

# Exvar: A gene expression and genetic variation data analysis and visualization R package

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Exvar is a novel R package for RNA sequencing data analysis. It includes functions for gene expression and genetic variant analysis and visualization.

The package consists of 6 data analysis functions (`processfastq()`; `counts()`; `expression()`; `callsnp()`; `callcnv()`; and `callindel()`) and 3 data visualization functions (`vizexp()`, `vizsnp()`, and `vizcnv()`).

## 1. requirements()

### Description

This function will install and call the required packages according to the data source species.

### Usage

```
requirements(species)
```

### Arguments

`species`      Select Specie number from list.

## 2. processfastq()

**Description:** This function takes in FASTQ files and performs quality control before aligning to a reference genome. It assumes paired-end samples are of the same file name with an underscore ( \_ ) and a number to signify different reads of the same sample. Each sample's outputs will be stored in a separate directory.

### Usage

```
processfastq( file = list_files_with_exts(dir = dir, exts =  
"fastq"), dir = getwd(), genome, genomedir, paired = FALSE, threads  
= 4L, molecule = "RNA" )
```

### Arguments

<code>file</code>	A list of paths to FASTQ files. If no paths are entered, it defaults to all fastq files in <code>dir</code> .
<code>dir</code>	Output directory.
<code>genome</code>	A BSgenome object, GmapGenome object, or a character string indicating the genome name eg. "hg19".

genomedir	A directory containing the reference genome. Otherwise, it is the parent directory of the reference genome where genome is a character string or BSgenome object.
paired	Indicates whether the samples are from paired-end or single-end reads.
threads	The number of cores to use in the process.
molecule	A character string indicating either DNA or RNA samples.

### Value

A list of file paths to created BAM files

## 3. counts()

### Description

This function counts reads of gene regions between sample groups. It assumes that sample BAM files are ordered in a directory structure such as "group/sample/" as processfastq() would order it. It outputs a CSV file showing gene counts. Works similarly to expression(), but outputs count data instead of differential expression data.

### Usage

```
counts( dir = getwd(), groups, TxDb, orgDb, outputdir = getwd(),
        threads = 4L, paired = FALSE )
```

### Arguments

dir	The parent directory of the sample groups.
groups	Folder names of the sample groups. The default is all folders in dir.
TxDb	A TxDb object upon which regions of the genome are counted.
orgDb	An orgDb object for annotating the CSV with gene symbols and Ensembl IDs.
outputdir	Output directory of CSV file.
threads	Number of cores to use.
paired	Indicates whether the samples are from paired-end reads.

### Value

A data frame containing gene counts.

## 4. expression()

### Description

This function analyzes differentially expressed genes between sample groups. It assumes that sample BAM files are ordered in a directory structure such as "group/sample/" as processfastq() would order it. There should be more than one sample per group or else differential expression analysis won't work. It outputs a CSV file showing differential expression (ordered by p-value). It works similarly to counts(), but then further analyzes those counts to obtain differential expression data.

### Usage

```
expression( dir = getwd(), groups, TxDb, orgDb, outputdir = getwd(),  
threads = 4L, paired = FALSE )
```

### Arguments

- dir        The parent directory of the sample groups.
- groups    Folder names of the sample groups. The default is all folders in dir.
- TxDb      A TxDb object upon which regions of the genome are counted.
- orgDb     An orgDb object for annotating the CSV with gene symbols and Ensembl IDs.
- outputdir Output directory of CSV file.
- threads   Number of cores to use.
- paired    Indicates whether the samples are from paired-end reads.

### Value

A data frame list containing all of the differential expression comparisons.

## 5.      callsnp()

### Description

This function calls single nucleotide polymorphism variants from BAM files. The results are formatted into a VCF file and the ID column is populated with dbSNP IDs.

### Usage

```
callsnp(bam, genome, genomedir, SNPlocs, threads = 4L, outputdir =  
getwd())
```

### Arguments

- bam        A list of paths to BAM files
- genome    A BSgenome object, a GmapGenome object, or a character string indicating the reference genome eg. "hg19"

genomedir	The directory containing the reference genome or, if genome is a character string, the parent directory of the reference genome directory.
SNPlocs	An SNPlocs object containing dbSNP IDs.
threads	The number of cores to use.
outputdir	The output directory for the VCF file.

### Value

A list of file paths to the VCF files.

## 6. callcnv()

### Description

This function calls copy number variants from sample BAM files compared to control BAM files. It assumes that BAM files are stored in separate folders as is created by processfastq(). This function requires that control BAM files are provided. Once complete, it creates a CSV file containing copy number information.

### Usage

```
callcnv( controldir, control = NULL, experimentdir, experiment =
NULL, bed, outputdir = getwd() )
```

### Arguments

controldir	The parent directory of the sample directories.
control	The names of the folders in which control BAM files are. If NULL, all folders in controldir will be checked for BAM files.
experimentdir	The parent directory of the sample on which to investigate copy numbers.
experiment	The names of the folders in which sample BAM files are. If NULL, all folders in experimentdir will be checked for BAM files.
bed	A character string indicating BED file path or a TxDb object from which to extract a BED file.
outputdir	The directory in which to place the copy number call.

### Value

A data frame containing copy number calls.

## 7. callindel()

### Description

This function calls indel variants from BAM files. The results are formatted into a VCF file and the ID column is populated with dbSNP IDs.

### Usage

```
callindel(bam, genome, genomedir, SNPlocs, threads, outputdir =  
getwd())
```

### Arguments

bam	A list of paths to BAM files
genome	A BSgenome object, a GmapGenome object, or a character string indicating the reference genome eg. "hg19"
genomedir	The directory containing the reference genome or, if genome is a character string, the parent directory of the reference genome directory.
SNPlocs	An SNPlocs object containing dbSNP IDs. For indels, this may be an XtraSNPlocs object.
threads	The number of cores to use. Cores should equal a factor of reference genome sequence levels ie. chromosome contigs should be equally divisible between cores.
outputdir	The output directory for the VCF file.

### Value

A list of file paths to the VCF files.

## 8. vizexp()

### Description

This function visualizes expression data from a CSV file.

### Usage

```
vizexp(csvfile, metadata, species)
```

### Arguments

csvfile	The count data csv file
metadata	The metadata excel file.
species	Select Specie number from list.

## 9. vizsnp()

### Description

This function visualizes SNPs data from a VCF file.

### Usage

```
vizsnp(dir = getwd())
```

### Arguments

**dir**            The parent directory of the files (expected to include two folders, one including the control group vcf files and another including the patients group files)

## 10. vizcnv()

### Description

This function visualizes CNVs data from a VCF file.

### Usage

```
vizcnv( csvfile, species)
```

### Arguments

**csvfile**        The variant data vcf file.

**species**        Select Specie number from list.