Spectra-trait PLSR example using leaf-level spectra and leaf mass per area (LMA) data from more than 40 species grassland species comprising both herbs and graminoids.

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Overview

This is an R Markdown Notebook to illustrate how to retrieve a dataset from the EcoSIS spectral database, choose the "optimal" number of plsr components, and fit a plsr model for leaf-mass area (LMA). In this example, the plants were cultivated in an outdoor setting in the botanical garden of the KIT using 40x40 cm pots with an standardized substrate. The data was measured on a weekly basis (the timestamp is included in the dataset).

Getting Started

Installation

```
### Install and load required R packages
list.of.packages <- c("devtools","readr","RCurl","httr","pls","dplyr","reshape2","here",
                      "ggplot2","gridExtra")  # packages needed for script
# check for dependencies and install if needed
new.packages <- list.of.packages[!(list.of.packages %in% installed.packages()[,"Package"])]
if(length(new.packages)) install.packages(new.packages)

# Load libraries
invisible(lapply(list.of.packages, library, character.only = TRUE))
```

```
## Loading required package: usethis
```

```
##
## Attaching package: 'pls'
```

```
## The following object is masked from 'package:stats':
##
##     loadings
```

```
##
## Attaching package: 'dplyr'
```

```
## The following objects are masked from 'package:stats':
##
##     filter, lag
```

```
## The following objects are masked from 'package:base':
##
##     intersect, setdiff, setequal, union
```

```
## here() starts at /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction
```

```
##
## Attaching package: 'gridExtra'
```

```
## The following object is masked from 'package:dplyr':
##
##     combine
```

Setup other functions and options

```
### Setup other functions and options
github_dir <- file.path(here::here(),"R_Scripts")
source_from_gh <- FALSE
if (source_from_gh) {
  # Source helper functions from GitHub
  devtools::source_url("https://raw.githubusercontent.com/TESTgroup-BNL/PLSR_for_plant_trait_prediction/
} else {
  functions <- file.path(github_dir,"functions.R")
  source(functions)
}

# not in
`%notin%` <- Negate(`%in%`)

# Script options
pls::pls.options(plsralg = "oscorespls")
pls::pls.options("plsralg")
```

```
## $plsralg
## [1] "oscorespls"
```

```
# Default par options
opar <- par(no.readonly = T)

# What is the target variable?
inVar <- "SLA_g_cm"

# What is the source dataset from EcoSIS?
ecosis_id <- "3cf6b27e-d80e-4bc7-b214-c95506e46daa"
```

Set working directory (scratch space)

```
## [1] "Output directory: /private/var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjq/T/Rtmp5X9H9j"
```

Grab data from EcoSIS

```
print(paste0("Output directory: ",getwd()))  # check wd
```

URL: https://ecosis.org/package/fresh-leaf-spectra-to-estimate-lma-over-neon-domains-in-eastern-united-states

```
## [1] "Output directory: /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction/vignettes"
```

```
### Get source dataset from EcoSIS
dat_raw <- get_ecosis_data(ecosis_id = ecosis_id)
```

```
## [1] "**** Downloading Ecosis data ****"
```

```
## Downloading data...
```

```
## Parsed with column specification:
## cols(
##   .default = col_double(),
##   `growth form` = col_character(),
##   species = col_character(),
##   timestamp = col_character()
## )
```

```
## See spec(...) for full column specifications.
```

```
## Download complete!
```

```
head(dat_raw)
```

```
## # A tibble: 6 x 2,114
##   `Anthocyanin co~ `Anthocyanin co~ `Carotenoid con~ `Carotenoid con~
##              <dbl>            <dbl>            <dbl>            <dbl>
## 1          0.00106            0.997          0.00799             7.49
## 2          0.00357            1.22           0.0221              7.53
## 3          0.00252            1.14           0.0188              8.55
## 4          0.00310            2.26           0.0158             11.5
## 5          0.00412            1.73           0.0216              9.08
## 6          0.00397            1.02           0.0336              8.66
## # ... with 2,110 more variables: `Chlorophyll concentration (mg/g)` <dbl>,
## #   `Chlorophyll content ( g/cm )` <dbl>, `LDMC (g/g)` <dbl>, `LFA (mg/cm
## #   )` <dbl>, `LWC (mg/cm )` <dbl>, `SLA (g/cm )` <dbl>, `growth form` <chr>,
## #   species <chr>, timestamp <chr>, `400` <dbl>, `401` <dbl>, `402` <dbl>,
## #   `403` <dbl>, `404` <dbl>, `405` <dbl>, `406` <dbl>, `407` <dbl>,
## #   `408` <dbl>, `409` <dbl>, `410` <dbl>, `411` <dbl>, `412` <dbl>,
## #   `413` <dbl>, `414` <dbl>, `415` <dbl>, `416` <dbl>, `417` <dbl>,
## #   `418` <dbl>, `419` <dbl>, `420` <dbl>, `421` <dbl>, `422` <dbl>,
## #   `423` <dbl>, `424` <dbl>, `425` <dbl>, `426` <dbl>, `427` <dbl>,
## #   `428` <dbl>, `429` <dbl>, `430` <dbl>, `431` <dbl>, `432` <dbl>,
## #   `433` <dbl>, `434` <dbl>, `435` <dbl>, `436` <dbl>, `437` <dbl>,
## #   `438` <dbl>, `439` <dbl>, `440` <dbl>, `441` <dbl>, `442` <dbl>,
## #   `443` <dbl>, `444` <dbl>, `445` <dbl>, `446` <dbl>, `447` <dbl>,
## #   `448` <dbl>, `449` <dbl>, `450` <dbl>, `451` <dbl>, `452` <dbl>,
## #   `453` <dbl>, `454` <dbl>, `455` <dbl>, `456` <dbl>, `457` <dbl>,
## #   `458` <dbl>, `459` <dbl>, `460` <dbl>, `461` <dbl>, `462` <dbl>,
## #   `463` <dbl>, `464` <dbl>, `465` <dbl>, `466` <dbl>, `467` <dbl>,
## #   `468` <dbl>, `469` <dbl>, `470` <dbl>, `471` <dbl>, `472` <dbl>,
## #   `473` <dbl>, `474` <dbl>, `475` <dbl>, `476` <dbl>, `477` <dbl>,
## #   `478` <dbl>, `479` <dbl>, `480` <dbl>, `481` <dbl>, `482` <dbl>,
## #   `483` <dbl>, `484` <dbl>, `485` <dbl>, `486` <dbl>, `487` <dbl>,
## #   `488` <dbl>, `489` <dbl>, `490` <dbl>, ...
```

```
names(dat_raw)[1:40]
```

```
##  [1] "Anthocyanin concentration (mg/g)" "Anthocyanin content ( g/cm )"
##  [3] "Carotenoid concentration (mg/g)"  "Carotenoid content ( g/cm )"
```

```
##  [5] "Chlorophyll concentration (mg/g)" "Chlorophyll content ( g/cm )"
##  [7] "LDMC (g/g)"                       "LFA (mg/cm )"
##  [9] "LWC (mg/cm )"                     "SLA (g/cm )"
## [11] "growth form"                      "species"
## [13] "timestamp"                        "400"
## [15] "401"                              "402"
## [17] "403"                              "404"
## [19] "405"                              "406"
## [21] "407"                              "408"
## [23] "409"                              "410"
## [25] "411"                              "412"
## [27] "413"                              "414"
## [29] "415"                              "416"
## [31] "417"                              "418"
## [33] "419"                              "420"
## [35] "421"                              "422"
## [37] "423"                              "424"
## [39] "425"                              "426"
```

Create full plsr dataset

```
### Create plsr dataset
## cleanup any missing
if (any(is.na(dat_raw))) {
  dat_raw <- na.omit(dat_raw)
}
# remove suspect high values
dat_raw <- dat_raw %>%
  filter(`SLA (g/cm )`<=500)

Start.wave <- 500
End.wave <- 2400
wv <- seq(Start.wave,End.wave,1)
Spectra <- as.matrix(dat_raw[,names(dat_raw) %in% wv])
colnames(Spectra) <- c(paste0("Wave_",wv))
sample_info <- dat_raw[,names(dat_raw) %notin% seq(350,2500,1)]
head(sample_info)
```

```
## # A tibble: 6 x 13
##   `Anthocyanin co~ `Anthocyanin co~ `Carotenoid con~ `Carotenoid con~
##              <dbl>            <dbl>            <dbl>            <dbl>
## 1          0.00106            0.997          0.00799             7.49
## 2          0.00357            1.22           0.0221              7.53
## 3          0.00252            1.14           0.0188              8.55
## 4          0.00310            2.26           0.0158             11.5
## 5          0.00412            1.73           0.0216              9.08
## 6          0.00397            1.02           0.0336              8.66
## # ... with 9 more variables: `Chlorophyll concentration (mg/g)` <dbl>,
## #   `Chlorophyll content ( g/cm )` <dbl>, `LDMC (g/g)` <dbl>, `LFA (mg/cm
## #   )` <dbl>, `LWC (mg/cm )` <dbl>, `SLA (g/cm )` <dbl>, `growth form` <chr>,
## #   species <chr>, timestamp <chr>
```

```
sample_info2 <- sample_info %>%
  select(Plant_Species=species,Growth_Form=`growth form`,timestamp,SLA_g_cm=`SLA (g/cm )`)
head(sample_info2)
```

```
## # A tibble: 6 x 4
##   Plant_Species          Growth_Form timestamp       SLA_g_cm
##   <chr>                  <chr>       <chr>              <dbl>
## 1 Calamagrostis epigejos graminoid   5/25/2016 12:20     107.
## 2 Anthoxanthum odoratum  graminoid   5/27/2016 8:40      293.
## 3 Alopecurus pratensis   graminoid   5/27/2016 9:23      220.
## 4 Festuca ovina          graminoid   5/27/2016 9:23      137.
## 5 Agrostis capillaris    graminoid   5/27/2016 9:42      237.
## 6 Aegopodium podagraria  forb        5/25/2016 12:20     388.
```

```
plsr_data <- data.frame(sample_info2,Spectra)
rm(sample_info,sample_info2,Spectra)
```

Create cal/val datasets

```
### Create cal/val datasets
## Make a stratified random sampling in the strata USDA_Species_Code and Domain

method <- "base" #base/dplyr
# base R - a bit slow
# dplyr - much faster
split_data <- create_data_split(approach=method, split_seed=2356812, prop=0.8,
                                group_variables="Plant_Species")
```

```
## Calamagrostis epigejos   Cal: 80%
## Anthoxanthum odoratum   Cal: 80%
## Alopecurus pratensis   Cal: 80%
## Festuca ovina   Cal: 78.9473684210526%
## Agrostis capillaris   Cal: 82.3529411764706%
## Aegopodium podagraria   Cal: 80%
## Arrhenatherum elatius   Cal: 82.3529411764706%
## Arctium lappa   Cal: 83.3333333333333%
## Urtica dioica   Cal: 78.9473684210526%
## Cirsium arvense   Cal: 80%
## Geranium pratense   Cal: 81.25%
## Geum urbanum   Cal: 80%
## Digitalis purpurea   Cal: 81.25%
## Stellaria media   Cal: 77.7777777777778%
## Trisetum flavescens   Cal: 80%
## Trifolium pratense   Cal: 80.9523809523809%
## Geranium robertianum   Cal: 78.5714285714286%
```

```
## Plantago major   Cal: 85.7142857142857%
## Nardus stricta   Cal: 78.9473684210526%
## Lamium purpureum   Cal: 77.7777777777778%
## Clinopodium vulgare   Cal: 78.5714285714286%
## Poa annua   Cal: 75%
## Campanula rotundifolia   Cal: 78.5714285714286%
## Taraxacum spec.   Cal: 80%
## Digitaria sanguinalis   Cal: 85.7142857142857%
## Holcus lanatus   Cal: 82.3529411764706%
## Lapsana communis   Cal: 75%
## Apera spica-venti   Cal: 80%
## Alopecurus geniculatus   Cal: 75%
## Bromus hordeaceus   Cal: 80%
## Phalaris arundinaceae   Cal: 81.25%
## Thlaspi arvense Not enough observations
## Origanum vulgare   Cal: 77.7777777777778%
## Pulicaria dysenterica   Cal: 79.1666666666667%
## Deschampsia cespitosa   Cal: 80%
## Cirsium acaule   Cal: 80%
## Brachypodium sylvaticum   Cal: 80%
## Centaurium erythraea   Cal: 77.7777777777778%
## Luzula multiflora   Cal: 78.5714285714286%
## Filipendula ulmaria   Cal: 78.5714285714286%
## Anthyllis vulneraria   Cal: 75%
## Medicago lupulina   Cal: 75%
## Succisa pratensis   Cal: 83.3333333333333%
## Scirpus sylvaticus   Cal: 77.7777777777778%
## Molinia caerulea   Cal: 83.3333333333333%
```

```
names(split_data)
```

```
## [1] "cal_data" "val_data"
```

```
cal.plsr.data <- split_data$cal_data
head(cal.plsr.data)[1:8]
```

```
##            Plant_Species Growth_Form       timestamp SLA_g_cm   Wave_500
## 1 Calamagrostis epigejos   graminoid 5/25/2016 12:20 106.6500 0.09180559
## 2  Anthoxanthum odoratum   graminoid  5/27/2016 8:40 293.3565 0.09022668
## 3   Alopecurus pratensis   graminoid  5/27/2016 9:23 220.2703 0.07998340
## 4          Festuca ovina   graminoid  5/27/2016 9:23 137.1220 0.05205080
## 5    Agrostis capillaris   graminoid  5/27/2016 9:42 237.4237 0.06695127
```

```
## 6  Aegopodium podagraria        forb 5/25/2016 12:20 388.2384 0.04091566
##     Wave_501   Wave_502   Wave_503
## 1 0.09293251 0.09417092 0.09552863
## 2 0.09125158 0.09237300 0.09359694
## 3 0.08109460 0.08231389 0.08365015
## 4 0.05256869 0.05314560 0.05378355
## 5 0.06766205 0.06845248 0.06932220
## 6 0.04169865 0.04257613 0.04355737
```

```
val.plsr.data <- split_data$val_data
head(val.plsr.data)[1:8]
```

```
##            Plant_Species Growth_Form       timestamp SLA_g_cm   Wave_500
## 9          Urtica dioica        forb 5/25/2016 12:37 284.6788 0.04716736
## 14       Stellaria media        forb 5/25/2016 13:21 418.4284 0.05694278
## 21  Alopecurus pratensis   graminoid  6/1/2016 11:32 218.2117 0.08135086
## 41  Alopecurus pratensis   graminoid   6/8/2016 8:37 216.7568 0.10062342
## 43   Agrostis capillaris   graminoid   6/8/2016 9:05 231.5292 0.08099724
## 44 Aegopodium podagraria        forb   6/7/2016 9:05 311.4018 0.03778815
##      Wave_501   Wave_502   Wave_503
## 9  0.04781633 0.04854276 0.04935320
## 14 0.05811729 0.05940497 0.06080936
## 21 0.08249180 0.08373915 0.08509719
## 41 0.10190706 0.10330054 0.10480538
## 43 0.08178586 0.08265099 0.08360108
## 44 0.03845043 0.03919155 0.04001581
```

```
rm(split_data)

# Datasets:
print(paste("Cal observations: ",dim(cal.plsr.data)[1],sep=""))
```

```
## [1] "Cal observations: 490"
```

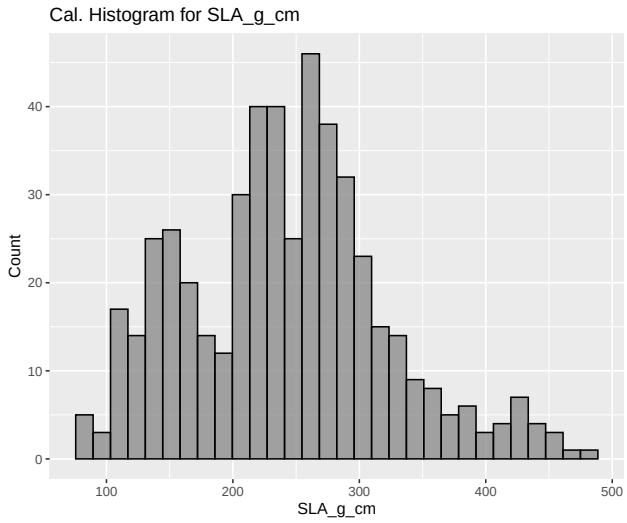
```
print(paste("Val observations: ",dim(val.plsr.data)[1],sep=""))
```

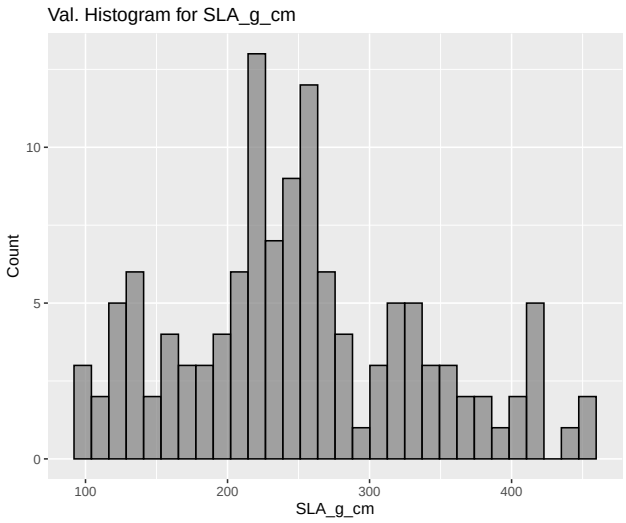
```
## [1] "Val observations: 124"
```

```
cal_hist_plot <- qplot(cal.plsr.data[,paste0(inVar)],geom="histogram",
                       main = paste0("Cal. Histogram for ",inVar),
                       xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),
                       col=I("black"),alpha=I(.7))
val_hist_plot <- qplot(val.plsr.data[,paste0(inVar)],geom="histogram",
                       main = paste0("Val. Histogram for ",inVar),
                       xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),
                       col=I("black"),alpha=I(.7))
grid.arrange(cal_hist_plot, val_hist_plot, ncol=2)
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```





Create calibration and validation PLSR datasets

```
### Format PLSR data for model fitting
cal_spec <- as.matrix(cal.plsr.data[, which(names(cal.plsr.data) %in% paste0("Wave_",wv))])
cal.plsr.data <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% paste0("Wave_",wv))],
                            Spectra=I(cal_spec))
head(cal.plsr.data)[1:5]
```

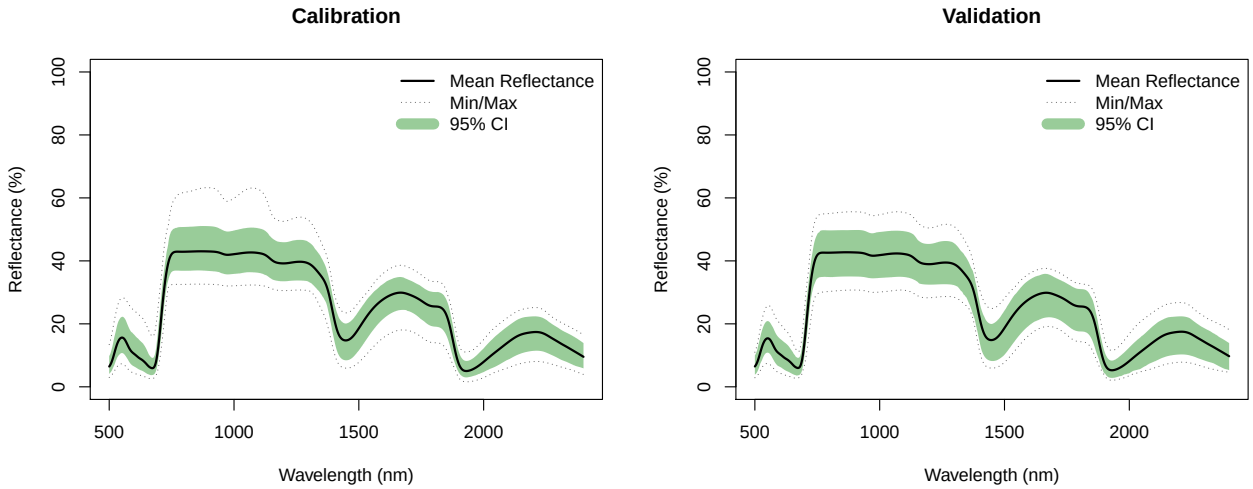
```
##            Plant_Species Growth_Form       timestamp SLA_g_cm CalVal
## 1 Calamagrostis epigejos   graminoid 5/25/2016 12:20 106.6500    Cal
## 2  Anthoxanthum odoratum   graminoid  5/27/2016 8:40 293.3565    Cal
## 3   Alopecurus pratensis   graminoid  5/27/2016 9:23 220.2703    Cal
## 4          Festuca ovina   graminoid  5/27/2016 9:23 137.1220    Cal
## 5    Agrostis capillaris   graminoid  5/27/2016 9:42 237.4237    Cal
## 6  Aegopodium podagraria        forb 5/25/2016 12:20 388.2384    Cal
```

```
val_spec <- as.matrix(val.plsr.data[, which(names(val.plsr.data) %in% paste0("Wave_",wv))])
val.plsr.data <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% paste0("Wave_",wv))],
                            Spectra=I(val_spec))
head(val.plsr.data)[1:5]
```

```
##            Plant_Species Growth_Form       timestamp SLA_g_cm CalVal
## 9          Urtica dioica        forb 5/25/2016 12:37 284.6788    Val
## 14       Stellaria media        forb 5/25/2016 13:21 418.4284    Val
## 21  Alopecurus pratensis   graminoid  6/1/2016 11:32 218.2117    Val
## 41  Alopecurus pratensis   graminoid   6/8/2016 8:37 216.7568    Val
## 43   Agrostis capillaris   graminoid   6/8/2016 9:05 231.5292    Val
## 44 Aegopodium podagraria        forb   6/7/2016 9:05 311.4018    Val
```

plot cal and val spectra

```
par(mfrow=c(1,2)) # B, L, T, R
f.plot.spec(Z=cal.plsr.data$Spectra,wv=seq(Start.wave,End.wave,1),plot_label="Calibration")
f.plot.spec(Z=val.plsr.data$Spectra,wv=seq(Start.wave,End.wave,1),plot_label="Validation")
```



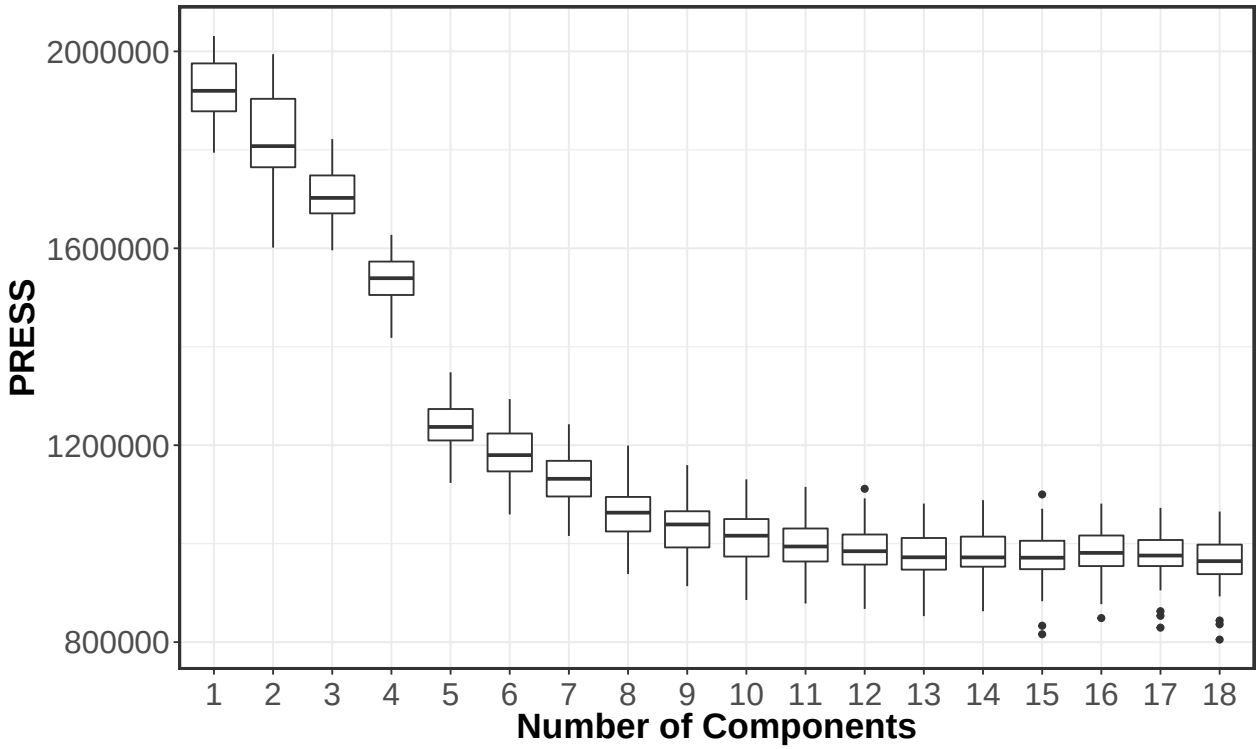

```
par(mfrow=c(1,1))
```

Use Jackknife permutation to determine optimal number of components

```
### Use permutation to determine the optimal number of components
if(grepl("Windows", sessionInfo()$running)){
  pls.options(parallel = NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

method <- "custom" #pls/custom
random_seed <- 2356812
seg <- 100
maxComps <- 18
iterations <- 30
if (method=="pls") {
  # pls package approach - faster but estimates more components....
  nComps <- find_optimal_components(method=method, maxComps=maxComps, seg=seg,
                                    random_seed=random_seed)
} else {
  # custom method - slow but generally finds the smallest number of components
  nComps <- find_optimal_components(method=method, maxComps=maxComps, iterations=iterations,
                                    seg=seg, prop=0.70,
                                    random_seed=random_seed)
}
```

```
## [1] "*** Running jacknife permutation test.  Please hang tight, this can take awhile ***"
## Running interation 1
## Running interation 2
## Running interation 3
## Running interation 4
## Running interation 5
## Running interation 6
```

```
## Running interation 7
## Running interation 8
## Running interation 9
## Running interation 10
## Running interation 11
## Running interation 12
## Running interation 13
## Running interation 14
## Running interation 15
## Running interation 16
## Running interation 17
## Running interation 18
## Running interation 19
## Running interation 20
## Running interation 21
## Running interation 22
## Running interation 23
## Running interation 24
## Running interation 25
## Running interation 26
## Running interation 27
## Running interation 28
## Running interation 29
## Running interation 30
## No id variables; using all as measure variables
```




```
## [1] "*** Optimal number of components based on t.test: 9"
```

Fit final model

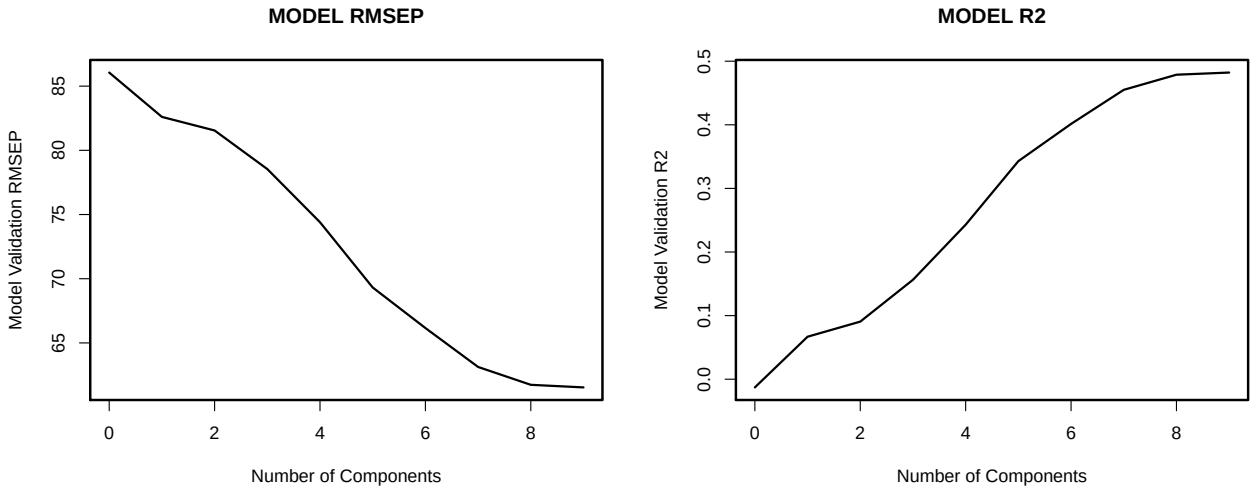
```
segs <- 100
plsr.out <- plsr(as.formula(paste(inVar,"~","Spectra")),scale=FALSE,ncomp=nComps,validation="CV",
                 segments=segs, segment.type="interleaved",trace=FALSE,data=cal.plsr.data)
fit <- plsr.out$fitted.values[,1,nComps]
pls.options(parallel = NULL)

# External validation fit stats
par(mfrow=c(1,2)) # B, L, T, R
RMSEP(plsr.out, newdata = val.plsr.data)
```

```
## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##       86.06        82.60        81.55        78.54        74.40        69.32
##     6 comps      7 comps      8 comps      9 comps
##       66.16        63.13        61.74        61.53
```

```
plot(RMSEP(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL RMSEP",
     xlab="Number of Components",ylab="Model Validation RMSEP",lty=1,col="black",cex=1.5,lwd=2)
box(lwd=2.2)

R2(plsr.out, newdata = val.plsr.data)
```

```
## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##    -0.01288      0.06681      0.09056      0.15636      0.24295      0.34288
##     6 comps      7 comps      8 comps      9 comps
##     0.40138      0.45499      0.47875      0.48216
```

```
plot(R2(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL R2",
     xlab="Number of Components",ylab="Model Validation R2",lty=1,col="black",cex=1.5,lwd=2)
box(lwd=2.2)
```




```
par(opar)
```

PLSR fit observed vs. predicted plot data

```
#calibration
cal.plsr.output <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% "Spectra")],
                              PLSR_Predicted=fit,
                              PLSR_CV_Predicted=as.vector(plsr.out$validation$pred[,,nComps]))
cal.plsr.output <- cal.plsr.output %>%
  mutate(PLSR_CV_Residuals = PLSR_CV_Predicted-get(inVar))
head(cal.plsr.output)
```

```
##            Plant_Species Growth_Form       timestamp SLA_g_cm CalVal
## 1 Calamagrostis epigejos   graminoid 5/25/2016 12:20 106.6500    Cal
## 2  Anthoxanthum odoratum   graminoid  5/27/2016 8:40 293.3565    Cal
## 3   Alopecurus pratensis   graminoid  5/27/2016 9:23 220.2703    Cal
## 4          Festuca ovina   graminoid  5/27/2016 9:23 137.1220    Cal
## 5    Agrostis capillaris   graminoid  5/27/2016 9:42 237.4237    Cal
## 6  Aegopodium podagraria        forb 5/25/2016 12:20 388.2384    Cal
##   PLSR_Predicted PLSR_CV_Predicted PLSR_CV_Residuals
## 1       229.2730          231.4534         124.80345
## 2       237.3005          236.4479         -56.90865
## 3       262.9303          264.5936          44.32334
## 4       134.4899          151.8084          14.68640
## 5       251.9651          251.2404          13.81676
## 6       274.5014          271.5183        -116.72010
```

```
cal.R2 <- round(pls::R2(plsr.out)[[1]][nComps],2)
cal.RMSEP <- round(sqrt(mean(cal.plsr.output$PLSR_CV_Residuals^2)),2)

val.plsr.output <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% "Spectra")],
                              PLSR_Predicted=as.vector(predict(plsr.out,
                                                               newdata = val.plsr.data,
```

```
                                                    ncomp=nComps, type="response")[,,1]))
val.plsr.output <- val.plsr.output %>%
  mutate(PLSR_Residuals = PLSR_Predicted-get(inVar))
head(val.plsr.output)
```

```
##           Plant_Species Growth_Form       timestamp SLA_g_cm CalVal
## 1         Urtica dioica        forb 5/25/2016 12:37 284.6788    Val
## 2       Stellaria media        forb 5/25/2016 13:21 418.4284    Val
## 3  Alopecurus pratensis   graminoid  6/1/2016 11:32 218.2117    Val
## 4  Alopecurus pratensis   graminoid   6/8/2016 8:37 216.7568    Val
## 5   Agrostis capillaris   graminoid   6/8/2016 9:05 231.5292    Val
## 6 Aegopodium podagraria        forb   6/7/2016 9:05 311.4018    Val
##   PLSR_Predicted PLSR_Residuals
## 1       235.7095     -48.969233
## 2       245.7077    -172.720735
## 3       214.8754      -3.336342
## 4       274.1596      57.402724
## 5       287.2400      55.710777
## 6       275.1441     -36.257634
```

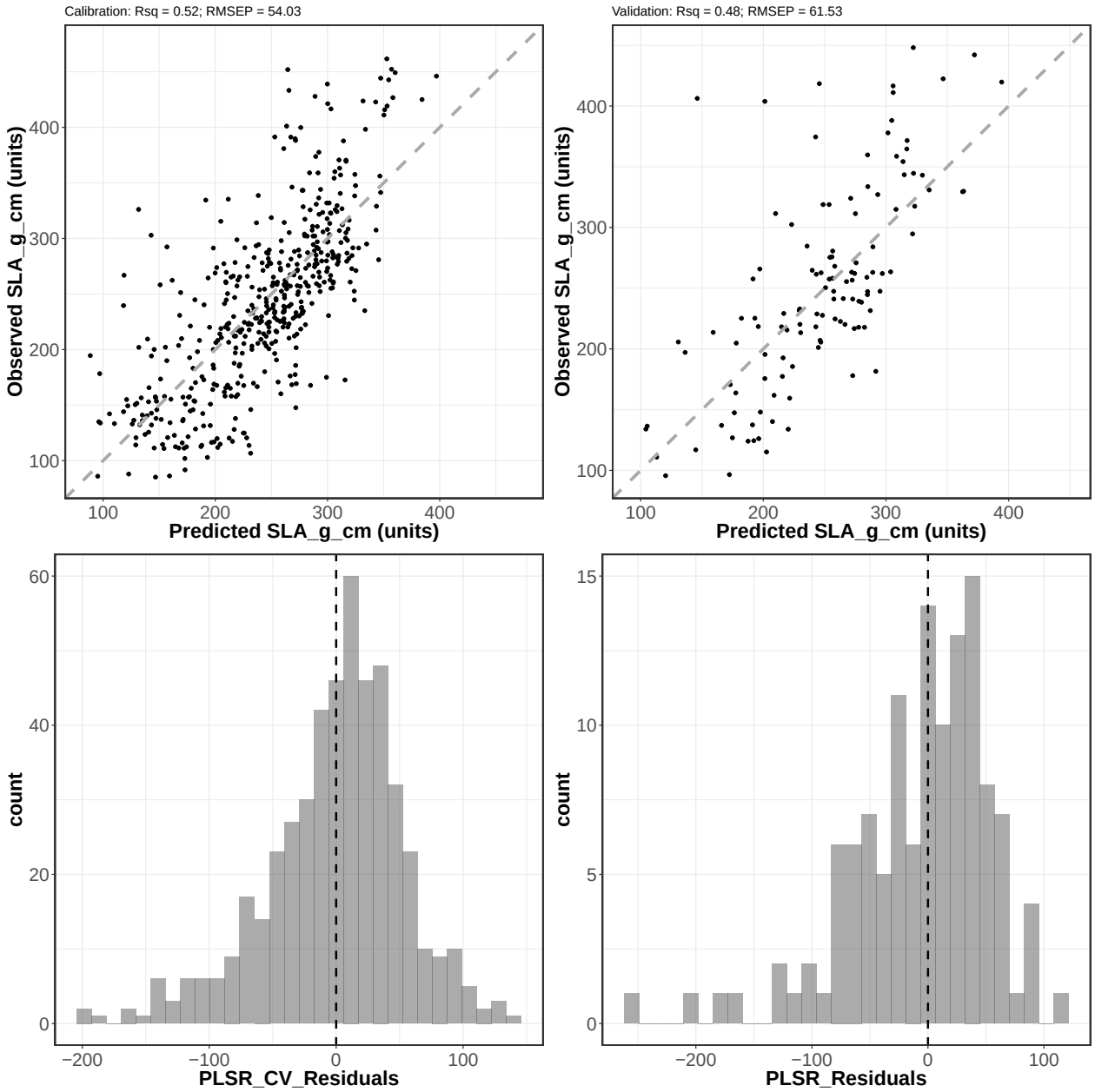
```
val.R2 <- round(pls::R2(plsr.out,newdata=val.plsr.data)[[1]][nComps],2)
val.RMSEP <- round(sqrt(mean(val.plsr.output$PLSR_Residuals^2)),2)

rng_quant <- quantile(cal.plsr.output[,inVar], probs = c(0.001, 0.999))
cal_scatter_plot <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                              rng_quant[2]) +
  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Calibration: ", paste0("Rsq = ", cal.R2), "; ", paste0("RMSEP = ",
                                                                            cal.RMSEP))) +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

cal_resid_histogram <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +
  geom_vline(xintercept = 0, color="black",
             linetype="dashed", size=1) + theme_bw() +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

rng_quant <- quantile(val.plsr.output[,inVar], probs = c(0.001, 0.999))
val_scatter_plot <- ggplot(val.plsr.output, aes(x=PLSR_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                              rng_quant[2]) +
  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
```

```
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Validation: ", paste0("Rsq = ", val.R2), "; ", paste0("RMSEP = ",
                                                                           val.RMSEP))) +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

val_resid_histogram <- ggplot(val.plsr.output, aes(x=PLSR_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +
  geom_vline(xintercept = 0, color="black",
             linetype="dashed", size=1) + theme_bw() +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

# plot cal/val side-by-side
grid.arrange(cal_scatter_plot, val_scatter_plot, cal_resid_histogram, val_resid_histogram,
             nrow=2,ncol=2)
```

```
## Warning: Removed 5 rows containing missing values (geom_point).
```

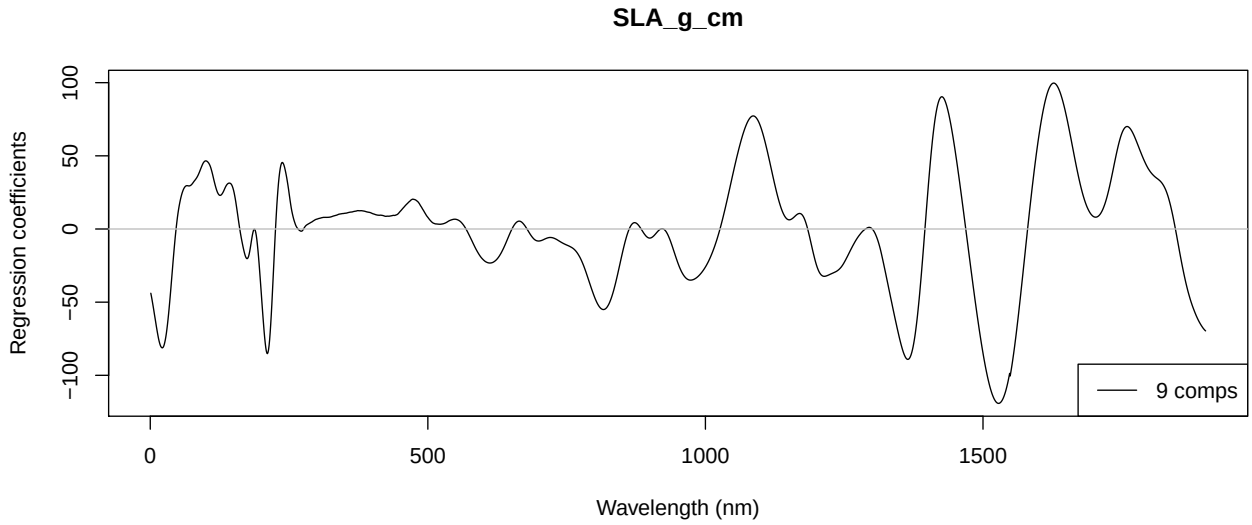
```
## Warning: Removed 3 rows containing missing values (geom_point).
```

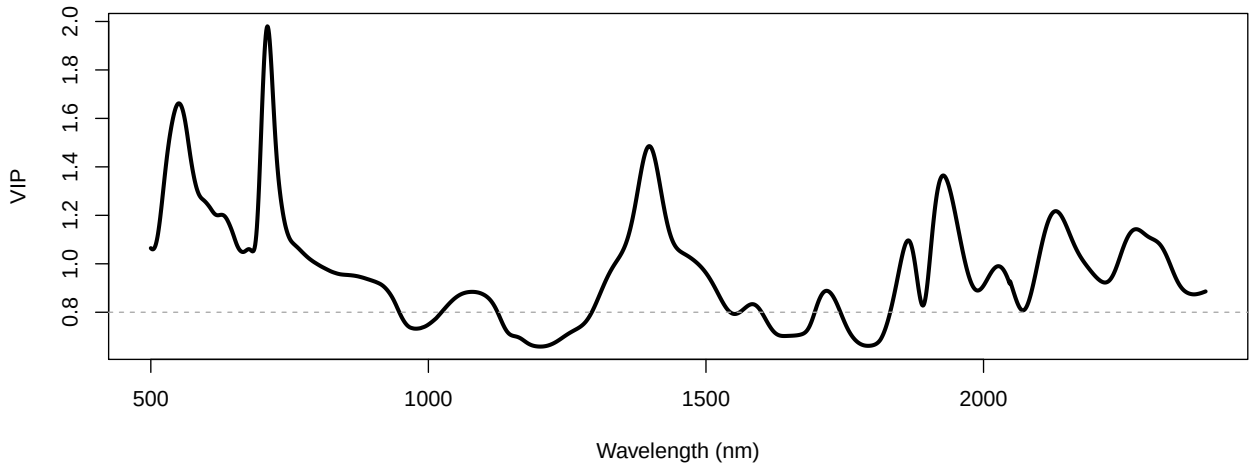
```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



Generate Coefficient and VIP plots

```
vips <- VIP(plsr.out)[nComps,]
par(mfrow=c(2,1))
plot(plsr.out, plottype = "coef",xlab="Wavelength (nm)",
     ylab="Regression coefficients",legendpos = "bottomright",ncomp=nComps)

plot(seq(Start.wave,End.wave,1),vips,xlab="Wavelength (nm)",ylab="VIP",cex=0.01)
lines(seq(Start.wave,End.wave,1),vips,lwd=3)
abline(h=0.8,lty=2,col="dark grey")
```





Jackknife validation

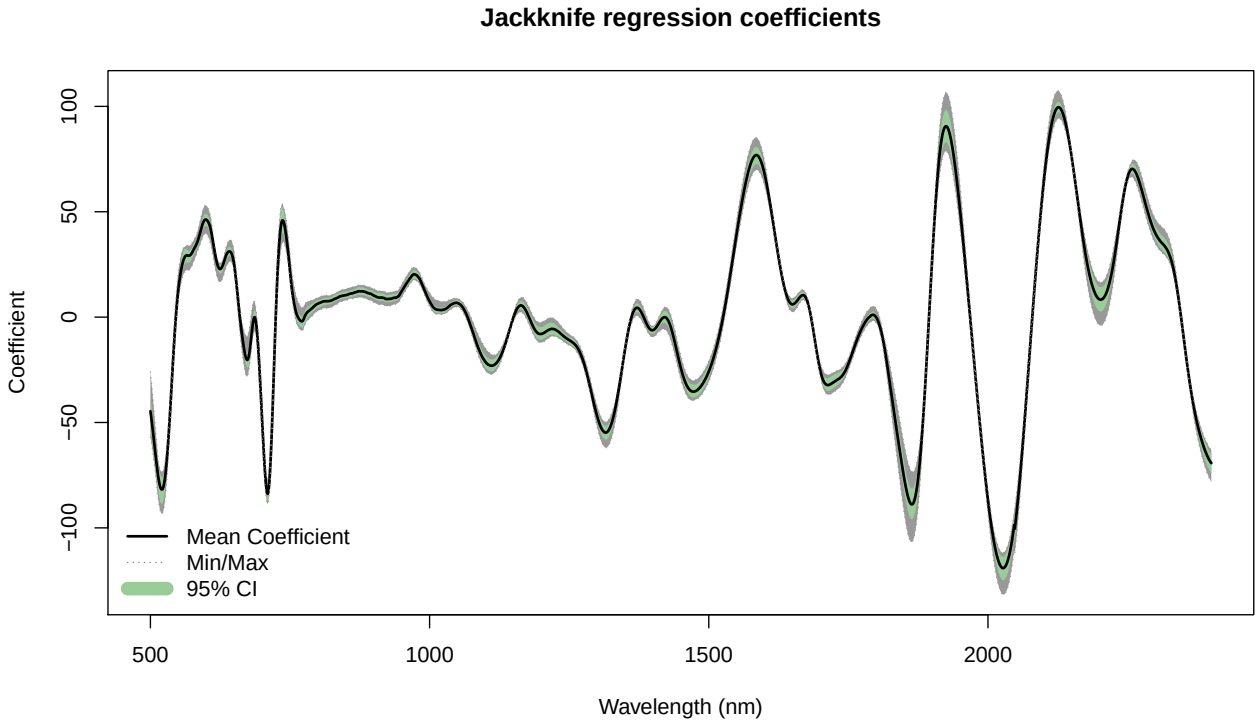
```
if(grepl("Windows", sessionInfo()$running)){
  pls.options(parallel =NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

seg <- 100
jk.plsr.out <- pls::plsr(as.formula(paste(inVar,"~","Spectra")), scale=FALSE, center=TRUE, ncomp=nComps
                         validation="CV", segments = seg, segment.type="interleaved", trace=FALSE,
                         jackknife=TRUE, data=cal.plsr.data)
pls.options(parallel = NULL)

Jackknife_coef <- f.coef.valid(plsr.out = jk.plsr.out,
                               data_plsr = cal.plsr.data, ncomp = nComps)
Jackknife_intercept <- Jackknife_coef[1,,,]
```

```
Jackknife_coef <- Jackknife_coef[2:dim(Jackknife_coef)[1],,,]

#interval <- c(0.025,0.975)
interval <- c(0.05,0.95)
Jackknife_Pred <- val.plsr.data$Spectra%*%Jackknife_coef+Jackknife_intercept
Interval_Conf <- apply(X = Jackknife_Pred, MARGIN = 1, FUN = quantile,
                       probs=c(interval[1], interval[2]))
Interval_Pred <- apply(X = Jackknife_Pred, MARGIN = 1, FUN = quantile,
                       probs=c(interval[1], interval[2]))
sd_mean <- apply(X = Jackknife_Pred, MARGIN = 1, FUN =sd)
sd_res <- sd(val.plsr.output$PLSR_Residuals)
sd_tot <- sqrt(sd_mean^2+sd_res^2)
val.plsr.output$LCI <- Interval_Pred[1,]
val.plsr.output$UCI <- Interval_Pred[2,]
val.plsr.output$LPI <- val.plsr.output$PLSR_Predicted-1.96*sd_tot
val.plsr.output$UPI <- val.plsr.output$PLSR_Predicted+1.96*sd_tot
head(val.plsr.output)
```

```
##           Plant_Species Growth_Form       timestamp SLA_g_cm CalVal
## 1         Urtica dioica        forb 5/25/2016 12:37 284.6788    Val
## 2       Stellaria media        forb 5/25/2016 13:21 418.4284    Val
## 3  Alopecurus pratensis   graminoid  6/1/2016 11:32 218.2117    Val
## 4  Alopecurus pratensis   graminoid   6/8/2016 8:37 216.7568    Val
## 5   Agrostis capillaris   graminoid   6/8/2016 9:05 231.5292    Val
## 6 Aegopodium podagraria        forb   6/7/2016 9:05 311.4018    Val
##   PLSR_Predicted PLSR_Residuals      LCI      UCI       LPI      UPI
## 1       235.7095     -48.969233 222.2974 253.5497 113.78223 357.6368
## 2       245.7077    -172.720735 229.3466 261.7874 123.88508 367.5303
## 3       214.8754      -3.336342 201.4821 231.2104  93.31679 336.4339
## 4       274.1596      57.402724 260.3334 286.7905 152.48278 395.8363
## 5       287.2400      55.710777 270.3968 305.3789 164.95946 409.5205
## 6       275.1441     -36.257634 258.2812 288.5656 153.30680 396.9814
```

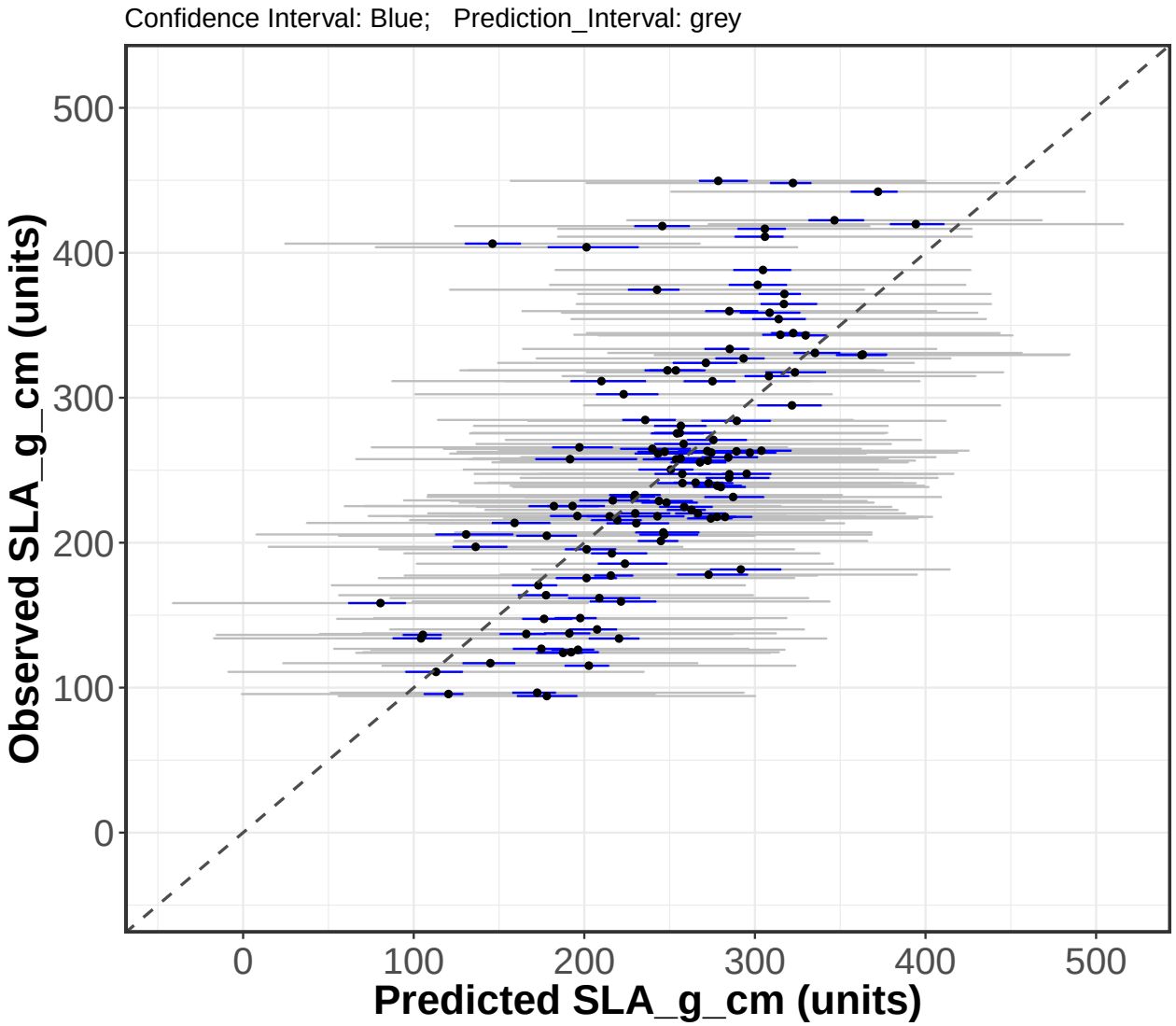
Jackknife coefficient plot

```
f.plot.coef(Z = t(Jackknife_coef), wv = seq(Start.wave,End.wave,1),
            plot_label="Jackknife regression coefficients",
            position = 'bottomleft')
```



Jackknife validation plot

```
#rng_vals <- quantile(val.plsr.output[,inVar], probs = c(0.001, 0.999))
rng_vals <- c(min(val.plsr.output$LPI), max(val.plsr.output$UPI))
jk_val_scatterplot <- ggplot(val.plsr.output, aes(x=PLSR_Predicted,
                                                  y=get(inVar))) +
  theme_bw()+ geom_errorbar(aes(xmin = LPI,xmax = UPI),color='grey',
                            width=0.2) +
  geom_errorbar(aes(xmin = LCI,xmax = UCI),color='blue',width=0.2) +
  geom_point(size=1.3)  +
  geom_abline(intercept = 0, slope = 1, color="grey30",
              linetype="dashed", size=0.7) +
  xlim(rng_vals[1], rng_vals[2]) +
  ylim(rng_vals[1], rng_vals[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Confidence Interval: Blue;   Prediction_Interval: grey")) +
  theme(axis.text=element_text(size=18),legend.position = 'right',
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0,vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA,
                                    size=1.5))
jk_val_scatterplot
```



Output jackknife results

```
out.jk.coefs <- data.frame(Iteration=seq(1,seg,1),
                           Intercept=Jackknife_intercept,t(Jackknife_coef))
head(out.jk.coefs)[1:6]
```

```
##       Iteration Intercept  Wave_500  Wave_501  Wave_502  Wave_503
## Seg 1         1  264.5504 -46.86435 -49.05338 -51.29486 -53.71050
## Seg 2         2  272.1996 -47.47939 -49.32410 -51.21585 -53.28765
## Seg 3         3  265.2322 -50.37124 -52.20291 -54.06858 -56.09309
## Seg 4         4  267.4766 -47.37086 -49.38918 -51.45968 -53.70623
## Seg 5         5  277.2098 -42.98897 -44.99306 -47.04947 -49.28996
## Seg 6         6  265.5952 -37.12018 -38.96434 -40.86150 -42.94944
```

```
write.csv(out.jk.coefs,file=file.path(outdir,
                                      paste0(inVar,
                                             '_Jackkife_PLSR_Coefficients.csv')),
          row.names=FALSE)
```

Create core PLSR outputs

```
print(paste("Output directory: ", getwd()))
```

```
## [1] "Output directory:  /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction/vignettes"
```

```
# Observed versus predicted
write.csv(cal.plsr.output,file=file.path(outdir,
                                         paste0(inVar,'_Observed_PLSR_CV_Pred_',
                                                nComps,'comp.csv')),
          row.names=FALSE)

# Validation data
write.csv(val.plsr.output,file=file.path(outdir,
                                         paste0(inVar,'_Validation_PLSR_Pred_',
                                                nComps,'comp.csv')),
          row.names=FALSE)

# Model coefficients
coefs <- coef(plsr.out,ncomp=nComps,intercept=TRUE)
write.csv(coefs,file=file.path(outdir,
                               paste0(inVar,'_PLSR_Coefficients_',
                                      nComps,'comp.csv')),
          row.names=TRUE)

# PLSR VIP
write.csv(vips,file=file.path(outdir,
                              paste0(inVar,'_PLSR_VIPs_',
                                     nComps,'comp.csv')))
```

Confirm files were written to temp space

```
print("**** PLSR output files: ")
```

```
## [1] "**** PLSR output files: "
```

```
list.files(outdir)[grep(pattern = inVar, list.files(outdir))]
```

```
## [1] "SLA_g_cm_Jackkife_PLSR_Coefficients.csv"
## [2] "SLA_g_cm_Observed_PLSR_CV_Pred_9comp.csv"
## [3] "SLA_g_cm_PLSR_Coefficients_9comp.csv"
## [4] "SLA_g_cm_PLSR_VIPs_9comp.csv"
## [5] "SLA_g_cm_Validation_PLSR_Pred_9comp.csv"
```