Contents

Ch	napter 1 - General Introduction:]
	Taxonomic complexity in plants:	
	The role of hybridisation in taxonomic complexity:	-
	The role of polyploidy in taxonomic complexity:	•
	Novel features of TCGs; parasitism and plasticity:	ļ
	The genus Euphrasia:	,
	The British flora:	1:
	New methods for investigating taxonomic complexity:	1:
	Thesis aims:	1:

Chapter 1 - General Introduction:

Taxonomic complexity in plants:

Much of biology is built upon the idea that the diversity we see in nature can be categorised into discrete units, which at its heart is the concept of a species (Simpson 1951). As many species tend to hybridise, this precludes a strict biological species concept being applicable – where species are recognised as a community of interfertile individuals (Mayr 1963). Current species concepts tend to reflect underlying evolutionary processes, that can often be clearly represented in a bifurcating phylogenetic tree. There is still no single, universal species concept however, and each one requires some subjective judgement (Coyne and Orr 2004). If we use a relaxed biological species concept, how much gene flow do we tolerate? If we use a phylogenetic species concept, which collection of genes do we use to define a species?

Ever since this task of discriminating and categorising species has been embarked upon, difficulties have emerged as to how to delimit certain species, mainly because the processes that give rise to species are in constant motion (Luo et al. 2018). Arguably some of the most difficult groups to classify are collectively placed under the umbrella term of 'taxonomically complex groups' (TCGs; Ennos, French, and Hollingsworth (2005)). A TCG is here defined as one where it is difficult to categorise the biodiversity present due to underlying processes which blur species boundaries. TCGs are problematic, not only in terms of classification but also as to how to best conserve them (Ennos et al. 2012; Federici et al. 2013). TCGs are present across the tree of life, from fungi (Leavitt et al. 2011), to fish (Garcia-Melo et al. 2019), and arthropods (Stenberg et al. 2003). TCGs are remarkably frequent in plants however, and particularly common in certain families such as Poaceae (Roodt and Spies 2003), Rosaceae (Dickinson, Lo, and Talent 2007), and Asteraceae (Czapik 1996). TCGs can be the result of plant populations that no longer undergo sexual, random mating (Hollingsworth et al. 2006), but other TCGs are the result of phenotypic plasticity and recent speciation blurring species boundaries (Belton et al. 2014; Wang et al. 2018).

Factors commonly contributing to TCGs include selfing, apomixis (or agamospermy), hybridisation, and polyploidy. Selfing and apomixis disturb random mating and restrict gene flow between populations, and often interact with hybridisation and polyploidy (Hollingsworth et al. 2006). For example, apomixis coupled with rare hybridisation can produce arrays of microspecies, characteristically seen in genera such as *Rubus* (330+ microspecies in Britain), *Hieracium* (400+) and *Taraxacum* (240+; Stace (2019)). Mating system shifts from outcrossing to selfing or apomixis, drive rapid reproductive isolation from progenitor lineages through founder effects, genetic drift, and selection of advantageous recessive alleles (Hollingsworth et al. 2006). This leads to strong population structure, characterised by many and varied subtle morphological changes between populations. Similar phenomena result from ploidy level variation (Raggi et al. 2015; Spaniel et al. 2011), however hybridisation can assist in moving genetic material between populations, which can produce yet more

morphological variation (Alix et al. 2017). Species concepts usually break down in TCGs, as the evolutionary processes creating new variation overwhelm any stable pattern of species, and the relationships between species become subtle, finely divided, and overlap.

The role of hybridisation in taxonomic complexity:

Hybridisation, defined here as the mating between different species, is an important factor driving taxonomic complexity (Ennos, French, and Hollingsworth 2005; Campbell and Wright 1996). The role of hybridisation is not simple however, as many different outcomes are possible. Hybridisation can be destructive, where rare species may lose their genetic integrity, resulting in populations of entirely hybrid genotypes, and can eventually lead to the extinction of the rarer species (Rhymer and Simberloff 1996; Brochmann 1984). On the other hand hybridisation can be creative, by allowing adaptive traits to move between species (Chapman and Abbott 2010). Hybridisation can also lead to introgression, where there is the incorporation of genetic material from one species in the genetic background of another (Twyford and Ennos 2012). In the extreme, new species can be formed in a process known as hybrid speciation (Mallet 2007). There are two main pathways to speciation involving hybridisation: polyploid hybrid speciation where the hybrid species has duplicated its chromosome complement (allopolyploidy; Rieseberg and Willis (2007)), and homoploid hybrid speciation where the parental species and the hybrid remain at the same ploidy level (Rieseberg 1997). Many TCGs involve hybridisation which blurs species boundaries, coupled with processes that may allow the hybrid derivatives to persist (Ennos, French, and Hollingsworth 2005). Three categories in which TCGs can be placed include agamic complexes, in which hybridisation is combined with a mode of asexual reproduction (e.g. apomixis) to propagate lineages (Hersh, Grimm, and Whitton 2016), polyploid complexes, where the hybrid derivatives are sexual polyploids (Zohary and Nur 1959), and homogamic complexes (or homoploid complexes, e.g. Helianthus Rieseberg, Vanfossen, and Desrochers (1995)), where hybrid derivatives are mainly diploid and isolated from parental progenitors ecologically.

Some TCGs defy these three broad categories, by combining properties of different species complexes. A good example of this is the genus Sorbus in the British Isles, where there are 45 taxa, plus seven more which have been introduced (Pellicer et al. 2012). In Sorbus, hybridisation, polyploidy, and apomixis have interacted to form numerous endemic species in England, Scotland, and Wales (Ludwig et al. 2013). In England alone, this process happens because there are few sexual diploid species (S. torminalis, S. aria, and S. aucuparia, but not S. domestica) which hybridise with apomictic polyploid derivatives of the species S. aria (at least 20 taxa; (Pellicer et al. 2012; Robertson et al. 2010)). Apomictic Sorbus species require pollen to achieve successful asexual reproduction, however at low frequencies the pollen can fertilise the maternal embryo and this leads to a stable, new, polyploid, apomictic taxon (Ludwig et al. 2013). In all, more than 31 apomictic species have arisen in this way (C. A. Stace, Preston, and Pearman 2015). The apomictic condition ensures that any new hybrid genotype is frozen in stasis, and can lead to complex reticulate evolutionary histories, which is seen in many other apomictic plant systems (Wittzell 1999; Sochor et al. 2015). Not only is hybridisation important in generating biological diversity and complexity, it is also a common phenomenon both geographically, and phylogenetically (Ellstrand, Whitkus, and Rieseberg 1996). The clear and widespread abundance of hybridisation means it has had, and continues to have, a profound effect on the evolution of plants – especially in conjunction with selfing, apomixis, and polyploidy.

The role of polyploidy in taxonomic complexity:

Polyploidy is the condition where a cell contains more than two sets of chromosomes as a result of whole genome duplication (WGD), and is featured in almost all TCGs in plants (Ennos, French, and Hollingsworth 2005). The two major routes to polyploidy are either through WGD of a single species chromosome complement, known

as autopolyploidy, or through hybridisation between two species followed by WGD, known as allopolyploidy (however, there are other mechanisms; see Ramsey and Schemske (1998); Figure 1). As polyploid individuals tend to be larger, more vigorous, and quicker growing, it is no surprise that many crop species are polyploid (e.g. wheat, rice, potato, maize Renny-Byfield and Wendel (2014)). Although the majority of extant plant species are diploid (~67%, Rice et al. (2019)), extensive variability in ploidy levels exist across flowering plants, at all taxonomic levels (Kolar et al. 2017; Soltis et al. 2010). Most if not all plant species have experienced historical WGD a number of times (Clark and Donoghue 2018). Over evolutionary time the process of diploidisation reduces the size of these previously polyploid genomes through loss of repetitive sequence, chromosomal rearrangements, and reductions in chromosome number (Wendel 2015). Both the spatial and phylogenetic distribution of ploidy variation are unlikely to be uniform due to climatic and clade specific effects on unreduced gamete formation, which is the main driver in the creation of polyploid organisms (Kreiner, Kron, and Husband 2017; Bretagnolle and Thompson 1995; Rice et al. 2019).

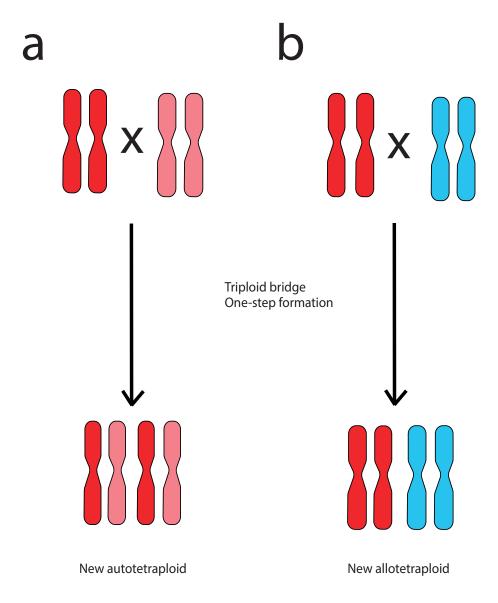


Figure 1: A simplified diagram of the formation of tetraploids from diploid progenitors. a) the formation of an autotetraploid from two diploid related individuals (red and pink) and b) the formation of an allotetraploid from two divergent diploid species (red and blue). Here, triploid intermediates can be formed through the combination of unreduced and reduced gametes. These triploids can then either self-fertilise, or backcross with parental species to produce tetraploids. Alternatively the union of unreduced gametes from both parents can produce a tetraploid in one step. Note: the distinction between auto and allopolyploids are blurred as the parental species may only be partially differentiated (Spoelhof, Soltis, and Soltis 2017), and there are other rarer mechanisms to produce auto and allopolyploids involving genome doubling.

Polyploidy is a contributing factor to taxonomic complexity, with recurrent polyploid formation followed by hybridisation within and between ploidy levels being the main driver of some TCGs (Brochmann et al.

2004). Further, there is evidence to suggest that polyploidy facilitates hybridisation as ploidy level increases, i.e. tetraploid x tetraploid crosses are more successful than diploid x diploid crosses (Novikova et al. 2020; Monnahan et al. 2019). Shifts in mating system from outcrossing to selfing or apomixis are also correlated with polyploid formation, due to the breakdown of self-incompatibility, selection against minority cytotype exclusion, and the capability of polyploids to alleviate inbreeding depression (Barringer 2007). Recurrent polyploidisation has been shown to generate taxa of ever higher ploidy levels, some of which may persist to form new species, or backcross with parental species to form a complex reticulate group (Brochmann, Soltis, and Soltis 1992). Polyploid type (auto or allopolyploid) is important in determining their role in taxonomic complexity. Autopolyploids and allopolyploids are both frequent in nature; current estimates for both kinds of polyploids are around 10% (Barker et al. 2016). Autopolyploids form complexes within species where ecology can shape the distribution of cytotypes (e.g. Wilson et al. (2020)), however these cytotypes are rarely considered species (Ramsey and Ramsey, 2014). Allopolyploids contribute to taxonomic complexity by combining two diverged genotypes to form a usually distinct and isolated taxon from the parents (Qiu, Liu, and Liu 2020). Both kinds of polyploid are present within some TCGs (e.g. Sorbus and Cochlearia; Ludwig et al. (2013); Gill, McAllister, and Fearn (1978)).

Unusual cytogenetic features in some plant groups have generated TCGs that have defied classification for over a hundred years. Two examples are the dog roses in Rosa section Caninae and the evening primroses in section Oenothera (Lim et al. 2005; Cleland 1944). The dog roses present an unusual breeding system where the female parent contributes 3-5 copies of the genome and the male parent contributes only one (section Caninae species are usually pentaploid; Rowley (1967)). This results in the hybrids containing a genome which is mostly maternal, and has the added complication that reciprocal crosses entirely change the genomic constitution of the hybrid. Species are difficult to tell apart when this breeding system is combined with hybridisation between rose species, as hybrids are fertile and readily backcross, forming swarms of individuals that may be impossible to identify morphologically (C. A. Stace, Preston, and Pearman 2015). Another example of strange cytogenetic behaviour, but at the diploid level, are the evening primroses. Here, the species exhibit a breeding system where translocation hybrids and balanced lethals produce chains of chromosomes which are inherited as single units (Cleland 1944). While evening primroses breed true when selfing, different combinations of chromosome chains generated through hybridisation can produce completely new morphological taxa (Cleland 1972). This introduces taxonomic complexity as, like the roses, when two species come into contact they form hybrid swarms of intermediate genotypes, each of which would breed true in isolation (C. A. Stace, Preston, and Pearman 2015).

Novel features of TCGs; parasitism and plasticity:

Approximately 1% of all angiosperm species are parasitic, with some genera being particularly speciose such as *Pedicularis* (c.a. 650sp), *Euphrasia* (c.a. 260 sp), and *Thesium* (c.a. 300sp) (Nickrent 2020; Moore, Verboom, and Forest 2010; Twyford 2018). Parasitic plants are defined by the formation of a structure called the haustorium (Figure 2), which is used to extract water and soluble nutrients from the host plant (Twyford 2018). The haustorium can attach either to roots (e.g. *Orobanche*; Musselman (1980)) or shoots (e.g. *Viscum*; Becker (1986)), or rarely the haustorium is present inside the host plant itself (e.g. endoparasitic *Pilostyles*; Fernandes et al. (1998)). There are two types of parasitic plants – hemiparasites which retain photosynthetic competency, and holoparasites which are devoid of chlorophyll and entirely dependent on host plants (Joel, Gressel, and Musselman 2013). Host species range and identity varies widely between parasitic plants, and some generalists can parasitise more than 100 host plant species (e.g. *Amyema miquelii*; Clark, McComb, and Taylor-Robinson (2020)), while others specialise on a single or few host plants (e.g. *Epifagus virginiana* on *Fagus grandiflora*; Tsai and Manos (2010)).



Figure 2: A variety of parasitic plants, showing haustorial connections. a) Cuscuta europaea (dodder), showing flowers and twining red stems. b) Erianthemum ngamicum showing its large terminal haustorium. c) Orobanche hederae haustorial connection to the host root (light) d) Hydnora visseri forming multiple haustoria on host root (light). e) Cassytha pubescens twining around host forming many haustorial connections. f) Viscum album showing self-parasitism where two younger V. album individuals have established on an internode of an older specimen. g) Terminal (asterisk) and lateral (arrows) haustoria of Plicosepalus kalachariensis. h) flowers of Aqelanthus gracilis. Figure and text adapted from Joel, Gressel, and Musselman (2013).

How different host species may influence parasitic plant phenotypes however, has been little explored in relation to taxonomic complexity. Yet it is well known that different host species can dramatically impact the growth, development, and evolution of parasitic plant individuals (Matthies 2017; Rowntree et al. 2014).

There are two main ways in which the parasitic lifestyle may contribute to taxonomic complexity. Firstly, parasitic plant species may show phenotypic plasticity when utilising different host species, which may be substantial enough to confuse species identification (Wilkins 1963). For example, a suite of traits used in species discrimination in the Orobanchaceae, including corolla length, node to first flower, and plant height, all vary in relation to host quality (Matthies 2017; Jonstrup, Hedren, and Andersson 2016). Secondly, differential host use can drive the evolution of cryptic taxa, which has been seen in the plant genus Orobanche, which specialise on different host species (Thorogood, Rumsey, and Hiscock 2009).

The genus *Euphrasia*:

The genus Euphrasia (evebrights) are a large group of 260, mainly annual but sometimes perennial, hemiparasitic plant species in the Orobanchaceae (Nickrent 2020; Yeo 1978). A study of global Euphrasia species has established its bipolar distribution, and estimated the age of the genus to be around 20-30 Mya (Gussarova et al. 2008). Euphrasia species are found in Chile, Australia and New Zealand, parts of South East Asia, and widely across the northern hemisphere. It is here, in the northern hemisphere that Euphrasia species are most diverse and where the genus is a notorious TCG (Yeo 1978). Within Europe, Britain and Ireland contain the highest concentration of species, where 21 have been described (Metherell and Rumsey 2018). The number of described species in Britain may be an exaggeration however due to over-splitting. Here, Euphrasia is widespread and occupies a range of habitats, from coastal scrub to heather moorland, and damp grassland to mountain tops (Metherell and Rumsey 2018). While the split between diploid species and tetraploid species is unequivocal, within each ploidy level there is much uncertainty in species limits due to rapid recent divergence (Wang et al. 2018). Within tetraploids, some species groups are more distinct that others, for example E. micrantha and E. scottica form a natural group, as do the widespread outcrossing species E. arctica and E. nemorosa (French et al. 2008). Omitting these two species groups, it leaves a nebulous pool of widespread and more localised selfing species that may be genetically partitioned more by geography than by species limits. Within the diploid species, there is evidence to suggest that the endemic diploid species E. vigursii and E. rivularis are distinct from the other diploids (E. anglica and E. rostkoviana; French et al. (2008)).

The taxonomic complexity of Euphrasia in Britain is reflected in the unstable taxonomy over the past hundred years, in the great diversity of morphologies found in the field, and in the complex genetic structure of the species (Wettstein 1896; Metherell and Rumsey 2018). The main drivers of this diversity are recent postglacial divergence, the tendency to self-fertilise, and rampant hybridisation – 71 hybrid combinations have been reported to date (Figure 3; Wang et al. (2018); C. A. Stace, Preston, and Pearman (2015); Metherell and Rumsey (2018)). At least five species of the British species of Euphrasia have a putative hybrid origin, with two species having arisen from diploid-tetraploid hybridisation events (Yeo 1956; Silverside 1990). The evidence of cross-ploidy hybridisation began with early cytological work on a triploid hybrid which was intermediate between E. micrantha and E. anglica (= E. vigursii) in morphology (Yeo 1956). Further evidence was found by French et al. (2008) when tetraploid specific AFLP bands were seen in diploid samples. Lastly, Becher et al. (2020) in a genomic analysis of British Euphrasia, found some evidence for gene flow between E. arctica and the sampled diploid species. These cross-ploidy hybrid species are of particular interest, as they combine three important features of TCGs – hybridisation, polyploidy, and mating system variability.

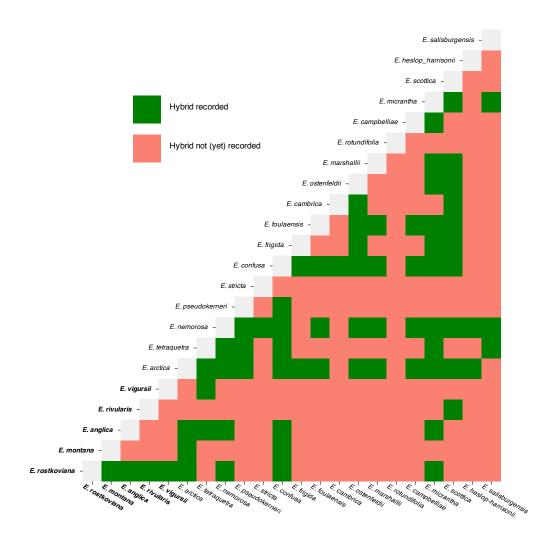


Figure 3: A matrix showing interspecific crossing barriers in British *Euphrasia* species. Grey squares indicate intraspecific crosses, pink squares show crosses not known to produce hybrids in the wild, and green squares show those crosses to have produced hybrids in the wild. Diploid species are emphasised in bold type. Data taken from Metherell and Rumsey (2018).

Euphrasia in Britain and Ireland have two different ploidy levels, with diploids (2n = 2x = 22) that have a southern distribution and tetraploids (2n = 4x = 44) more predominant in the north (Yeo 1978). The tetraploid species are allotetraploids, containing one subgenome which is closely related to extant diploids (0.2% divergent; Becher et al. (2020)). This low divergence has led to the hypothesis that pairing can occur between the chromosomes of diploid species and the diploid-like subgenomes of tetraploids, which could explain diploid-tetraploid hybridisation in the genus (see Figure 4; Yeo (1956)). The divergence between diploids and tetraploids is around 5% based on both ITS sequencing, and genome wide data, which corresponds to a split time of around 8 Mya (Wang et al. 2018; Becher et al. 2020). High divergence between diploids and tetraploids points to ploidy being an effective barrier to gene exchange, however there is mounting evidence that gene flow between ploidy levels is present, but rare (French et al. 2008; Becher et al. 2020). Hand crosses have generally failed to produce diploid-tetraploid hybrids, and only a single triploid hybrid has been

found in the field (Yeo 1954, 1956). Controlled crosses may yet yield useful insights into the biology of diploid-tetraploid hybrids in Euphrasia.

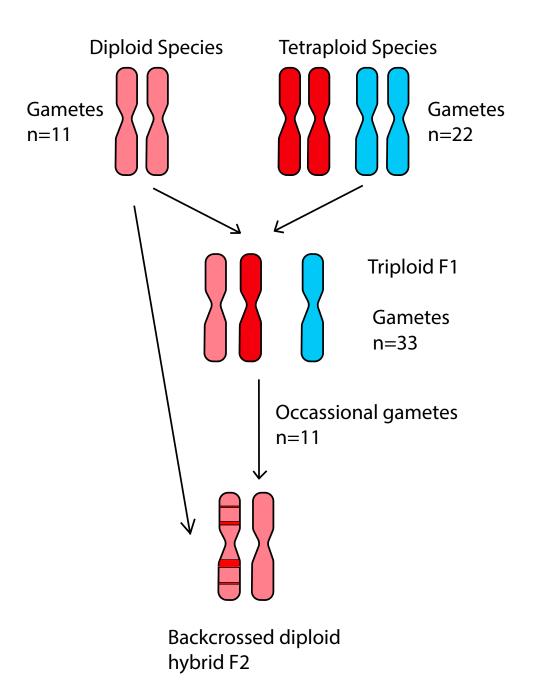


Figure 4: Schematic of diploid-tetraploid hybridisation in *Euphrasia*. Here, a diploid species hybridises with a tetraploid species to form a triploid intermediate. Note that pink and red colours indicate the low (0.2%; Becher et al. (2020)) divergence between diploid (pink) and diploid-like (red) tetraploid subgenomes. This triploid F1 can then backcross to the diploid parent through rare haploid gamete segregation to produce a backcrossed F2 individual. Note the striped chromosomes in the F2 backcross indicate recombination between the diploid (pink) and the diploid-like (red) subgenome of the tetraploid. It is thought this process has given rise to the hybrid species *Euphrasia vigursii* and *E. rivularis* (Yeo 1956; Silverside 1990).

10

The mating system in British Euphrasia is highly variable, and ranges from outcrossing to highly selfing (note apomixis has not been found in the genus). Outcrossing rate is correlated with flower size in Euphrasia (r = -0.89; French et al. (2005)), with smaller flowered species more likely to self than larger flowered species with showy corollas. For example, the tetraploid mountain specialist E. cambrica which is endemic to Wales has a corolla length of 4mm, while the diploid E. montana of wet grasslands has a corolla length of up to 12mm (and can be larger in cultivation; Figure 5; Metherell and Rumsey (2018)). Mating system may also impact the directionality of introgression in some cases, for example between tetraploid E. micrantha and diploid E. anglica which are the putative parental species for the diploid hybrid species E. vigursii. E. anglica has large flowers and is mainly outcrossing, therefore it is likely to be the pollen donor as its flowers are visited first with greater probability (Yeo 1968). Therefore the diploid species pollen will be more competitive on the stigma of the tetraploid E. micrantha (Ruhsam, Hollingsworth, and Ennos 2011). As pollen fitness is low in the triploid F1 hybrid, selfing is unlikely to occur and the diploid E. anglica is likely to fertilise the F1 (Ruhsam, Hollingsworth, and Ennos 2013). If this happens over many generations, introgression will occur from the tetraploid to the diploid species (as shown in Figure 5).



Figure 5: corolla size extremes in British Euphrasia species. a) shows diploid E. montana from the Lake District, England, which has a corolla size of ~ 12 mm. b) shows the Welsh endemic E. cambrica from Snowdon, with small flowers ~ 4 mm which rarely open fully (Metherell and Rumsey 2018). Photo credit: author.

Being generalist parasites, Euphrasia can gain benefit from a wide variety of plant species which in turn impact the morphology, growth, and fitness of the parasitic Euphrasia plants (Svensson and Carlsson 2004). Common garden experiments have allowed researchers to investigate these factors in different hemiparasitic plant systems, especially in the Orobanchaceae. For example, it has been shown in Rhinanthus that different host species employ different resistance mechanisms to haustorial attack, and this underlies the performance of hemiparasitic Rhinanthus (Cameron, Coats, and Seel 2007). Common garden experiments have also established that host species have strong effects on the biomass and morphology of Melampyrum (Matthies

2017). As shown from early work by Yeo (1964), Euphrasia can easily be brought into cultivation, and their annual life histories (like many other members of the hemiparasitic Orobanchaceae) facilitate experimental work within the timeframe of a PhD.

The British flora:

To create a synthesis of taxonomic complexity, it is useful to be able to frame it in a broad, comparative context. The British flora is an ideal study system, as it contains a manageable number of native plant species (~1,400), but with around 20% of all familial flowering plant diversity (Stace 2019). Alien species increase the total number of species over two-fold, and most of these are very well characterised (CA Stace and Crawley 2015). The British flora represents the most comprehensively studied flora to date, across a variety of disciplines due to the collaboration between amateur and professional botanists for over a century (Allen 1986). The Botanical Society of Britain and Ireland (BSBI) have played a leading role, holding large databases of plant distributions. The result of this, is that plants can be found and told apart easily in the field, and has led to a huge growth in knowledge of the system. Now for almost all native plants, there is detailed information on plant identification (Stace 2019), alien species (CA Stace and Crawley 2015), ecology and life history (Fitter and Peat 1994), chromosome numbers and ploidy level (BSBI Cytology Database), genome sizes (Kew C-value Database), hybridisation (C. A. Stace, Preston, and Pearman 2015), and most recently DNA barcoding (Jones, 2020 unpublished). This wealth of knowledge has been leveraged in parts of this thesis to gain a broader perspective on the topic at hand.

New methods for investigating taxonomic complexity:

In recent years, new methods have emerged that can help us to understand taxonomic complexity, and to resolve major questions. Older marker types such as allozymes, AFLPs, RAPD, and microsatellites had many limitations, including needing large amounts of DNA, problems with homology, detection of few polymorphisms, and lack of reproducibility (Lowe, Harris, and Ashton 2004). Next generation sequencing (NGS) is one tool to aid resolution of complex taxonomic relationships, for example - genotyping by sequencing (GBS) (Anderson et al. 2017), restriction associated digestion (RAD) sequencing (Zhou et al. 2020), target capture (Carter et al. 2019), and whole genome sequencing (Dupuis et al. 2017) are becoming common. These datasets can be used to infer relationships across the entire genome, to correlate morphology and genetics to reveal the genes underlying certain traits, and to compare large and diverse sample sets. DNA barcoding can now be deployed at scale across diverse taxa (Hollingsworth et al. 2009), which means large scale phylogenetic analyses that incorporate diverse information are now possible. New analytical models now provide a framework for accounting for different sources of variation, and are applicable to use for many different questions across evolutionary biology (e.g. (Generalised) Linear Mixed Models; (Bolker et al. 2009; Hadfield 2010)). These methods are used throughout this thesis to gain novel insights into taxonomic complexity.

Thesis aims:

The main aim of my thesis is to investigate taxonomic complexity in *Euphrasia*, and the role of hybridisation across the British flora, with a particular focus polyploidy, parasitism, and their interactions. It is not currently known what the extent and prevalence of such interactions between hybridisation and plants of different ploidy level are, and is important to explore because of the potentially large impact they can have on plant evolution. In addition, the parasitic habit of some plants and how this affects their phenotype has been little explored and opens up a new avenue of explanations for taxonomic complexity.

The thesis is split into two parts which target different aspects of taxonomic complexity, and draw on broader themes in evolutionary biology. The first part concentrates on the contributions of hybridisation to taxonomic complexity in plants (and to a lesser degree, animals), with a case study in the genus Euphrasia. Here, I firstly ask how prevalent hybridisation is between plants that differ in ploidy level, and whether it is a significant evolutionary phenomenon (Chapter 2). The interaction between ploidy level and hybridisation is a poorly explored topic and scattered across the literature. I bring together a comprehensive list of examples, and synthesise current knowledge on this topic. After understanding this global variation, I concentrate on the British flora, and model the probability of hybridisation across the flora (Chapter 3). Significantly, this model includes phylogenetic relationships, and genetic distances based on the first complete DNA barcode dataset across a flora, which previous studies have not been able to comprehensively address (e.g. Ellstrand, Whitkus, and Rieseberg (1996); Mitchell et al. (2019)). Then, I focus on the promiscuous genus Euphrasia and ask whether we see evidence of hybridisation between divergent species of different ploidy at a fine spatial scale. Using reduced representation sequencing of genome wide markers (genotyping by sequencing; GBS), I use a combination of classical population genetic tools, genome sequencing, and demographic simulation, to understand the pattern of hybridisation in a Euphrasia contact zone (Chapter 4).

In the second part of the thesis, two novel features of taxonomic complexity - phenotypic plasticity and parasitism - are explored in the parasitic plant genus *Euphrasia*. I use common garden experiments to ask firstly how different host species affect the morphology of *Euphrasia* and the ability to discriminate between *Euphrasia* species (Chapter 5). This was investigated using a single species of *Euphrasia* grown across eight different host species and multiple species of *Euphrasia* on a single host, where various morphological traits of *Euphrasia* were quantified. Finally, I look at the role of host parasite interactions to understand host specialisation in the genus, by growing multiple *Euphrasia* and multiple host species together (Chapter 6).

Alix, K., P. R. Gerard, T. Schwarzacher, and J. S. Heslop-Harrison. 2017. "Polyploidy and Interspecific Hybridization: Partners for Adaptation, Speciation and Evolution in Plants." Journal Article. *Annals of Botany* 120 (2): 183–94. https://doi.org/10.1093/aob/mcx079.

Allen, David Elliston. 1986. The Botanists: A History of the Botanical Society of the British Isles Through a Hundred and Fifty Years. Book. Winchester: St. Paul's Bibliographies.

Anderson, B. M., K. R. Thiele, S. L. Krauss, and M. D. Barrett. 2017. "Genotyping-by-Sequencing in a Species Complex of Australian Hummock Grasses (Triodia): Methodological Insights and Phylogenetic Resolution." Journal Article. *Plos One* 12 (1). https://doi.org/10.1371/journal.pone.0171053.

Barker, M. S., N. Arrigo, A. E. Baniaga, Z. Li, and D. A. Levin. 2016. "On the Relative Abundance of Autopolyploids and Allopolyploids." Journal Article. *New Phytologist* 210 (2): 391–98. https://doi.org/10.1111/nph.13698.

Barringer, B. C. 2007. "Polyploidy and Self-Fertilization in Flowering Plants." Journal Article. American Journal of Botany 94 (9): 1527–33. https://doi.org/10.3732/ajb.94.9.1527.

Becher, H., M. Brown, G. Powell, C. Metherell, N.J. Riddiford, and A.D. Twyford. 2020. "Maintenance of Species Differences in Closely Related Tetraploid Parasitic Euphrasia (Orobanchaceae) on an Isolated Island." Journal Article. *Biorxiv*.

Becker, H. 1986. "BOTANY of European Mistletoe (Viscum-Album L)." Journal Article. Oncology 43: 2–7. <Go to ISI>://WOS:A1986F476300002.

Belton, G. S., W. F. P. van Reine, J. M. Huisman, S. G. A. Draisma, and C. F. D. Gurgel. 2014. "RESOLVING Phenotypic Plasticity and Species Designation in the Morphologically Challenging Caulerpa Racemosa-Peltata Complex (Caulerpaceae, Chlorophyta)." Journal Article. *Journal of Phycology* 50 (1): 32–54. https://doi.org/10.1111/jpy.12132.

- Bolker, B. M., M. E. Brooks, C. J. Clark, S. W. Geange, J. R. Poulsen, M. H. H. Stevens, and J. S. S. White. 2009. "Generalized Linear Mixed Models: A Practical Guide for Ecology and Evolution." Journal Article. *Trends in Ecology & Evolution* 24 (3): 127–35. https://doi.org/10.1016/j.tree.2008.10.008.
- Bretagnolle, F., and J. D. Thompson. 1995. "TANSLEY Review No-78 Gametes with the Somatic Chromosome-Number Mechanisms of Their Formation and Role in the Evolution of Autopolyploid Plants." Journal Article. New Phytologist 129 (1): 1–22. https://doi.org/10.1111/j.1469-8137.1995.tb03005.x.
- Brochmann, C. 1984. "HYBRIDIZATION and Distribution of Argyranthemum-Coronopifolium (Asteraceae, Anthemideae) in the Canary Islands." Journal Article. *Nordic Journal of Botany* 4 (6): 729–36. https://doi.org/10.1111/j.1756-1051.1984.tb02001.x.
- Brochmann, C., A. K. Brysting, I. G. Alsos, L. Borgen, H. H. Grundt, A. C. Scheen, and R. Elven. 2004. "Polyploidy in Arctic Plants." Journal Article. *Biological Journal of the Linnean Society* 82 (4): 521–36. https://doi.org/10.1111/j.1095-8312.2004.00337.x.
- Brochmann, C., P. S. Soltis, and D. E. Soltis. 1992. "RECURRENT Formation and Polyphyly of Nordic Polyploids in Draba (Brassicaceae)." Journal Article. *American Journal of Botany* 79 (6): 673–88. https://doi.org/10.2307/2444883.
- Cameron, D. D., A. M. Coats, and W. E. Seel. 2007. "Differential Resistance Among Host and Non-Host Species Underlies the Variable Success of the Hemi-Parasitic Plant Rhinanthus Minor (Vol 98, Pg 1289, 2006)." Journal Article. *Annals of Botany* 99 (3): 563–63. https://doi.org/10.1093/aob/mcm020.
- Campbell, C. S., and W. A. Wright. 1996. "Apomixis, Hybridization, and Taxonomic Complexity in Eastern North American Amelanchier (Rosaceae)." Journal Article. Folia Geobotanica & Phytotaxonomica 31 (3): 345–54. https://doi.org/10.1007/bf02815379.
- Carter, K. A., A. Liston, N. V. Bassil, L. A. Alice, J. M. Bushakra, B. L. Sutherland, T. C. Mockler, D. W. Bryant, and K. E. Hummer. 2019. "Target Capture Sequencing Unravels Rubus Evolution." Journal Article. Frontiers in Plant Science 10. https://doi.org/10.3389/fpls.2019.01615.
- Chapman, M. A., and R. J. Abbott. 2010. "Introgression of Fitness Genes Across a Ploidy Barrier." Journal Article. New Phytologist 186 (1): 63–71. https://doi.org/10.1111/j.1469-8137.2009.03091.x.
- Clark, J. W., and P. C. J. Donoghue. 2018. "Whole-Genome Duplication and Plant Macroevolution." Journal Article. Trends in Plant Science 23 (10): 933–45. https://doi.org/10.1016/j.tplants.2018.07.006.
- Clark, N. F., J. A. McComb, and A. W. Taylor-Robinson. 2020. "Host Species of Mistletoes (Loranthaceae and Viscaceae) in Australia." Journal Article. *Australian Journal of Botany* 68 (1): 1–13. https://doi.org/10.1071/bt19137.
- Cleland, Ralph Erskine. 1972. Oenothera: Cytogenetics and Evolution. Book. London: Academic Press.
- Cleland, R. E. 1944. "The Problem of Species in Oenothera." Journal Article. *American Naturalist* 78: 5–28. https://doi.org/10.1086/281163.
- Coyne, Jerry A., and H. Allen Orr. 2004. Speciation. Book. Sunderland, Mass.; [Great Britain]: Sinauer Associates. Table of contents http://www.loc.gov/catdir/toc/ecip0417/2004009505.html.
- Czapik, R. 1996. "Problems of Apomictic Reproduction in the Families Compositae and Rosaceae." Journal Article. Folia Geobotanica & Phytotaxonomica 31 (3): 381–87. https://doi.org/10.1007/bf02815382.
- Dickinson, T. A., E. Lo, and N. Talent. 2007. "Polyploidy, Reproductive Biology, and Rosaceae: Understanding Evolution and Making Classifications." Journal Article. *Plant Systematics and Evolution* 266 (1-2): 59–78. https://doi.org/10.1007/s00606-007-0541-2.

- Dupuis, J. R., B. M. T. Brunet, H. M. Bird, L. M. Lumley, G. Fagua, B. Boyle, R. Levesque, M. Cusson, J. A. Powell, and F. A. H. Sperling. 2017. "Genome-Wide Snps Resolve Phylogenetic Relationships in the North American Spruce Budworm (Choristoneura Fumiferana) Species Complex." Journal Article. *Molecular Phylogenetics and Evolution* 111: 158–68. https://doi.org/10.1016/j.ympev.2017.04.001.
- Ellstrand, N. C., R. Whitkus, and L. H. Rieseberg. 1996. "Distribution of Spontaneous Plant Hybrids." Journal Article. *Proceedings of the National Academy of Sciences of the United States of America* 93 (10): 5090–3. https://doi.org/10.1073/pnas.93.10.5090.
- Ennos, R. A., G. C. French, and P. M. Hollingsworth. 2005. "Conserving Taxonomic Complexity." Journal Article. Trends in Ecology & Evolution 20 (4): 164–68. https://doi.org/10.1016/j.tree.2005.01.012.
- Ennos, R. A., R. Whitlock, M. F. Fay, B. Jones, L. E. Neaves, R. Payne, I. Taylor, N. De Vere, and P. M. Hollingsworth. 2012. "Process-Based Species Action Plans: An Approach to Conserve Contemporary Evolutionary Processes That Sustain Diversity in Taxonomically Complex Groups." Journal Article. *Botanical Journal of the Linnean Society* 168 (2): 194–203. https://doi.org/10.1111/j.1095-8339.2011.01206.x.
- Federici, S., A. Galimberti, F. Bartolucci, I. Bruni, F. De Mattia, P. Cortis, and M. Labra. 2013. "DNA Barcoding to Analyse Taxonomically Complex Groups in Plants: The Case of Thymus (Lamiaceae)." Journal Article. *Botanical Journal of the Linnean Society* 171 (4): 687–99. https://doi.org/10.1111/boj.12034.
- Fernandes, G. W., E. A. de Mattos, A. C. Franco, U. Luttge, and H. Ziegler. 1998. "Influence of the Parasite Pilostyles Ingae (Rafflesiaceae) on Some Physiological Parameters of the Host Plant, Mimosa Naguirei (Mimosaceae)." Journal Article. *Botanica Acta* 111 (1): 51–54. https://doi.org/10.1111/j.1438-8677.1998.tb00676.x.
- Fitter, A. H., and H. J. Peat. 1994. "THE Ecological Flora Database." Journal Article. *Journal of Ecology* 82 (2): 415–25. https://doi.org/10.2307/2261309.
- French, G. C., R. A. Ennos, A. J. Silverside, and P. M. Hollingsworth. 2005. "The Relationship Between Flower Size, Inbreeding Coefficient and Inferred Selfing Rate in British Euphrasia Species." Journal Article. *Heredity* 94 (1): 44–51. https://doi.org/10.1038/sj.hdy.6800553.
- French, G. C., P. M. Hollingsworth, A. J. Silverside, and R. A. Ennos. 2008. "Genetics, Taxonomy and the Conservation of British Euphrasia." Journal Article. *Conservation Genetics* 9 (6): 1547–62. https://doi.org/10.1007/s10592-007-9494-9.
- Garcia-Melo, J. E., C. Oliveira, G. J. D. Silva, L. E. Ochoa-Orrego, L. H. G. Pereira, and J. A. Maldonado-Ocampo. 2019. "Species Delimitation of Neotropical Characins (Stevardiinae): Implications for Taxonomy of Complex Groups." Journal Article. *Plos One* 14 (6). https://doi.org/10.1371/journal.pone.0216786.
- Gill, J.J.B., H.A. McAllister, and G.M. Fearn. 1978. "Cytotaxonomic Studies on the Cochlearia Officinalis L. Group from Inland Stations in Britain." Journal Article. Watsonia 12: 15–21.
- Gussarova, G., M. Popp, E. Vitek, and C. Brochmann. 2008. "Molecular Phylogeny and Biogeography of the Bipolar Euphrasia (Orobanchaceae): Recent Radiations in an Old Genus." Journal Article. *Molecular Phylogenetics and Evolution* 48 (2): 444–60. https://doi.org/10.1016/j.ympev.2008.05.002.
- Hadfield, J. D. 2010. "MCMC Methods for Multi-Response Generalized Linear Mixed Models: The Mcmcglmm R Package." Journal Article. *Journal of Statistical Software* 33 (2): 1-22. <Go to ISI>://WOS:000275203300001.
- Hersh, E., J. Grimm, and J. Whitton. 2016. "Attack of the Clones: Reproductive Interference Between Sexuals and Asexuals in the Crepis Agamic Complex." Journal Article. *Ecology and Evolution* 6 (18): 6473–83. https://doi.org/10.1002/ece3.2353.

- Hollingsworth, P. M., L. L. Forrest, J. L. Spouge, M. Hajibabaei, S. Ratnasingham, M. van der Bank, M. W. Chase, et al. 2009. "A Dna Barcode for Land Plants." Journal Article. *Proceedings of the National Academy of Sciences of the United States of America* 106 (31): 12794–7. https://doi.org/10.1073/pnas.0905845106.
- Hollingsworth, P. M., J. Squirrell, M. L. Hollingsworth, A.J. Richards, and R. M. Bateman. 2006. "Taxonomic Complexity, Conservation and Recurrent Origins of Self-Pollination in Epipactis." Journal Article. *Current Taxonomic Research on the British & European Flora*, BSBI, 27–44. http://www.rbge.org.uk/assets/files/science/Genetics Conservation/EpipactisTaxonomicComplexity.pdf.
- Joel, JM, J Gressel, and LJ Musselman. 2013. Parasitic Orobanchaceae. Book. Berlin, New York: Springer.
- Jonstrup, A., M. Hedren, and S. Andersson. 2016. "Host Environment and Local Genetic Adaptation Determine Phenotype in Parasitic Rhinanthus Angustifolius." Journal Article. *Botanical Journal of the Linnean Society* 180 (1): 89–103. https://doi.org/10.1111/boj.12352.
- Kolar, F., M. Certner, J. Suda, P. Schonswetter, and B. C. Husband. 2017. "Mixed-Ploidy Species: Progress and Opportunities in Polyploid Research." Journal Article. *Trends in Plant Science* 22 (12): 1041–55. https://doi.org/10.1016/j.tplants.2017.09.011.
- Kreiner, J. M., P. Kron, and B. C. Husband. 2017. "Evolutionary Dynamics of Unreduced Gametes." Journal Article. *Trends in Genetics* 33 (9): 583–93. https://doi.org/10.1016/j.tig.2017.06.009.
- Leavitt, S. D., L. A. Johnson, T. Goward, and L. L. St Clair. 2011. "Species Delimitation in Taxonomically Difficult Lichen-Forming Fungi: An Example from Morphologically and Chemically Diverse Xanthoparmelia (Parmeliaceae) in North America." Journal Article. *Molecular Phylogenetics and Evolution* 60 (3): 317–32. https://doi.org/10.1016/j.ympev.2011.05.012.
- Lim, K. Y., G. Werlemark, R. Matyasek, J. B. Bringloe, V. Sieber, H. El Mokadem, J. Meynet, J. Hemming, A. R. Leitch, and A. V. Roberts. 2005. "Evolutionary Implications of Permanent Odd Polyploidy in the Stable Sexual, Pentaploid of Rosa Canina L." Journal Article. *Heredity* 94 (5): 501–6. https://doi.org/10.1038/sj.hdy.6800648.
- Lowe, Andrew Dr, Stephen Harris, and Paul Dr Ashton. 2004. *Ecological Genetics : Design, Analysis, and Application*. Book. Malden, Mass. ; Oxford: Blackwell Pub.
- Ludwig, S., A. Robertson, T. C. G. Rich, M. Djordjevic, R. Cerovic, L. Houston, S. A. Harris, and S. J. Hiscock. 2013. "Breeding Systems, Hybridization and Continuing Evolution in Avon Gorge Sorbus." Journal Article. *Annals of Botany* 111 (4): 563–75. https://doi.org/10.1093/aob/mct013.
- Luo, A., C. Ling, S. Y. W. Ho, and C. D. Zhu. 2018. "Comparison of Methods for Molecular Species Delimitation Across a Range of Speciation Scenarios." Journal Article. *Systematic Biology* 67 (5): 830–46. https://doi.org/10.1093/sysbio/syy011.
- Mallet, J. 2007. "Hybrid Speciation." Journal Article. Nature 446 (7133): 279–83. https://doi.org/10.1038/nature05706.
- Matthies, D. 2017. "Interactions Between a Root Hemiparasite and 27 Different Hosts: Growth, Biomass Allocation and Plant Architecture." Journal Article. *Perspectives in Plant Ecology Evolution and Systematics* 24: 118–37. https://doi.org/10.1016/j.ppees.2016.12.006.
- Mayr, Ernst. 1963. Animal Species and Evolution. Book. Cambridge, Belknap Press of Harvard University Press.
- Metherell, C., and F. Rumsey. 2018. Eyebrights (Euphrasia) of the Uk and Ireland. Book. Botanical Society of Britain; Ireland.

- Mitchell, N., L. G. Campbell, J. R. Ahern, K. C. Paine, A. B. Giroldo, and K. D. Whitney. 2019. "Correlates of Hybridization in Plants." Journal Article. *Evolution Letters*, 16. https://doi.org/10.1002/evl3.146.
- Monnahan, P., F. Kolar, P. Baduel, C. Sailer, J. Koch, R. Horvath, B. Laenen, et al. 2019. "Pervasive Population Genomic Consequences of Genome Duplication in Arabidopsis Arenosa." Journal Article. *Nature Ecology & Evolution* 3 (3): 457-+. https://doi.org/10.1038/s41559-019-0807-4.
- Moore, T. E., G. A. Verboom, and F. Forest. 2010. "Phylogenetics and Biogeography of the Parasitic Genus Thesium L. (Santalaceae), with an Emphasis on the Cape of South Africa." Journal Article. *Botanical Journal of the Linnean Society* 162 (3): 435–52. https://doi.org/10.1111/j.1095-8339.2010.01032.x.
- Musselman, L. J. 1980. "THE Biology of Striga, Orobanche, and Other Root-Parasitic Weeds." Journal Article. Annual Review of Phytopathology 18: 463–89. https://doi.org/10.1146/annurev.py.18.090180.002335.
- Nickrent, D. L. 2020. "Parasitic Angiosperms: How Often and How Many?" Journal Article. *Taxon* 69 (1): 5–27. https://doi.org/10.1002/tax.12195.
- Novikova, P. Y., I. G. Brennan, W. Booker, M. Mahony, P. Doughty, A. R. Lemmon, E. M. Lemmon, et al. 2020. "Polyploidy Breaks Speciation Barriers in Australian Burrowing Frogs Neobatrachus." Journal Article. *Plos Genetics* 16 (5). https://doi.org/10.1371/journal.pgen.1008769.
- Pellicer, J., S. Clermont, L. Houston, T. C. G. Rich, and M. F. Fay. 2012. "Cytotype Diversity in the Sorbus Complex (Rosaceae) in Britain: Sorting Out the Puzzle." Journal Article. *Annals of Botany* 110 (6): 1185–93. https://doi.org/10.1093/aob/mcs185.
- Qiu, T., Z. Y. Liu, and B. Liu. 2020. "The Effects of Hybridization and Genome Doubling in Plant Evolution via Allopolyploidy." Journal Article. *Molecular Biology Reports* 47 (7): 5549–58. https://doi.org/10.1007/s11033-020-05597-y.
- Raggi, L., E. Bitocchi, L. Russi, G. Marconi, T. F. Sharbel, F. Veronesi, and E. Albertini. 2015. "Understanding Genetic Diversity and Population Structure of a Poa Pratensis Worldwide Collection Through Morphological, Nuclear and Chloroplast Diversity Analysis." Journal Article. *Plos One* 10 (4). https://doi.org/10.1371/journal.pone.0124709.
- Ramsey, J., and D. W. Schemske. 1998. "Pathways, Mechanisms, and Rates of Polyploid Formation in Flowering Plants." Journal Article. Annual Review of Ecology and Systematics 29: 467-501. https://doi.org/10.1146/annurev.ecolsys.29.1.467.
- Renny-Byfield, S., and J. F. Wendel. 2014. "DOUBLING down on Genomes: POLYPLOIDY and Crop Plants." Journal Article. *American Journal of Botany* 101 (10): 1711–25. https://doi.org/10.3732/ajb.1400119.
- Rhymer, J. M., and D. Simberloff. 1996. "Extinction by Hybridization and Introgression." Journal Article. *Annual Review of Ecology and Systematics* 27: 83–109. https://doi.org/10.1146/annurev.ecolsys.27.1.83.
- Rice, A., P. Smarda, M. Novosolov, M. Drori, L. Glick, N. Sabath, S. Meiri, J. Belmaker, and I. Mayrose. 2019. "The Global Biogeography of Polyploid Plants." Journal Article. *Nature Ecology & Evolution* 3 (2): 265–+. https://doi.org/10.1038/s41559-018-0787-9.
- Rieseberg, L. H. 1997. "Hybrid Origins of Plant Species." Journal Article. Annual Review of Ecology and Systematics 28: 359–89. https://doi.org/10.1146/annurev.ecolsys.28.1.359.
- Rieseberg, L. H., C. Vanfossen, and A. M. Desrochers. 1995. "HYBRID Speciation Accompanied by Genomic Reorganization in Wild Sunflowers." Journal Article. *Nature* 375 (6529): 313–16. https://doi.org/10.1038/375313a0.
- Rieseberg, L. H., and J. H. Willis. 2007. "Plant Speciation." Journal Article. Science 317 (5840): 910–14. https://doi.org/10.1126/science.1137729.

Robertson, A., T. C. G. Rich, A. M. Allen, L. Houston, C. Roberts, J. R. Bridle, S. A. Harris, and S. J. Hiscock. 2010. "Hybridization and Polyploidy as Drivers of Continuing Evolution and Speciation in Sorbus." Journal Article. *Molecular Ecology* 19 (8): 1675–90. https://doi.org/10.1111/j.1365-294X.2010.04585.x.

Roodt, R., and J. J. Spies. 2003. "Chromosome Studies in the Grass Subfamily Chloridoideae. II. An Analysis of Polyploidy." Journal Article. *Taxon* 52 (4): 736–46. https://doi.org/10.2307/3647348.

Rowley, G.D. 1967. "Chromosome Studies and Evolution in Rosa." Journal Article. Bulletin Du Jardin Botanique National de Belgique 37 (1): 45–52.

Rowntree, J. K., D. F. Barham, A. J. A. Stewart, and S. E. Hartley. 2014. "The Effect of Multiple Host Species on a Keystone Parasitic Plant and Its Aphid Herbivores." Journal Article. Functional Ecology 28 (4): 829–36. https://doi.org/10.1111/1365-2435.12281.

Ruhsam, M., P. M. Hollingsworth, and R. A. Ennos. 2011. "Early Evolution in a Hybrid Swarm Between Outcrossing and Selfing Lineages in Geum." Journal Article. *Heredity* 107 (3): 246–55. https://doi.org/10.1038/hdy.2011.9.

———. 2013. "PATTERNS of Mating, Generation of Diversity, and Fitness of Offspring in a Geum Hybrid Swarm." Journal Article. *Evolution* 67 (9): 2728–40. https://doi.org/10.1111/evo.12147.

Silverside, A. 1990. "A Guide to Eyebrights (Euphrasia) (Ii) the E. Rostkoviana Group." Journal Article. Wild Flower Magazine 418: 31–34.

Simpson, G. G. 1951. "THE Species Concept." Journal Article. Evolution 5 (4): 285–98. https://doi.org/10.2307/2405675.

Sochor, M., R. J. Vasut, T. F. Sharbel, and B. Travnicek. 2015. "How Just a Few Makes a Lot: Speciation via Reticulation and Apomixis on Example of European Brambles (Rubus Subgen. Rubus, Rosaceae)." Journal Article. *Molecular Phylogenetics and Evolution* 89: 13–27. https://doi.org/10.1016/j.ympev.2015.04.007.

Soltis, D. E., R. J. A. Buggs, J. J. Doyle, and P. S. Soltis. 2010. "What We Still Don't Know About Polyploidy." Journal Article. *Taxon* 59 (5): 1387–1403. https://doi.org/10.1002/tax.595006.

Spaniel, S., K. Marhold, B. Filova, and J. Zozomova-Lihova. 2011. "Genetic and Morphological Variation in the Diploid-Polyploid Alyssum Montanum in Central Europe: Taxonomic and Evolutionary Considerations." Journal Article. *Plant Systematics and Evolution* 294 (1-2): 1–25. https://doi.org/10.1007/s00606-011-0438-y.

Spoelhof, J. P., P. S. Soltis, and D. E. Soltis. 2017. "Pure Polyploidy: Closing the Gaps in Autopolyploid Research." Journal Article. *Journal of Systematics and Evolution* 55 (4): 340–52. https://doi.org/10.1111/jse. 12253.

Stace, C.A. 2019. New Flora of the British Isles. Book. 4th ed. C & M Floristics.

Stace, CA, and MJ Crawley. 2015. Alien Plants. Book. William Collins.

Stace, C A., C D. Preston, and D Pearman. 2015. *Hybrid Flora of the British Isles*. Book. Bristol::Botanical Society of Britain & Ireland.

Stenberg, P., M. Lundmark, S. Knutelski, and A. Saura. 2003. "Evolution of Clonality and Polyploidy in a Weevil System." Journal Article. *Molecular Biology and Evolution* 20 (10): 1626–32. https://doi.org/10.1093/molbev/msg180.

Svensson, B. M., and B. A. Carlsson. 2004. "Significance of Time of Attachment, Host Type, and Neighbouring Hemiparasites in Determining Fitness in Two Endangered Grassland Hemiparasites." Journal Article. *Annales Botanici Fennici* 41 (1): 63-75. <Go to ISI>://WOS:000220328700007.

- Thorogood, C. J., F. J. Rumsey, and S. J. Hiscock. 2009. "Host-Specific Races in the Holoparasitic Angiosperm Orobanche Minor: Implications for Speciation in Parasitic Plants." Journal Article. *Annals of Botany* 103 (7): 1005–14. https://doi.org/10.1093/aob/mcp034.
- Tsai, Y. H. E., and P. S. Manos. 2010. "Host Density Drives the Postglacial Migration of the Tree Parasite, Epifagus Virginiana." Journal Article. *Proceedings of the National Academy of Sciences of the United States of America* 107 (39): 17035–40. https://doi.org/10.1073/pnas.1006225107.
- Twyford, A. D. 2018. "Parasitic Plants." Journal Article. Current Biology 28 (16): R857–R859. https://doi.org/10.1016/j.cub.2018.06.030.
- Twyford, A. D., and R. A. Ennos. 2012. "Next-Generation Hybridization and Introgression." Journal Article. *Heredity (Edinb)* 108 (3): 179–89. https://doi.org/10.1038/hdy.2011.68.
- Wang, X. M., G. Gussarova, M. Ruhsam, N. de Vere, C. Metherell, P. M. Hollingsworth, and A. D. Twyford. 2018. "DNA Barcoding a Taxonomically Complex Hemiparasitic Genus Reveals Deep Divergence Between Ploidy Levels but Lack of Species-Level Resolution." Journal Article. *Aob Plants* 10 (3): 13. https://doi.org/10.1093/aobpla/ply026.
- Wendel, J. F. 2015. "The Wondrous Cycles of Polyploidy in Plants." Journal Article. *American Journal of Botany* 102 (11): 1753–6. https://doi.org/10.3732/ajb.1500320.
- Wettstein, R. von. 1896. "Zur Systematik Der Europäischen Euphrasia-Arten." Journal Article. Österreichische Botanische Zeitschrift 46: 381–86.
- Wilkins, D. A. 1963. "PLASTICITY and Establishment in Euphrasia." Journal Article. *Annals of Botany* 27 (107): 533 –&. <Go to ISI>://WOS:A19634440A00013.
- Wilson, J., A. Perry, J. R. Shepherd, M. Duran-Castillo, C. E. Jeffree, and S. Cavers. 2020. "Invasion, Isolation and Evolution Shape Population Genetic Structure in Campanula Rotundifolia." Journal Article. *Aob Plants* 12 (2). https://doi.org/10.1093/aobpla/plaa011.
- Wittzell, H. 1999. "Chloroplast Dna Variation and Reticulate Evolution in Sexual and Apomictic Sections of Dandelions." Journal Article. *Molecular Ecology* 8 (12): 2023–35. https://doi.org/10.1046/j.1365-294x.1999.00807.x.
- Yeo, P.F. 1954. "The Cytology of British Species of Euphrasia." Journal Article. Watsonia 3 (2): 101-8.
- ———. 1956. "Hybridisation Between Diploid and Tetraploid Species of Euphrasia." Journal Article. Watsonia 3: 253–69.
- ——. 1978. "A Taxonomic Revision of Euphrasia in Europe." Journal Article. *Botanical Journal of the Linnean Society* 77: 223–334.
- Yeo, PF. 1964. "The Growth of Euphrasia in Cultivation." Journal Article. Watsonia 6 (1): 1–24.
- ——. 1968. "The Evolutionary Significance of the Speciation of Euphrasia in Europe ." Journal Article. Evolution 22 (4): 736–47.
- Zhou, M. M., G. Q. Yang, G. L. Sun, Z. H. Guo, X. Gong, and Y. Z. Pan. 2020. "Resolving Complicated Relationships of the Panax Bipinnati Fi Dus Complex in Southwestern China by Rad-Seq Data." Journal Article. *Molecular Phylogenetics and Evolution* 149. https://doi.org/10.1016/j.ympev.2020.106851.
- Zohary, D., and U. Nur. 1959. "NATURAL Triploids in the Orchard Grass, Dactylis-Glomerata L Polyploid Complex and Their Significance for Gene Flow from Diploid to Tetraploid Levels." Journal Article. *Evolution* 13 (3): 311–17. https://doi.org/10.2307/2406108.