



M1 ISDD (Feb 2018)
PROTEIN DOCKING

Lesson 6

**Interface prediction, hot-spots and
biomedical applications**

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- **Interface prediction**
- Hot-spot identification
- Application to drug design targeting PPIs
- Pathological mutations affecting PPIs

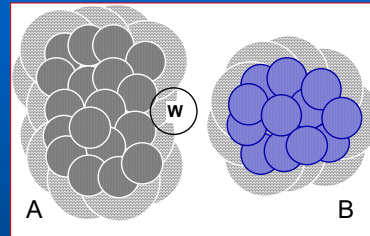
Protein-protein interface size

Defining macromolecular interfaces based on solvent accessibility

ASA accessible surface area

measures molecule-solvent contacts with the rolling ball algorithm

Lee & Richards, JMB, 1971



Protein-protein interface size

Defining macromolecular interfaces based on solvent accessibility

ASA accessible surface area

measures molecule-solvent contacts with the rolling ball algorithm

Lee & Richards, JMB, 1971

BSA buried surface area

measures molecule-molecule contacts

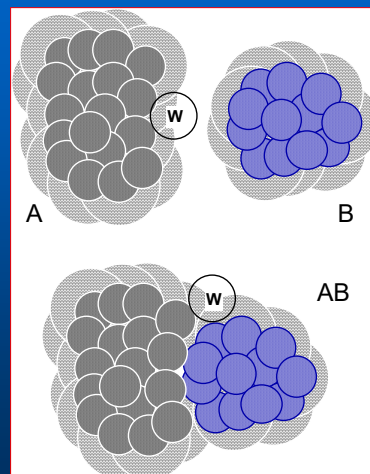
Chothia & Janin, Nature, 1975

Interface atoms or residues

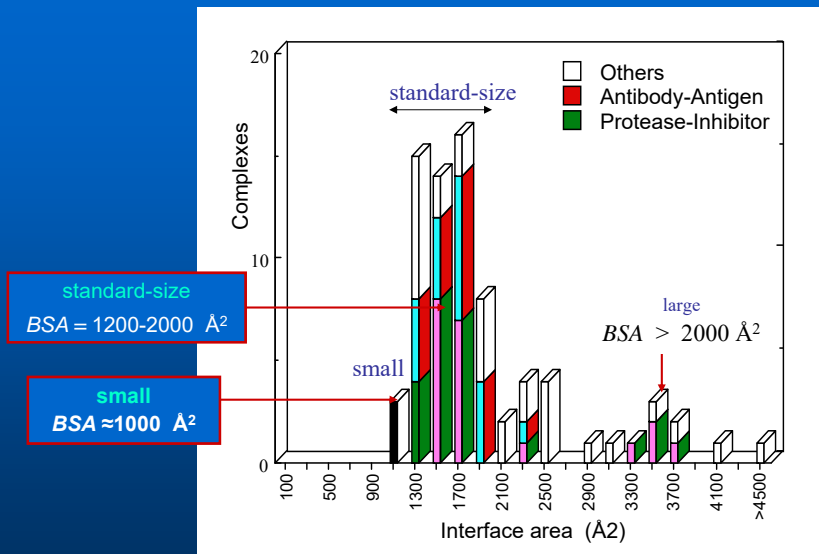
are all atoms or residues that contribute to the BSA. On average, each interface atom contributes $\approx 10 \text{ \AA}^2$

Hydrophobic effect

$$BSA = ASA_A + ASA_B - ASA_{AB}$$

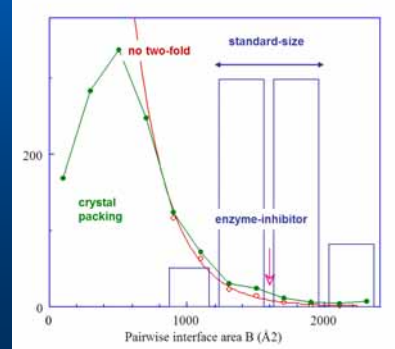
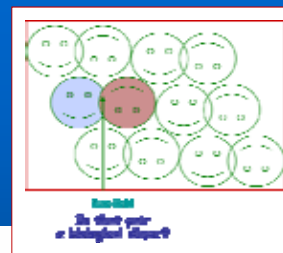
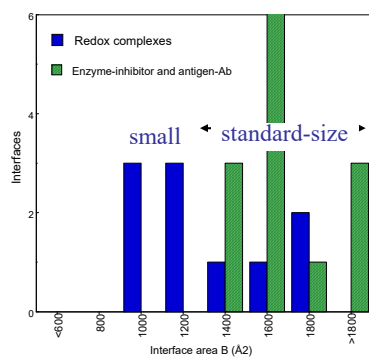


Protein-protein interface size

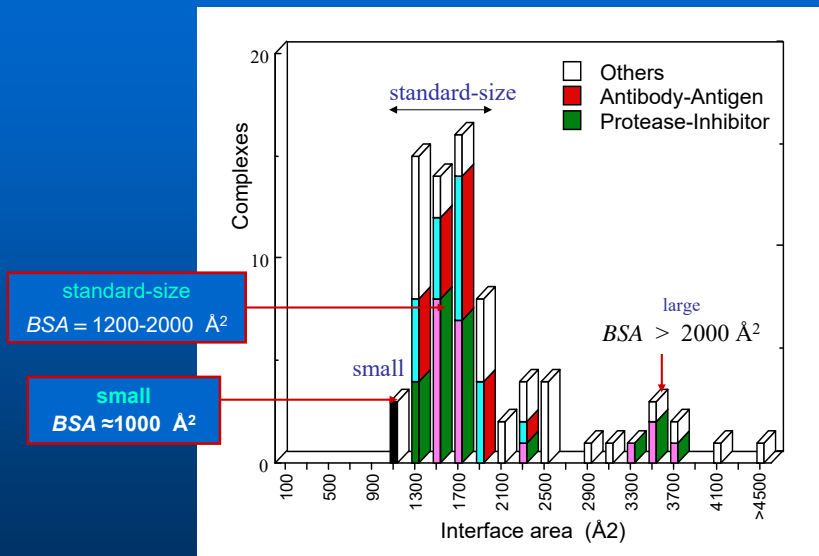


Protein-protein interface size

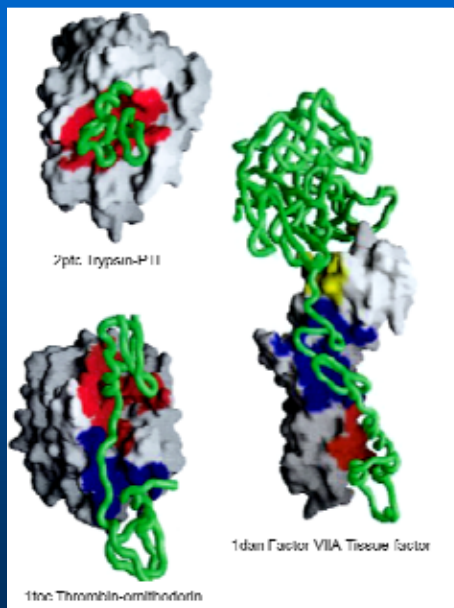
Electron transfer complexes
have small interfaces
BSA = 900 1200 Å²
0-3 H bonds
Crawley & Carrondo (2004)



Protein-protein interface size

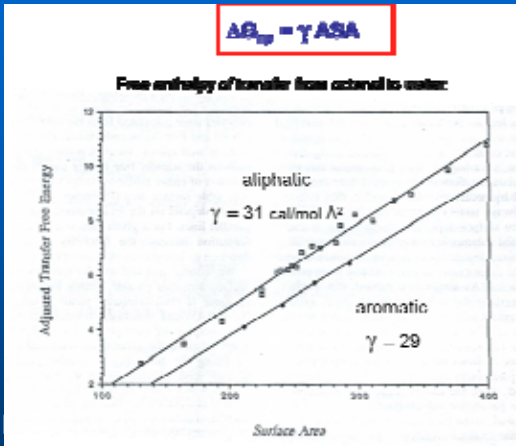
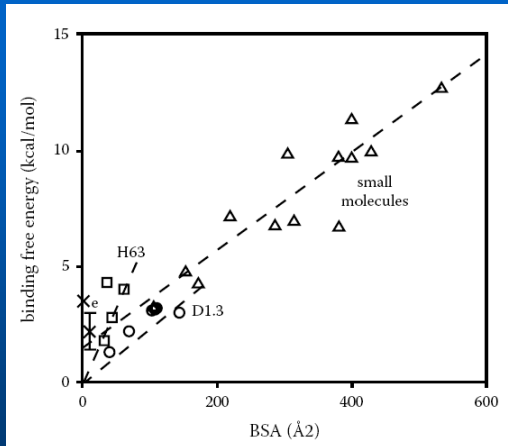


Protein-protein interface size



Large interfaces are multi-patch

Protein-protein interface size and desolvation energy



Protein-protein interface prediction

Common Descriptors Used for Prediction of Interaction Sites

Descriptor	Comments
Features Derived from 3-D Structure	
Neighbor list: Residues in spatial vicinity to the residue in question	9–20 residues
B-factor	A crystallographic measure that approximates the flexibility of a residue.
Solvent accessibility (ASA)	Measured in Å².
Relative solvent accessibility	Measured as a fraction of the overall surface of the residue that is exposed to solvent.
Shape index/curvedness	Three state (helix, strand, loop) or more.
Secondary structure	The separation in sequence between residues within the same patch. Some results indicate that structurally contiguous residues that are not adjacent in sequence are more likely to form interaction sites.
Sequence distance	
Planarity	
Predicted/Approximate Structural Features	
Predicted secondary structure	Methods relying only on sequence can use computational tools to generate predicted solvent accessibility/secondary structure. This improves performance without limiting applicability to proteins with known 3-D structure.
Predicted solvent accessibility	
Sequence neighbor list	Can be used instead of neighbor list to approximate the environment of the analyzed residue. Nine to fifteen residues around the residue in question. Four to seven on each side of the residue. Some structure-based methods use this in addition to neighbor list.

Protein-protein interface prediction

Common Descriptors Used for Prediction of Interaction Sites

Descriptor	Comments
Sequence profile	Evolutionary Features Extracted from a multiple sequence alignment, a profile reveals patterns of evolutionary conservation.
Conservation score	A quantification of the level of conservation of an individual position.
Conservation of physicochemical traits	If the position is not conserved, scoring conservation of traits such as charge, hydrophobicity, or size may improve prediction.
Hydrophobicity	Physicochemical Features Several different scales are available.
Electrostatic potential	Measured for individual residue or for a patch. Requires 3-D structure.
Atom propensities	Serves as a way to sum physicochemical properties across residues in the patch.
Desolvation energy	Used mostly in predictions for rigid-body docking.
Protein-protein interaction	External Knowledge Can be used to: (1) identify sequence or structural elements that are significantly overrepresented in interacting pairs, and (2) to assess coevolution of positions in interacting pairs.
Functional annotation of the protein	Enzyme-inhibitor and antigen-antibody have different types of interfaces than other complexes. Adding this information may improve prediction.

AG - YASA

Protein-protein interface prediction

Method name	Input data	Method	Details	Web
ISIS ⁵⁸	sequence	Neural network	Predicted structural features, evolutionary information	http://cubic.bioc.columbia.edu/services/isis/
TreeDet ⁷¹	sequence, structure	Scoring function	sequence and structural alignments	http://treedetv2.bioinfo.cnio.es/treedet/index.html
Promate ⁷³	structure	Scoring function	Secondary structure, sequence conservation, residue type	http://bioinfo41.weizmann.ac.il/promate/
PINUP ⁷⁶	structure	Scoring function	side-chain energy score, propensity, sequence conservation	http://sparks.informatics.iupui.edu/PINUP/
InterProSurf ⁵⁹	structure	Scoring function	solvent accessibility, propensities	http://curie.utmb.edu/
PRISM ⁷⁷	structure	Scoring function	geometric complementarity, conservation	http://prism.cccb.ku.edu.tr/prism/
ConSurf ⁶⁸	structure	Scoring function	conservation	http://consurf.tau.ac.il/
ET ⁶⁶	structure	Scoring function	multiple sequence alignments	http://mammoth.bcm.tmc.edu/traceview/
JET ⁷⁰	structure	Scoring function	structural and functional conservation	http://www.ihes.fr/~carbone/data.htm
WHISCY ⁷⁹	structure	Scoring function	conservation, surface properties	http://www.nmr.chem.uu.nl/Software/whiscy/startpage.htm
PIER ⁶¹	structure	Scoring function	atomic statistical propensities	http://abagyan.ucsd.edu/PIER/

Protein-protein interface prediction

Method name	Input data	Method	Details	Web
SiteEngines ⁶⁰	structure	Hierarchical scoring function	structural matching, physico-chemical properties	http://bioinfo3d.cs.tau.ac.il/SiteEngine/
PPI-Pred ⁸⁴	structure	SVM	surface shape, electrostatic potential	http://bioinformatics.leeds.ac.uk/ppi-pred
cons-PPISP ^{80,81}	structure	Neural network	PSI-Blast sequence profile and solvent accessibility	http://pipe.scs.fsu.edu/ppisp.html
SPPIDER ⁸⁵	structure	Neural Network	solvent accessibility and other features	http://sppider.cchmc.org/
Patch Finder Plus ⁸²	structure	Neural Network	conservation, concavity, area, H-bond, residue frequency	http://pfp.technion.ac.il/
meta-PPISP ⁸⁷	structure	Meta web server	cons-PPISP, Promate and PINUP	http://pipe.scs.fsu.edu/meta-ppisp.html
PI ² PE ⁸⁸	structure	Meta web server	cons-PPISP, WESA, DISPLAR	http://pipe.scs.fsu.edu/
SHARP ²⁹⁰	structure	Energy-based, scoring function	Desolvation, hydrophobicity, ASA, propensity, surface shape	http://www.bioinformatics.sussex.ac.uk/SHARP2/sharp2.html
ODA ⁶²	structure	Energy-based	Desolvation energy	http://www.molsoft.com/oda.html
NIP ¹⁰⁰	structure	Energy-based	Docking simulations	https://life.bsc.es/pid/pydock/

Promate

<http://bioinfo41.weizmann.ac.il/promate/>

The screenshot shows the ProMate web interface. The title is "ProMate" with the subtitle "Predicting the location of potential protein-protein binding sites for unbound proteins". The interface includes a sidebar with links for "ProMate 2.0", "MultiProMate", "ProMateus", "ProMate's help", "ProMateus' help", "ProMateus' FAQ", "Check out our visitors around the world!", and "Memorial". The main content area has the following sections:

- Upload your pdb file:** A "Choose File" button and "No file chosen" text.
- And/Or enter a 4 letters PDB-ID:** A text input field and a "Search the PDB" link.
- Which chain to use?** A dropdown menu currently showing "none".
- Please choose the scores configuration:** Radio buttons for "use default" (selected) and "let me choose".
- Configuration options:**
 - Single amino acids distribution: ☐ use ☒ don't use
 - Atom distribution: ☒ use ☐ don't use
 - Chemical character: ☒ use ☐ don't use
 - Amino acid pairs distribution: ☒ use ☐ don't use
 - Evolutionary conserved residues: ☒ use ☐ don't use

Promate

<http://bioinfo41.weizmann.ac.il/promate/>

Please choose the scores configuration: ☒ use default ☐ let me choose

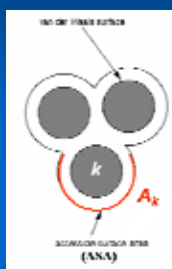
Single amino acids distribution: ☐ use ☒ don't use
 Atoms distribution: ☒ use ☐ don't use
 Chemical character: ☒ use ☐ don't use
 Amino acid pairs distribution: ☒ use ☐ don't use
 Evolutionary conserved positions: ☒ use ☐ don't use
 Non-regular secondary structure length: ☒ use ☐ don't use
 Sequence distances within a circle: ☒ use ☐ don't use
 Secondary structure: ☒ use ☐ don't use
 Domains: ☐ use ☒ don't use
 Hydrophobic patch rank: ☒ use ☐ don't use
 Hydrophobic patch size: ☐ use ☒ don't use
 Temperature factor (B-factor): ☐ use ☒ don't use
 Water molecules: ☒ use ☐ don't use
 Initial probabilities file: ☐ use ☒ don't use

Please select the output files to be produced:

- ☐ A text file with the surface dots probabilities
- ☒ A pdb file with the AAs colored by their interface probability (full range of colors)
- ☒ A pdb file with the surface atoms colored by their interface probability (full range of colors)
- ☒ A pdb file with the predicted interface patch colored in red

Optimal Docking Areas (ODA)

Desolvation Energy for Protein-Protein Docking



$$\Delta G_{\text{sol}} = \sum_{k=1}^N \sigma_k A_k$$

ASP are optimized
for protein
docking

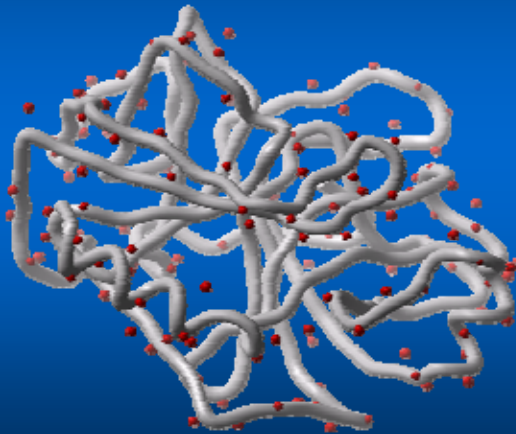
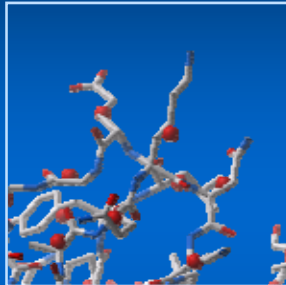
ASPs from
vacuum/water

ASPs from
octanol/water

ASPs for
interface/water

σ (cal/(mol Å ²))	Radius (Å)	Atom type
15.1	1.95	C aliphatic
17.7	1.8	C aromatic
-17.0	1.7	N uncharged
-54.8	1.7	N ⁺ , N ^ε in Lys ⁺
-27.3	1.7	N ^{η1} , N ^{η2} in Arg ⁺
-18.5	1.6	O hydroxyl
-13.6	1.4	O carbonyl
-29.9	1.4	O ⁻ in Glu, Asp
11.2	2.0	S in SH
2.2	1.85	S in Met or S-S

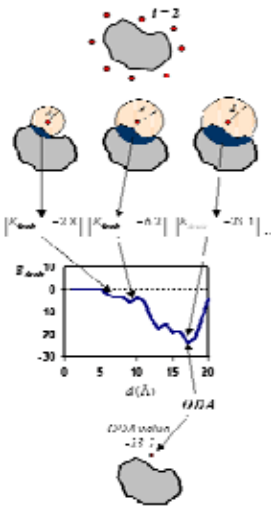
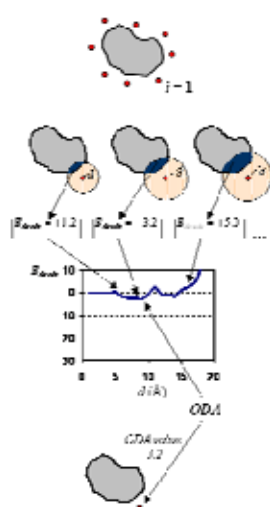
Optimal Docking Areas (ODA)



Optimal Docking Areas (ODA)

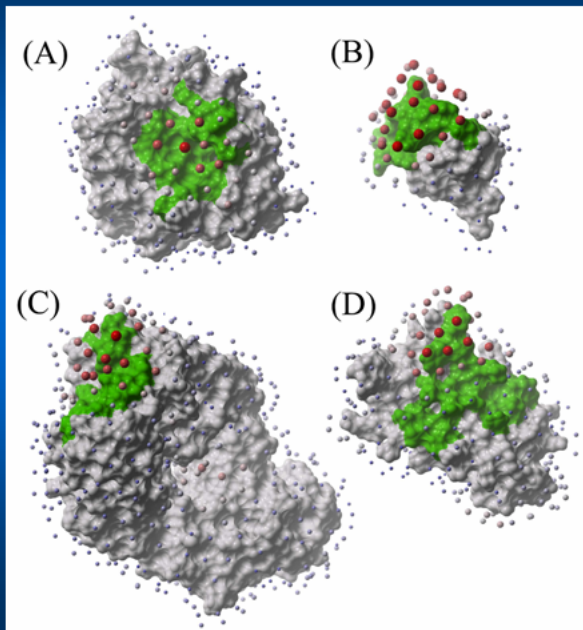
$$\Delta E_j^{desolv} \approx ASA_j \sigma_j$$

$$\Delta E_i^{ODA} = \min \left(\sum_j \Delta E_j^{desolv} \right)$$



Fernández-Recio et al. (2005) *Proteins* 58, 134-43

Optimal Docking Areas (ODA)



Benchmark: 66 unbound cases

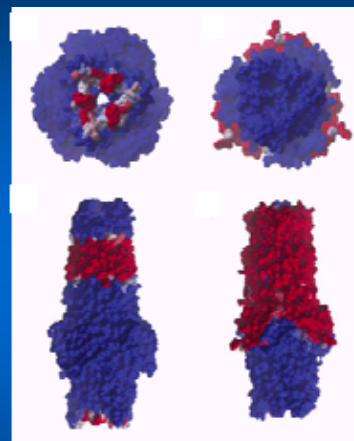
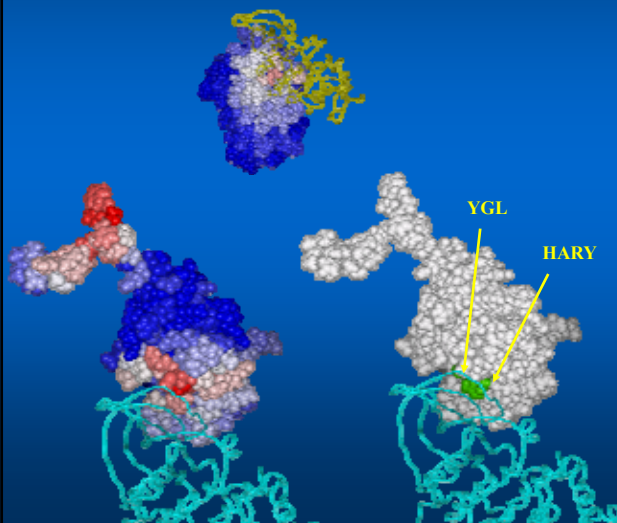
- ODAs in 50% proteins
- 80% correctly located in binding sites
- pyDockODA (centered in residues);



Fernández-Recio et al. (2005)
Proteins 58, 134-143

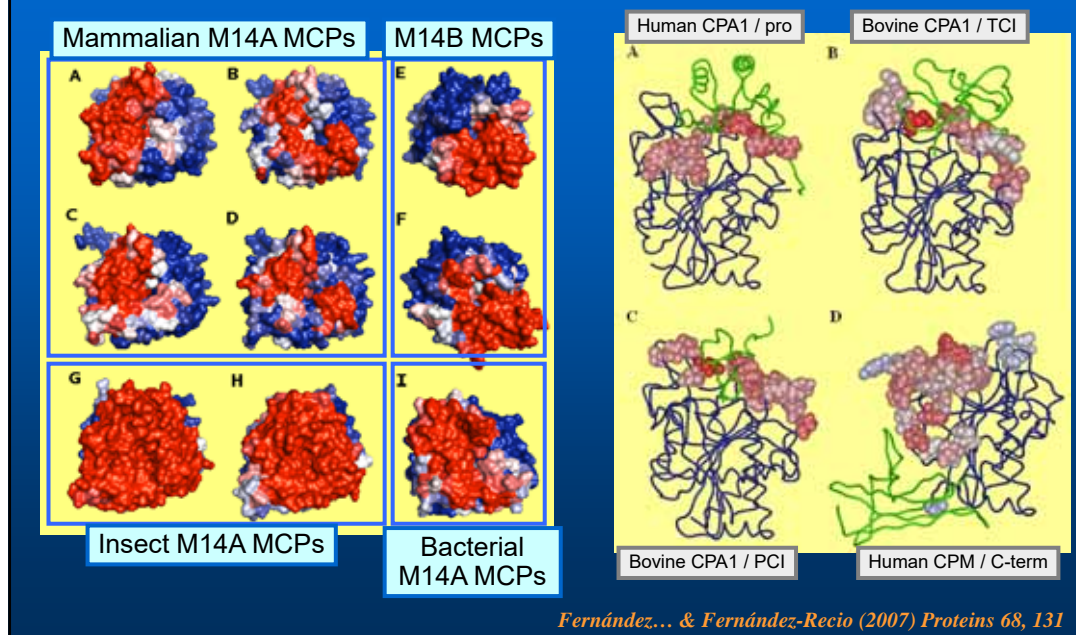
ODA - applications

Bolanos-García, Fernández-Recio et al.
TiBS (2006) 31, 654

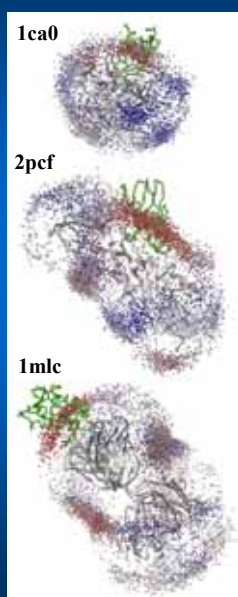


Federici, ..., Fernández-Recio et al.
(2005) J.Biol.Chem. 280, 15307-14

ODA - applications

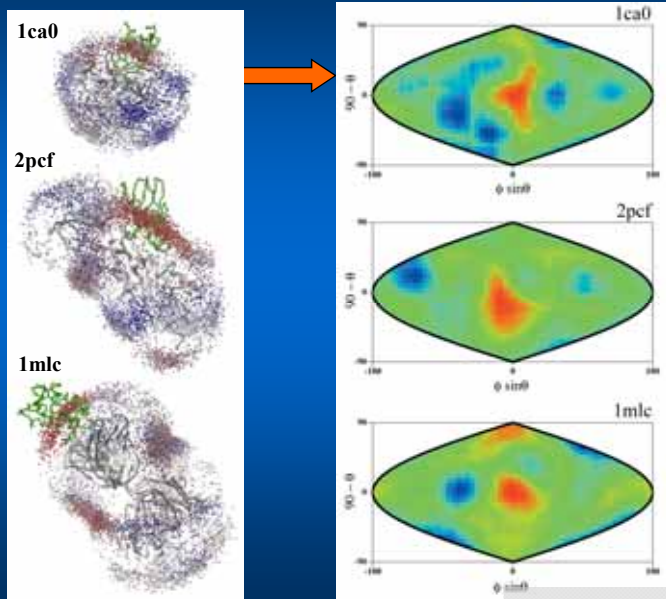


Interface predictions from docking results



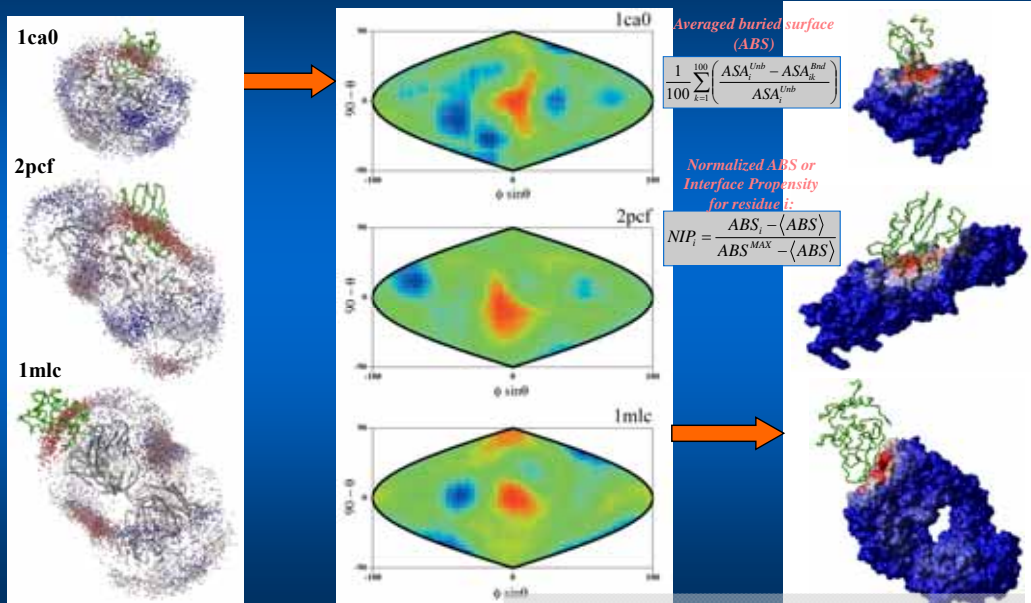
Fernandez-Recio et al. (2004) JMB 335, 843-865

Interface predictions from docking results



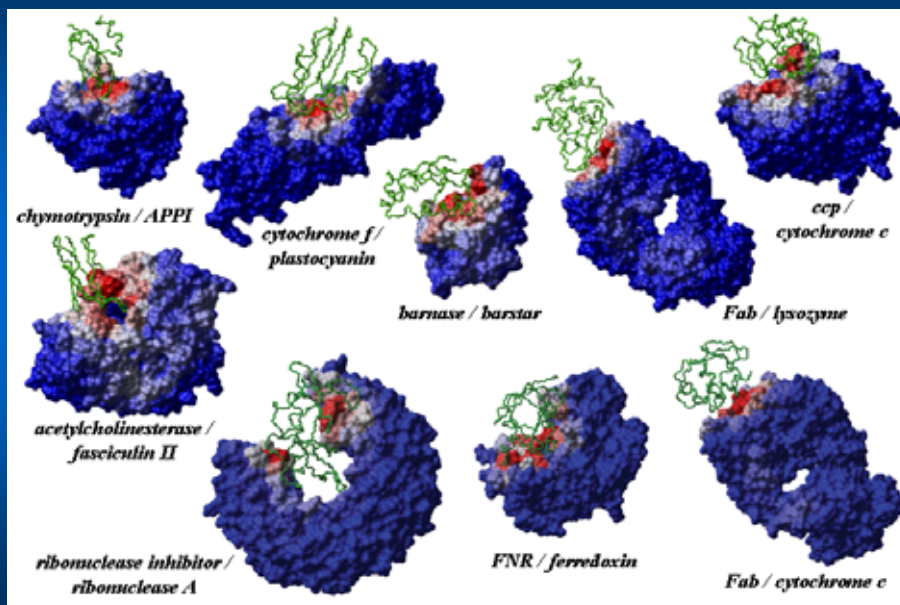
Fernandez-Recio et al. (2004) JMB 335, 843-865

Interface predictions from docking results

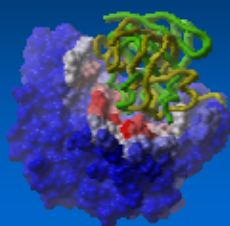


Fernandez-Recio et al. (2004) JMB 335, 843-865

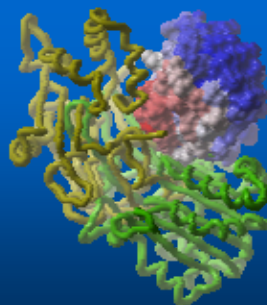
Interface predictions from docking results



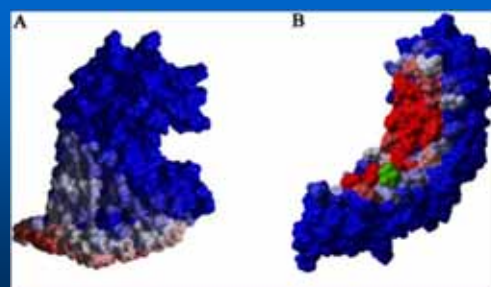
Interface predictions from docking results



Medina, Abagyan, Gómez-Moreno,
Fernández-Recio (2008) *Proteins* 72, 848



Sicilia et al. (2005) *Plant Physiology* 578, 5
Federici et al., *Trends in Plant Sciences* (2006)



- Interface prediction
- **Hot-spot identification**
- Application to drug design targeting PPIs
- Pathological mutations affecting PPIs

Hot-Spots

The **O-ring model**: alanine-scanning **hotspots** tend to be located at the center of the interface and surrounded by energetically unimportant residues that occlude solvent from them.

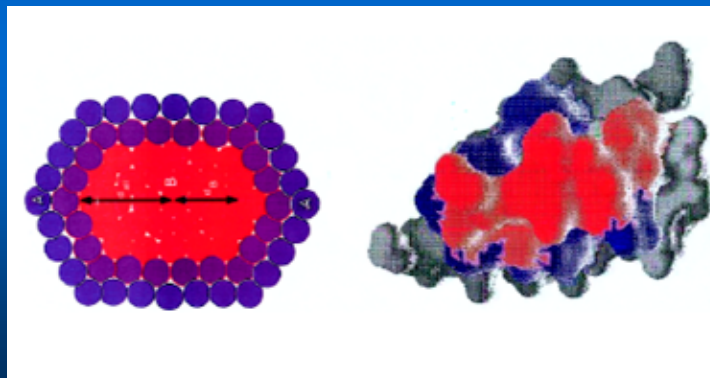
Bogan & Thorn (1998)

A sketch of the **solvent accessible and fully buried** atoms in a standard-size protein-protein interface. Each protein contributes about 70 atoms, and 1/3 are buried.

Lo Conte, Chothia & Janin (1999)

The **core/rim model** is a realistic implementation of that sketch. (Chakrabarti & Janin 2002)

The **core** and **rim** of the CI2 inhibitor interface with subtilisin (1sni)

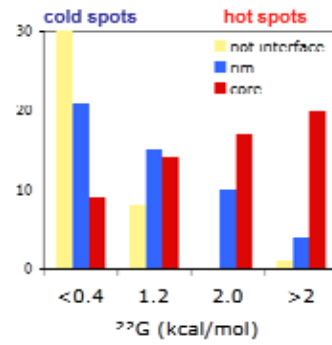


Hot-Spots

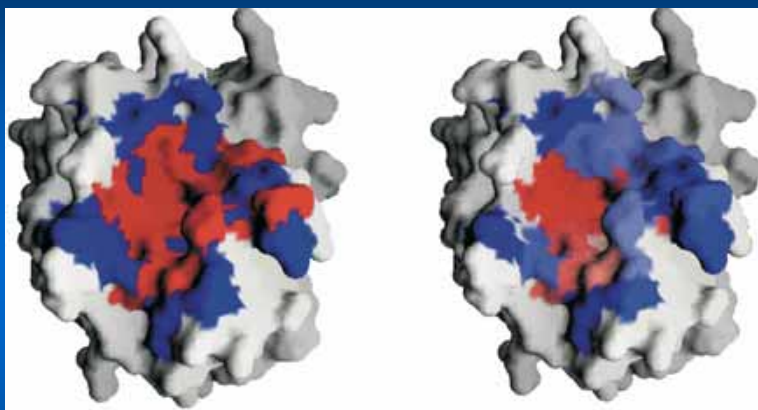
Alanine-scanning experiments

- The residues at the interface of a protein-protein complex are mutated to Ala one by one.
- The dissociation constant K_d of the mutated complex is compared K_d of the wild type.
- The mutation changes the free enthalpy of dissociation by:

$$\Delta G_d = RT \ln K'_d/K_d$$



Hot-Spots



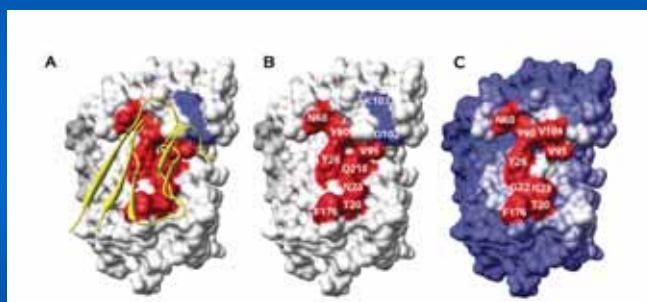
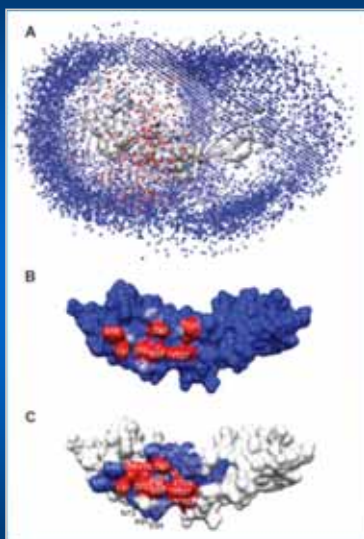
Core and **rim** in the recognition patch of the ovomucoid inhibitor (1cho)

S=0 (red) fully conserved residues.
S>1 (purple) variable residues

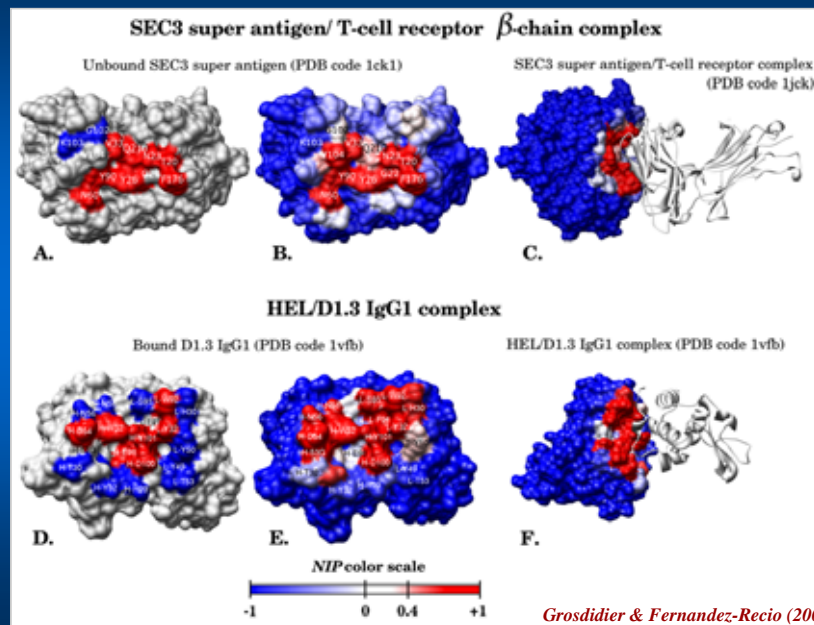
Prediction of Binding Hot-Spots

Method name	Input data	Method	Details	Sensitivity	PPV	availability
ISIS ⁵⁸	sequence	Neural network	Predicted structural features, evolutionary information	15%	89%	http://cubic.bioc.columbia.edu/services/isis/
FOLDEF ¹²⁶	complex structure	Energy-based	Alanine scanning	45-72% ^a	61-73% ^a	http://foldx.org.es/
ROBETTA ¹²²	complex structure	Energy-based	Alanine scanning	28-69% ^b	60-71% ^b	http://rosetta.org/submit.jsp
K-FADE ¹²⁵ / K-CON/ ROBETTA	complex structure	Machine learning algorithm	Physical-biochemical features	48%	53%	http://kfc.mitchell-lab.org
MAPPIS ¹¹⁹	complex structure	Evolutionary conservation	Multiple alignments, 3D clustering	66%	63%	http://bioinfo3d.cs.tau.ac.il/MAPPIS
HotPoint ¹²¹	complex structure	Empirical model	Accessibility, knowledge-based potentials	59%	70%	http://prism.ccbb.ku.edu.tr/hotpoint
pyDockNIP ¹²⁷	unbound protein structure	Energy-based	Docking simulations	42-43%	68-75%	http://mmb.pcb.ub.es/PyDock

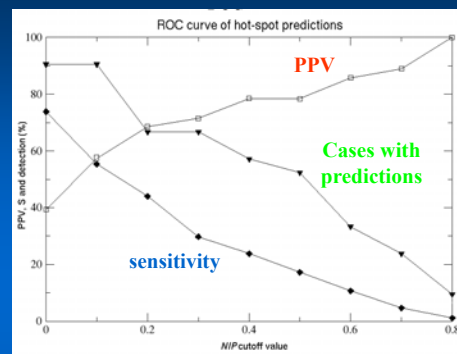
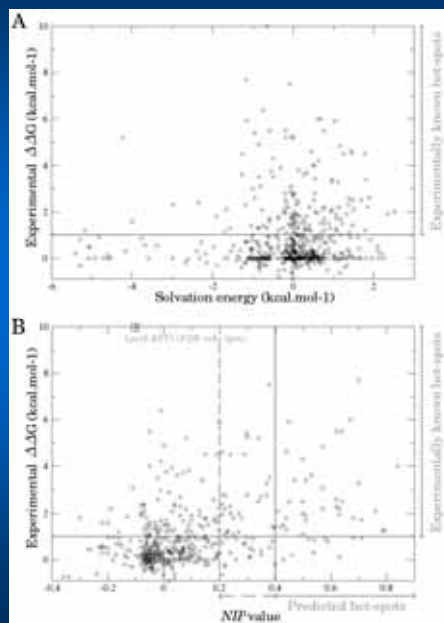
Prediction of Binding Hot-Spots



Prediction of Binding Hot-Spots



Prediction of Binding Hot-Spots



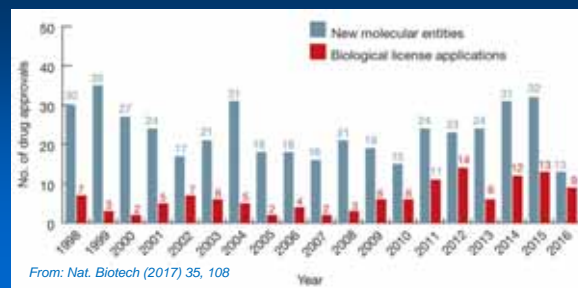
Hot-spot prediction benchmark

	PPV	S
Standard benchmark (21 cases)		
NIP ≥ 0.2	68%	43%
NIP ≥ 0.4	78%	24%
FOLDEF	73%	46%
ROBETTA ^b	71%	69%
Benchmark on modeled subunits (11 cases)		
NIP ≥ 0.2	59%	33%
NIP ≥ 0.4	75%	15%

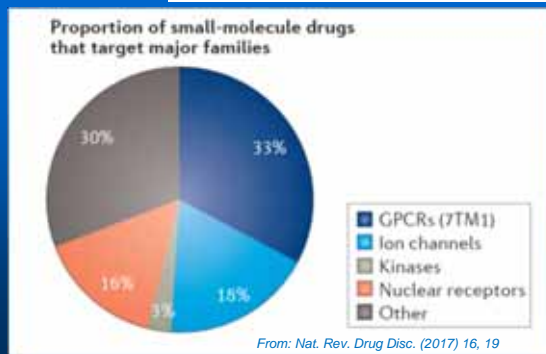
Grosdidier & Fernandez-Recio (2008) BMC Bioinf

- Interface prediction
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- Pathological mutations affecting PPIs

Current limitations in drug discovery



Drug discovery: new targets are strongly needed!



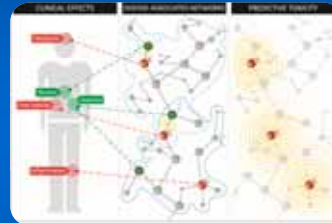
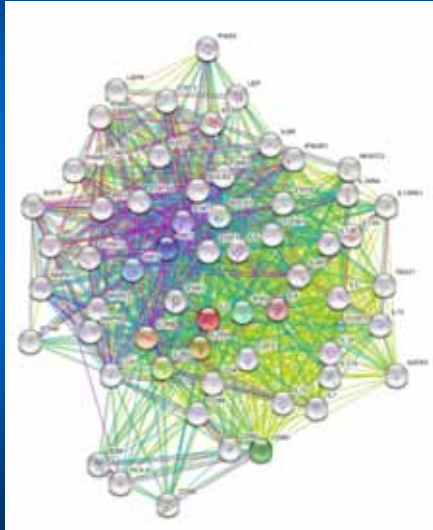
>70% FDA-approved drugs target only 4 gene families, basically cell-surface receptors and enzymes

Most of the available drugs target individual proteins

Only 10–15% of all human proteins are currently «druggable»

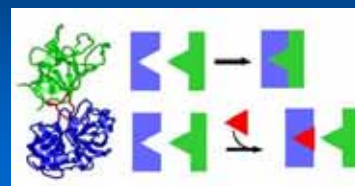
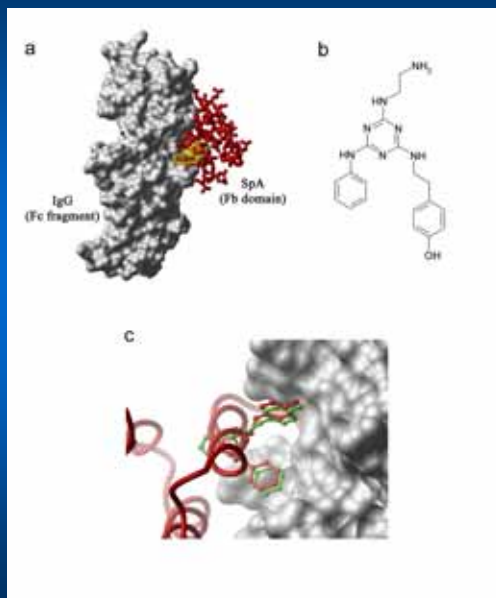
Importance of protein-protein interactions in biomedicine

Drug discovery: new targets are strongly needed!

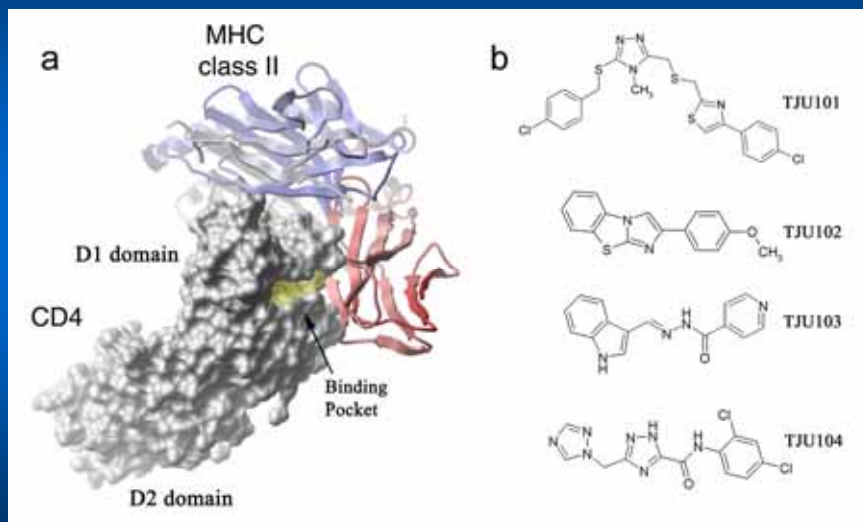


New approaches:
drugs targeting protein-protein
interactions

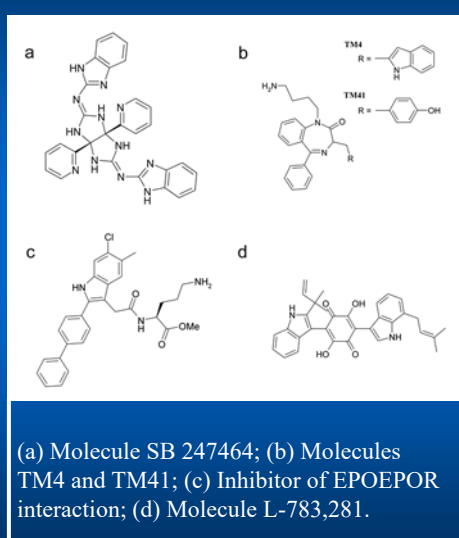
Inhibiting protein-protein interactions



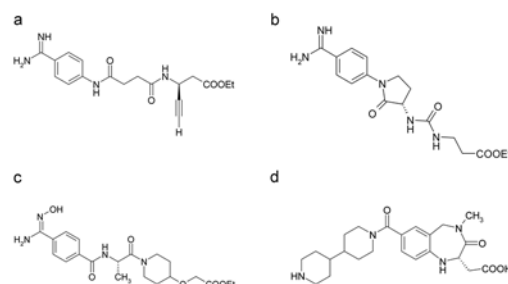
Inhibiting protein-protein interactions



Inhibiting protein-protein interactions



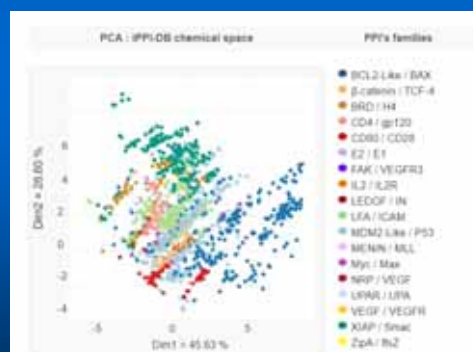
Antagonists of GPIIb-IIIa currently in clinical trials



Inhibiting protein-protein interactions



<http://www.ippidb.cdithem.fr/>

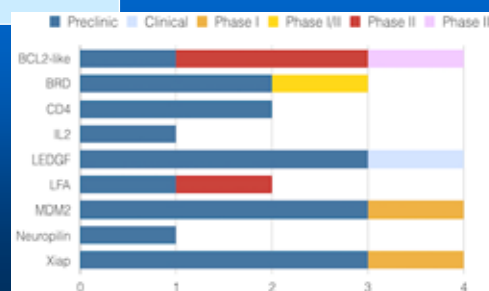


Inhibiting protein-protein interactions

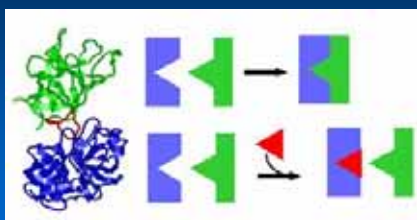
Drug discovery targeting protein interactions

PPI target	Disease	Clinical
BCL-2 family	Cancer	Phase III (disc.)
MDM2-p53	Cancer	Phase III
LFA1-ICAM1	Dry eye	Pre-reg.
α IIb β 3	Cardiovascular	Approved
α L β 2	Psoriasis	Phase II (disc.)
α 4 β 1	Ulcerative colitis	Phase III
α 5 β 1	Macular degeneration	Phase I (disc.)
IAP	Cancer	Phase II
Bromodomain family	Cardiovascular, cancer	Phase III, II

From: Nat. Rev. Drug Disc. (2016) 15, 533



Rational design of protein-protein inhibitors

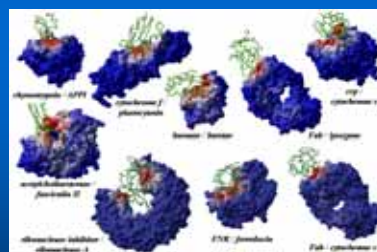
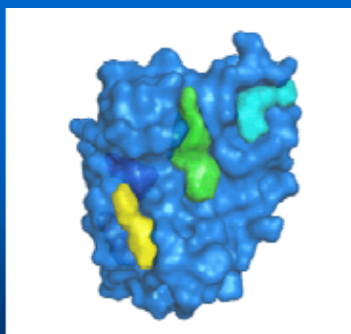


We need a suitable cavity

Rational design of protein-protein inhibitors

Major difficulties:

Where to search?
(In most cases protein-protein interface is not known)

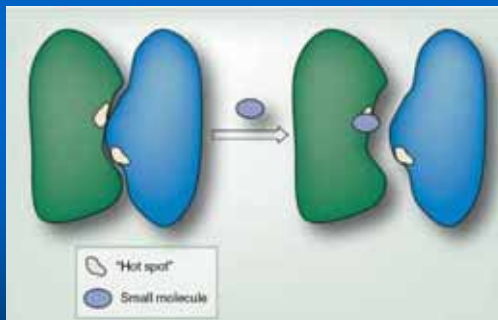


- Protein-protein docking
- Interface predictions

Rational design of protein-protein inhibitors

Major difficulties:

Absence of binding pockets in natural interfaces

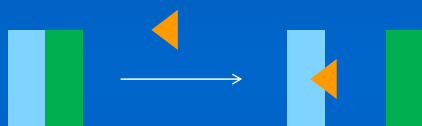


- In standard drug design:
3D structure with bound ligands,
known active site
- In PPI inhibition:
no clear pockets (difficult to
identify computationally)

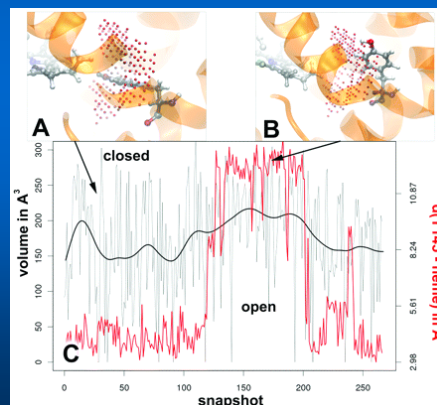
Rational design of protein-protein inhibitors

Major difficulties:

Absence of binding pockets in natural interfaces



Molecular Dynamics
+
Pocket predictors (Fpocket, CASTp,...)

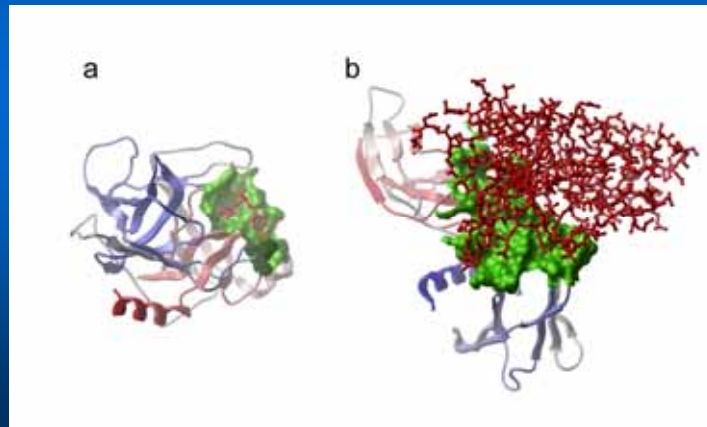


<https://bioinformatictools.wordpress.com/tag/pocket-finder/>

Rational design of protein-protein inhibitors

Major difficulties:

How to compete with a large interface?

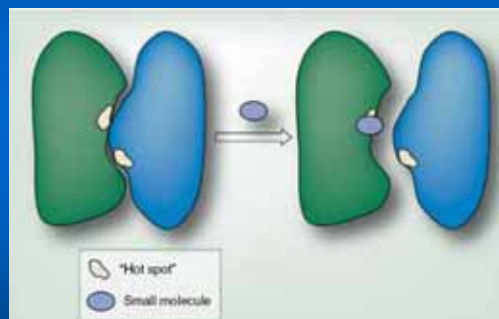


Rational design of protein-protein inhibitors

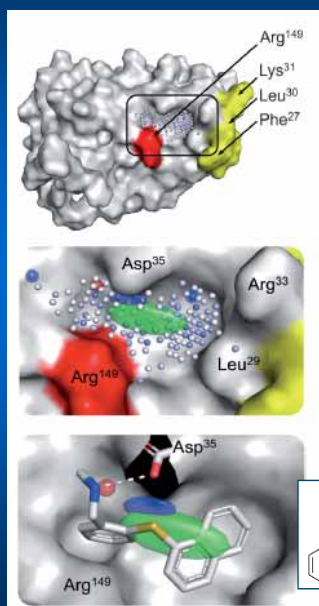
Major difficulties:

How to compete with a large interface?

- Strategy: Targeting hot-spots.
- Hot-spot experimental identification: alanine-scanning (costly)
- Hot-spot computational prediction: ROBETTA, FOLDEF...
(need 3D structure of complex)



Rational design of protein-protein inhibitors

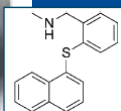


Blocking IFN- α / receptor
Geppert et al. (2012) *Angew.Chem*

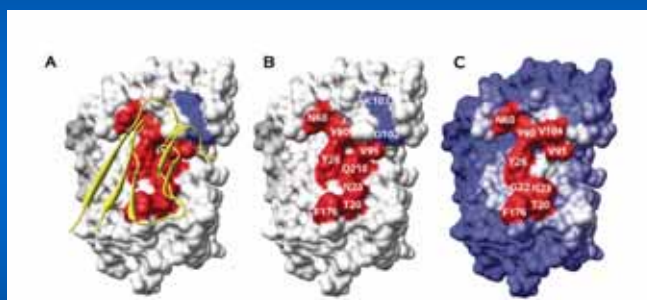
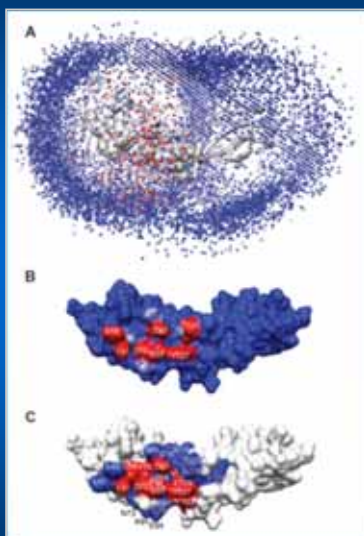
Predicted hot-spots
(iPred)

Pharmacophore
(VirtualLigand)

Docking pose (Gold)



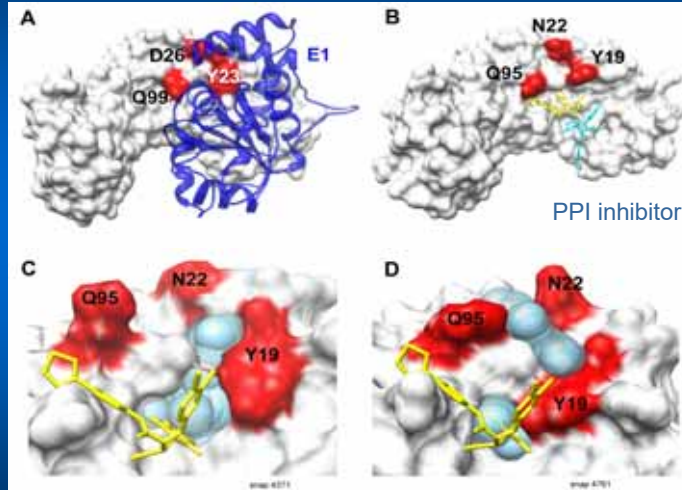
Docking-based prediction of Binding Hot-Spots



NIP (no need complex structure)

Transient cavities in interfaces for ligand docking

HPV E2 protein - predicted hot-pots

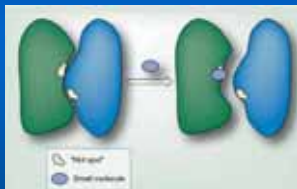


Combine MD, pocket identification and docking-based hot-spot predictions
(no need complex structure)

Rational design of protein-protein inhibitors

Major difficulties:

Are current ligand libraries appropriate?



PPI-HitProfiler

<http://www.cdithem.fr/getPPIHitProfiler.php>



Rational design of protein-protein inhibitors

Major difficulties:

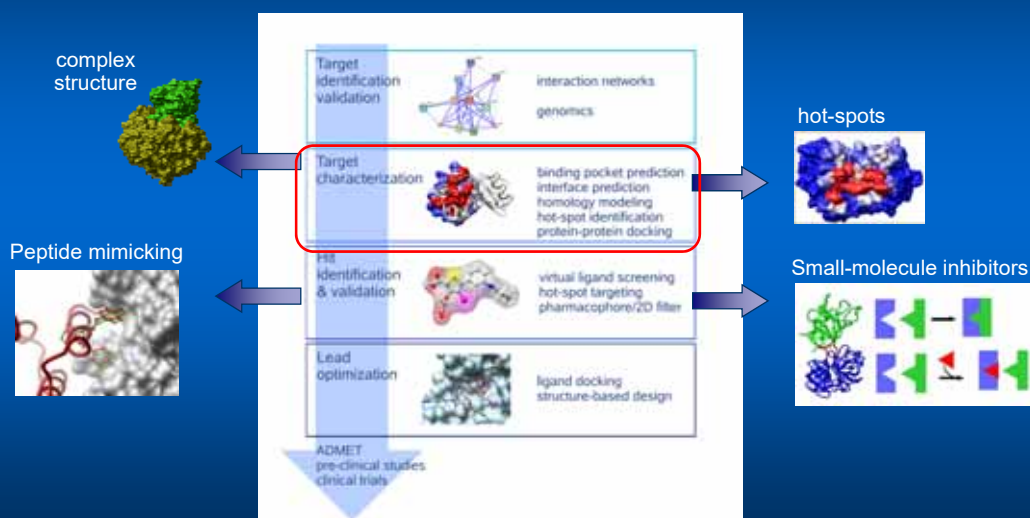


Problem	Possible solution
No natural cavities in PP interfaces	MD to generate transient cavities
Transient cavities difficult to identify (even bound-to-inhibitor cavities difficult)	Specialized pocket prediction tools
Where to search (in most cases, PP interface not known)	PP docking
Find best binding site (to compete with a large interface)	Target hot-spots
Find best PP inhibitor (chemically different from traditional ligands)	Specific libraries for PP inhibitors

Docking applied to drug discovery targeting protein-protein interactions

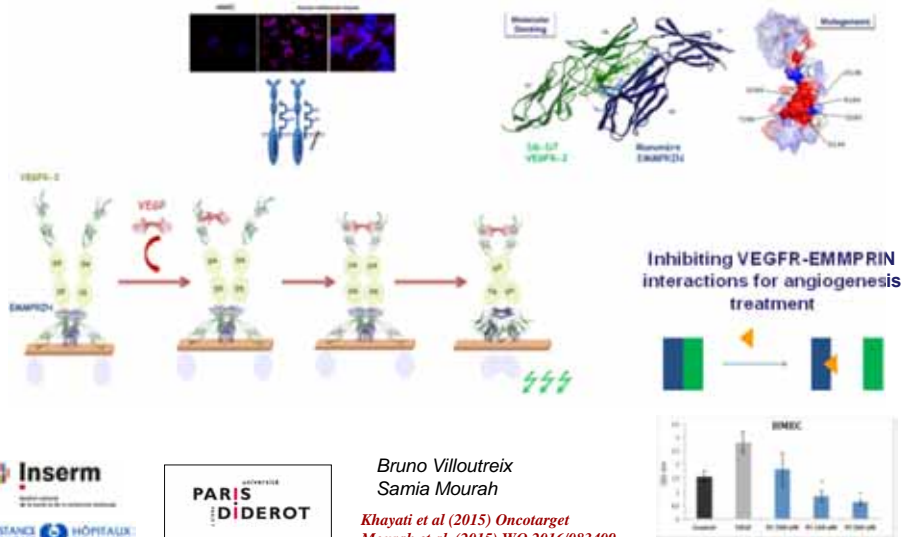
Protein-protein docking

Interface and hot-spot predictions



Drug discovery targeting protein interactions

- VEGF is the most relevant pro-angiogenic factor in cancer
- EMMPRIN is needed for VEGFR activation by VEGF



- Interface prediction
- Hot-spot identification
- Application to drug design targeting PPIs
- Pathological mutations affecting PPIs

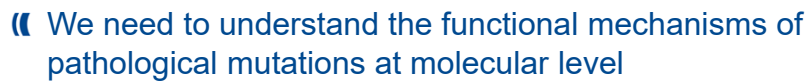
Personalized Medicine



Personalized medicine



ANALYSIS



CLINICAL EFFECTS

DISEASE ASSOCIATED NETWORKS

PREDICTIVE TOXICITY

The figure illustrates the SABiosciences network approach, which integrates clinical effects, disease-associated networks, and predictive toxicity modeling.

Clinical Effects: The top left panel shows a silhouette of a human figure with four highlighted regions: Head/Neck (red), Torso (green), Limbs (blue), and Internal Organs (yellow). These regions are linked to a central network of nodes and edges, representing disease-associated networks.

Disease Associated Networks: The top right panel displays a complex network diagram with numerous nodes (represented by colored circles) and connecting lines, illustrating the intricate relationships between various biological entities.

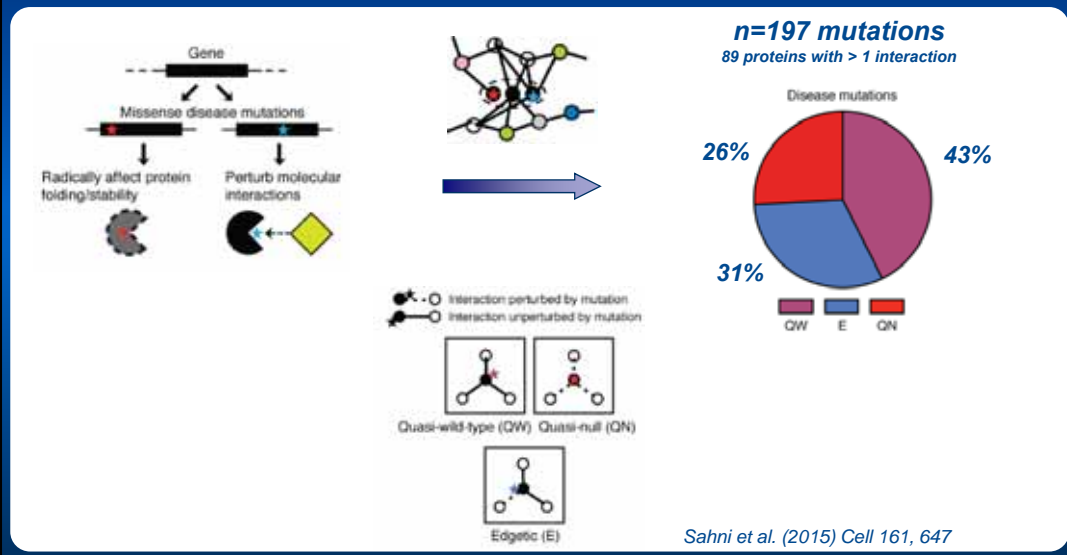
Predictive Toxicity Modeling: The bottom right panel features a large, detailed signaling pathway diagram. It depicts the flow of information from external stimuli (e.g., Epinephrine, Neurotransmitters, Hormones, Stress, Inflammatory Stimuli) through various receptors (GPCRs, RTKs, Ion Channels) and intracellular signaling molecules (G-proteins, Kinases, Phosphatases, etc.) leading to cellular responses such as Cell Survival, Proliferation, and Apoptosis. Key components include Ras, Raf, MEK, ERK, JAK, STAT, PI3K, Akt, mTOR, and others.

Imaging and Cellular Data: The bottom left panel includes three visual representations: a grayscale medical scan (likely MRI or CT), a colorful brain heatmap (showing regional activity), and a microscopic image of cells stained with purple dye.

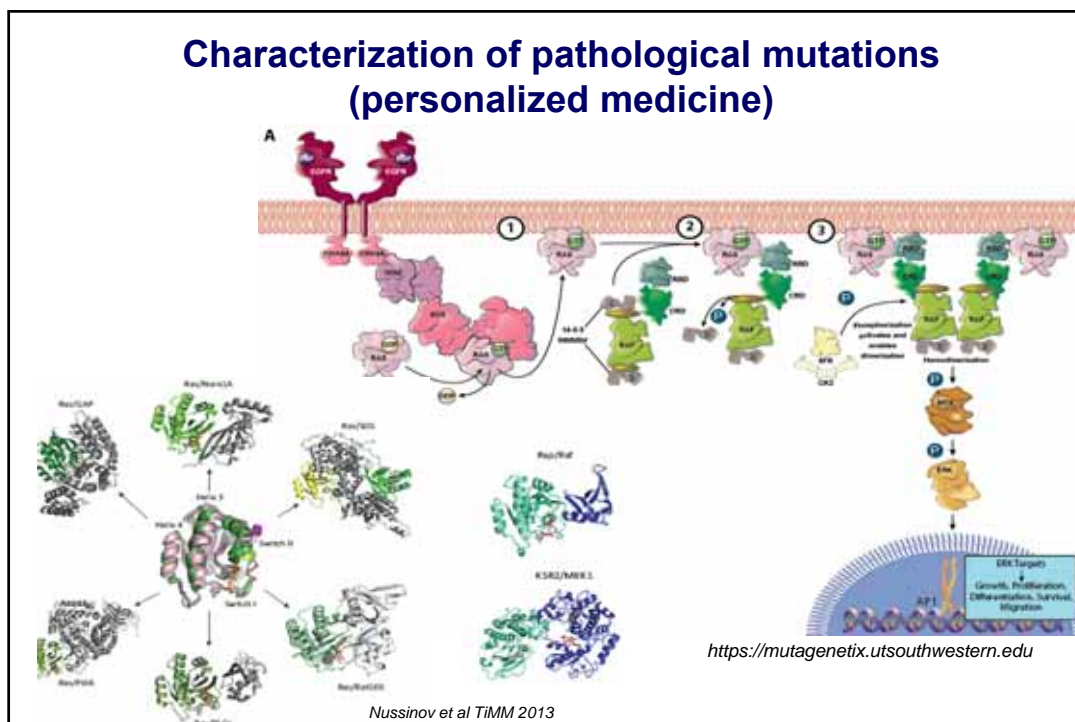
SABiosciences Logo: The logo is located at the bottom center, featuring a stylized 'A' followed by the text "SABiosciences" and "A CIAGEN Company".

Applications in Biomedicine: Prediction and interpretation of pathological mutations

Effect of pathological mutations on PPIs and networks

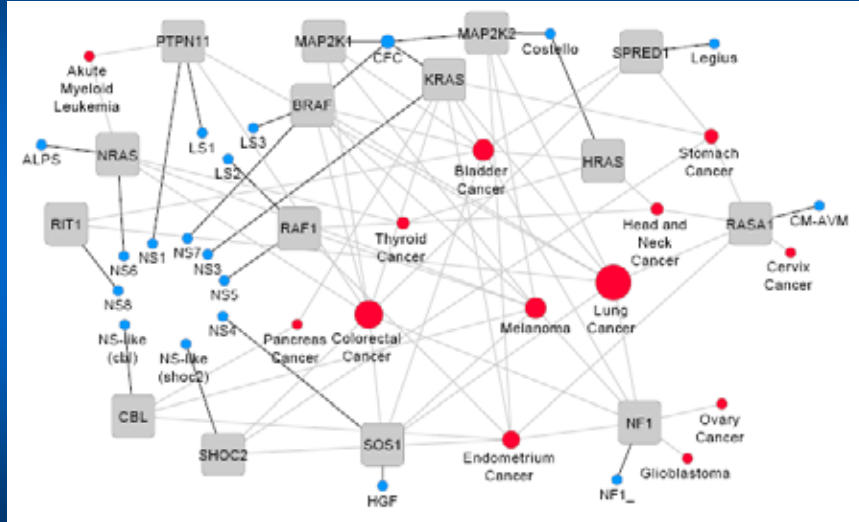


Characterization of pathological mutations (personalized medicine)



Characterization of pathological mutations

Mutations in MAPK/ERK pathway cause cancer or RASopathies



Kiel & Serrano (2014) Mol.Sys.Biol.

Characterization of pathological mutations

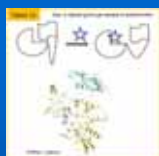
Folding/Stability



956 mutations

Germline mutations: RASopathies
Somatic mutations: cancer

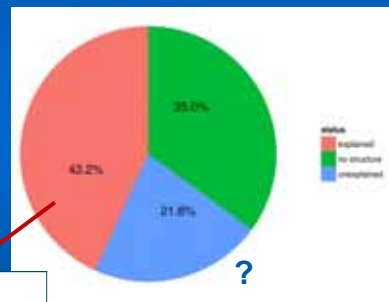
Activity / Function



PPI



Localization



We need more structural and dynamic models for PPIs

Kiel & Serrano (2014) Mol.Sys.Biol.

Pathological mutations affecting protein interactions

Disease nsSNPs in Protein Complexes
David et al. 2012 *Human Mut.* 33, 359

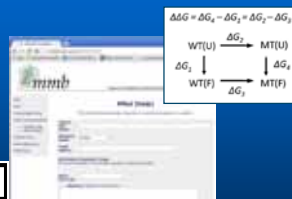
	Total residues	Observed	Expected	O/E percent ratio
Disease nsSNPs				
Core	42,342	781	516.1	1.5
Interface	43,381	620	528.8	1.2
Surface noninterface	112,805	1019	1375.1	0.7
Total	198,528	2420		

Affecting Protein Stability



FoldX destabilization

Kiel & Serrano 2014
Mol. Sys. Biol.



PMut
Ferrer-Costa et al. 2012 *JMB*

Pathological mutations affecting protein interactions

Disease nsSNPs in Protein Complexes
David et al. 2012 *Human Mut.* 33, 359

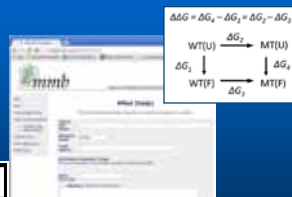
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Affecting Protein Stability



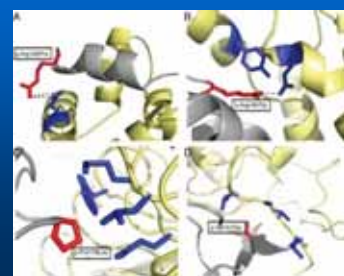
FoldX destabilization

Kiel & Serrano 2014
Mol. Sys. Biol.



PMut
Ferrer-Costa et al. 2012 *JMB*

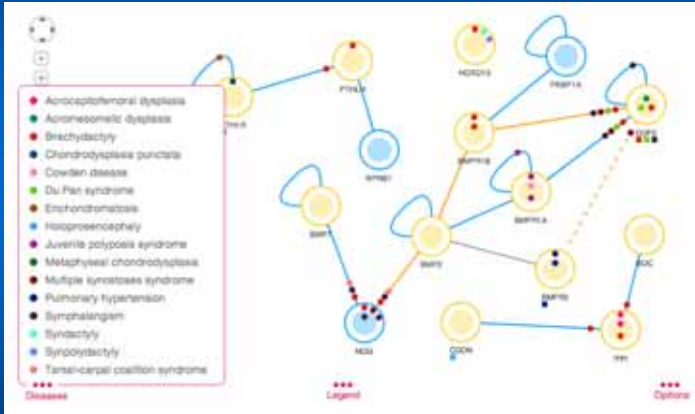
Affecting Binding Affinity



Pathological mutations affecting protein interactions



<http://dsysmap.irbbarcelona.org/>



Mosca et al. 2015 *Nature Methods* 12, 167

24K disease mutations
9.5K mapped on PPI structures
2.3K mapped on interfaces

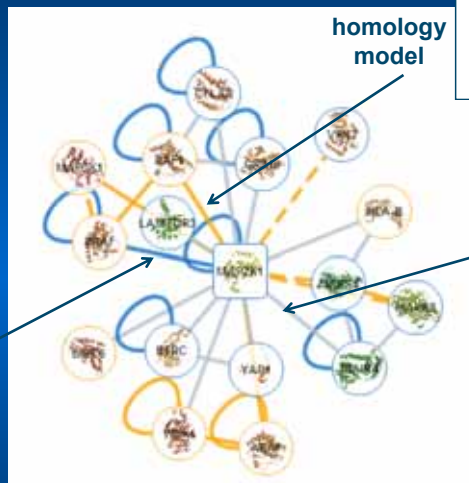
In 2012:
23K disease mutations
2.4K mapped on PPI structures
0.6K mapped on interfaces

Characterization of pathological mutations in PPIs

MEK1
structural
interaction network



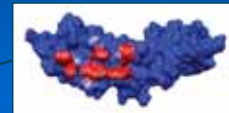
x-ray



homology
model



docking
binding site predictions



Applications in Biomedicine: Prediction and interpretation of pathological mutations

Pathological mutations in PPIs, personalized diagnosis

