

CMDA-3654

Homework 10

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Due as a .pdf upload

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## Problem 1: [100 pts] More PCA

Food technicians are interested in comparing pizza doughs from Naples, made with traditional methods, with doughs from other places. Even though they taste different, previous research has not found good univariate characteristics to distinguish between the different types of doughs. Therefore, the food technician collected a multivariate data set.

Doughs from six restaurants are investigated, with five samples from each restaurant. The first four restaurants are famous Naples restaurants and the last two are from other Italian cities. For each dough two mechanical test (measuring pressure load and deformation volume) and four microbiological/chemical tests (counting the bacteria in the dough, counting the yeast, measuring pH and measuring total titratable acidity) are performed. The data is given in `doughs.csv`.

- a. Produce a principal component analysis object called `myPCAfit` for the first 6 columns of the `doughs` dataset only (the last column is just a restaurant ID). Remember that you have to do either `scale. = TRUE` if using `prcomp()` or `cor = TRUE` if using `princomp()`.

```
#Reading csv file first 6 columns
```

```
doughs<-read.csv("/Users/eduardosalvador/Desktop/FINAL\ Spring\ Semester\ 2021/CMDA\ /Assignments/HW10/doughs.csv")
doughs[,1:6]
```

	PressureLoad	DefVol	Bacteria	Yeast	pH	TTA
1	130.02568	35.52756	5.757972	6.542892	6.097234	1.1110839
2	123.86851	40.34567	6.034986	6.317691	6.063427	1.1380931
3	115.46597	26.58752	5.410269	6.668726	5.784212	1.1991341
4	127.58992	41.19118	5.870146	6.534598	6.148323	0.9865285
5	132.26647	45.74070	5.709261	6.838558	5.837073	1.6194396
6	122.15912	56.40890	7.318165	6.382213	5.865991	1.2675547
7	120.19586	33.66885	5.467638	6.436234	5.811155	1.3182378
8	126.89051	46.73313	7.784137	7.380511	5.743203	1.5392976
9	115.51803	35.78070	6.235717	7.306624	5.511193	1.4904791
10	113.86556	50.23146	7.928312	7.503896	5.718868	1.3443432
11	103.65008	28.95107	4.732192	6.990209	5.528908	1.6071463
12	108.79718	35.04796	8.945713	7.221850	5.728546	1.4261939
13	115.14925	46.68753	6.096369	7.935406	5.671798	1.6282453
14	106.21542	34.87870	6.184451	7.346561	5.524667	1.4803353
15	109.43033	44.43017	5.293796	7.108836	5.854408	1.5413321
16	113.28606	37.58062	5.599988	5.434055	5.918052	1.2092090
17	114.27267	31.29681	5.271806	6.440685	5.998186	0.8948270
18	112.45257	30.09542	6.410794	6.071087	5.840823	0.8860809
19	132.13855	53.15689	7.256938	6.436790	5.904885	1.3014732
20	125.52017	52.56363	7.022526	5.907304	5.991819	1.1720496
21	97.05126	22.09524	7.141561	6.147562	6.075787	1.1939264
22	94.34469	40.03719	4.623785	7.830586	6.153740	1.0555925
23	107.19449	34.88862	6.691160	7.366451	6.104683	1.2118887
24	102.57482	46.07382	8.085106	7.755833	6.092374	1.3194596
25	93.07118	37.95756	6.063649	6.874420	6.204745	0.9398126
26	111.32353	36.67969	5.238570	6.921054	6.075195	1.1870484
27	96.58765	36.64397	6.341320	7.444028	5.858414	1.2855114
28	105.08312	44.87899	5.951783	7.109943	6.009198	1.2542399
29	103.23041	47.07211	6.545814	7.401993	5.847526	1.2924900
30	106.80567	26.13801	6.504370	6.443620	6.075960	1.2830062

```
#using princomp to standarize the data
```

```
myPCAfit<-prcomp(doughs[, -7], scale=T)
```

```
myPCAfit
```

Standard deviations (1, ..., p=6):

```
[1] 1.4562678 1.2277267 1.0585905 0.8569041 0.5263959 0.4898777
```

Rotation (n x k) = (6 x 6):

	PC1	PC2	PC3	PC4	PC5
PressureLoad	-0.1171101	0.6838728	-0.3728393	0.12391859	0.5569172
DefVol	-0.3326713	0.4931852	0.3674845	0.48092876	-0.5285789
Bacteria	-0.2791707	0.2758539	0.5205766	-0.74978124	0.1123330
Yeast	-0.4108043	-0.4192826	0.4097314	0.35965118	0.4658433
pH	0.5121716	0.1548093	0.4767097	0.24759902	0.3974856
TTA	-0.6054740	-0.1150096	-0.2445692	0.02292519	0.1510763

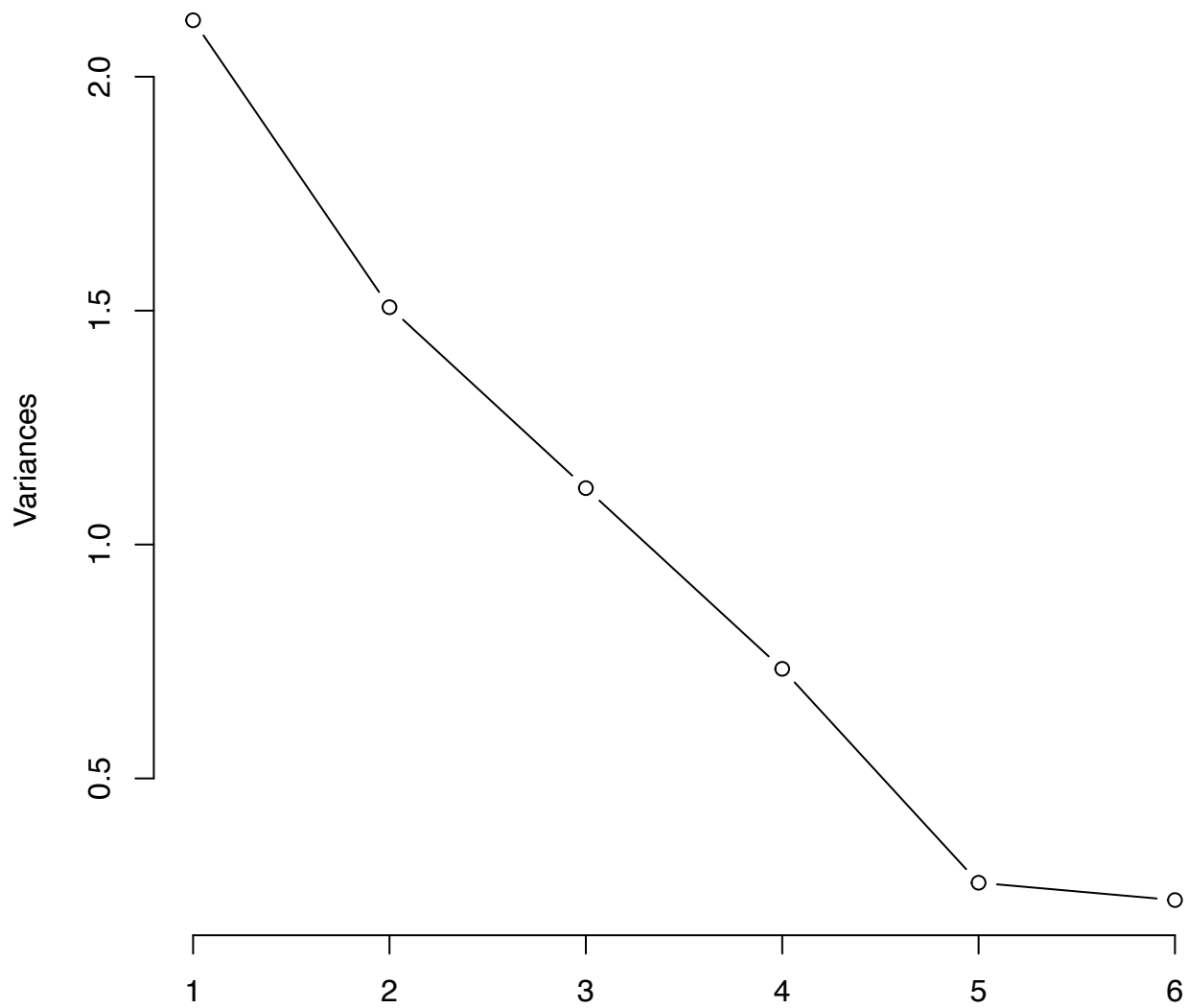
	PC6
PressureLoad	-0.23255427
DefVol	0.01910856
Bacteria	-0.01333191
Yeast	-0.37577017
pH	0.51687727
TTA	0.73280986

- i. Plot an associated screeplot using `screeplot(myPCAfit, type = "lines")`. You can use the elbow method to determine how many principal components seem sufficient for capturing the majority of the variation of the data.

```
#Plotting screeplot
```

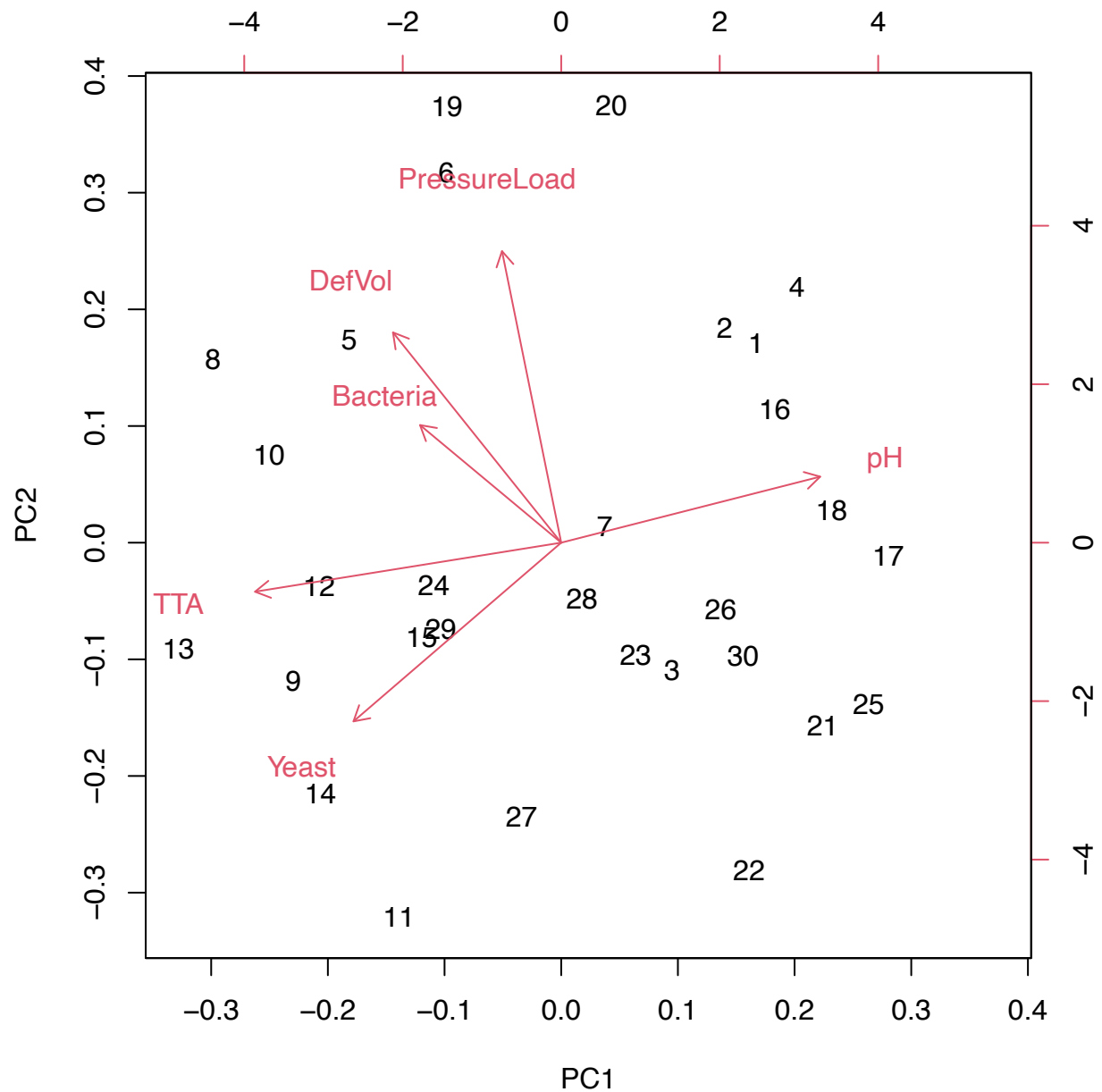
```
screeplot(myPCAfit, type="lines")
```

## myPCAfit



- ii. Construct a biplot for PC2 versus PC1. Based upon the loadings and the results of seen in biplot can you determine which variables are the most important for PC1? What about PC2?

```
biplot(myPCAfit)
```



The function `biplot(myPCAfit)` will easily make this plot for you.

Additionally if you install and enable the `ggfortify` library then you can also make this plot doing

```
library(ggfortify)
autoplot(myPCAfit, loadings = TRUE, loadings.label = TRUE)
```

- b. Find the first four principal components of the data (don't forget to scale the data – it's an option in the R functions). Do they seem to be sufficient to describe most of the variation in the data set (specifically report how much variation they describe individually and together)?

```
#Using summary function to find the four principal principal components
summary(myPCAfit)
```

Importance of components:

PC1	PC2	PC3	PC4	PC5	PC6
-----	-----	-----	-----	-----	-----

Standard deviation	1.4563	1.2277	1.0586	0.8569	0.52640	0.4899
Proportion of Variance	0.3534	0.2512	0.1868	0.1224	0.04618	0.0400
Cumulative Proportion	0.3534	0.6047	0.7914	0.9138	0.96000	1.0000

#PC1 has 33% variation, PC2 has 25%, PC3 has 19% and PC4 has around 12% variation. In total,  
 #all PC's together have around 91% variation suggesting that it is sufficient to describe most of the variation.

- c. Use `grid.arrange()` to plot scatter plots of the first three principal components versus each other. Judging from the plots, can the first three PCs be used to discriminate doughs from Naples and doughs from other places? Does this agree with your conclusion in (a)?

```
library(gridExtra)
library(tidyverse)
```

```
#Making changes to doughs dataframe using mutate
```

```
mdoughs<-mutate(doughs,Naple=
  case_when(Restaurant==1~1,
            Restaurant==2~1,
            Restaurant==3~1,
            Restaurant==4~1,
            Restaurant==5~0,
            Restaurant==6~0))
```

```
#Assigning first three principal components to mdoughs variable
```

```
mdoughs$PC1<-myPCAfit$x[,1]
mdoughs$PC2<-myPCAfit$x[,2]
mdoughs$PC3<-myPCAfit$x[,3]
```

```
#Using qplot to wrap for creating a number of different types of plots
```

```
#Assigning variable for PC1 vs PC2
```

```
scp1<-qplot(PC1,PC2,data=mdoughs,col=Naple)
```

```
#Assigning variable for PC1 vs PC3
```

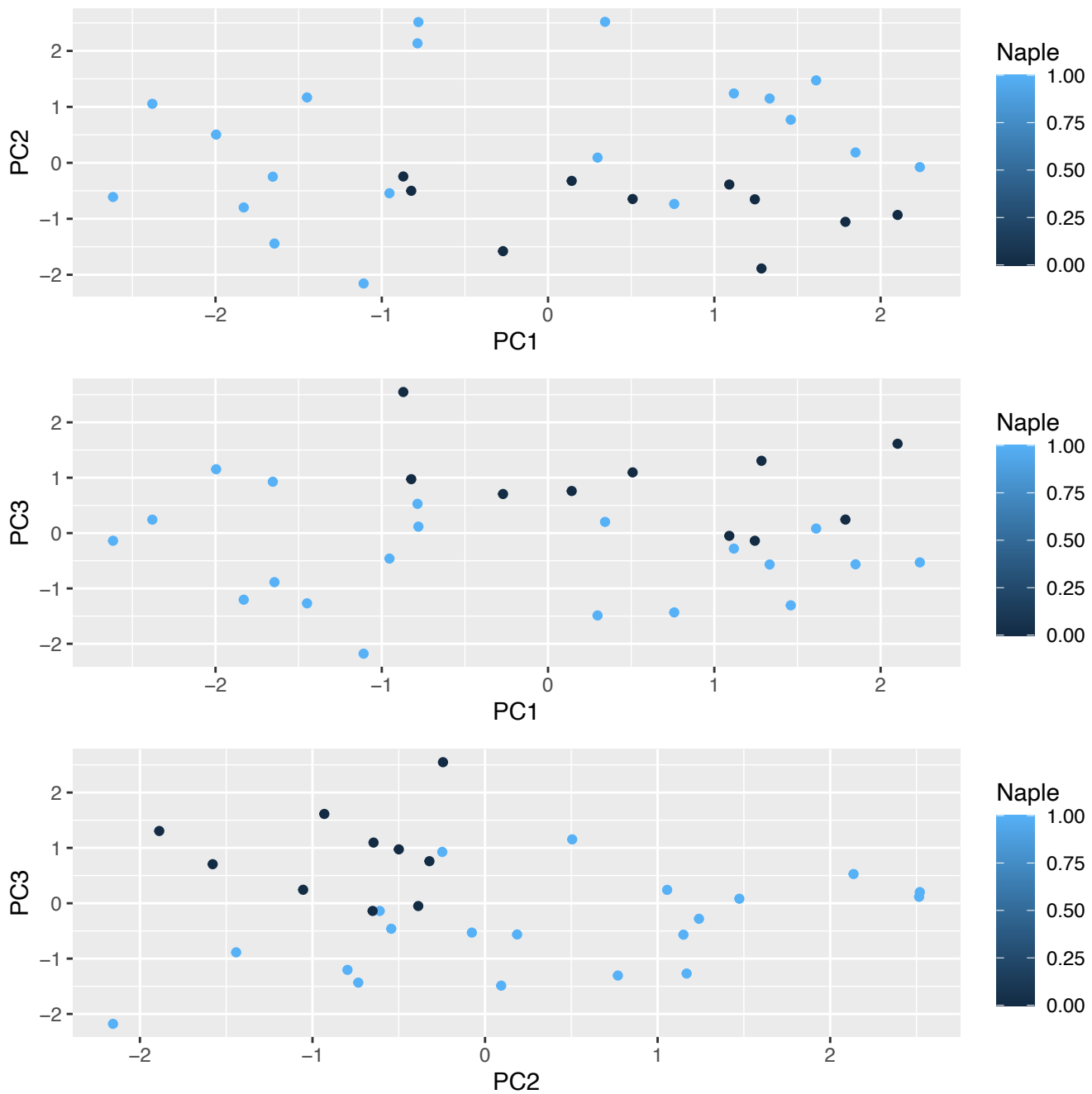
```
scp2<-qplot(PC1,PC3,data=mdoughs,col=Naple)
```

```
#Assigning variable for PC1 vs PC2
```

```
scp3<-qplot(PC2,PC3,data=mdoughs,col=Naple)
```

```
#Using grid.arrange to plot scatter plots
```

```
grid.arrange(scp1,scp2,scp3)
```



#Judging from the plots, the first 3 can be used to discriminate doughs from Naples and others  
 #It does agree

Hint: You will need to use ggplot multiple times to get plot objects p1, p2, etc for plotting the principal components versus each other and you need to be plotting the component scores for the different restaurant. You need to colorize the restaurants from Naples to be the same color and the restaurants that aren't from Naples a different color.

- d. If you had to perform a specific statistical learning method to classify the doughs based upon these features, which method would it be and why?

#If I had to perform a specific statistical method to classify the doughs based upon the features,  
 #I would use K means clustering because it can divide the dataset into non-overlapping data points.