

RWorksheet#4b_Parrenas

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#Using Loop Function

#1. Using the for loop, create an R script that will display a 5x5 matrix as shown in Figure 1. It must

```
vectorA <- c(1, 2, 3, 4, 5)
zeroMatrix <- matrix(0, nrow = 5, ncol = 5)
```

```
for (i in 1:5){
  for (j in 1:5){

    diff <- abs(vectorA[i] - j)

    cat(diff, " ")
  }
  cat("\n")
}
```

```
## 0  1  2  3  4
## 1  0  1  2  3
## 2  1  0  1  2
## 3  2  1  0  1
## 4  3  2  1  0
```

#2. Print the string "" using for() function. The output should be the same as shown in Figure*

```
for(i in 1:5){
  ejp <- rep("*", i)
  print(ejp)
}
```

```
## [1] "*"
## [1] "*" "*"
## [1] "*" "*" "*"
## [1] "*" "*" "*" "*"
## [1] "*" "*" "*" "*" "*"
```

#3. Get an input from the user to print the Fibonacci sequence starting from the 1st input up to 500. U

```
user_input <- as.numeric(readline("Enter a number to start the Fibonacci sequence: "))
```

```
## Enter a number to start the Fibonacci sequence:
```

```
a <- 0
b <- 1
```

```
cat("Fibonacci sequence starting from", user_input, ": ")
```

```
## Fibonacci sequence starting from NA :
```

```
cat(user_input, " ")
```

```
## NA
```

```
repeat {
```

```
  fiboSeq <- a + b
```

```
  if (fiboSeq > 500) {
```

```
    break
```

```
  }
```

```
  cat(fiboSeq, " ")
```

```
  a <- b
```

```
  b <- fiboSeq
```

```
}
```

```
## 1 2 3 5 8 13 21 34 55 89 144 233 377
```

```
#4a. Import the data set as shown in Figure 1 you have created previously.
```

```
library(readr)
```

```
shoeSizes <- read_csv("shoeSizes.csv")
```

```
## Rows: 28 Columns: 3
```

```
## -- Column specification -----
```

```
## Delimiter: ","
```

```
## chr (1): gender
```

```
## dbl (2): shoe_Size, height
```

```
##
```

```
## i Use `spec()` to retrieve the full column specification for this data.
```

```
## i Specify the column types or set `show_col_types = FALSE` to quiet this message.
```

```
View(shoeSizes)
```

```
## Warning in View(shoeSizes): unable to open display
```

```
## Error in .External2(C_dataviewer, x, title): unable to start data viewer
```

```
#b. Create a subset for gender(female and male). How many observations are there in Male? How about in Female?
```

```
male <- shoeSizes[shoeSizes$gender == "M",]
```

```
female <- shoeSizes[shoeSizes$gender == "F",]
```

```
maleCount <- nrow(male)
```

```
femaleCount <- nrow(female)
```

```
cat("Numbers of male: ", maleCount, "\n")
```

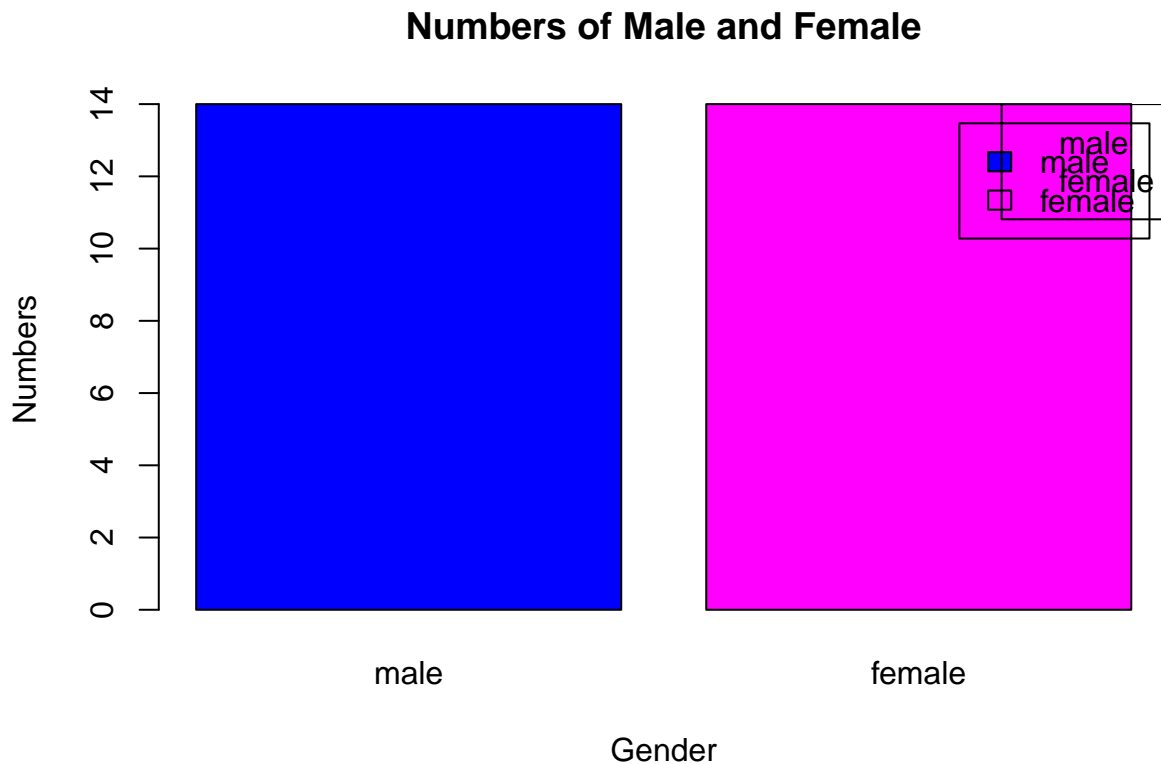
```
## Numbers of male: 14
```

```
cat("Numbers of female: ", femaleCount, "\n")
```

```
## Numbers of female: 14
```

```
#c. Create a graph for the number of males and females for Household Data. Use plot(), chart type = barplot  
genderCount <- c(male = maleCount, female = femaleCount)
```

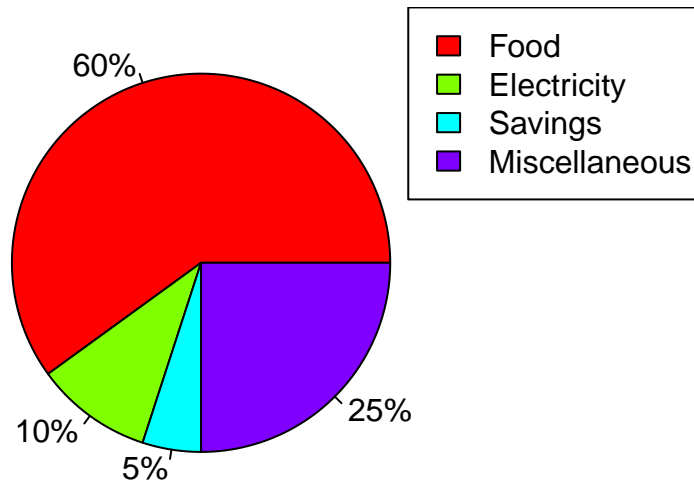
```
barplot(genderCount, main = "Numbers of Male and Female",  
        xlab = "Gender", ylab = "Numbers", col = c("blue", "magenta"),  
        legend.text = TRUE, width = c(0.2, 0.2))  
legend("topright", legend = names(genderCount), col = c("blue", "magenta"))
```



```
#5. The monthly income of Dela Cruz family was spent  
mypie <- c(60, 10, 5, 25)
```

```
pie(mypie,  
    main = "Monthly Income of Dela Cruz family",  
    col = rainbow(length(mypie)),  
    labels = c("60%", "10%", "5%", "25%"),  
)  
legend("topright", legend = c("Food", "Electricity", "Savings", "Miscellaneous"), fill = rainbow(length(mypie)))
```

Monthly Income of Dela Cruz family



#6. Use the iris data set

#a. Check for the structure of the data set using the str() function. Describe what you have seen in the output.

```
data(iris)
str(iris)
```

```
## 'data.frame': 150 obs. of 5 variables:
## $ Sepal.Length: num 5.1 4.9 4.7 4.6 5 5.4 4.6 5 4.4 4.9 ...
## $ Sepal.Width : num 3.5 3 3.2 3.1 3.6 3.9 3.4 3.4 2.9 3.1 ...
## $ Petal.Length: num 1.4 1.4 1.3 1.5 1.4 1.7 1.4 1.5 1.4 1.5 ...
## $ Petal.Width : num 0.2 0.2 0.2 0.2 0.2 0.4 0.3 0.2 0.2 0.1 ...
## $ Species : Factor w/ 3 levels "setosa","versicolor",...: 1 1 1 1 1 1 1 1 1 1 ...
```

#The output shows the data's structure, which includes the variables and objects as well as the length, class, and levels of each variable.

#b. Create an R object that will contain the mean of the sepal.length, sepal.width, petal.length, and petal.width.

```
sepaleng <- mean(iris$Sepal.Length)

sepalwid <- mean(iris$Sepal.Width)

petlen <- mean(iris$Petal.Length)

petwin <- mean(iris$Petal.Width)

print(sepaleng)
```

```
## [1] 5.843333
```

```
print(sepalwid)
```

```
## [1] 3.057333
```

```
print(petlen)
```

```
## [1] 3.758
```

```
print(petwin)
```

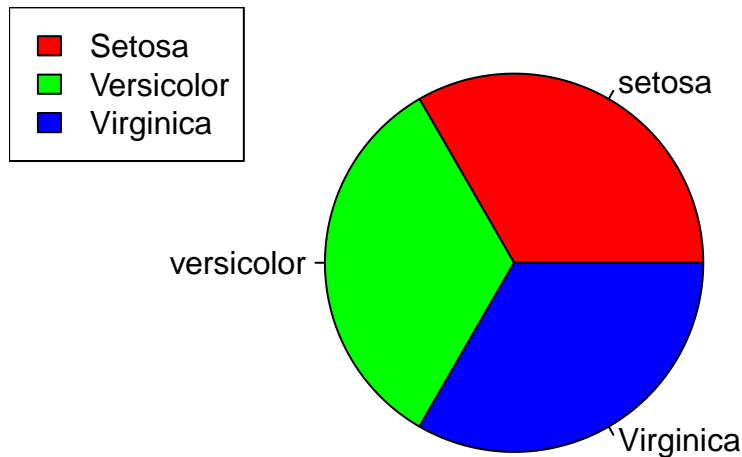
```
## [1] 1.199333
```

#c. Create a pie chart for the Species distribution. Add title, legends, and colors. Write the R script

```
specCount <- table(iris$Species)

pie(specCount,
    main = "Species",
    col = rainbow(length(specCount)),
    labels = c("setosa", "versicolor", "Virginica")
)
legend("topleft", legend = c("Setosa", "Versicolor", "Virginica"), fill = rainbow(length(specCount)))
```

Species



#d. Subset the species into setosa, versicolor, and virginica. Write the R scripts and show the last six

```
setsub <- iris[iris$Species == "setosa" | iris$Species == "Versicolor" | iris$Species == "virginica", ]
setsub
```

```
##      Sepal.Length Sepal.Width Petal.Length Petal.Width Species
## 1           5.1         3.5         1.4         0.2    setosa
## 2           4.9         3.0         1.4         0.2    setosa
## 3           4.7         3.2         1.3         0.2    setosa
## 4           4.6         3.1         1.5         0.2    setosa
## 5           5.0         3.6         1.4         0.2    setosa
## 6           5.4         3.9         1.7         0.4    setosa
## 7           4.6         3.4         1.4         0.3    setosa
## 8           5.0         3.4         1.5         0.2    setosa
## 9           4.4         2.9         1.4         0.2    setosa
## 10          4.9         3.1         1.5         0.1    setosa
## 11          5.4         3.7         1.5         0.2    setosa
## 12          4.8         3.4         1.6         0.2    setosa
## 13          4.8         3.0         1.4         0.1    setosa
## 14          4.3         3.0         1.1         0.1    setosa
## 15          5.8         4.0         1.2         0.2    setosa
## 16          5.7         4.4         1.5         0.4    setosa
```

## 17	5.4	3.9	1.3	0.4	setosa
## 18	5.1	3.5	1.4	0.3	setosa
## 19	5.7	3.8	1.7	0.3	setosa
## 20	5.1	3.8	1.5	0.3	setosa
## 21	5.4	3.4	1.7	0.2	setosa
## 22	5.1	3.7	1.5	0.4	setosa
## 23	4.6	3.6	1.0	0.2	setosa
## 24	5.1	3.3	1.7	0.5	setosa
## 25	4.8	3.4	1.9	0.2	setosa
## 26	5.0	3.0	1.6	0.2	setosa
## 27	5.0	3.4	1.6	0.4	setosa
## 28	5.2	3.5	1.5	0.2	setosa
## 29	5.2	3.4	1.4	0.2	setosa
## 30	4.7	3.2	1.6	0.2	setosa
## 31	4.8	3.1	1.6	0.2	setosa
## 32	5.4	3.4	1.5	0.4	setosa
## 33	5.2	4.1	1.5	0.1	setosa
## 34	5.5	4.2	1.4	0.2	setosa
## 35	4.9	3.1	1.5	0.2	setosa
## 36	5.0	3.2	1.2	0.2	setosa
## 37	5.5	3.5	1.3	0.2	setosa
## 38	4.9	3.6	1.4	0.1	setosa
## 39	4.4	3.0	1.3	0.2	setosa
## 40	5.1	3.4	1.5	0.2	setosa
## 41	5.0	3.5	1.3	0.3	setosa
## 42	4.5	2.3	1.3	0.3	setosa
## 43	4.4	3.2	1.3	0.2	setosa
## 44	5.0	3.5	1.6	0.6	setosa
## 45	5.1	3.8	1.9	0.4	setosa
## 46	4.8	3.0	1.4	0.3	setosa
## 47	5.1	3.8	1.6	0.2	setosa
## 48	4.6	3.2	1.4	0.2	setosa
## 49	5.3	3.7	1.5	0.2	setosa
## 50	5.0	3.3	1.4	0.2	setosa
## 101	6.3	3.3	6.0	2.5	virginica
## 102	5.8	2.7	5.1	1.9	virginica
## 103	7.1	3.0	5.9	2.1	virginica
## 104	6.3	2.9	5.6	1.8	virginica
## 105	6.5	3.0	5.8	2.2	virginica
## 106	7.6	3.0	6.6	2.1	virginica
## 107	4.9	2.5	4.5	1.7	virginica
## 108	7.3	2.9	6.3	1.8	virginica
## 109	6.7	2.5	5.8	1.8	virginica
## 110	7.2	3.6	6.1	2.5	virginica
## 111	6.5	3.2	5.1	2.0	virginica
## 112	6.4	2.7	5.3	1.9	virginica
## 113	6.8	3.0	5.5	2.1	virginica
## 114	5.7	2.5	5.0	2.0	virginica
## 115	5.8	2.8	5.1	2.4	virginica
## 116	6.4	3.2	5.3	2.3	virginica
## 117	6.5	3.0	5.5	1.8	virginica
## 118	7.7	3.8	6.7	2.2	virginica
## 119	7.7	2.6	6.9	2.3	virginica
## 120	6.0	2.2	5.0	1.5	virginica

```
## 121      6.9      3.2      5.7      2.3 virginica
## 122      5.6      2.8      4.9      2.0 virginica
## 123      7.7      2.8      6.7      2.0 virginica
## 124      6.3      2.7      4.9      1.8 virginica
## 125      6.7      3.3      5.7      2.1 virginica
## 126      7.2      3.2      6.0      1.8 virginica
## 127      6.2      2.8      4.8      1.8 virginica
## 128      6.1      3.0      4.9      1.8 virginica
## 129      6.4      2.8      5.6      2.1 virginica
## 130      7.2      3.0      5.8      1.6 virginica
## 131      7.4      2.8      6.1      1.9 virginica
## 132      7.9      3.8      6.4      2.0 virginica
## 133      6.4      2.8      5.6      2.2 virginica
## 134      6.3      2.8      5.1      1.5 virginica
## 135      6.1      2.6      5.6      1.4 virginica
## 136      7.7      3.0      6.1      2.3 virginica
## 137      6.3      3.4      5.6      2.4 virginica
## 138      6.4      3.1      5.5      1.8 virginica
## 139      6.0      3.0      4.8      1.8 virginica
## 140      6.9      3.1      5.4      2.1 virginica
## 141      6.7      3.1      5.6      2.4 virginica
## 142      6.9      3.1      5.1      2.3 virginica
## 143      5.8      2.7      5.1      1.9 virginica
## 144      6.8      3.2      5.9      2.3 virginica
## 145      6.7      3.3      5.7      2.5 virginica
## 146      6.7      3.0      5.2      2.3 virginica
## 147      6.3      2.5      5.0      1.9 virginica
## 148      6.5      3.0      5.2      2.0 virginica
## 149      6.2      3.4      5.4      2.3 virginica
## 150      5.9      3.0      5.1      1.8 virginica
```

```
tail(setsub, 6)
```

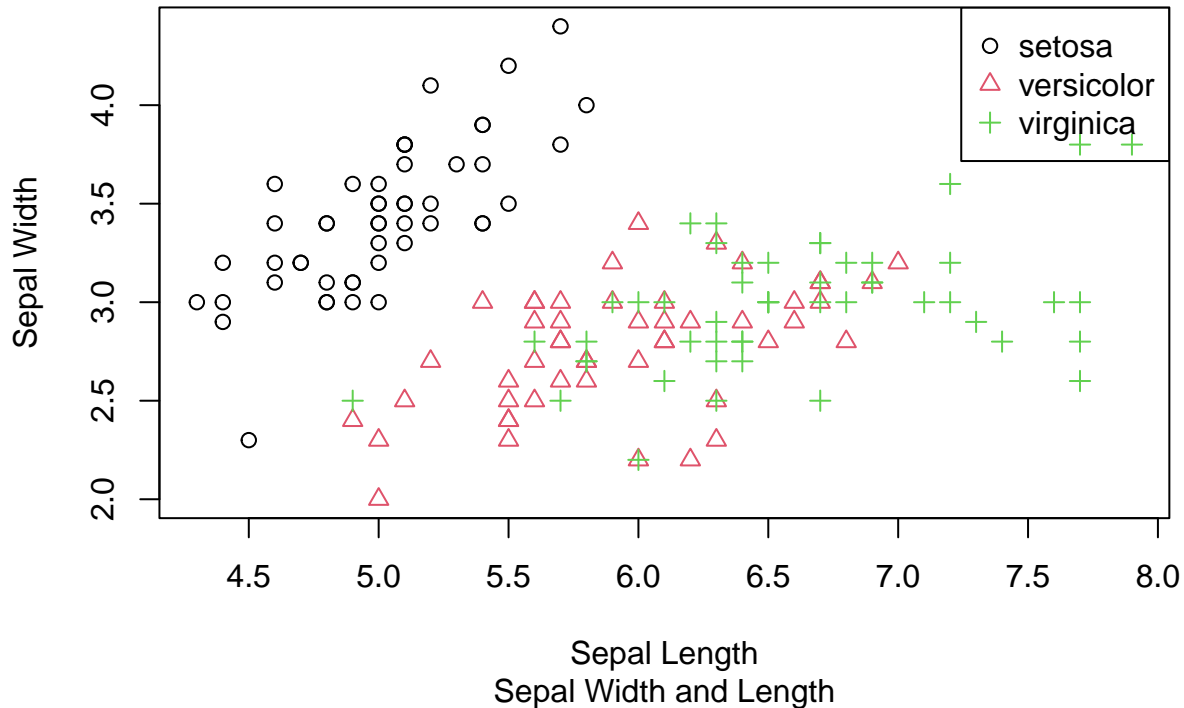
```
##      Sepal.Length Sepal.Width Petal.Length Petal.Width  Species
## 145      6.7      3.3      5.7      2.5 virginica
## 146      6.7      3.0      5.2      2.3 virginica
## 147      6.3      2.5      5.0      1.9 virginica
## 148      6.5      3.0      5.2      2.0 virginica
## 149      6.2      3.4      5.4      2.3 virginica
## 150      5.9      3.0      5.1      1.8 virginica
```

#e. Create a scatterplot of the sepal.length and sepal.width using the different species(setosa,versicolour)

```
plot(iris$Sepal.Length, iris$Sepal.Width,
     col = as.numeric(iris$Species),
     pch = as.numeric(iris$Species),
     main = "Iris Dataset",
     sub = "Sepal Width and Length",
     xlab = "Sepal Length", ylab = "Sepal Width"
)

legend("topright", legend = levels(iris$Species), col = unique(as.numeric(iris$Species)), pch = unique(
```

Iris Dataset



#6f. The scatterplot displays the relationship between the sepal length and width.

#Basic Cleaning and Transformation of Objects

#7. Import the alexa-file.xlsx. Check on the variations. Notice that there are extra whitespaces among

```
library(readxl)
```

```
alexa_file <- read_excel("alexa_file.xlsx")
```

```
## Error: `path` does not exist: 'alexa_file.xlsx'
```

```
View(alexa_file)
```

```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
```

```
library(readxl)
```

```
alexa_file$variation <- gsub("Black Dot", "BlackDot", alexa_file$variation)
```

```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
```

```
alexa_file$variation <- gsub("Black Plus", "BlackPlus", alexa_file$variation)
```

```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
```

```
alexa_file$variation <- gsub("Black Show", "BlackShow", alexa_file$variation)
```

```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
```

```
alexa_file$variation <- gsub("Black Spot", "BlackSpot", alexa_file$variation)
```

```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
```

```
alexa_file$variation <- gsub("White Dot", "WhiteDot", alexa_file$variation)
```



```
## Error in eval(expr, envir, enclos): object 'alexa_file' not found
alexa_file$variation <- gsub("White Plus", "WhitePlus", alexa_file$variation)

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
alexa_file$variation <- gsub("White Show", "WhiteShow", alexa_file$variation)

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
alexa_file$variation <- gsub("White Spot", "WhiteSpot", alexa_file$variation)

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
alexa_file

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
#b. Get the total number of each variations and save it into another object. Save the object as variations.RData

library("dplyr")

##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##   filter, lag
## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
vari <- alexa_file

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
count(alexa_file$variation)

## Error in eval(expr, envir, enclos): object 'alexa_file' not found
vari

## Error in eval(expr, envir, enclos): object 'vari' not found
save(vari, file = "variations.RData")

## Error in save(vari, file = "variations.RData"): object 'vari' not found
#c. From the variations.RData, create a barplot(). Complete the details of the chart which include the
load("variations.RData")
vari

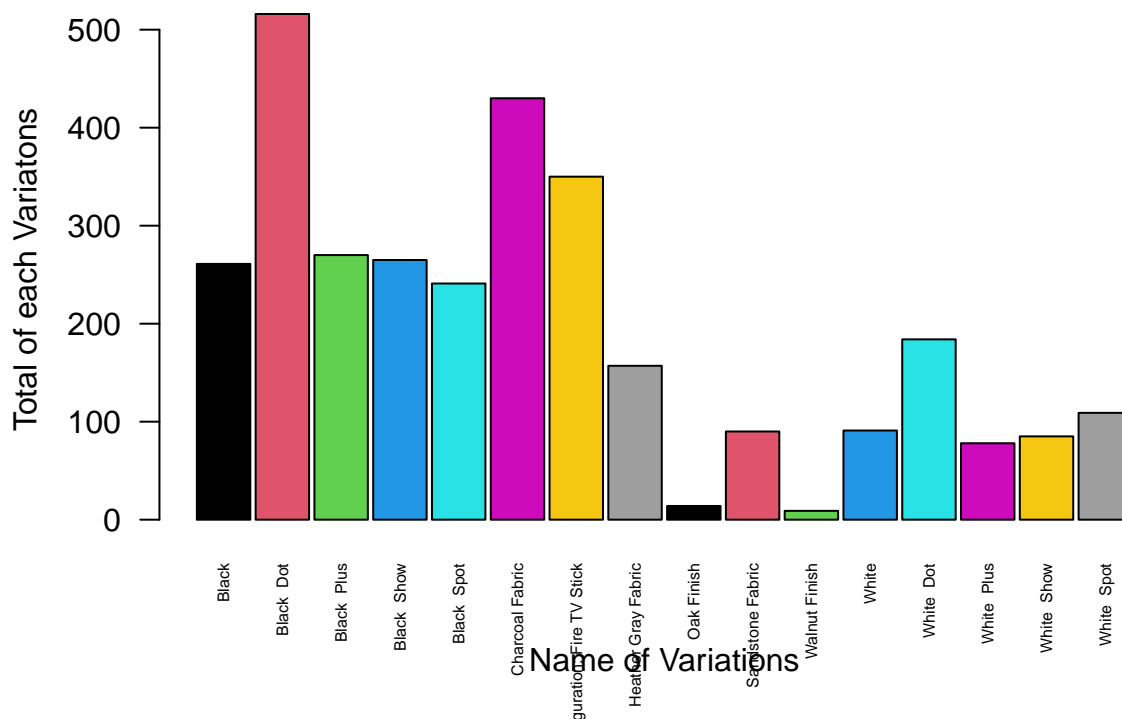
## # A tibble: 16 x 2
##   `alexa_file$variation`      n
##   <chr>                  <int>
## 1 Black                  261
## 2 Black Dot              516
## 3 Black Plus             270
## 4 Black Show             265
## 5 Black Spot             241
## 6 Charcoal Fabric        430
## 7 Configuration: Fire TV Stick 350
```

```
## 8 Heather Gray Fabric      157
## 9 Oak Finish               14
## 10 Sandstone Fabric       90
## 11 Walnut Finish          9
## 12 White                  91
## 13 White Dot              184
## 14 White Plus             78
## 15 White Show             85
## 16 White Spot            109
```

```
namevari <- vari$`alexa_file$variation`

ploto <- barplot(vari$,
                 names.arg = namevari,
                 main = "Total number of each variations",
                 xlab = "Name of Variations",
                 ylab = "Total of each Variatons",
                 col = 1:16,
                 space = 0.1,
                 cex.names = 0.5,
                 las = 2)
```

Total number of each variations



#d. Create a barplot() for the black and white variations. Plot it in 1 frame, side byside. Complete th

```
library(ggplot2)
library(magrittr)

#layout one frame
par(mfrow = c(1, 2), mar = c(2, 2, 2, 2))
```

```

Blackplot <- vari[vari$`alexa_file$variation` %in% c("Black", "BlackDot", "BlackShow", "BlackPlus", "Bl

Whiteplot <- vari[vari$`alexa_file$variation` %in% c("White", "WhiteDot", "WhiteShow", "WhitePlus", "Wh

# Barplot for Black variations
Black <- barplot(height = Blackplot$n,
                 names.arg = Blackplot$`alexa_file$variation`,
                 col = "Black",
                 main = "Black Variations",
                 xlab = "Number of Variation",
                 ylab = "Count",
                 las = 0.0,
                 cex.names = 0.4,
                 space = 0.2
)

# Barplot for White variations
White <- barplot(height = Whiteplot$n,
                 names.arg = Whiteplot$`alexa_file$variation`,
                 col = "White",
                 main = "White Variations",
                 xlab = "Number of Variation",
                 ylab = "Count",
                 las = 0.0,
                 cex.names = 0.4,
                 space = 0.2
)

```

