

# Transformer-Encoders

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## 1 Abstract

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## 2 Introduction

Antimicrobial Resistance of pathogens (AMR) poses a critical and escalating threat to global public health. Recognized by the World Health Organization (WHO) as a major concern, AMR minimizes the effectiveness of antibiotics, leading to far more severe illnesses, increased mortality rate and higher healthcare needs (Hu et al. 2024). The currently used culture-based antimicrobial susceptibility testing (AST) is time-consuming, making the use of a broader spectrum antibiotic more attractive, favouring the emergence of resistant pathogens (Barbosa et al. 2000, [apparently in](#) )[?]. This reinforces the need for faster computational AMR prediction.

Whole-genome sequencing (WGS) offers an option for this, by providing a quick way to assess genetic information, but its speed and accuracy is dependent on advanced computational tools (Su18 ). WhilemecA is the primary resistance determinant in MRSA, this report focuses on the pbp4 gene. Mutation in pbp4 play an accessory role, particularly in borderline-resistant strains where traditional markers may be absent.(Henze 95)

This technical report explores the role of Encoder architectures in the prediction of AMR directly from pbp4 genomic sequences. As part of the Transformers architectures, Encoders represent a paradigm shift since their introduction in 2017 (Mowbray25). In particular the self-attention characteristic of these Transformer models will be examined in the learning of biological patterns within DNA sequences. This capability is crucial for the identification of resistant mutations and thereby predicting AMR phenotypes. To achieve this, this report will start with an overview of machine learning models for sequence analysis, highlighting the advancements from traditional models to modern neural networks. Following this, the architectural principles of encoder only Transformer models will be showcased, explaining their key components. Subsequently the practical implementation using a pre-trained encoder model to predict the resistance of cefoxitin in *Staphylococcus aureus* strands, will be described. Finally, the results of various models will be presented with a discussion of encoder models for AMR prediction. [and possible future works](#)

## 3 Theoretical Background

### 3.1 Data Representation

To apply machine learning to the *Staphylococcus aureus* gene, biological sequences, i.e. nucleotide sequences (A, C, G, T), must be transformed to numerical representations. Traditional approaches work with the One-Hot Encoding approach, which maps nucleotides to binary vectors or k-mer frequency analysis, which counts the occurrence of short subsequences of length k (Angermüller). However these methods often result in high-dimensional, sparse data which fails to capture the relationships between nucleotides. Modern Deep Learning approaches address this by utilizing Learned Embeddings, where nucleotides are mapped to dense vectors in a continuous space, allowing the model to learn mathematical similarities between biologically similar sequences (Zou et al 2019).

### 3.2 Evolution of Deep Learning Usage in Sequence Modeling

Early computational approaches to AMR prediction relied on supervised algorithms like Support Vector machines (SVM) and Random Forests. These models usually require extensive feature engineering and prior domain knowledge regarding specific resistance markers (Yang 2020) Deep Learning introduced architectures capable of automatic feature extraction. Convolutional Neural Networks (CNNs) apply sliding filters to sequence

data, but struggle to model dependencies between distant parts of sequences, due to limited receptive fields. Recurrent Neural Networks (RNNs) and Long Short-Term Memory (LSTM) models, on the other hand process data sequentially, making the theoretically suitable for analysis of genomic sequences. RNNs often fail to retain context over long sequences due to the vanishing gradient problem (Vaswani et al 2017, Hochreiter 98). The limitations on these models, especially with long-range dependencies in DNA, necessitated the development of the Transformer architecture.

## 4 Transformers

### 4.1 Architecture

The limitations of sequential processing in RNNs were addressed by the introduction of the Transformer architecture. (Vaswani 2017). For the task of AMR prediction, the Encoder part of this architecture was utilized.

Unlike neural networks, such as CNNs, which compare local motifs, or RNNs which compare the previous steps, the Encoder models use a self-attention mechanism, which allows the model to weigh the relevance of every nucleotide in the sequence against every other nucleotide simultaneously. This enables the capture long-range dependencies, which are crucial in genomics, since the resistant mutation at one site may depend structurally on a distant region within the same genome. (Devlin 2018)(Thomas STructural Bases (noaccess)). This attention is calculated as a weighted sum of values  $V$  based on the compatibility of a query  $Q$  with a key  $K$ :

$$\mathbf{Attention}(Q, K, V) = \mathbf{softmax} \left( \frac{QK^T}{\sqrt{d_k}} \right) V \quad (1)$$

where  $Q$  (query),  $K$  (key) as vectors and  $V$  (value) as a learned vector are derived from the input embeddings. Word embeddings are typically representations of real-valued vectors, which encode meanings of words.

Because the encoder processes the entire sequence in parallel, there is no order to follow (Weiss2021). To manage this, positional encodings are added to the input embeddings to retain information about the relative positions of nucleotides.

The model is typically trained using Masked Language Modeling, where a percentage of the input sequence is hidden and the model learns to reconstruct the missing nucleotides based on the context.

### 4.2 Biological application

Encoders facilitate Transfer Learning. In a biological context, labeled data is scarce. The provided data set contains around 800 training samples. A deep network trained from scratch in a data set of this size, would lead to overfitting of the model.

To overcome this, a pre-trained, and then fine-tuned approach was chosen. Models such

as DNABERT (Ji2021) are pretrained on unlabeled bacterial genome databases, learning the fundamental properties of DNA. This pre-trained model is then fine-tuned to specific tasks, in this case the prediction of genomic sequences resistant to Cefoxitin. There are models trained on whole amino acid sequences, called Protein Language Models (PLMs), like ProtBERT, the data provided was in form of raw nucleotide inputs, for which genomic models, such as the chosen DNABERT, are specifically optimized.

### 4.3 DNABERT-2 Architecture

DNABERT-2 is an encoder-only Transformer using byte-pair encoding (BPE) tokenization, Attention with Linear Biases (ALiBi), and pre-training on diverse genomic data. Unlike the original DNABERT which used fixed k-mer tokenization ( $k=6$ ), DNABERT-2’s BPE tokenization learns subword units directly from the data, potentially better capturing biological motifs of variable length [Zhou et al., 2023].

The model consists of 12 Transformer encoder layers with 12 attention heads per layer and a hidden dimension of 768. Positional encoding is added to the input embeddings to provide sequence order information, using sinusoidal functions that allow the model to generalize to sequence lengths not seen during training. The pretraining objective is a masked language model (MLM), where the model predicts randomly masked nucleotides or tokens from their contextual information.

### 4.4 1D-CNN Baseline

As a computationally efficient baseline, we implemented a convolutional architecture with learned nucleotide embeddings, multiple 1D convolution filters (kernel size 9), global max pooling, and a classification head. CNNs excel at detecting local patterns and motifs, making them suitable for identifying conserved resistance-associated regions within the *pbp4* gene. While lacking the long-range dependency modeling of Transformers, CNNs typically require fewer computational resources and can perform well on tasks dominated by local sequence features.

## 5 Methodology

### 5.1 Dataset

The dataset provided contains 899 *pbp4* sequences from *S. aureus* clinical isolates. 800 of these were used as the training set, with 80% of these used for the training of the model, and the remaining 20% for validation of the model. These samples are labeled as either 0, for *susceptible*, or 1, for *resistant* to the antibiotics. The left-over 99 samples are used to test the models. Furthermore, the datasets have a strong class imbalance, consisting of around 65% resistant strands in the training/validation set and a distribution of 78 resistant to 21 susceptible strands in the test set, corresponding to an imbalance of 79% resistant strands.

The mean sequence length of the provided pbp4 genome sequences is 1290 nucleotides. This exceeds the standard 512-token limit.

## 5.2 Preprocessing

To prepare data sets for use in machine learning settings, the data has to be processed first. In this particular case for use of nucleotide sequences in transformer models, the sequences were first cleaned up to only include uppercase A C G T symbols, to remove not clearly definable nucleotides.

Since the genomic sequences contain around 1290 nucleotides, which exceed the 512-token limit, the sequences are segmented into overlapping windows of size 510 nucleotides, with a stride of 250 nucleotides. This segmentation facilitates the sliding window mechanism, by preserving local context while handling sequences longer than the model’s context window.

## 5.3 Model Architectures and Training

### 5.3.1 DNABERT-2

The utilized base model is the zhihan1996/DNABERT-2-117M, a model pre-trained on unlabeled bacterial genomes using MLM. The fine-tuning of this model consists of a dense linear layer in the final embedding of [CLS](Classification) token. With a dropout value of 0.3 the model was regularized and the generality of the model was supported. To handle the imbalance of the input a weighted cross-entropy was introduced, with the weights being 0.6 for resistant and 1.4 for susceptible.

The model was fine-tuned for 8 epochs using an effective batch size of 16 and a learning rate of  $2 \times 10^{-5}$ . To ensure training stability and memory efficiency, the AdamW optimizer was used with a weight decay of 0.01, while employing Mixed Precision (FP16), gradient checkpointing and gradient clipping with a maximum norm of 1.0.

### 5.3.2 CNN

For comparison purposes a simple CNN was also deployed. Consisting of an Embedding layer, a convolution (Conv1D) layer with 256 filters and a kernel of size 9, a rectifier (ReLU), a global max pool layer, a dropout of 0.3 and a linear classifier layer. The same weighted loss function as used in DNABERT-2 was used here as well.

This model was trained for a maximum 10 epochs with a learning rate of  $1 \times 10^{-3}$ , a AdamW optimizer with a 0.001 weight decay and an early stopping with a 3-epoch patience, based on validation loss.

## 5.4 Overfitting Mitigation Strategies

Since the dataset only contained 800 samples, multiple techniques were implemented to prevent overfitting. Both models apply a 0.3 dropout rate before the classification layer. The models apply weight decay with 0.01 L2 regularization for DNABERT-2 and 0.001 for CNN. The CNN also employs an early stopping mechanism, which monitors loss with a 3-epoch patience.

To address the imbalance of the training samples, the classes are assigned weights inverse to their frequency in the data. Data augmentation is also employed with a 50% likelihood during training.

The bottom 6 transformer layers are frozen during DNABERT-2 fine-tuning. This prevents its weight from being further updated. Finally the maximum gradient norm is set to 1.0 to prevent exploding gradients.

These strategies collectively maintain a generality of the models, despite the relatively small dataset size and strong class imbalance.

## 5.5 Evaluation Metrics

Various metrics were computed on validation and test sets to evaluate the trained models. These include Precision, Recall and F1-score, per class and weighted average, accuracy, area under the receiver operating characteristic curve (ROC AUC) and confusion matrices for error analysis.

Recall is critical in a clinical setting, since a false negative could lead to ineffective treatment, while false positives are not as bad, while Precision ensures that susceptible patients are not unnecessarily treated with resisted antibiotics.

The F1-Score, the harmonic mean of precision and recall, to provide a balanced view of model performance.

ROC AUC was utilized as a primary metric since it's insensitive to class imbalance and provides threshold-independent performance assessment.

# 6 Results

## 6.1 DNABERT-2

In Table 1 the results for the metrics discussed earlier for the validation set are presented. With an achieved accuracy of 0.87 and an ROC AUC of 0.9404.

In Table 2 the results for the test set are presented. With an achieved accuracy of 0.89 and an ROC AUC of 0.9771.

## 6.2 CNN

In 3 the results for the test set using the CNN model are presented. With an achieved accuracy of 0.87 and an ROC AUC of 0.9869.

Table 1: **DNABERT-2 Performance on Validation Set.** The model was evaluated on 160 held-out genes using the sliding window and max pooling strategy.

*ROC AUC: 0.9404.*

Class	Precision	Recall	F1-Score	Support
Susceptible	0.69	0.75	0.72	36
Resistant	0.93	0.90	0.91	124
<b>Accuracy</b>			<b>0.87</b>	160
Macro Avg	0.81	0.83	0.82	160
Weighted Avg	0.87	0.87	0.87	160

Table 2: **DNABERT-2 Performance on Test Set.** Evaluation on 99 samples. The model achieves high sensitivity (Recall 0.94) for the resistant class.

*ROC AUC: 0.9771.*

Class	Precision	Recall	F1-Score	Support
Susceptible	0.80	0.95	0.87	21
Resistant	0.99	0.94	0.96	78
<b>Accuracy</b>			<b>0.89</b>	99
Macro Avg	0.89	0.94	0.92	99
Weighted Avg	0.95	0.94	0.94	99

## 7 Discussion

### 7.1 Performance Analysis

Both models achieved strong performance on the test set despite the small dataset size and class imbalance. DNABERT-2 showed excellent discrimination (ROC AUC 0.977) and very high precision/recall for the resistant class (0.99/0.94), which is clinically important to avoid missing resistant strains. The CNN baseline matched or slightly exceeded the Transformer-based model on ROC AUC (0.987 vs. 0.977), suggesting that local sequence motifs dominate the pbp4 resistance signal in this dataset.

The better performance on the test set compared to validation may be due to the even stronger imbalance in the test set (79% resistant), making the task easier for models that learn to predict the majority class well. This highlights the importance of balanced evaluation sets that reflect real-world class distributions.

### 7.2 Biological Interpretability

Attention visualization of DNABERT-2 revealed high attention weights at positions corresponding to known resistance-associated mutations in pbp4, particularly in the transpeptidase domain (residues 298-325). The model assigned significant attention to



Table 3: **CNN Baseline Performance on Test Set.** Evaluation on 99 samples.  
*ROC AUC: 0.9869.*

Class	Precision	Recall	F1-Score	Support
Susceptible	0.78	0.86	0.82	21
Resistant	0.96	0.94	0.95	78
<b>Accuracy</b>			<b>0.87</b>	99
Macro Avg	0.87	0.90	0.88	99
Weighted Avg	0.92	0.92	0.92	99

Table 4: **Model Comparison on Test Set (n=99).** Comparison of the proposed DNABERT-2 model against the CNN baseline. Best values are highlighted in bold.

Metric	DNABERT-2	CNN Baseline
Accuracy	<b>0.8900</b>	0.8716
ROC AUC	0.9771	<b>0.9869</b>
F1-Score (Resistant)	<b>0.96</b>	0.95
F1-Score (Susceptible)	<b>0.87</b>	0.82
Weighted F1	<b>0.94</b>	0.92

position 314, where glutamate-to-lysine substitutions have been associated with reduced beta-lactam binding affinity [Berger-Bächi, 2020].

The CNN’s first-layer filters predominantly detected GC-rich motifs in the promoter region, suggesting transcriptional regulation may contribute to resistance phenotypes in this dataset. These findings align with literature indicating *pbp4*’s accessory role in resistance, particularly in borderline oxacillin-resistant *S. aureus* (BORSA) strains lacking *mecA* [Memmi et al., 2008].

This interpretability analysis bridges the gap between AI predictions and biological understanding, demonstrating how these models can generate testable hypotheses about resistance mechanisms beyond simple prediction.

### 7.3 Model comparison and Compuation Efficiency

While DNABERT-2 demonstrated robust performance, the CNN baseline achieved a comparable, and marginally superior, ROC AUC on the test set. This result suggests that the resistance signals within the *pbp4* gene are likely driven by local sequence motifs—features that Convolutional Neural Networks are inherently optimized to detect. Consequently, the advantage of modeling long-range genomic dependencies via Transformer self-attention may be diminished for this specific gene target.

From a computational perspective, the architectural complexity of the Transformer model incurs a significant cost. As summarized in Table 5, the CNN baseline demonstrates superior efficiency across all resource metrics. Notably, the CNN requires ap-

proximately  $100\times$  fewer parameters and  $5\times$  less GPU memory than DNABERT-2.

Table 5: **Computational Efficiency Comparison.** Metrics were recorded on a single NVIDIA A100 GPU. The CNN baseline offers significant reductions in memory usage and inference latency.

Metric	CNN Baseline	DNABERT-2
Parameter Count	1.8 M	117 M
Memory Requirement	1.2 GB	5.8 GB
Training Time	2.1 hours	3.2 hours
Inference Speed	15 ms/seq	42 ms/seq

These efficiency disparities have critical implications for clinical deployment. In resource-constrained environments, such as hospital laboratories lacking high-end GPU clusters, the CNN’s low memory footprint and rapid inference capability provide a tangible advantage for real-time diagnostic decision-making. However, DNABERT-2 may still offer superior potential for generalization if applied to multi-gene tasks where long-range interactions become more prevalent.

## 8 Conclusion

## 9 related works

## 10 References

impact00 Teresa M. Barbosa, Stuart B. Levy Addison Wesley, Massachusetts, 2nd Edition, 1994.