

Spectra-trait PLSR example using leaf-level spectra and leaf mass per area (LMA) data from 36 species growing in *Rosa rugosa* invaded coastal grassland communities in Belgium

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Overview

This is an R Markdown Notebook to illustrate how to retrieve a dataset from the EcoSIS spectral database, choose the “optimal” number of pls components, and fit a pls model for leaf-mass area (LMA)

Getting Started

Step 1. Load libraries to run example script

```
list.of.packages <- c("pls", "dplyr", "reshape2", "here", "plotrix", "ggplot2", "gridExtra",
                      "spectratrait")
invisible(lapply(list.of.packages, library, character.only = TRUE))

##
## Attaching package: 'pls'
## The following object is masked from 'package:stats':
##
##   loadings
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##   filter, lag
## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
## here() starts at /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction
##
## Attaching package: 'gridExtra'
## The following object is masked from 'package:dplyr':
##
##   combine
```

Step 2. Setup other functions and options

```
### Setup other functions and options
# not in
`%notin%` <- Negate(`%in%`)

# Script options
pls::pls.options(plsralg = "oscorespls")
pls::pls.options("plsralg")

## $plsralg
## [1] "oscorespls"

# Default par options
opar <- par(no.readonly = T)

# What is the target variable?
inVar <- "LMA_g_m2"

# What is the source dataset from EcoSIS?
ecosis_id <- "9db4c5a2-7eac-4e1e-8859-009233648e89"

# Specify output directory, output_dir
# Options:
# tempdir - use a OS-specified temporary directory
# user defined PATH - e.g. "~/scratch/PLSR"
output_dir <- "tempdir"
```

Step 3. Set working directory (scratch space)

```
## [1] "/private/var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjQ/T/RtmpHuvmyI"
```

Step 4. Pull example dataset from EcoSIS (ecosis.org)

```
print(paste0("Output directory: ",getwd())) # check wd

## [1] "Output directory: /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction/vignettes"
### Get source dataset from EcoSIS
dat_raw <- spectratrait::get_ecosis_data(ecosis_id = ecosis_id)

## [1] "**** Downloading Ecosis data ****"

## Downloading data...

##
## -- Column specification -----
## cols(
##   .default = col_double(),
##   `Latin Species` = col_character(),
##   ids = col_character(),
##   `plot code` = col_character(),
##   `species code` = col_character()
## )
```

```
## i Use `spec()` for the full column specifications.
```

```
## Download complete!
```

```
head(dat_raw)
```

```
## # A tibble: 6 x 2,164
##   `Cw/EWT (cm3/cm2` `Latin Species`   `Leaf area (mm2` `Leaf calcium content pe~
##         <dbl> <chr>                   <dbl>                <dbl>
## 1      0.00887 Arrhenatherum el~      696.                0.0291
## 2      0.00824 Bromus sterilis        447.                0.0230
## 3      0.0280  Jacobaea vulgaris      2418.                0.0950
## 4      0.0106  Rubus caesius        5719.                0.0700
## 5      0.00851 Arrhenatherum el~      671.                0.0286
## 6      0.0153  Crepis capillaris      1401.                0.0470
## # ... with 2,160 more variables:
## #   Leaf magnesium content per leaf area (mg/mm2) <dbl>,
## #   Leaf mass per area (g/cm2) <dbl>,
## #   Leaf nitrogen content per leaf area (mg/mm2) <dbl>,
## #   Leaf phosphorus content per leaf area (mg/mm2) <dbl>,
## #   Leaf potassium content per leaf area (mg/mm2) <dbl>,
## #   Plant height vegetative (cm) <dbl>, ids <chr>, plot code <chr>,
## #   species code <chr>, 350 <dbl>, 351 <dbl>, 352 <dbl>, 353 <dbl>, 354 <dbl>,
## #   355 <dbl>, 356 <dbl>, 357 <dbl>, 358 <dbl>, 359 <dbl>, 360 <dbl>,
## #   361 <dbl>, 362 <dbl>, 363 <dbl>, 364 <dbl>, 365 <dbl>, 366 <dbl>,
## #   367 <dbl>, 368 <dbl>, 369 <dbl>, 370 <dbl>, 371 <dbl>, 372 <dbl>,
## #   373 <dbl>, 374 <dbl>, 375 <dbl>, 376 <dbl>, 377 <dbl>, 378 <dbl>,
## #   379 <dbl>, 380 <dbl>, 381 <dbl>, 382 <dbl>, 383 <dbl>, 384 <dbl>,
## #   385 <dbl>, 386 <dbl>, 387 <dbl>, 388 <dbl>, 389 <dbl>, 390 <dbl>,
## #   391 <dbl>, 392 <dbl>, 393 <dbl>, 394 <dbl>, 395 <dbl>, 396 <dbl>,
## #   397 <dbl>, 398 <dbl>, 399 <dbl>, 400 <dbl>, 401 <dbl>, 402 <dbl>,
## #   403 <dbl>, 404 <dbl>, 405 <dbl>, 406 <dbl>, 407 <dbl>, 408 <dbl>,
## #   409 <dbl>, 410 <dbl>, 411 <dbl>, 412 <dbl>, 413 <dbl>, 414 <dbl>,
## #   415 <dbl>, 416 <dbl>, 417 <dbl>, 418 <dbl>, 419 <dbl>, 420 <dbl>,
## #   421 <dbl>, 422 <dbl>, 423 <dbl>, 424 <dbl>, 425 <dbl>, 426 <dbl>,
## #   427 <dbl>, 428 <dbl>, 429 <dbl>, 430 <dbl>, 431 <dbl>, 432 <dbl>,
## #   433 <dbl>, 434 <dbl>, 435 <dbl>, 436 <dbl>, 437 <dbl>, 438 <dbl>,
## #   439 <dbl>, 440 <dbl>, ...
```

```
names(dat_raw)[1:40]
```

```
## [1] "Cw/EWT (cm3/cm2)"
## [2] "Latin Species"
## [3] "Leaf area (mm2)"
## [4] "Leaf calcium content per leaf area (mg/mm2)"
## [5] "Leaf magnesium content per leaf area (mg/mm2)"
## [6] "Leaf mass per area (g/cm2)"
## [7] "Leaf nitrogen content per leaf area (mg/mm2)"
## [8] "Leaf phosphorus content per leaf area (mg/mm2)"
## [9] "Leaf potassium content per leaf area (mg/mm2)"
## [10] "Plant height vegetative (cm)"
## [11] "ids"
## [12] "plot code"
## [13] "species code"
## [14] "350"
## [15] "351"
```

```
## [16] "352"
## [17] "353"
## [18] "354"
## [19] "355"
## [20] "356"
## [21] "357"
## [22] "358"
## [23] "359"
## [24] "360"
## [25] "361"
## [26] "362"
## [27] "363"
## [28] "364"
## [29] "365"
## [30] "366"
## [31] "367"
## [32] "368"
## [33] "369"
## [34] "370"
## [35] "371"
## [36] "372"
## [37] "373"
## [38] "374"
## [39] "375"
## [40] "376"
```

Step 5. Create full plsr dataset

```
### Create plsr dataset
Start.wave <- 500
End.wave <- 2400
wv <- seq(Start.wave,End.wave,1)
Spectra <- as.matrix(dat_raw[,names(dat_raw) %in% wv])
colnames(Spectra) <- c(paste0("Wave_",wv))
sample_info <- dat_raw[,names(dat_raw) %notin% seq(350,2500,1)]
head(sample_info)

## # A tibble: 6 x 13
##   `Cw/EWT (cm3/cm2~` `Latin Species`   `Leaf area (mm2~` `Leaf calcium content pe~
##           <dbl> <chr>                   <dbl>                <dbl>
## 1         0.00887 Arrhenatherum el~         696.                0.0291
## 2         0.00824 Bromus sterilis          447.                0.0230
## 3         0.0280  Jacobaea vulgaris        2418.                0.0950
## 4         0.0106  Rubus caesius          5719.                0.0700
## 5         0.00851 Arrhenatherum el~         671.                0.0286
## 6         0.0153  Crepis capillaris        1401.                0.0470
## # ... with 9 more variables:
## #   Leaf magnesium content per leaf area (mg/mm2) <dbl>,
## #   Leaf mass per area (g/cm2) <dbl>,
## #   Leaf nitrogen content per leaf area (mg/mm2) <dbl>,
## #   Leaf phosphorus content per leaf area (mg/mm2) <dbl>,
## #   Leaf potassium content per leaf area (mg/mm2) <dbl>,
## #   Plant height vegetative (cm) <dbl>, ids <chr>, plot code <chr>,
```

```
## # species code <chr>
sample_info2 <- sample_info %>%
  select(Plant_Species=`Latin Species`,Species_Code=`species code`,Plot=`plot code`,
         LMA_g_cm2=`Leaf mass per area (g/cm2)`)
sample_info2 <- sample_info2 %>%
  mutate(LMA_g_m2=LMA_g_cm2*10000)
head(sample_info2)
```

```
## # A tibble: 6 x 5
##   Plant_Species      Species_Code Plot  LMA_g_cm2 LMA_g_m2
##   <chr>             <chr>      <chr>    <dbl>    <dbl>
## 1 Arrhenatherum elatius Arrela      DC1     0.00342    34.2
## 2 Bromus sterilis      Broste      DC1     0.00282    28.2
## 3 Jacobaea vulgaris     Jacvul      DC1     0.00417    41.7
## 4 Rubus caesius         Rubcae      DC1     0.00566    56.6
## 5 Arrhenatherum elatius Arrela      DC2     0.00361    36.1
## 6 Crepis capillaris     Creves      DC2     0.00283    28.3

plsr_data <- data.frame(sample_info2,Spectra)
rm(sample_info,sample_info2,Spectra)
```

Step 6. Example data cleaning.

```
#### Example data cleaning. End user needs to do what's appropriate for their
#### data. This may be an iterative process.
# Keep only complete rows of inVar and spec data before fitting
plsr_data <- plsr_data[complete.cases(plsr_data[,names(plsr_data) %in%
                                     c(inVar,paste0("Wave_",wv))])],]
```

Step 7. Create cal/val datasets

```
method <- "dplyr" #base/dplyr
# base R - a bit slow
# dplyr - much faster
split_data <- spectratrait::create_data_split(dataset=plsr_data, approach=method, split_seed=7529075,
                                              prop=0.8, group_variables="Species_Code")
names(split_data)
```

```
## [1] "cal_data" "val_data"
```

```
cal.plsr.data <- split_data$cal_data
head(cal.plsr.data)[1:8]
```

```
##      Plant_Species Species_Code Plot  LMA_g_cm2 LMA_g_m2 Wave_500 Wave_501
## 1 Ammophila arenaria      Ammare MC2  0.01679492 167.9492 0.135785 0.13685
## 2 Ammophila arenaria      Ammare WC3  0.01844376 184.4376 0.151750 0.15275
## 3 Ammophila arenaria      Ammare MC4  0.02030190 203.0190 0.156830 0.15790
## 4 Ammophila arenaria      Ammare ZC2  0.01591894 159.1894 0.144450 0.14525
## 5 Ammophila arenaria      Ammare ZC1  0.01483469 148.3469 0.147665 0.14910
## 6 Ammophila arenaria      Ammare ZC3  0.01802409 180.2409 0.130885 0.13175
##      Wave_502
## 1 0.138150
```

```
## 2 0.154150
## 3 0.159065
## 4 0.146220
## 5 0.150330
## 6 0.132750
```

```
val.plsr.data <- split_data$val_data
head(val.plsr.data)[1:8]
```

```
##      Plant_Species Species_Code Plot   LMA_g_cm2 LMA_g_m2   Wave_500
## 184  Jacobaea vulgaris      Jacvul  WC2 0.003551614 35.51614 0.06736887
## 185  Potentilla reptans      Potrep  WC2 0.005586320 55.86320 0.07125000
## 186    Rubus caesius      Rubcae  WC2 0.005803902 58.03902 0.05993560
## 187    Urtica dioica      Urtdio  WC2 0.005215705 52.15705 0.06508300
## 188  Ammophila arenaria    Ammare  WC3 0.018443757 184.43757 0.15175000
## 189  Jacobaea vulgaris      Jacvul  WC3 0.004980002 49.80002 0.06805547
##      Wave_501   Wave_502
## 184 0.06870667 0.07014220
## 185 0.07235000 0.07368350
## 186 0.06162000 0.06352233
## 187 0.06625000 0.06758350
## 188 0.15275000 0.15415000
## 189 0.06938000 0.07093553
```

```
rm(split_data)
```

```
# Datasets:
```

```
print(paste("Cal observations: ",dim(cal.plsr.data)[1],sep=""))
```

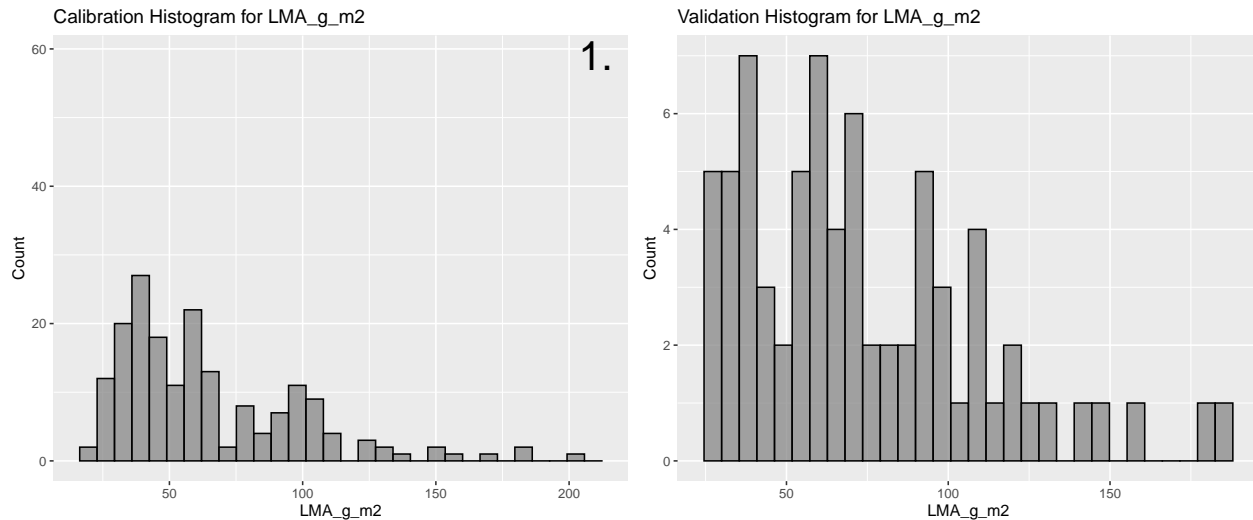
```
## [1] "Cal observations: 183"
```

```
print(paste("Val observations: ",dim(val.plsr.data)[1],sep=""))
```

```
## [1] "Val observations: 73"
```

```
text_loc <- c(max(hist(cal.plsr.data[,paste0(inVar)], plot=FALSE)$counts),
              max(hist(cal.plsr.data[,paste0(inVar)], plot=FALSE)$mids))
cal_hist_plot <- qplot(cal.plsr.data[,paste0(inVar)],geom="histogram",
                      main = paste0("Calibration Histogram for ",inVar),
                      xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),col=I("black"),
                      alpha=I(.7)) +
  annotate("text", x=text_loc[2], y=text_loc[1], label= "1.",size=10)
val_hist_plot <- qplot(val.plsr.data[,paste0(inVar)],geom="histogram",
                      main = paste0("Validation Histogram for ",inVar),
                      xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),col=I("black"),
                      alpha=I(.7))
histograms <- grid.arrange(cal_hist_plot, val_hist_plot, ncol=2)
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
ggsave(filename = file.path(outdir,paste0(inVar,"_Cal_Val_Histograms.png")), plot = histograms,
        device="png", width = 30, height = 12, units = "cm", dpi = 300)
# output cal/val data
write.csv(cal.plsr.data,file=file.path(outdir,paste0(inVar,'_Cal_PLSR_Dataset.csv')),
          row.names=FALSE)
write.csv(val.plsr.data,file=file.path(outdir,paste0(inVar,'_Val_PLSR_Dataset.csv')),
          row.names=FALSE)
```

Step 8. Create calibration and validation PLSR datasets

```
### Format PLSR data for model fitting
cal_spec <- as.matrix(cal.plsr.data[, which(names(cal.plsr.data) %in% paste0("Wave_",wv))])
cal.plsr.data <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% paste0("Wave_",wv))],
                           Spectra=I(cal_spec))
head(cal.plsr.data)[1:5]

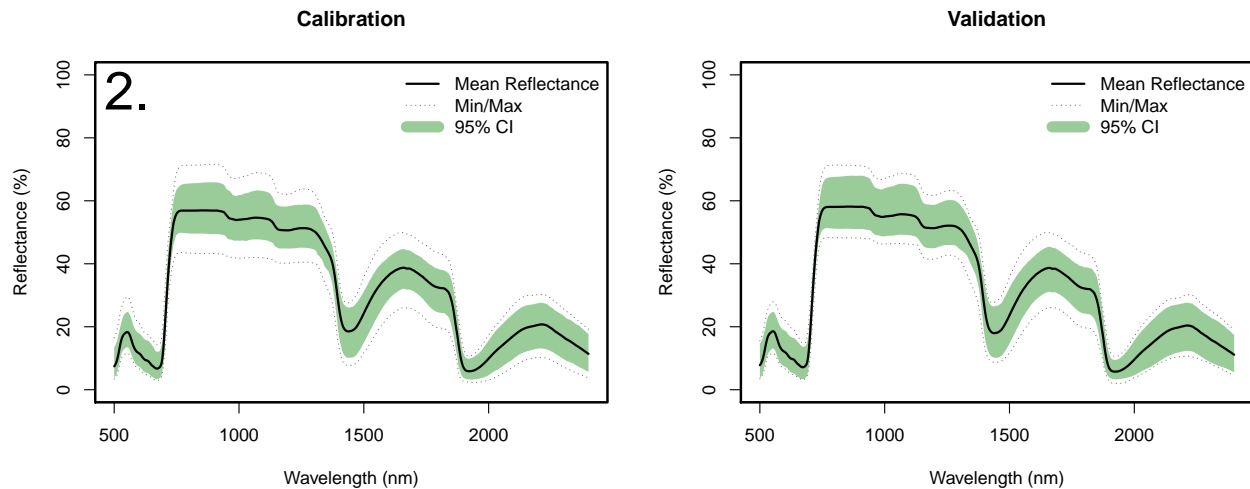
##      Plant_Species Species_Code Plot  LMA_g_cm2 LMA_g_m2
## 1 Ammophila arenaria      Ammare MC2 0.01679492 167.9492
## 2 Ammophila arenaria      Ammare WC3 0.01844376 184.4376
## 3 Ammophila arenaria      Ammare MC4 0.02030190 203.0190
## 4 Ammophila arenaria      Ammare ZC2 0.01591894 159.1894
## 5 Ammophila arenaria      Ammare ZC1 0.01483469 148.3469
## 6 Ammophila arenaria      Ammare ZC3 0.01802409 180.2409

val_spec <- as.matrix(val.plsr.data[, which(names(val.plsr.data) %in% paste0("Wave_",wv))])
val.plsr.data <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% paste0("Wave_",wv))],
                           Spectra=I(val_spec))
head(val.plsr.data)[1:5]

##      Plant_Species Species_Code Plot  LMA_g_cm2 LMA_g_m2
## 184 Jacobaea vulgaris      Jacvul WC2 0.003551614 35.51614
## 185 Potentilla reptans      Potrep WC2 0.005586320 55.86320
## 186 Rubus caesius          Rubcae WC2 0.005803902 58.03902
## 187 Urtica dioica          Urtdio WC2 0.005215705 52.15705
## 188 Ammophila arenaria      Ammare WC3 0.018443757 184.43757
## 189 Jacobaea vulgaris      Jacvul WC3 0.004980002 49.80002
```

Step 9. Calibration and Validation spectra plot

```
par(mfrow=c(1,2)) # B, L, T, R
spectratrait::f.plot.spec(Z=cal.plsr.data$Spectra,wv=wv,
  plot_label="Calibration")
text(550,95,labels = "2.",cex=3)
spectratrait::f.plot.spec(Z=val.plsr.data$Spectra,wv=wv,
  plot_label="Validation")
```



```
dev.copy(png,file.path(outdir,paste0(inVar,'_Cal_Val_Spectra.png')),
  height=2500,width=4900, res=340)
```

```
## quartz_off_screen
## 3
```

```
dev.off();
```

```
## pdf
## 2
```

```
par(mfrow=c(1,1))
```

Step 10. Use permutation to determine the optimal number of components

```
### Use permutation to determine the optimal number of components
if(grepl("Windows", sessionInfo()$running)){
  pls.options(parallel = NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

method <- "firstMin" #pls, firstPlateau, firstMin
random_seed <- 7529075
seg <- 80
maxComps <- 16
iterations <- 50
prop <- 0.70
if (method=="pls") {
```



```

nComps <- spectratrait::find_optimal_components(dataset=cal.plsr.data, method=method,
                                                maxComps=maxComps, seg=seg,
                                                random_seed=random_seed)

print(paste0("*** Optimal number of components: ", nComps))
} else {
  nComps <- spectratrait::find_optimal_components(dataset=cal.plsr.data, method=method,
                                                maxComps=maxComps, iterations=iterations,
                                                seg=seg, prop=prop,
                                                random_seed=random_seed)
}

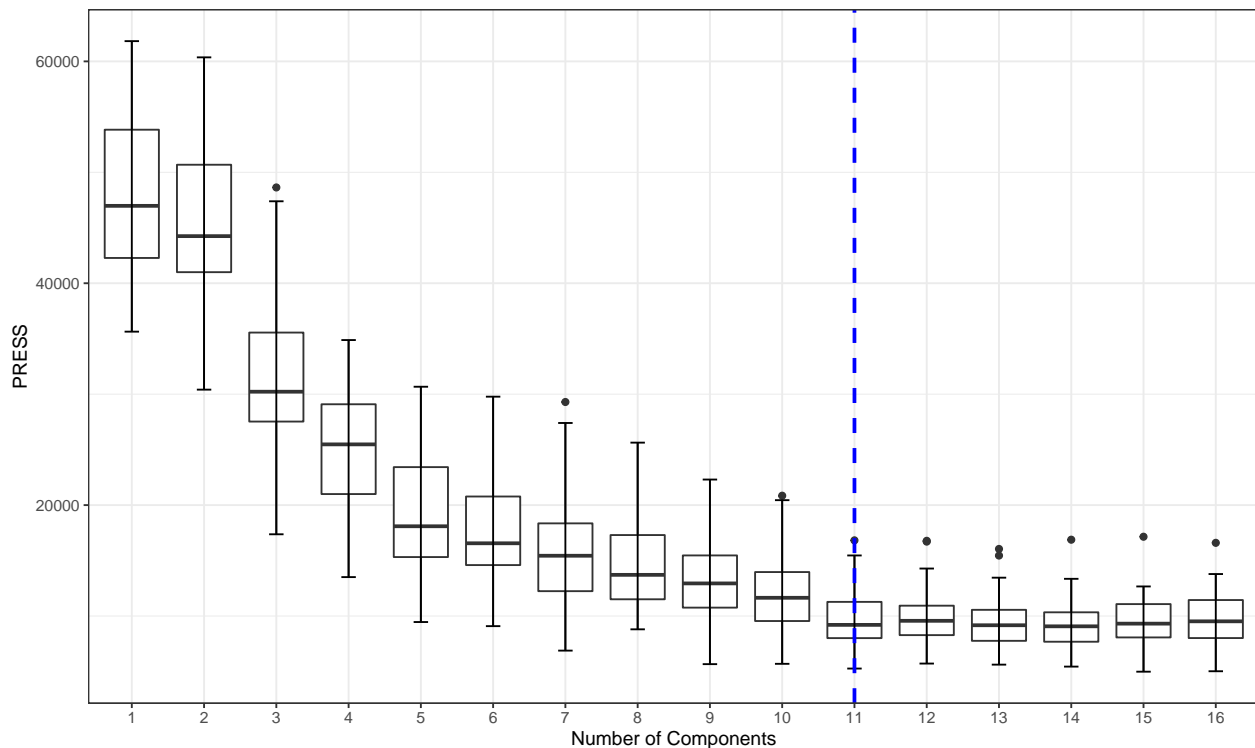
```

```

## [1] "*** Running permutation test. Please hang tight, this can take awhile ***"
## [1] "Options:"
## [1] "Max Components: 16 Iterations: 50 Data Proportion (percent): 70"
## [1] "*** Providing PRESS and coefficient array output ***"

## No id variables; using all as measure variables
## [1] "*** Optimal number of components based on t.test: 11"

```



```

print("*** Figure 3. Optimal PLSR component selection ***")

```

```

## [1] "*** Figure 3. Optimal PLSR component selection ***"

```

```

dev.copy(png,file.path(outdir,paste0(paste0("Figure_3_",inVar,
                                                "_PLSR_Component_Selection.png"))),
        height=2800, width=3400, res=340)

```

```

## quartz_off_screen

```

```

## 3

```

```

dev.off();

```

```
## pdf
## 2
```

Step 11. Fit final model

```
### Fit final model - using leave-one-out cross validation
plsr.out <- plsr(as.formula(paste(inVar,"~","Spectra")),scale=FALSE,ncomp=nComps,validation="LOO",
                trace=FALSE,data=cal.plsr.data)
fit <- plsr.out$fitted.values[,1,nComps]
pls.options(parallel = NULL)

# External validation fit stats
text_loc <- c(max(RMSEP(plsr.out, newdata = val.plsr.data)$comps),
              RMSEP(plsr.out, newdata = val.plsr.data)$val[1])
par(mfrow=c(1,2)) # B, L, T, R
pls::RMSEP(plsr.out, newdata = val.plsr.data)
```

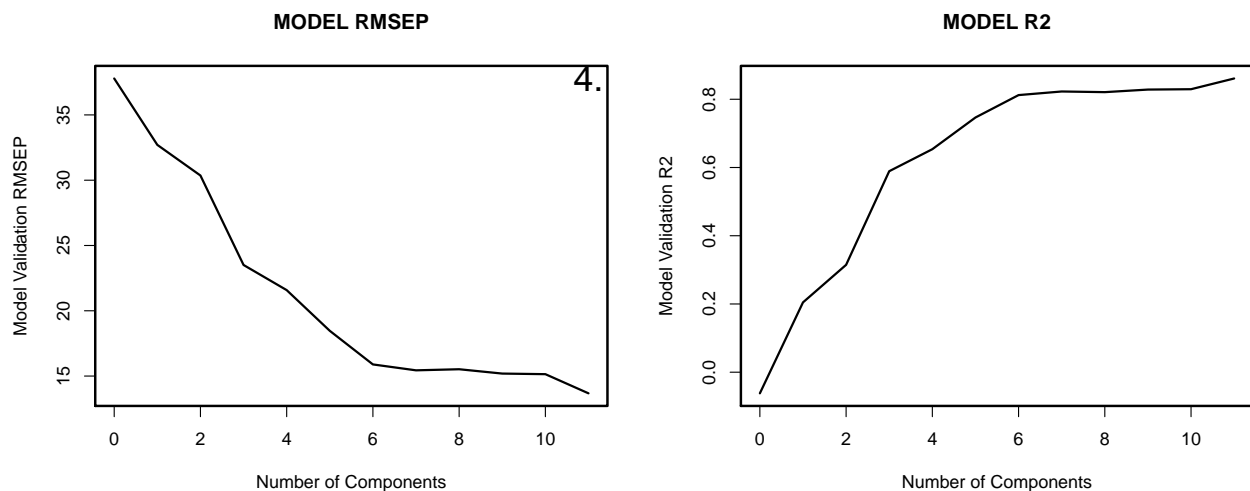
```
## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##      37.79      32.71      30.36      23.51      21.58      18.46
##      6 comps      7 comps      8 comps      9 comps     10 comps     11 comps
##      15.89      15.44      15.52      15.19      15.14      13.68
```

```
plot(pls::RMSEP(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL RMSEP",
     xlab="Number of Components",ylab="Model Validation RMSEP",lty=1,col="black",cex=1.5,lwd=2)
text(text_loc[1],text_loc[2],labels = "4.", cex=2)
box(lwd=2.2)
```

```
pls::R2(plsr.out, newdata = val.plsr.data)
```

```
## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##     -0.06195     0.20461     0.31467     0.58911     0.65365     0.74649
##      6 comps      7 comps      8 comps      9 comps     10 comps     11 comps
##      0.81222     0.82276     0.82084     0.82841     0.82945     0.86090
```

```
plot(pls::R2(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL R2",
     xlab="Number of Components",ylab="Model Validation R2",lty=1,col="black",cex=1.5,lwd=2)
box(lwd=2.2)
```



```

dev.copy(png, file.path(outdir, paste0(paste0(inVar, "_Validation_RMSEP_R2_by_Component.png"))),
         height=2800, width=4800, res=340)

## quartz_off_screen
##      3
dev.off();

## pdf
##      2
par(opar)

```

Step 12. PLSR fit observed vs. predicted plot data

```

#calibration
cal.plsr.output <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% "Spectra")],
                             PLSR_Predicted=fit,
                             PLSR_CV_Predicted=as.vector(plsr.out$validation$pred[, , nComps]))
cal.plsr.output <- cal.plsr.output %>%
  mutate(PLSR_CV_Residuals = PLSR_CV_Predicted-get(inVar))
head(cal.plsr.output)

##      Plant_Species Species_Code Plot  LMA_g_cm2 LMA_g_m2 PLSR_Predicted
## 1 Ammophila arenaria      Ammare MC2 0.01679492 167.9492      154.1892
## 2 Ammophila arenaria      Ammare WC3 0.01844376 184.4376      147.0878
## 3 Ammophila arenaria      Ammare MC4 0.02030190 203.0190      153.8674
## 4 Ammophila arenaria      Ammare ZC2 0.01591894 159.1894      161.6047
## 5 Ammophila arenaria      Ammare ZC1 0.01483469 148.3469      144.9268
## 6 Ammophila arenaria      Ammare ZC3 0.01802409 180.2409      148.2100
##      PLSR_CV_Predicted PLSR_CV_Residuals
## 1          151.7161          -16.233027
## 2          137.3863          -47.051273
## 3          144.2584          -58.760574
## 4          162.6250           3.435614
## 5          142.9101          -5.436767
## 6          142.5160          -37.724928

cal.R2 <- round(pls::R2(plsr.out)[[1]][nComps], 2)
cal.RMSEP <- round(sqrt(mean(cal.plsr.output$PLSR_CV_Residuals^2)), 2)

val.plsr.output <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% "Spectra")],
                             PLSR_Predicted=as.vector(predict(plsr.out,
                                                             newdata = val.plsr.data,
                                                             ncomp=nComps, type="response")[, , 1]))
val.plsr.output <- val.plsr.output %>%
  mutate(PLSR_Residuals = PLSR_Predicted-get(inVar))
head(val.plsr.output)

##      Plant_Species Species_Code Plot  LMA_g_cm2 LMA_g_m2 PLSR_Predicted
## 184 Jacobaea vulgaris      Jacvul WC2 0.003551614 35.51614      43.51586
## 185 Potentilla reptans      Potrep WC2 0.005586320 55.86320      61.41726
## 186 Rubus caesius          Rubcae WC2 0.005803902 58.03902      45.55789
## 187 Urtica dioica          Urtdio WC2 0.005215705 52.15705      46.65139
## 188 Ammophila arenaria      Ammare WC3 0.018443757 184.43757      147.08781

```

```

## 189  Jacobaea vulgaris      Jacvul  WC3 0.004980002  49.80002      53.09532
##      PLSR_Residuals
## 184      7.999719
## 185      5.554059
## 186     -12.481126
## 187     -5.505664
## 188     -37.349758
## 189      3.295298

val.R2 <- round(pls::R2(plsr.out,newdata=val.plsr.data)[[1]][nComps],2)
val.RMSEP <- round(sqrt(mean(val.plsr.output$PLSR_Residuals^2)),2)

rng_quant <- quantile(cal.plsr.output[,inVar], probs = c(0.001, 0.999))
cal_scatter_plot <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                                      rng_quant[2]) +

  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Calibration: ", paste0("Rsqr = ", cal.R2), "; ", paste0("RMSEP = ",
                                                                                      cal.RMSEP))) +

  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5)) +
  annotate("text", x=rng_quant[1], y=rng_quant[2], label= "5.",size=10)

cal_resid_histogram <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +
  geom_vline(xintercept = 0, color="black",
            linetype="dashed", size=1) + theme_bw() +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

rng_quant <- quantile(val.plsr.output[,inVar], probs = c(0.001, 0.999))
val_scatter_plot <- ggplot(val.plsr.output, aes(x=PLSR_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                                      rng_quant[2]) +

  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Validation: ", paste0("Rsqr = ", val.R2), "; ", paste0("RMSEP = ",
                                                                                      val.RMSEP))) +

  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

val_resid_histogram <- ggplot(val.plsr.output, aes(x=PLSR_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +

```

```

geom_vline(xintercept = 0, color="black",
           linetype="dashed", size=1) + theme_bw() +
theme(axis.text=element_text(size=18), legend.position="none",
      axis.title=element_text(size=20, face="bold"),
      axis.text.x = element_text(angle = 0,vjust = 0.5),
      panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

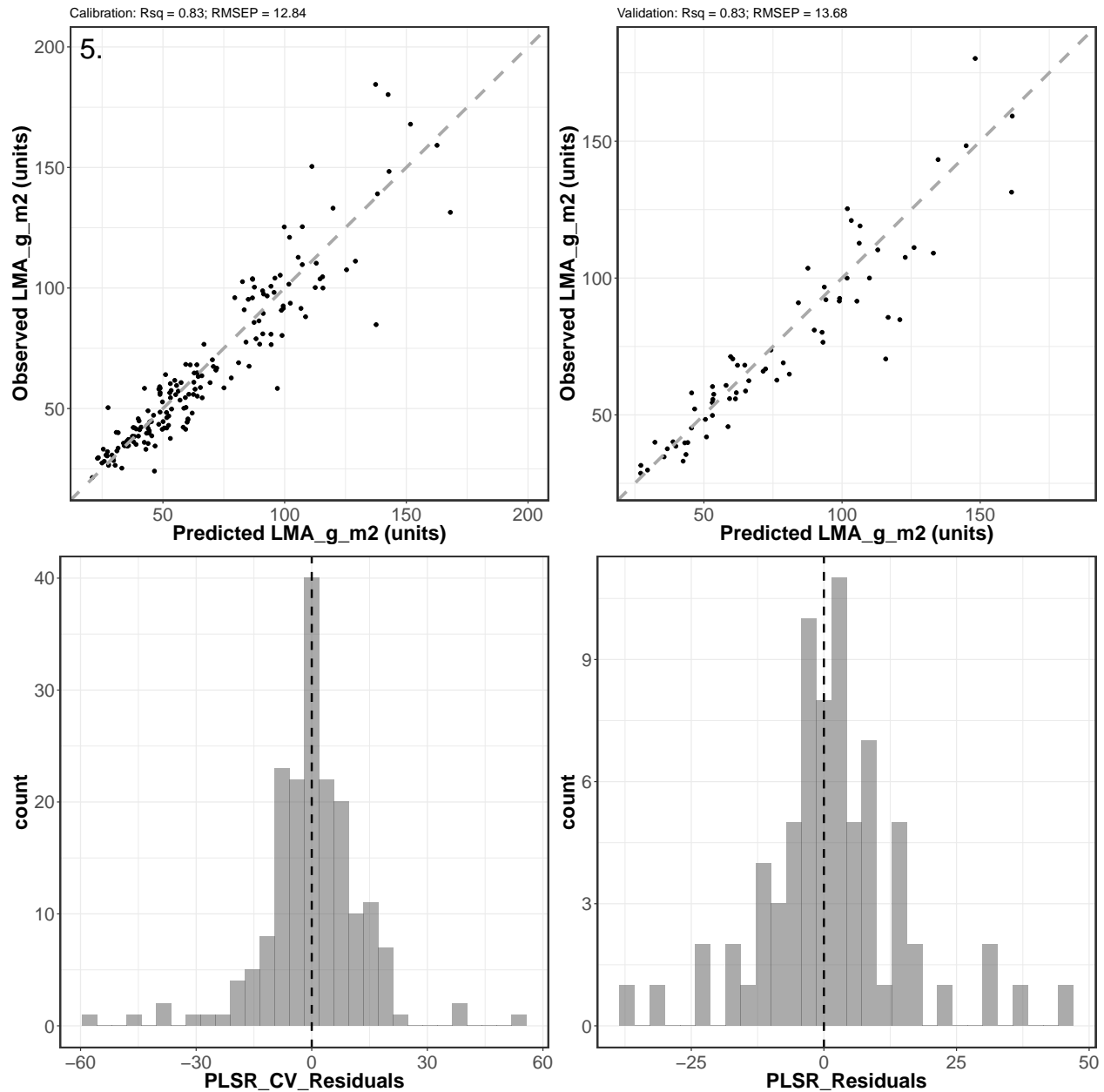
# plot cal/val side-by-side
scatterplots <- grid.arrange(cal_scatter_plot, val_scatter_plot, cal_resid_histogram,
                             val_resid_histogram, nrow=2, ncol=2)

## Warning: Removed 6 rows containing missing values (geom_point).

## Warning: Removed 6 rows containing missing values (geom_point).

## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.

```



```
ggsave(filename = file.path(outdir,paste0(inVar,"_Cal_Val_Scatterplots.png")),
  plot = scatterplots, device="png", width = 32, height = 30, units = "cm",
  dpi = 300)
```

Step 13. Generate Coefficient and VIP plots

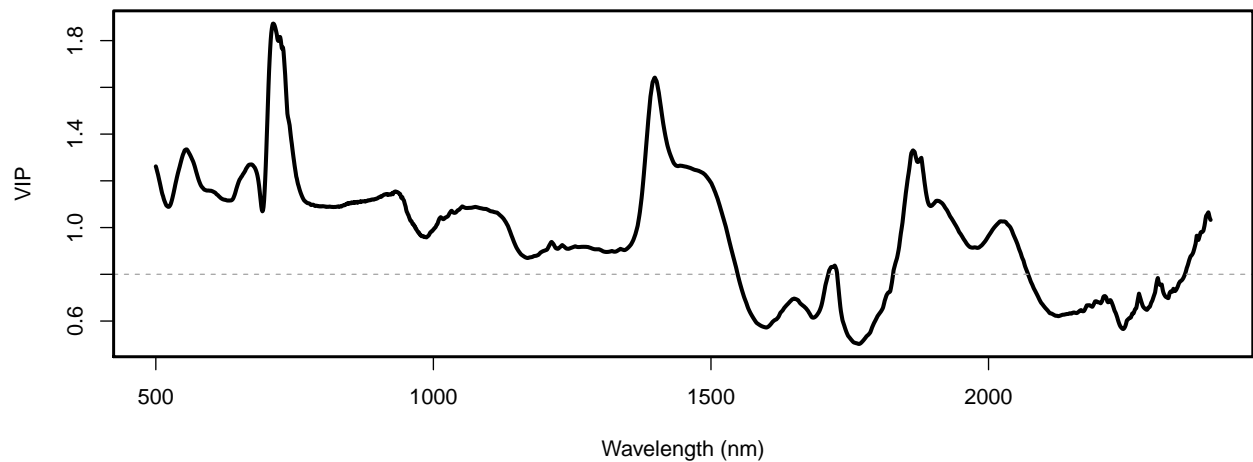
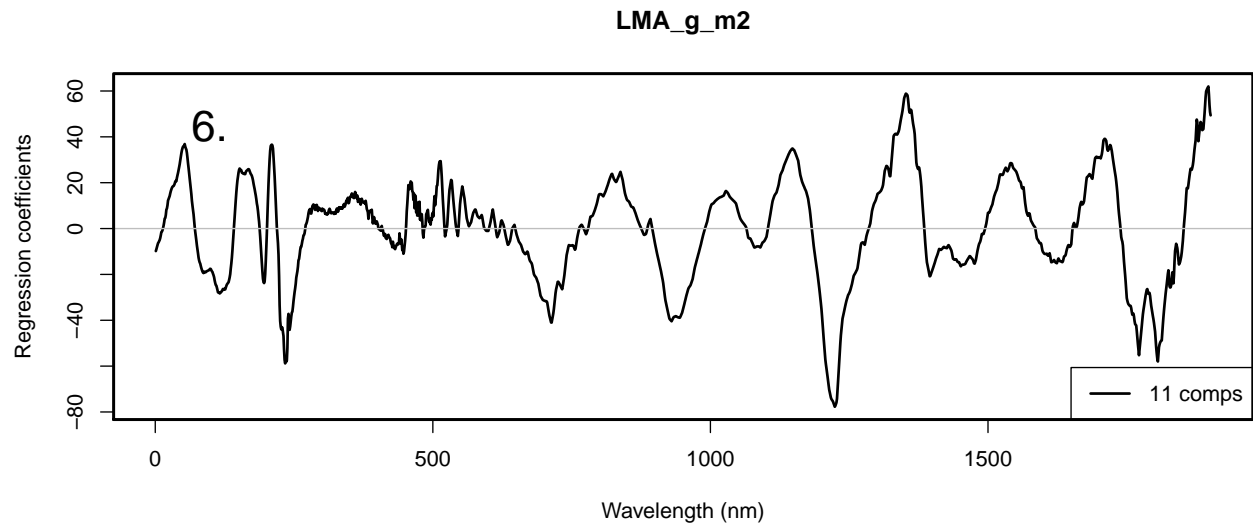
```
vips <- spectratrait::VIP(plsr.out)[nComps,]

par(mfrow=c(2,1))
plot(plsr.out, plottype = "coef",xlab="Wavelength (nm)",
  ylab="Regression coefficients",legendpos = "bottomright",
  ncomp=nComps,lwd=2)
legend("topleft",legend = "6.", cex=2, bty="n")
```

```

box(lwd=2.2)
plot(seq(Start.wave,End.wave,1),vips,xlab="Wavelength (nm)",ylab="VIP",cex=0.01)
lines(seq(Start.wave,End.wave,1),vips,lwd=3)
abline(h=0.8,lty=2,col="dark grey")
box(lwd=2.2)

```



```

dev.copy(png,file.path(outdir,paste0(inVar,'_Coefficient_VIP_plot.png')),
         height=3100, width=4100, res=340)

```

```

## quartz_off_screen
##           3

```

```

dev.off();

```

```

## pdf
##    2

```

Step 14. Permutation analysis to derive uncertainty estimates

```

if(grepl("Windows", sessionInfo()$running)){
  pls.options(parallel=NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

jk.plsr.out <- pls::plsr(as.formula(paste(inVar,"~","Spectra")), scale=FALSE,
                        center=TRUE, ncomp=nComps, validation="LOO", trace=FALSE,
                        jackknife=TRUE,
                        data=cal.plsr.data)
pls.options(parallel = NULL)

Jackknife_coef <- spectratrait::f.coef.valid(plsr.out = jk.plsr.out, data_plsr = cal.plsr.data,
                                             ncomp = nComps, inVar=inVar)
Jackknife_intercept <- Jackknife_coef[1,,]
Jackknife_coef <- Jackknife_coef[2:dim(Jackknife_coef)[1],,,]

interval <- c(0.025,0.975)
Jackknife_Pred <- val.plsr.data$Spectra %*% Jackknife_coef +
  matrix(rep(Jackknife_intercept, length(val.plsr.data[,inVar])), byrow=TRUE,
          ncol=length(Jackknife_intercept))
Interval_Conf <- apply(X = Jackknife_Pred, MARGIN = 1, FUN = quantile,
                      probs=c(interval[1], interval[2]))
sd_mean <- apply(X = Jackknife_Pred, MARGIN = 1, FUN = sd)
sd_res <- sd(val.plsr.output$PLSR_Residuals)
sd_tot <- sqrt(sd_mean^2+sd_res^2)
val.plsr.output$LCI <- Interval_Conf[1,]
val.plsr.output$UCI <- Interval_Conf[2,]
val.plsr.output$LPI <- val.plsr.output$PLSR_Predicted-1.96*sd_tot
val.plsr.output$UPI <- val.plsr.output$PLSR_Predicted+1.96*sd_tot
head(val.plsr.output)

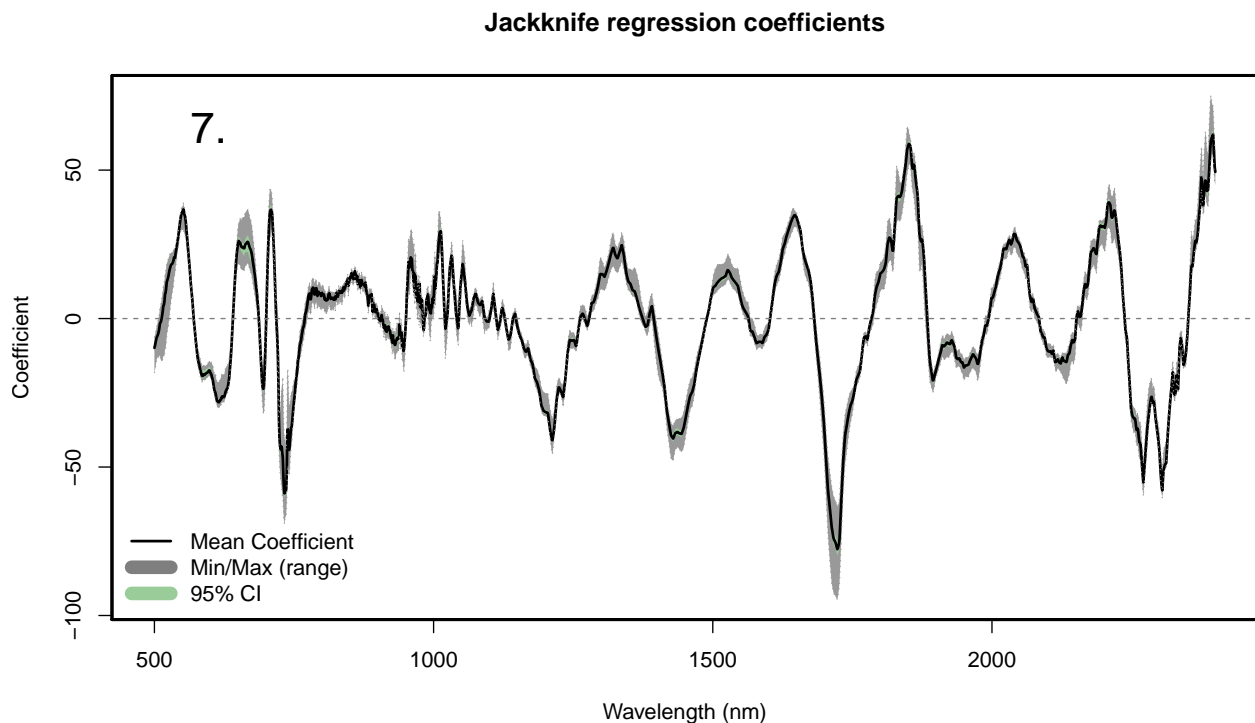
##          Plant_Species Species_Code Plot   LMA_g_cm2  LMA_g_m2 PLSR_Predicted
## 184  Jacobaea vulgaris      Jacvul  WC2 0.003551614  35.51614      43.51586
## 185  Potentilla reptans      Potrep  WC2 0.005586320  55.86320      61.41726
## 186    Rubus caesius      Rubcae  WC2 0.005803902  58.03902      45.55789
## 187    Urtica dioica      Urtdio  WC2 0.005215705  52.15705      46.65139
## 188  Ammophila arenaria      Ammare  WC3 0.018443757 184.43757     147.08781
## 189  Jacobaea vulgaris      Jacvul  WC3 0.004980002  49.80002      53.09532
##      PLSR_Residuals      LCI      UCI      LPI      UPI
## 184      7.999719  42.58086  44.15724  16.70642  70.32530
## 185      5.554059  60.10507  62.52674  34.59536  88.23916
## 186     -12.481126  44.66849  48.22967  18.70489  72.41090
## 187      -5.505664  45.70375  47.84938  19.82512  73.47765
## 188     -37.349758 145.09309 148.61694 120.18052 173.99510
## 189      3.295298  52.40880  53.97806  26.28498  79.90565

### Permutation coefficient plot
spectratrait::f.plot.coef(Z = t(Jackknife_coef), ww = ww,
                          plot_label="Jackknife regression coefficients",position = 'bottomleft')
abline(h=0,lty=2,col="grey50")
legend("topleft",legend = "7.", cex=2, bty="n")

```



```
box(lwd=2.2)
```



```
dev.copy(png,file.path(outdir,paste0(inVar,'_Jackknife_Regression_Coefficients.png')),
         height=2100, width=3800, res=340)
```

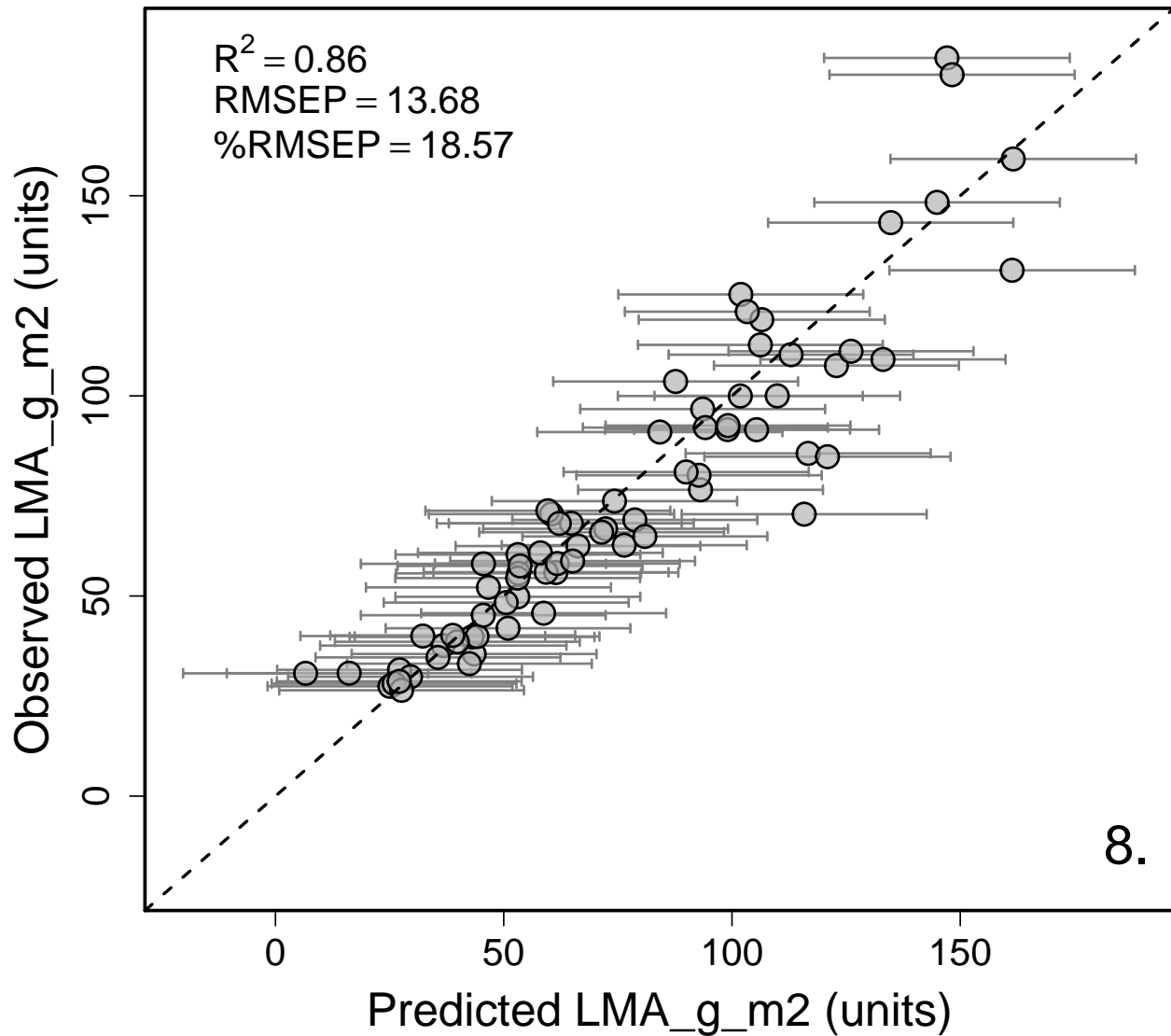
```
## quartz_off_screen
##           3
```

```
dev.off();
```

```
## pdf
##    2
```

```
#### Permutation validation plot
RMSEP <- sqrt(mean(val.plsr.output$PLSR_Residuals^2))
pecr_RMSEP <- RMSEP/mean(val.plsr.output[,inVar])*100
r2 <- round(pls::R2(plsr.out, newdata = val.plsr.data)$val[nComps+1],2)
expr <- vector("expression", 3)
expr[[1]] <- bquote(R^2==.(r2))
expr[[2]] <- bquote(RMSEP==.(round(RMSEP,2)))
expr[[3]] <- bquote("%RMSEP"==.(round(pecr_RMSEP,2)))
rng_vals <- c(min(val.plsr.output$LPI), max(val.plsr.output$UPI))
par(mfrow=c(1,1), mar=c(4.2,5.3,1,0.4), oma=c(0, 0.1, 0, 0.2))
plotrix::plotCI(val.plsr.output$PLSR_Predicted,val.plsr.output[,inVar],
  li=val.plsr.output$LPI, ui=val.plsr.output$UPI, gap=0.009,sfrac=0.004,
  lwd=1.6, xlim=c(rng_vals[1], rng_vals[2]), ylim=c(rng_vals[1], rng_vals[2]),
  err="x", pch=21, col="black", pt.bg=scales::alpha("grey70",0.7), scol="grey50",
  cex=2, xlab=paste0("Predicted ", paste(inVar), " (units)"),
  ylab=paste0("Observed ", paste(inVar), " (units)"),
  cex.axis=1.5,cex.lab=1.8)
abline(0,1,lty=2,lw=2)
legend("topleft", legend=expr, bty="n", cex=1.5)
```

```
legend("bottomright", legend="8.", bty="n", cex=2.2)
box(lwd=2.2)
```



```
dev.copy(png, file.path(outdir, paste0(inVar, "_PLSR_Validation_Scatterplot.png")),
         height=2800, width=3200, res=340)
```

```
## quartz_off_screen
## 3
```

```
dev.off();
```

```
## pdf
## 2
```

Step 15. Output permutation coefficients for later use

```
out.jk.coefs <- data.frame(Iteration=seq(1, length(Jackknife_intercept), 1),
                          Intercept=Jackknife_intercept, t(Jackknife_coef))
```

```
head(out.jk.coefs)[1:6]
```

```
##      Iteration Intercept   Wave_500   Wave_501   Wave_502   Wave_503
## Seg 1          1  18.33909  -7.580446  -6.724083  -5.886226  -4.984744
## Seg 2          2  21.22164  -8.574931  -7.084795  -6.255716  -5.384000
## Seg 3          3  19.63843 -18.104491 -17.260522 -16.154983 -14.960119
## Seg 4          4  15.90905 -10.715594  -9.874766  -8.926979  -8.007834
## Seg 5          5  17.51805  -8.952143  -8.305344  -7.136167  -6.221407
## Seg 6          6  12.18563  -7.702160  -7.128890  -6.532276  -5.840220
```

```
write.csv(out.jk.coefs,file=file.path(outdir,
                                     paste0(inVar,'_Jackkife_PLSR_Coefficients.csv')),
          row.names=FALSE)
```

Step 16. Output remaining core PLSR outputs

```
print(paste("Output directory: ", outdir))
```

```
## [1] "Output directory: /var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjgq/T//RtmpHuvmyI"
```

```
# Observed versus predicted
```

```
write.csv(cal.plsr.output,file=file.path(outdir,
                                     paste0(inVar,'_Observed_PLSR_CV_Pred_',
                                             nComps,'comp.csv')),
          row.names=FALSE)
```

```
# Validation data
```

```
write.csv(val.plsr.output,file=file.path(outdir,
                                     paste0(inVar,'_Validation_PLSR_Pred_',
                                             nComps,'comp.csv')),
          row.names=FALSE)
```

```
# Model coefficients
```

```
coefs <- coef(plsr.out,ncomp=nComps,intercept=TRUE)
write.csv(coefs,file=file.path(outdir,
                              paste0(inVar,'_PLSR_Coefficients_',
                                      nComps,'comp.csv')),
          row.names=TRUE)
```

```
# PLSR VIP
```

```
write.csv(vips,file=file.path(outdir,
                              paste0(inVar,'_PLSR_VIPs_',
                                      nComps,'comp.csv')))
```

Step 17. Confirm files were written to temp space

```
print("**** PLSR output files: ")
```

```
## [1] "**** PLSR output files: "
```

```
print(list.files(outdir)[grep(pattern = inVar, list.files(outdir))])
```

```
## [1] "Figure_3_LMA_g_m2_PLSR_Component_Selection.png"
```

```
## [2] "LMA_g_m2_Cal_PLSR_Dataset.csv"
## [3] "LMA_g_m2_Cal_Val_Histograms.png"
## [4] "LMA_g_m2_Cal_Val_Scatterplots.png"
## [5] "LMA_g_m2_Cal_Val_Spectra.png"
## [6] "LMA_g_m2_Coefficient_VIP_plot.png"
## [7] "LMA_g_m2_Jackknife_PLSR_Coefficients.csv"
## [8] "LMA_g_m2_Jackknife_Regression_Coefficients.png"
## [9] "LMA_g_m2_Observed_PLSR_CV_Pred_11comp.csv"
## [10] "LMA_g_m2_PLSR_Coefficients_11comp.csv"
## [11] "LMA_g_m2_PLSR_Validation_Scatterplot.png"
## [12] "LMA_g_m2_PLSR_VIPs_11comp.csv"
## [13] "LMA_g_m2_Val_PLSR_Dataset.csv"
## [14] "LMA_g_m2_Validation_PLSR_Pred_11comp.csv"
## [15] "LMA_g_m2_Validation_RMSEP_R2_by_Component.png"
```