very inaccurate signal fitting and the estimated quantities do not resemble the known brain anatomy. Future work will still be required to investigate the benefits of these global approaches for brain connectivity analyses.

REFERENCES. [1] Jones et al, *NeuroImage* 73:239-54 (2012) [2] Daducci et al, *IEEE TMI* (2014) [3] Pestilli et al, *Nat Methods* (2014) [4] Daducci et al, *Proc. ISBI* (2013) [5] Sherbondy et al, *Proc. MICCAI* (2010) [6] Smith et al, *NeuroImage* 67:298-312 (2012) [7] Panagiotaki et al, *NeuroImage* 3:2241-54 (2010) [8] Assaf et al, *NeuroImage* 27:48-58 (2008) [9] Alexander et al, *NeuroImage* 52:1374-89 (2010) [10] Zhang et al, *NeuroImage* 61:1000-16 (2012) [11] Jespersen et al, *NeuroImage* 49:205-16 (2010) [12] Van Essen et al, *NeuroImage* 62:2222-31 (2012) [13] Reisert et al, *NeuroImage* 54:955-62 (2011)

