

# STRUCTURAL BIOINFORMATICS REPORT

## Introduction

In this study, we are interested in a family of proteins belonging to the Major Facilitator Superfamily (MFS)[1]. This Family is the largest family of secondary carriers.

More specifically, we are interested in the Mammalian Sugar Transporters. This study case we concentrate on the protein named Solute carrier family 2, facilitated glucose transporter member 4 (GLUT4). This protein (UniProt id : P14672) is coded by a gene named SLC2A4 from Homo sapiens (Human)[2].

This gene is part of the GLUT family protein which is composed of 14 isoforms [3]. The GLUT protein family is classified into three different families that can be distinguished based on their protein sequence homologies (Fig 1).

Class	1	2	3
GLUT	1,2,3,4,1 4	5,7,9,1 1	6,8,10,12,1 3

FIG 1. Repartition of the different GLUT protein into classes. a class groups proteins with strong homology

The protein GLUT 4 is part of the first family and it's an Insulin-regulated facilitative glucose transporter.

This protein is intensively studied among the glucose transporters because it has an important physiological role. It regulates the glucose uptake of skeletal and cardiac muscle, brown and white adipose tissue and is regulated by the insulin level . Moreover, impaired GLUT4 translocation is bind to insulin resistance[4]. Indeed, default of the insulin-mediated translocation of GLUT4 to the plasma membrane is known to be a peripheral insulin resistance.

Because most of the blood glucose is absorbed by skeletal muscle in the presence of high insulin and the GLUT4 transport activity is limiting for this process, GLUT4 plays a vital role in the regulation of body glucose homeostasis.

The 3D structure of GLUT4 has yet to be resolved but some homological structure has already been resolved like GLUT1 (Uniprot id:P11166) and GLUT3 (Uniprot id : P11169).

Plus GLUT1, GLUT3, GLUT4 have a really similar sequence. So we proposed to use an homology approach to try to model the GLUT4 structure and if possible find two different conformations of the receptor. After the modeling, we are going to use normal mode analysis to find the transition between the open and closed conformation. Because a receptor have different structure according to its conformation[5].

## Materials and methods

### METHODS

The whole process to obtain an accurate model need a lot of online or desktop software (Fig 2). Mostly it is more convenient to use online tools, it allows to speed up the process of modeling except for the main task which is the modeling itself that requires a lot of computing power.

steps	Software or server used
<b>Structure and transmembrane prediction</b>	T-Coffee[6], PsiPred[7], Blastp[8]
<b>Modeling</b>	Swiss-Model[9], Memoir[10], Medeller[11]
<b>refining</b>	Galaxyweb refine[12], sidepro2[13]
<b>Quality check</b>	ProQ[14], ProQM, Verify 3D, Oremp[15], Qmean[16]
<b>After modeling</b>	Bio3D(NMA)[17], Swiss-dock (protein ligand docking)[18]

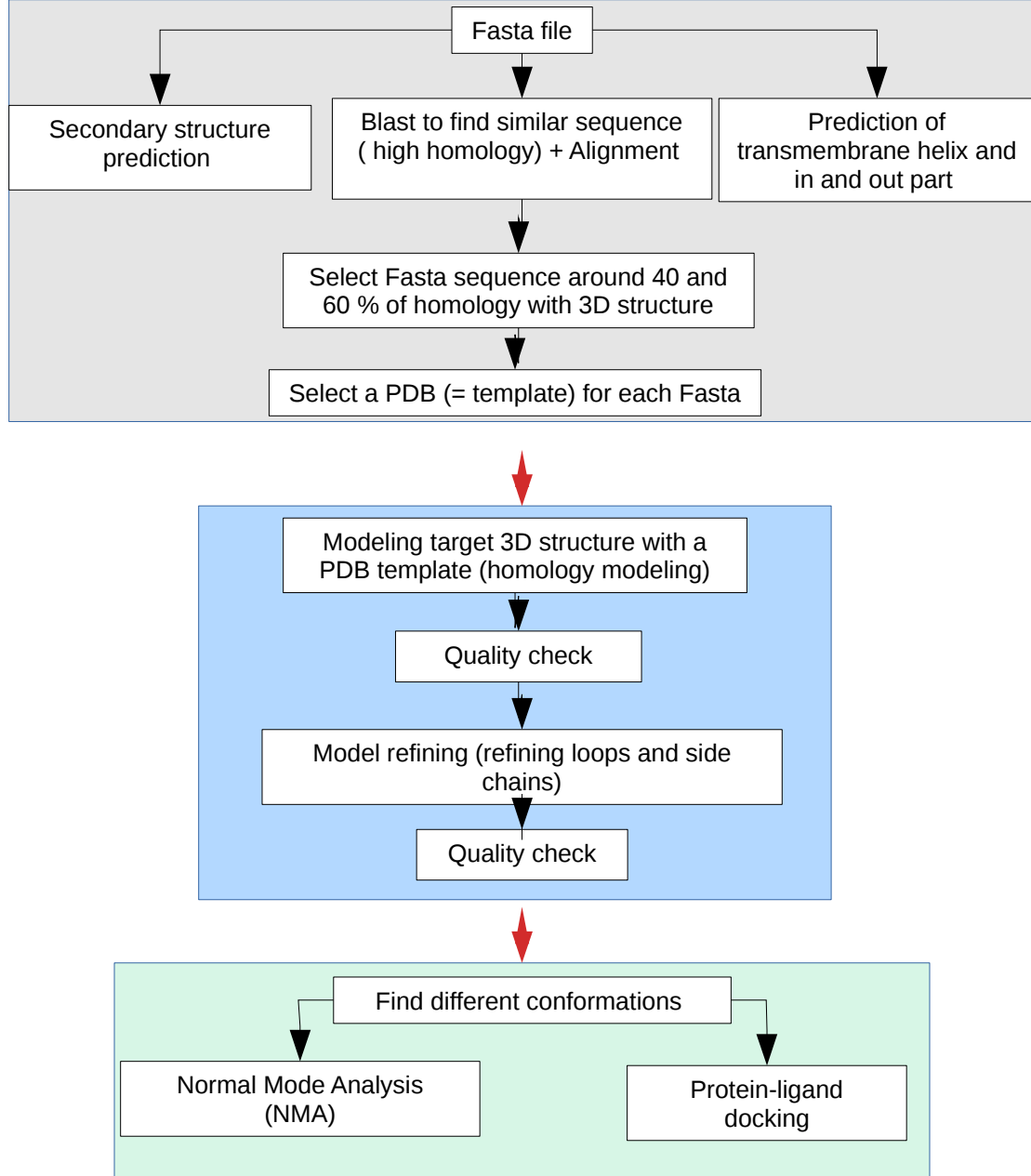
*Fig 2. Different online or desktop software use during this study*

This study has required the use of Fasta files to select sequences that have a similarity between 40% and 60%. After this selection, we use one PDB for each Fasta. This PDB is going to be the template used in order to model the secondary and tertiary structure of our reference Fasta. At the same time, we predict the secondary structure and the putative position of the transmembrane helix.

Each selected PDB is used as a template that is going to provide the secondary and the tertiary structure. Then the modeling can begin, using mostly Memoir. This software is going to provide a full modeling pipeline, using Medeller for the coordinate generation and Completionist and Galaxyweb refine for loop modeling. After each step, a quality check is performed with tools detailed in figure 2. The most Useful has been Oremp and Verify

3D which seems to be more compatible with our protein which is a transmembrane protein.

Then we select different conformation like open conformation and closed conformation. After these steps, we use the Bio3D tools to analyse the normal modes and Swiss-Dock to find a potential binding site for potential ligands like D-glucose.



*Figure 3. Workflow of steps for the protein modeling.*

*The steps before the modeling are in the grey rectangle the modeling is represented by the blue rectangle and the analysis step are on green*

## Results

### Details of the fasta sequence to model

The details about the Fasta sequence are in Fig 5. But it is important to insist on the fact that it is from the GLUT family

Features	Data
Uniprot id	P14672
name	Solute carrier family 2, facilitated glucose transporter member 4 (GLUT4)
Gene id	SLC2A4
function	Insulin-regulated facilitative glucose transporter.
Localization	Cellular membrane
length	519

Figure 4. Details of the sequence selected to be 3D models.

## Secondary structure prediction

The Psipred analysis predcit 21 helix and some strand. Instead the Uniprot web site predict 12 transmembrane helix. T-coffe predict 12 helix and it's position in the membrane (Annexe 1).

Finally, It is important to find the domain related to the molecular function of P14672. Especially the domain related to the sugar and glucose transporter. On Uniprot the position 333 (N) and 404 (W) are highlighted as binding site for Monosaccharide. The figure 6 precise these position and the other great domain like th MFS positions on P14672.

## Detailed signature matches

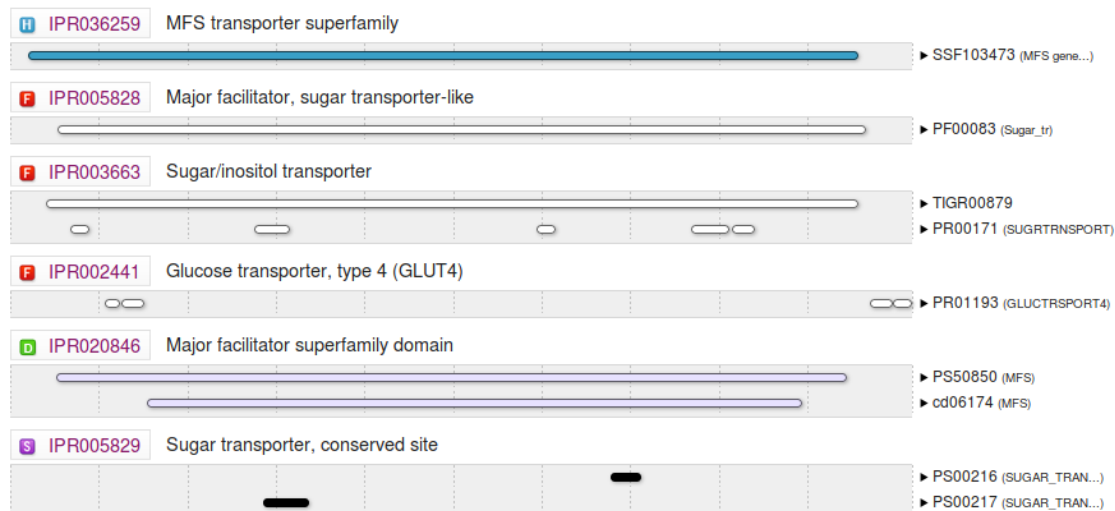


Figure 5. Principal domain found for the GLUT4

## Selecting the Fasta sequence and alignment

According to the homology method, I choose 4 Fasta that have a high homology with P14672. They also have a similar sequence, a similar origin, and function. The first three sequence share the same group and the next two are from a different group. It is going to be interesting to see if the modeling is going to be more difficult for the last two.

The figure 6 details the choose sequences. In annexe 2, the align provided by the T-Coffee server is quite good ( annexe 2).

It show that there is a gap at the beginning between position 3 to 13. But after that one the sequence fits well. All the selected sequence provide a very large cover as seen in the figure 6.

Pdb number	Gene	function	% of Identity	3D structure	organism
P14672	SLC2A4	Insulin-regulated facilitative glucose transporter.	100,00 %	0	Homo sapiens
P11166	SLC2A1	Facilitative glucose transporter	65,9	4	Homo sapiens
P11169	SLC2A3	Facilitative glucose transporter	60,9	4	<i>Homo sapiens</i>
P43427	SLC2A5	fructose transporter	43.4	1	<i>Rattus norvegicus (Rat)</i>
P58353	SLC2A5	fructose transporter	41.8	1	<i>Bos taurus (Bovine)</i>

Figure 6. Fasta sequence with 3D sequence choose

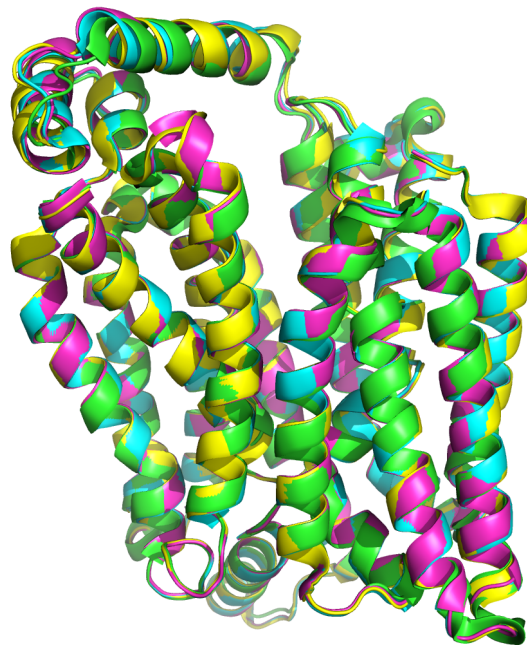
Entry	Alignment overview	Info	3D
4 result(s) selected. (Clear selection)			
<input checked="" type="checkbox"/>	Query: sp P14672 GTR4_HUMAN B201711268A530B6CA0138AFAA6D2B97CE8C2A924FB22FED		
<input checked="" type="checkbox"/>	P11166 GTR1_HUMAN - Solute carrier family 2, facilitate... Homo sapiens (Human) - View alignment	E-value: 0.0 Score: 1,653 Ident.: 65.5%	Model (1) X-ray crystallography (4)
<input checked="" type="checkbox"/>	P11169 GTR3_HUMAN - Solute carrier family 2, facilitate... Homo sapiens (Human) - View alignment	E-value: 0.0 Score: 1,440 Ident.: 60.9%	X-ray crystallography (4)
<input type="checkbox"/>	P22732 GTR5_HUMAN - Solute carrier family 2, facilitate... Homo sapiens (Human) - View alignment	E-value: 2.7e-128 Score: 995 Ident.: 42.7%	Model (1)
<input checked="" type="checkbox"/>	P43427 GTR5_RAT - Solute carrier family 2, facilitate... - Rattus norvegicu... - View alignment	E-value: 1.1e-127 Score: 991 Ident.: 43.4%	X-ray crystallography (1)
<input type="checkbox"/>	Q6PXP3 GTR7_HUMAN - Solute carrier family 2, facilitate... Homo sapiens (Human) - View alignment	E-value: 3.3e-126 Score: 982 Ident.: 40.6%	Model (1)
<input checked="" type="checkbox"/>	P58353 GTR5_BOVIN - Solute carrier family 2, facilitate... Bos taurus (Bovine) - View alignment	E-value: 1.3e-122 Score: 957 Ident.: 41.8%	X-ray crystallography (1)

Figure 7. Covering provided by the Fasta sequence on the original sequence

## Selecting the PDB template

For each Fasta sequence, a PDB has been chosen. I took four PDB, two PDB have an inward-open conformation and the other two have an outward-closed conformation. Each template was carefully selected. I align all the PDB and compute the RMSD in order to see if there is a big difference (annex 3). I only align the PDB that have the same conformation.

For P11166, all the conformation are inward open, with pymol the 3D structure are all very close (Fig 8) and the average RMSD is around 0.3 Å (annex 3) which show that all the 3D structure are really similar.



*Figure 8. All the PDB of P11166 align with pymol to show their similarity*

The same work has been made with the three PDB of P11169 and shows similar results. All the 3D structure are well aligned and the average RMSD is around 0.3 Å.

For both P58353 and P43427, only one structure is available.

So we can conclude that only a PDB for each Fasta is necessary. Figure 9 regroups chosen PDB a provide more information about each structure

<b>PDB entry</b>	<b>Method</b>	<b>Resolution (Å)</b>	<b>Chain</b>	<b>Positions</b>	<b>conformation</b>
<a href="#">4PYP</a>	X-ray	3.17	A	<a href="#">1-492</a>	inward-open
<a href="#">4ZW9</a>	X-ray	1.50	A	<a href="#">1-496</a>	outward-occluded
<a href="#">4YB9</a>	X-ray	3.20	D	<a href="#">1-473</a>	open inward-facing
<a href="#">4YBQ</a>	X-ray	3.27	A/B	<a href="#">1-502</a>	outward-open

*Figure 9. The four PDB chosen to be the template in order to model GLUT4's 3D structure*

## Modeling the PDB and quality check

As precise in the materials and methods, a huge part of all models have been realized with Memoir, a full membrane protein modeling pipeline. To compare the result I have built another model with Swiss-Model but the best model was built with the default option of memoir. Also memoir provide a tool that refine the model quality but to be sure of the best quality

Model	Template from	helix number	Predicted LGscore	Predicted MaxSub	PRO QM	Qmean score	Verify 3D( %)	Valid Model
1	4pyp	12	6.022	0.405	0.775	-6.67	85.46	OK
2	4zw9	10	6.128	0.377	0.785	-3.26 -2.22	90.21	NON
3	4yby	15	4.469	0.277	0.762	-5.40	73.00	NON
4	4yb9	19	5.085	0.333	0.774	-5.40	63.72	NON

*Figure 10. Details of each model and the quality score obtain*

To check the validity of the models after the modeling and before refining I use a lot of tools to validate the quality. Some of them were supposed to be more suitable for membrane protein like PROQM and verify 3D.

The ProQM score was the most useless because the difference between the value is too little to be able to find the real good conformation. The Qmean score was also not very adapted.

Instead the Lgscore, MAXsub and verify 3D score were the best and the most suitable to determine the best model.

So we can see that just with the Lgscore, the MaxSub and the verify3D score the last two model are not valid

After checking the number of the helix with Pymol and with Orempo I realize that only the first model was really a valid one.

So after that step, I use Galaxy-Web refine then SidePro2 to refine the model and the side chain. Galaxy-web gave me five model different models. So I use the two best tools to check the quality.

But the five models' quality was below the quality of the raw model obtain with memoir. Indeed, the models obtained are below 2 points than my previous model and the verify 3D score is also below (Figure 11).

Model	1	2	3	4	5
LGscore	4.247	4.114	4.430	4.425	4.378
MaxSub	0.217	0.226	0.244	0.264	0.240
verify 3D	70	68	76	82.10	75



*Figure 11. Quality score obtain after the refining step*

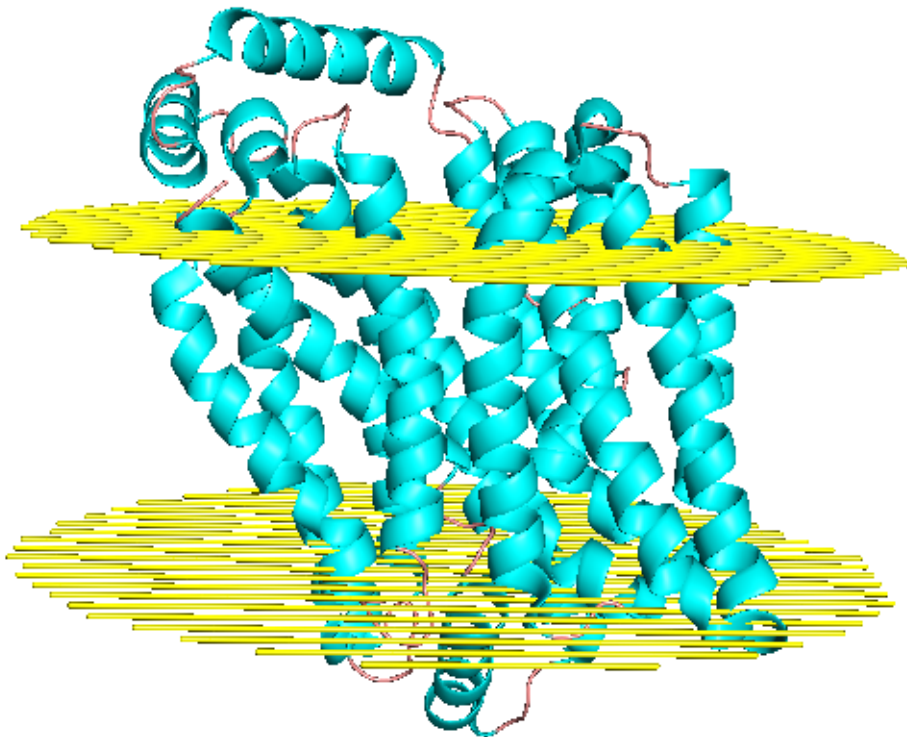
## **The best Model**

At the beginning, I had planned to built two model one with a inward open conformation and another one with an closed outward conformation. The first model is supposed to be in inward open conformation.

If the second model had been valid, I would also had a model with a outward closed which should have been useful to use with the normal modes analysis and to perform some molecular dynamic.

The best model has an Hydrophobic thickness of 29.1 Å and 12 helix as predicted by Uniprot and T-coffee.

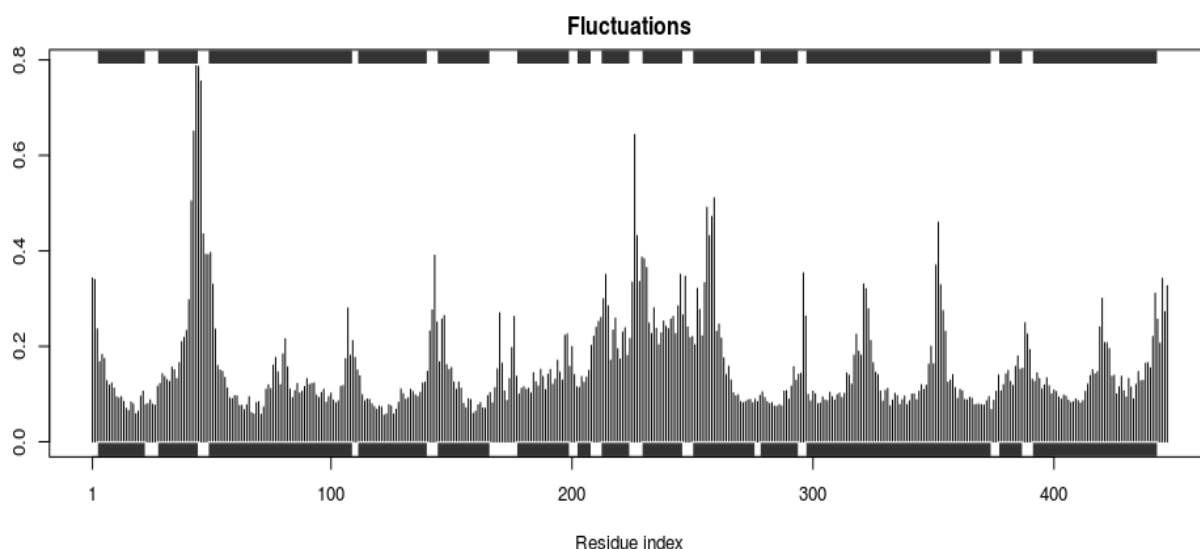
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*Figure 13. The best 3D model for GLUT4. the membrane is in yellow, the helix in blue and the loop in red*

## Normal Mode Analysis

This analysis shows that the parts that move the most are in the peripheric regions and most of the time the moving structure are loops (Fig 14). But some helix in the membrane has a translation move. The size of the barrel is changing in the same time as the entire structure move.



*Figure14. Fluctuation 's analysis of the range of the secondary structure. Helix are represented in Black and loop in white.*

## Conclusion

In this study we manage to find one of the putative 3D structure of the GLUT4 family. The next step would be to realize a molecular dynamic to confirm the move of the whole protein and its stability. After these steps a docking can be the best way to confirm that the protein really binds it's ligand.

# **ANNEX**

## 12

The multiple sequence alignment result as produced by T-coffee.

IN HEL OUT

sp	P14672	GTR4	EYLGPDEN---D
sp	P11166	GTR1	HPLGADSO---V
sp	P11169	GTR3	NSIEPAKETTTNV
sp	P43427	GTR5	NDLPATRE---Q
sp	P58353	GTR5	KEFPSTAR--Q

## 13

## Annexe 3

Fasta origin	PDB entry	Method	Resolution (Å)	Chain	Positions	conformation	RMSD (ref 5EQG)
P11166	<a href="#">4PYP</a>	X-ray	3.17	A	<a href="#">1-492</a>	inward-open	0.38
P11166	<a href="#">5EQG</a>	X-ray	2.90	A	<a href="#">1-492</a>	inward-open	0
P11166	<a href="#">5EQH</a>	X-ray	2.99	A	<a href="#">1-492</a>	inward-open	0.22
P11166	<a href="#">5EQI</a>	X-ray	3.00	A	<a href="#">1-492</a>	inward-open	0.323

Fasta origin	PDB entry	Method	Resolution (Å)	Chain	Positions	conformation	RMSD (ref 5EQG)
P11169	<a href="#">4ZW9</a>	X-ray	1.50	A	<a href="#">1-496</a>	outward-occluded	0.261
P11169	<a href="#">4ZWB</a>	X-ray	2.40	A	<a href="#">1-496</a>	outward-occluded	0
P11169	<a href="#">5C65</a>	X-ray	2.65	A/B	<a href="#">1-474</a>	outward-occluded	0.503

Fasta origin	PDB entry	Method	Resolution (Å)	Chain	Positions	conformation
P58353	<a href="#">4YB9</a>	X-ray	3.20	D	<a href="#">1-473</a>	Open inward-facing
P43427	<a href="#">4YBQ</a>	X-ray	3.27	A/B	<a href="#">1-502</a>	outward-open

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