

# Spectra-trait PLSR example using leaf-level spectra and leaf nitrogen content (Narea, g/m<sup>2</sup>) data from 36 species growing in Rosa rugosa invaded coastal grassland communities in Belgium

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## Overview

This is an R Markdown Notebook to illustrate how to retrieve a dataset from the EcoSIS spectral database, choose the “optimal” number of pls components, and fit a pls model for leaf nitrogen content (Narea, g/m<sup>2</sup>)

## Getting Started

### Load libraries

```
list.of.packages <- c("pls", "dplyr", "reshape2", "here", "plotrix", "ggplot2", "gridExtra",
                      "spectratrait")
invisible(lapply(list.of.packages, library, character.only = TRUE))

##
## Attaching package: 'pls'
## The following object is masked from 'package:stats':
##
##   loadings
##
## Attaching package: 'dplyr'
## The following objects are masked from 'package:stats':
##
##   filter, lag
## The following objects are masked from 'package:base':
##
##   intersect, setdiff, setequal, union
## here() starts at /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction
##
## Attaching package: 'gridExtra'
## The following object is masked from 'package:dplyr':
##
##   combine
```

## Setup other functions and options

```
### Setup other functions and options
# not in
`%notin%` <- Negate(`%in%`)

# Script options
pls::pls.options(plsralg = "oscorespls")
pls::pls.options("plsralg")

## $plsralg
## [1] "oscorespls"

# Default par options
opar <- par(no.readonly = T)

# What is the target variable?
inVar <- "Narea_g_m2"

# What is the source dataset from EcoSIS?
ecosys_id <- "9db4c5a2-7eac-4e1e-8859-009233648e89"

# Specify output directory, output_dir
# Options:
# tempdir - use a OS-specified temporary directory
# user defined PATH - e.g. "~/scratch/PLSR"
output_dir <- "tempdir"
```

## Set working directory (scratch space)

```
## [1] "/private/var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjq/T/Rtmpi5zkMP"
```

## Grab data from EcoSIS

```
print(paste0("Output directory: ",getwd())) # check wd

## [1] "Output directory: /Users/sserbin/Data/GitHub/PLSR_for_plant_trait_prediction/vignettes"
dat_raw <- spectratrait::get_ecosis_data(ecosis_id = ecosys_id)

## [1] "**** Downloading Ecosis data ****"

## Downloading data...

##
## -- Column specification -----
## cols(
##   .default = col_double(),
##   `Latin Species` = col_character(),
##   ids = col_character(),
##   `plot code` = col_character(),
##   `species code` = col_character()
## )
## i Use `spec()` for the full column specifications.
```

```
## Download complete!
```

```
head(dat_raw)
```

```
## # A tibble: 6 x 2,164
##   `Cw/EWT (cm3/cm2~` `Latin Species`   `Leaf area (mm2~` `Leaf calcium content pe~
##           <dbl> <chr>                <dbl>                <dbl>
## 1      0.00887 Arrhenatherum el~        696.                0.0291
## 2      0.00824 Bromus sterilis          447.                0.0230
## 3      0.0280  Jacobaea vulgaris        2418.                0.0950
## 4      0.0106  Rubus caesius          5719.                0.0700
## 5      0.00851 Arrhenatherum el~        671.                0.0286
## 6      0.0153  Crepis capillaris       1401.                0.0470
## # ... with 2,160 more variables:
## #   Leaf magnesium content per leaf area (mg/mm2) <dbl>,
## #   Leaf mass per area (g/cm2) <dbl>,
## #   Leaf nitrogen content per leaf area (mg/mm2) <dbl>,
## #   Leaf phosphorus content per leaf area (mg/mm2) <dbl>,
## #   Leaf potassium content per leaf area (mg/mm2) <dbl>,
## #   Plant height vegetative (cm) <dbl>, ids <chr>, plot code <chr>,
## #   species code <chr>, 350 <dbl>, 351 <dbl>, 352 <dbl>, 353 <dbl>, 354 <dbl>,
## #   355 <dbl>, 356 <dbl>, 357 <dbl>, 358 <dbl>, 359 <dbl>, 360 <dbl>,
## #   361 <dbl>, 362 <dbl>, 363 <dbl>, 364 <dbl>, 365 <dbl>, 366 <dbl>,
## #   367 <dbl>, 368 <dbl>, 369 <dbl>, 370 <dbl>, 371 <dbl>, 372 <dbl>,
## #   373 <dbl>, 374 <dbl>, 375 <dbl>, 376 <dbl>, 377 <dbl>, 378 <dbl>,
## #   379 <dbl>, 380 <dbl>, 381 <dbl>, 382 <dbl>, 383 <dbl>, 384 <dbl>,
## #   385 <dbl>, 386 <dbl>, 387 <dbl>, 388 <dbl>, 389 <dbl>, 390 <dbl>,
## #   391 <dbl>, 392 <dbl>, 393 <dbl>, 394 <dbl>, 395 <dbl>, 396 <dbl>,
## #   397 <dbl>, 398 <dbl>, 399 <dbl>, 400 <dbl>, 401 <dbl>, 402 <dbl>,
## #   403 <dbl>, 404 <dbl>, 405 <dbl>, 406 <dbl>, 407 <dbl>, 408 <dbl>,
## #   409 <dbl>, 410 <dbl>, 411 <dbl>, 412 <dbl>, 413 <dbl>, 414 <dbl>,
## #   415 <dbl>, 416 <dbl>, 417 <dbl>, 418 <dbl>, 419 <dbl>, 420 <dbl>,
## #   421 <dbl>, 422 <dbl>, 423 <dbl>, 424 <dbl>, 425 <dbl>, 426 <dbl>,
## #   427 <dbl>, 428 <dbl>, 429 <dbl>, 430 <dbl>, 431 <dbl>, 432 <dbl>,
## #   433 <dbl>, 434 <dbl>, 435 <dbl>, 436 <dbl>, 437 <dbl>, 438 <dbl>,
## #   439 <dbl>, 440 <dbl>, ...
```

```
names(dat_raw)[1:40]
```

```
## [1] "Cw/EWT (cm3/cm2)"
## [2] "Latin Species"
## [3] "Leaf area (mm2)"
## [4] "Leaf calcium content per leaf area (mg/mm2)"
## [5] "Leaf magnesium content per leaf area (mg/mm2)"
## [6] "Leaf mass per area (g/cm2)"
## [7] "Leaf nitrogen content per leaf area (mg/mm2)"
## [8] "Leaf phosphorus content per leaf area (mg/mm2)"
## [9] "Leaf potassium content per leaf area (mg/mm2)"
## [10] "Plant height vegetative (cm)"
## [11] "ids"
## [12] "plot code"
## [13] "species code"
## [14] "350"
## [15] "351"
## [16] "352"
```

```
## [17] "353"
## [18] "354"
## [19] "355"
## [20] "356"
## [21] "357"
## [22] "358"
## [23] "359"
## [24] "360"
## [25] "361"
## [26] "362"
## [27] "363"
## [28] "364"
## [29] "365"
## [30] "366"
## [31] "367"
## [32] "368"
## [33] "369"
## [34] "370"
## [35] "371"
## [36] "372"
## [37] "373"
## [38] "374"
## [39] "375"
## [40] "376"
```

## Create full pls dataset

### ### Create pls dataset

```
Start.wave <- 500
End.wave <- 2400
wv <- seq(Start.wave,End.wave,1)
Spectra <- as.matrix(dat_raw[,names(dat_raw) %in% wv])
colnames(Spectra) <- c(paste0("Wave_",wv))
sample_info <- dat_raw[,names(dat_raw) %notin% seq(350,2500,1)]
head(sample_info)
```

```
## # A tibble: 6 x 13
##   `Cw/EWT (cm3/cm2~` `Latin Species`   `Leaf area (mm2~` `Leaf calcium content pe~
##           <dbl> <chr>                   <dbl>                <dbl>
## 1         0.00887 Arrhenatherum el~         696.                0.0291
## 2         0.00824 Bromus sterilis         447.                0.0230
## 3         0.0280  Jacobaea vulgaris       2418.                0.0950
## 4         0.0106  Rubus caesius        5719.                0.0700
## 5         0.00851 Arrhenatherum el~         671.                0.0286
## 6         0.0153  Crepis capillaris       1401.                0.0470
## # ... with 9 more variables:
## #   Leaf magnesium content per leaf area (mg/mm2) <dbl>,
## #   Leaf mass per area (g/cm2) <dbl>,
## #   Leaf nitrogen content per leaf area (mg/mm2) <dbl>,
## #   Leaf phosphorus content per leaf area (mg/mm2) <dbl>,
## #   Leaf potassium content per leaf area (mg/mm2) <dbl>,
## #   Plant height vegetative (cm) <dbl>, ids <chr>, plot code <chr>,
## #   species code <chr>
```

```
sample_info2 <- sample_info %>%
  select(Plant_Species=`Latin Species`,Species_Code=`species code`,Plot=`plot code`,
         Narea_mg_mm2=`Leaf nitrogen content per leaf area (mg/mm2)`)
sample_info2 <- sample_info2 %>%
  # mutate(Narea_g_m2=Narea_mg_mm2*(0.001/1e-6)) # based on orig units should be this but conversion wro
  mutate(Narea_g_m2=Narea_mg_mm2*100) # this assumes orig units were g/mm2 or mg/cm2
head(sample_info2)
```

```
## # A tibble: 6 x 5
##   Plant_Species      Species_Code Plot  Narea_mg_mm2 Narea_g_m2
##   <chr>             <chr>      <chr>      <dbl>      <dbl>
## 1 Arrhenatherum elatius Arrela      DC1         0.0126        1.26
## 2 Bromus sterilis      Broste      DC1         0.00682       0.682
## 3 Jacobaea vulgaris    Jacvul      DC1         0.0102       1.02
## 4 Rubus caesius        Rubcae      DC1         0.0121       1.21
## 5 Arrhenatherum elatius Arrela      DC2         0.0117       1.17
## 6 Crepis capillaris    Creves      DC2         0.00877       0.877
```

```
plsr_data <- data.frame(sample_info2,Spectra)
rm(sample_info,sample_info2,Spectra)
```

```
#### End user needs to do what's appropriate for their data. This may be an iterative process.
# Keep only complete rows of inVar and spec data before fitting
plsr_data <- plsr_data[complete.cases(plsr_data[,names(plsr_data) %in%
                                     c(inVar,paste0("Wave_",wv))]),]
```

Example data cleaning.

Create cal/val datasets

```
#### Create cal/val datasets
## Make a stratified random sampling in the strata USDA_Species_Code and Domain

method <- "dplyr" #base/dplyr
# base R - a bit slow
# dplyr - much faster
split_data <- spectratrait::create_data_split(dataset=plsr_data, approach=method, split_seed=1245565,
                                              prop=0.8, group_variables="Species_Code")
names(split_data)
```

```
## [1] "cal_data" "val_data"
cal.plsr.data <- split_data$cal_data
head(cal.plsr.data)[1:8]
```

```
##      Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2 Wave_500
## 1 Ammophila arenaria   Ammare   ZC3   0.03240495   3.240495 0.130885
## 2 Ammophila arenaria   Ammare   MC2   0.02806279   2.806279 0.135785
## 3 Ammophila arenaria   Ammare   ZC1   0.02041612   2.041612 0.147665
## 4 Ammophila arenaria   Ammare   MC1   0.02426549   2.426549 0.142765
## 5 Ammophila arenaria   Ammare   WC3   0.02807281   2.807281 0.151750
## 6 Ammophila arenaria   Ammare   WR3   0.02286678   2.286678 0.150850
```

```
## Wave_501 Wave_502
## 1 0.13175 0.132750
## 2 0.13685 0.138150
## 3 0.14910 0.150330
## 4 0.14390 0.145200
## 5 0.15275 0.154150
## 6 0.15185 0.152815
```

```
val.plsr.data <- split_data$val_data
head(val.plsr.data)[1:8]
```

```
##      Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2 Wave_500
## 184 Jacobaea vulgaris Jacvul WC2 0.008756996 0.8756996 0.06736887
## 185 Potentilla reptans Potrep WC2 0.010313464 1.0313464 0.07125000
## 186 Rubus caesius Rubcae WC2 0.007968454 0.7968454 0.05993560
## 187 Urtica dioica Urtdio WC2 0.012737560 1.2737560 0.06508300
## 188 Ammophila arenaria Ammare WC3 0.028072806 2.8072806 0.15175000
## 189 Jacobaea vulgaris Jacvul WC3 0.010251687 1.0251687 0.06805547
##      Wave_501 Wave_502
## 184 0.06870667 0.07014220
## 185 0.07235000 0.07368350
## 186 0.06162000 0.06352233
## 187 0.06625000 0.06758350
## 188 0.15275000 0.15415000
## 189 0.06938000 0.07093553
```

```
rm(split_data)
```

```
# Datasets:
```

```
print(paste("Cal observations: ",dim(cal.plsr.data)[1],sep=""))
```

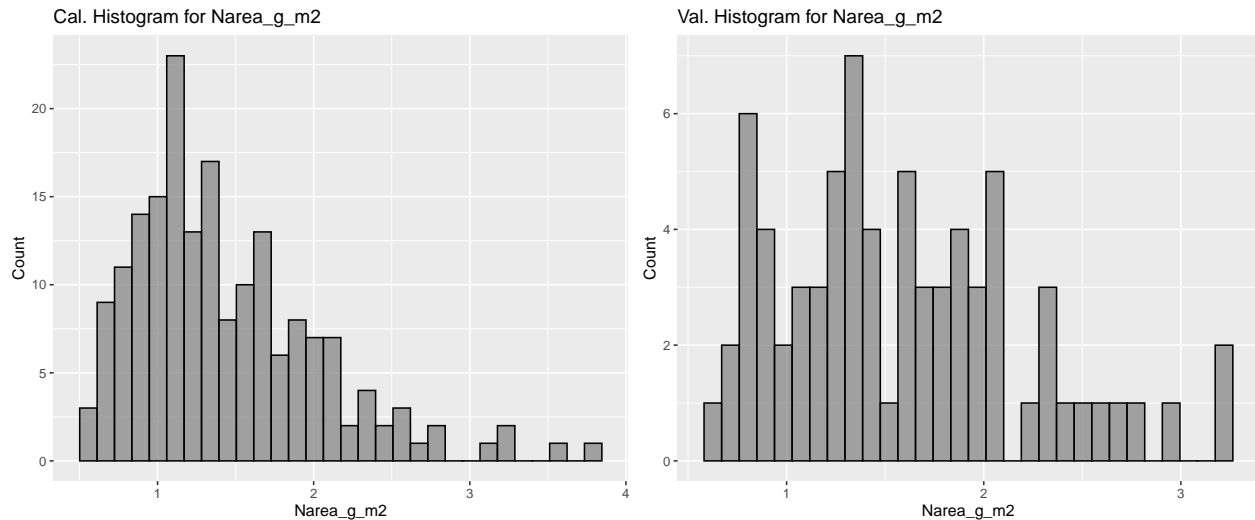
```
## [1] "Cal observations: 183"
```

```
print(paste("Val observations: ",dim(val.plsr.data)[1],sep=""))
```

```
## [1] "Val observations: 73"
```

```
cal_hist_plot <- qplot(cal.plsr.data[,paste0(inVar)],geom="histogram",
                        main = paste0("Cal. Histogram for ",inVar),
                        xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),col=I("black"),
                        alpha=I(.7))
val_hist_plot <- qplot(val.plsr.data[,paste0(inVar)],geom="histogram",
                       main = paste0("Val. Histogram for ",inVar),
                       xlab = paste0(inVar),ylab = "Count",fill=I("grey50"),col=I("black"),
                       alpha=I(.7))
histograms <- grid.arrange(cal_hist_plot, val_hist_plot, ncol=2)
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
ggsave(filename = file.path(outdir,paste0(inVar,"_Cal_Val_Histograms.png")), plot = histograms,
        device="png", width = 30,
        height = 12, units = "cm",
        dpi = 300)
# output cal/val data
write.csv(cal.plsr.data,file=file.path(outdir,paste0(inVar,'_Cal_PLSR_Dataset.csv')),
          row.names=FALSE)
write.csv(val.plsr.data,file=file.path(outdir,paste0(inVar,'_Val_PLSR_Dataset.csv')),
          row.names=FALSE)
```

## Create calibration and validation PLSR datasets

```
### Format PLSR data for model fitting
cal_spec <- as.matrix(cal.plsr.data[, which(names(cal.plsr.data) %in% paste0("Wave_",wv))])
cal.plsr.data <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% paste0("Wave_",wv))],
                           Spectra=I(cal_spec))
head(cal.plsr.data)[1:5]

##      Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2
## 1 Ammophila arenaria      Ammare ZC3  0.03240495  3.240495
## 2 Ammophila arenaria      Ammare MC2  0.02806279  2.806279
## 3 Ammophila arenaria      Ammare ZC1  0.02041612  2.041612
## 4 Ammophila arenaria      Ammare MC1  0.02426549  2.426549
## 5 Ammophila arenaria      Ammare WC3  0.02807281  2.807281
## 6 Ammophila arenaria      Ammare WR3  0.02286678  2.286678

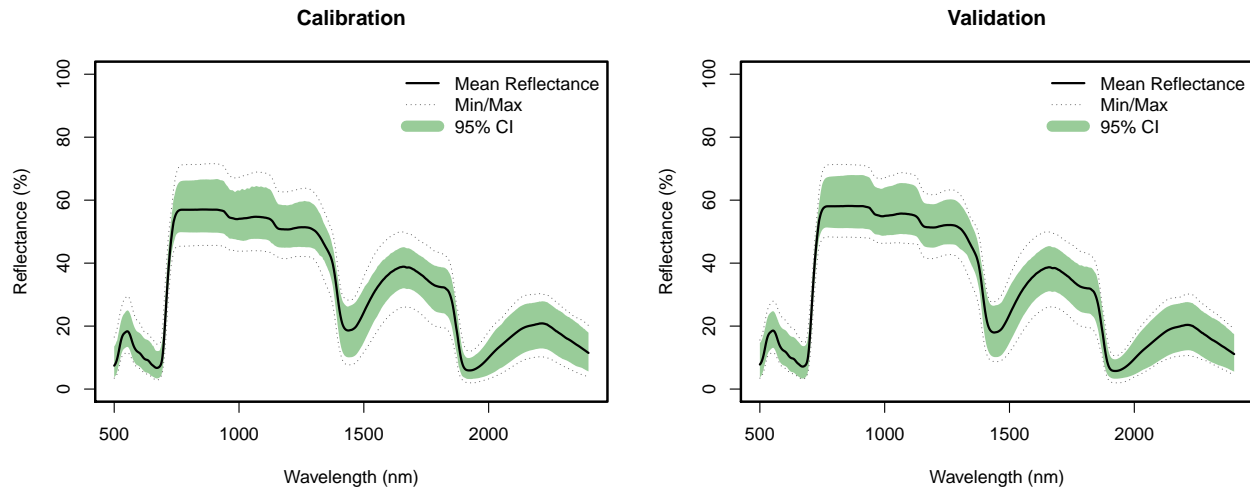
val_spec <- as.matrix(val.plsr.data[, which(names(val.plsr.data) %in% paste0("Wave_",wv))])
val.plsr.data <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% paste0("Wave_",wv))],
                           Spectra=I(val_spec))
head(val.plsr.data)[1:5]

##      Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2
## 184 Jacobaea vulgaris      Jacvul WC2  0.008756996  0.8756996
## 185 Potentilla reptans      Potrep WC2  0.010313464  1.0313464
## 186 Rubus caesius          Rubcae WC2  0.007968454  0.7968454
## 187 Urtica dioica          Urtdio WC2  0.012737560  1.2737560
## 188 Ammophila arenaria      Ammare WC3  0.028072806  2.8072806
```

```
## 189  Jacobaea vulgaris      Jacvul  WC3  0.010251687  1.0251687
```

plot cal and val spectra

```
par(mfrow=c(1,2)) # B, L, T, R
spectratrait::f.plot.spec(Z=cal.plsr.data$Spectra,wv=wv,plot_label="Calibration")
spectratrait::f.plot.spec(Z=val.plsr.data$Spectra,wv=wv,plot_label="Validation")
```



```
dev.copy(png,file.path(outdir,paste0(inVar,'_Cal_Val_Spectra.png')),
         height=2500,width=4900, res=340)
```

```
## quartz_off_screen
## 3
```

```
dev.off();
```

```
## pdf
## 2
```

```
par(mfrow=c(1,1))
```

Use Jackknife permutation to determine optimal number of components

```
### Use permutation to determine the optimal number of components
if(grepl("Windows", sessionInfo())$running){
  pls.options(parallel = NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

method <- "pls" #pls, firstPlateau, firstMin
random_seed <- 1245565
seg <- 50
maxComps <- 16
iterations <- 80
prop <- 0.70
if (method=="pls") {
```



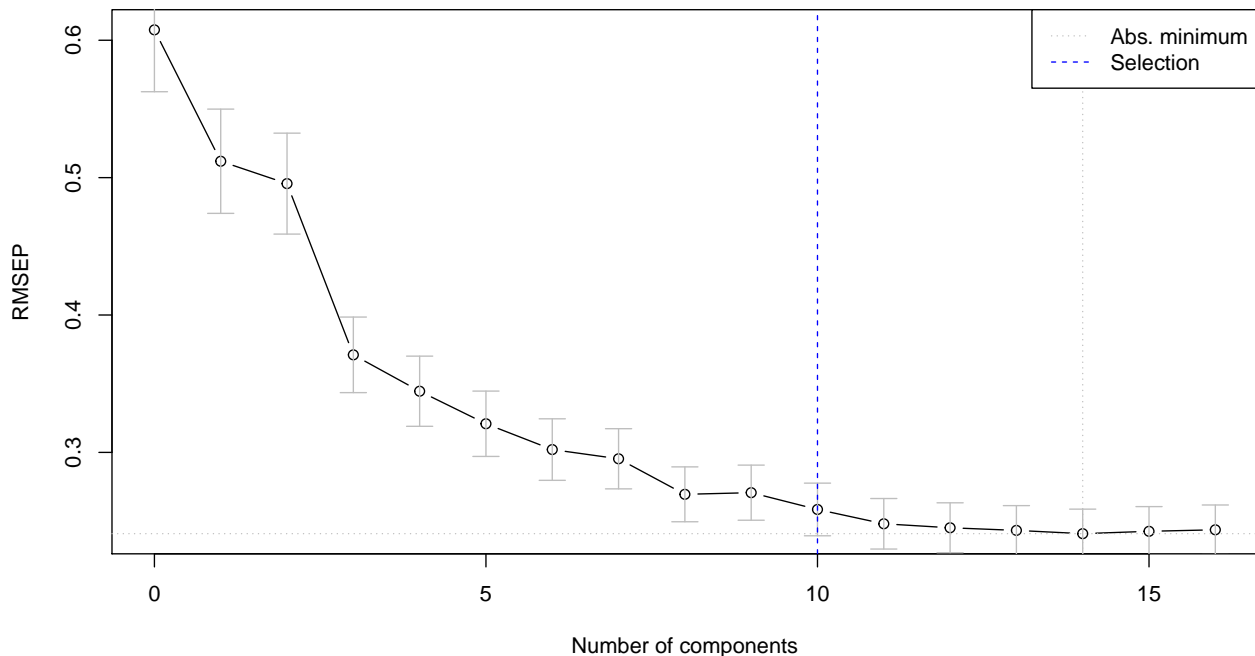
```

# pls package approach - faster but estimates more components....
nComps <- spectratrait::find_optimal_components(dataset=cal.plsr.data, method=method,
                                              maxComps=maxComps, seg=seg,
                                              random_seed=random_seed)

print(paste0("*** Optimal number of components: ", nComps))
} else {
  nComps <- spectratrait::find_optimal_components(dataset=cal.plsr.data, method=method,
                                              maxComps=maxComps, iterations=iterations,
                                              seg=seg, prop=prop,
                                              random_seed=random_seed)
}

```

```
## [1] "*** Running PLS permutation test ***"
```



```
## [1] "*** Optimal number of components: 10"
```

```

dev.copy(png,file.path(outdir,paste0(paste0(inVar,"_PLSR_Component_Selection.png"))),
        height=2800, width=3400, res=340)

```

```
## quartz_off_screen
```

```
## 3
```

```
dev.off();
```

```
## pdf
```

```
## 2
```

Fit final model

```

plsr.out <- plsr(as.formula(paste(inVar,"~","Spectra")),scale=FALSE,ncomp=nComps,validation="L00",
               trace=FALSE,data=cal.plsr.data)
fit <- plsr.out$fitted.values[,1,nComps]
pls.options(parallel = NULL)

```

```

# External validation fit stats
par(mfrow=c(1,2)) # B, L, T, R
pls::RMSEP(plsr.out, newdata = val.plsr.data)

## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##      0.6346      0.5045      0.4645      0.3415      0.3296      0.3037
##      6 comps      7 comps      8 comps      9 comps     10 comps
##      0.2703      0.2659      0.2524      0.2450      0.2452

plot(pls::RMSEP(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL RMSEP",
     xlab="Number of Components",ylab="Model Validation RMSEP",lty=1,col="black",cex=1.5,lwd=2)
box(lwd=2.2)

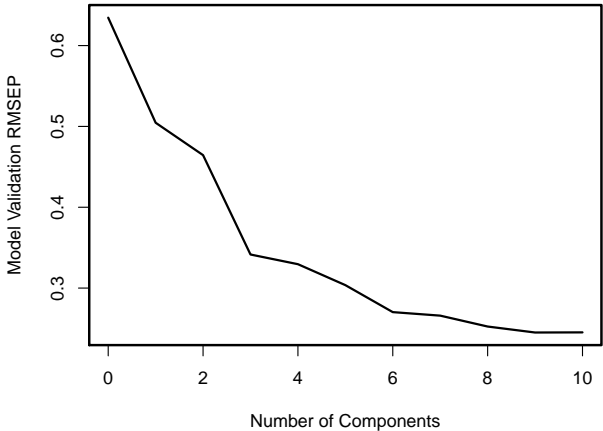
pls::R2(plsr.out, newdata = val.plsr.data)

## (Intercept)      1 comps      2 comps      3 comps      4 comps      5 comps
##     -0.05977      0.33000      0.43217      0.69298      0.71415      0.75732
##      6 comps      7 comps      8 comps      9 comps     10 comps
##      0.80776      0.81389      0.83228      0.84198      0.84176

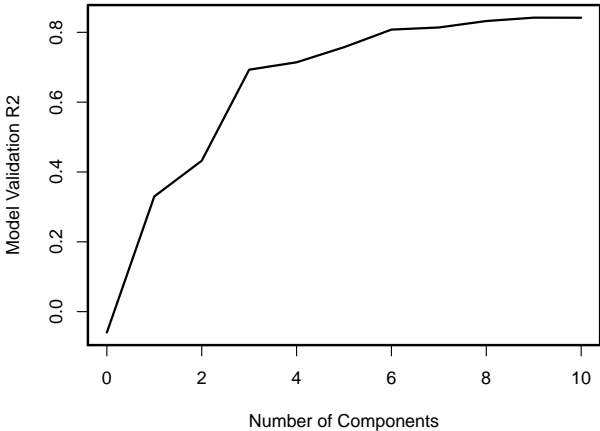
plot(pls::R2(plsr.out,estimate=c("test"),newdata = val.plsr.data), main="MODEL R2",
     xlab="Number of Components",ylab="Model Validation R2",lty=1,col="black",cex=1.5,lwd=2)
box(lwd=2.2)

```

**MODEL RMSEP**



**MODEL R2**



```

dev.copy(png,file.path(outdir,paste0(paste0(inVar,"_Validation_RMSEP_R2_by_Component.png"))),
         height=2800, width=4800, res=340)

## quartz_off_screen
##      3

dev.off();

## pdf
##      2

par(opar)

```

## PLSR fit observed vs. predicted plot data

```
#calibration
cal.plsr.output <- data.frame(cal.plsr.data[, which(names(cal.plsr.data) %notin% "Spectra")],
                             PLSR_Predicted=fit,
                             PLSR_CV_Predicted=as.vector(plsr.out$validation$pred[, ,nComps]))
cal.plsr.output <- cal.plsr.output %>%
  mutate(PLSR_CV_Residuals = PLSR_CV_Predicted-get(inVar))
head(cal.plsr.output)
```

	Plant_Species	Species_Code	Plot	Narea_mg_mm2	Narea_g_m2	PLSR_Predicted
## 1	Ammophila arenaria	Ammare	ZC3	0.03240495	3.240495	2.672029
## 2	Ammophila arenaria	Ammare	MC2	0.02806279	2.806279	2.651863
## 3	Ammophila arenaria	Ammare	ZC1	0.02041612	2.041612	2.178056
## 4	Ammophila arenaria	Ammare	MC1	0.02426549	2.426549	2.412013
## 5	Ammophila arenaria	Ammare	WC3	0.02807281	2.807281	2.452711
## 6	Ammophila arenaria	Ammare	WR3	0.02286678	2.286678	2.792340

```
## PLSR_CV_Predicted PLSR_CV_Residuals
## 1 2.598245 -0.642250440
## 2 2.652066 -0.154212969
## 3 2.200588 0.158975634
## 4 2.435784 0.009234491
## 5 2.384049 -0.423231444
## 6 2.943186 0.656508493

cal.R2 <- round(pls::R2(plsr.out)[[1]][nComps],2)
cal.RMSEP <- round(sqrt(mean(cal.plsr.output$PLSR_CV_Residuals^2)),2)

val.plsr.output <- data.frame(val.plsr.data[, which(names(val.plsr.data) %notin% "Spectra")],
                             PLSR_Predicted=as.vector(predict(plsr.out,
                                                                newdata = val.plsr.data,
                                                                ncomp=nComps, type="response")[, ,1]))
val.plsr.output <- val.plsr.output %>%
  mutate(PLSR_Residuals = PLSR_Predicted-get(inVar))
head(val.plsr.output)
```

	Plant_Species	Species_Code	Plot	Narea_mg_mm2	Narea_g_m2	PLSR_Predicted
## 184	Jacobaea vulgaris	Jacvul	WC2	0.008756996	0.8756996	0.9462916
## 185	Potentilla reptans	Potrep	WC2	0.010313464	1.0313464	1.5386676
## 186	Rubus caesius	Rubcae	WC2	0.007968454	0.7968454	0.8790482
## 187	Urtica dioica	Urtdio	WC2	0.012737560	1.2737560	1.1241560
## 188	Ammophila arenaria	Ammare	WC3	0.028072806	2.8072806	2.4527108
## 189	Jacobaea vulgaris	Jacvul	WC3	0.010251687	1.0251687	1.1553688

```
## PLSR_Residuals
## 184 0.07059201
## 185 0.50732119
## 186 0.08220284
## 187 -0.14959995
## 188 -0.35456980
## 189 0.13020008

val.R2 <- round(pls::R2(plsr.out,newdata=val.plsr.data)[[1]][nComps],2)
val.RMSEP <- round(sqrt(mean(val.plsr.output$PLSR_Residuals^2)),2)

rng_quant <- quantile(cal.plsr.output[,inVar], probs = c(0.001, 0.999))
```

```

cal_scatter_plot <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                                   rng_quant[2]) +

  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Calibration: ", paste0("Rsqr = ", cal.R2), "; ", paste0("RMSEP = ",
                                                                                   cal.RMSEP))) +

  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0, vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

cal_resid_histogram <- ggplot(cal.plsr.output, aes(x=PLSR_CV_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +
  geom_vline(xintercept = 0, color="black",
            linetype="dashed", size=1) + theme_bw() +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0, vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

rng_quant <- quantile(val.plsr.output[,inVar], probs = c(0.001, 0.999))
val_scatter_plot <- ggplot(val.plsr.output, aes(x=PLSR_Predicted, y=get(inVar))) +
  theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",
                                          linetype="dashed", size=1.5) + xlim(rng_quant[1],
                                                                                   rng_quant[2]) +

  ylim(rng_quant[1], rng_quant[2]) +
  labs(x=paste0("Predicted ", paste(inVar), " (units)"),
       y=paste0("Observed ", paste(inVar), " (units)"),
       title=paste0("Validation: ", paste0("Rsqr = ", val.R2), "; ", paste0("RMSEP = ",
                                                                                   val.RMSEP))) +

  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0, vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

val_resid_histogram <- ggplot(val.plsr.output, aes(x=PLSR_Residuals)) +
  geom_histogram(alpha=.5, position="identity") +
  geom_vline(xintercept = 0, color="black",
            linetype="dashed", size=1) + theme_bw() +
  theme(axis.text=element_text(size=18), legend.position="none",
        axis.title=element_text(size=20, face="bold"),
        axis.text.x = element_text(angle = 0, vjust = 0.5),
        panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))

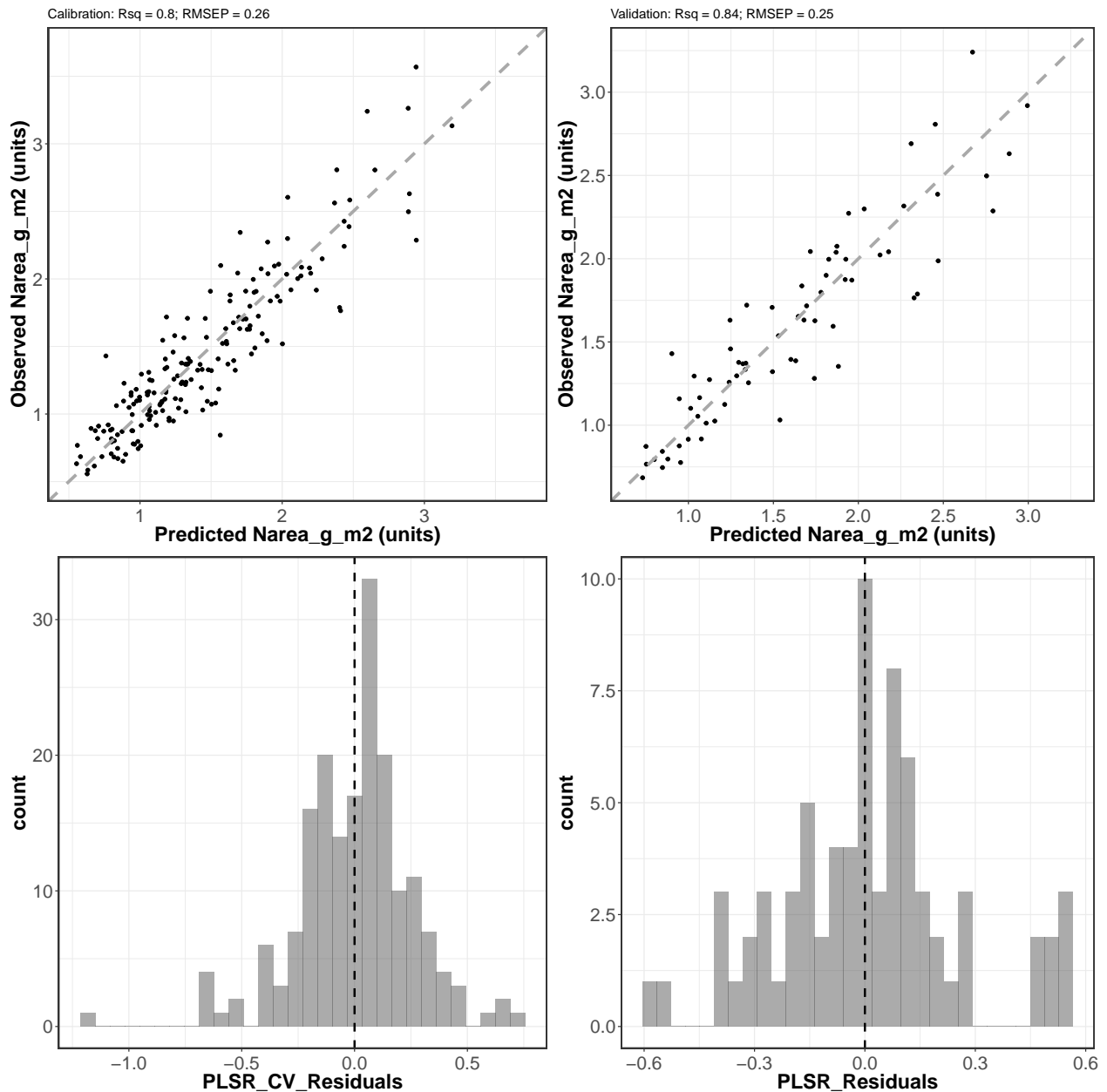
# plot cal/val side-by-side
scatterplots <- grid.arrange(cal_scatter_plot, val_scatter_plot, cal_resid_histogram,
                             val_resid_histogram, nrow=2, ncol=2)

```

```
## Warning: Removed 2 rows containing missing values (geom_point).
```

```
## Warning: Removed 3 rows containing missing values (geom_point).
```

```
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
## `stat_bin()` using `bins = 30`. Pick better value with `binwidth`.
```



```
ggsave(filename = file.path(outdir,paste0(inVar,"_Cal_Val_Scatterplots.png")),
  plot = scatterplots, device="png",
  width = 32,
  height = 30, units = "cm",
  dpi = 300)
```

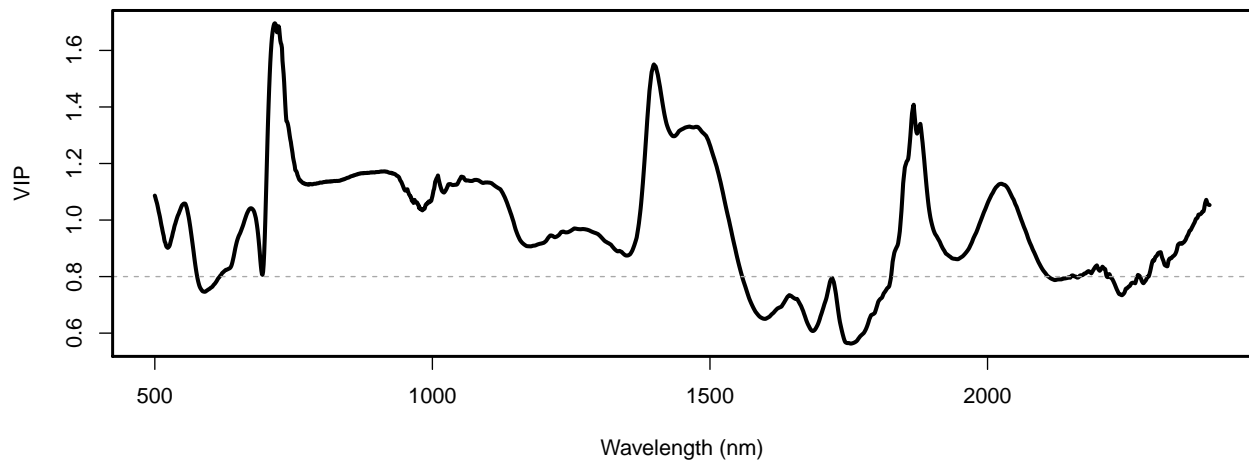
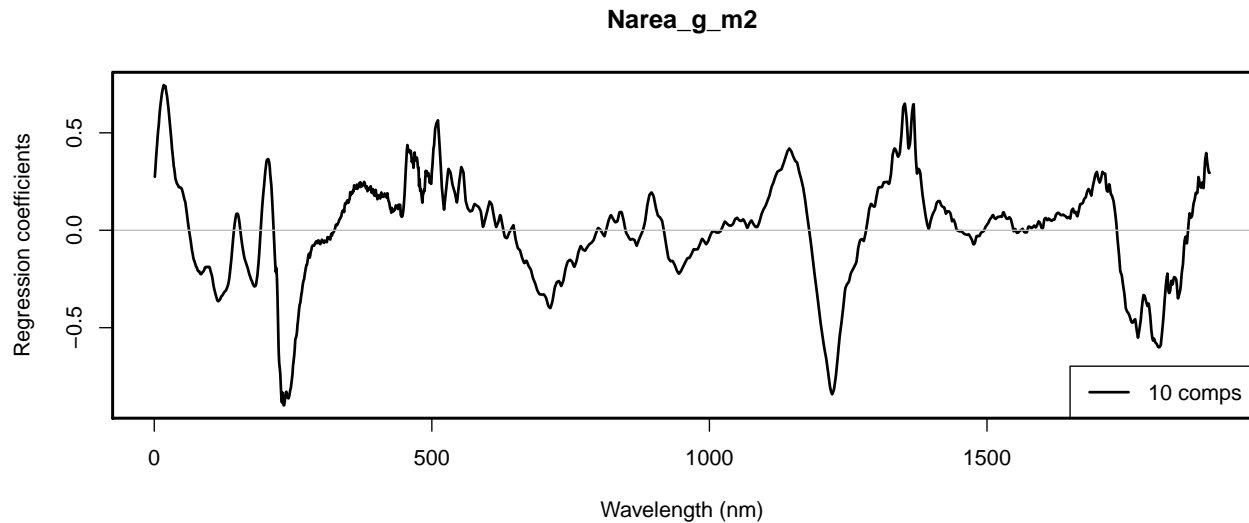
## Generate Coefficient and VIP plots

```
vips <- spectratrait::VIP(plsr.out)[nComps,]
par(mfrow=c(2,1))
```

```

plot(plsr.out, plotype = "coef", xlab="Wavelength (nm)",
     ylab="Regression coefficients", legendpos = "bottomright",
     ncomp=nComps, lwd=2)
box(lwd=2.2)
plot(seq(Start.wave, End.wave, 1), vips, xlab="Wavelength (nm)", ylab="VIP", cex=0.01)
lines(seq(Start.wave, End.wave, 1), vips, lwd=3)
abline(h=0.8, lty=2, col="dark grey")
box(lwd=2.2)

```



```

dev.copy(png, file.path(outdir, paste0(inVar, '_Coefficient_VIP_plot.png')),
         height=3100, width=4100, res=340)

```

```

## quartz_off_screen
##           3

```

```

dev.off();

```

```

## pdf
##    2

```

## Jackknife validation

```

if(grepl("Windows", sessionInfo()$running)){
  pls.options(parallel=NULL)
} else {
  pls.options(parallel = parallel::detectCores()-1)
}

jk.plsr.out <- pls::plsr(as.formula(paste(inVar,"~","Spectra")), scale=FALSE,
                        center=TRUE, ncomp=nComps, validation="LOO", trace=FALSE,
                        jackknife=TRUE,
                        data=cal.plsr.data)
pls.options(parallel = NULL)

Jackknife_coef <- spectratrait::f.coef.valid(plsr.out = jk.plsr.out, data_plsr = cal.plsr.data,
                                             ncomp = nComps, inVar=inVar)
Jackknife_intercept <- Jackknife_coef[1,,]
Jackknife_coef <- Jackknife_coef[2:dim(Jackknife_coef)[1],,,]

interval <- c(0.025,0.975)
Jackknife_Pred <- val.plsr.data$Spectra %*% Jackknife_coef +
  matrix(rep(Jackknife_intercept, length(val.plsr.data[,inVar])), byrow=TRUE,
          ncol=length(Jackknife_intercept))
Interval_Conf <- apply(X = Jackknife_Pred, MARGIN = 1, FUN = quantile,
                      probs=c(interval[1], interval[2]))
sd_mean <- apply(X = Jackknife_Pred, MARGIN = 1, FUN =sd)
sd_res <- sd(val.plsr.output$PLSR_Residuals)
sd_tot <- sqrt(sd_mean^2+sd_res^2)
val.plsr.output$LCI <- Interval_Conf[1,]
val.plsr.output$UCI <- Interval_Conf[2,]
val.plsr.output$LPI <- val.plsr.output$PLSR_Predicted-1.96*sd_tot
val.plsr.output$UPI <- val.plsr.output$PLSR_Predicted+1.96*sd_tot
head(val.plsr.output)

##          Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2 PLSR_Predicted
## 184  Jacobaea vulgaris      Jacvul  WC2  0.008756996  0.8756996      0.9462916
## 185  Potentilla reptans     Potrep  WC2  0.010313464  1.0313464      1.5386676
## 186    Rubus caesius       Rubcae  WC2  0.007968454  0.7968454      0.8790482
## 187    Urtica dioica       Urtdio  WC2  0.012737560  1.2737560      1.1241560
## 188  Ammophila arenaria     Ammare  WC3  0.028072806  2.8072806      2.4527108
## 189  Jacobaea vulgaris      Jacvul  WC3  0.010251687  1.0251687      1.1553688
##          PLSR_Residuals      LCI      UCI      LPI      UPI
## 184    0.07059201 0.9154961 0.9532972 0.4623162 1.430267
## 185    0.50732119 1.4875834 1.5528063 1.0540777 2.023258
## 186    0.08220284 0.8472007 0.9329303 0.3936085 1.364488
## 187   -0.14959995 1.1075928 1.1743800 0.6395189 1.608793
## 188   -0.35456980 2.4248448 2.5638131 1.9651833 2.940238
## 189    0.13020008 1.1262731 1.1615642 0.6713762 1.639361

val.plsr.output$LPI <- val.plsr.output$PLSR_Predicted-1.96*sd_tot
val.plsr.output$UPI <- val.plsr.output$PLSR_Predicted+1.96*sd_tot
head(val.plsr.output)

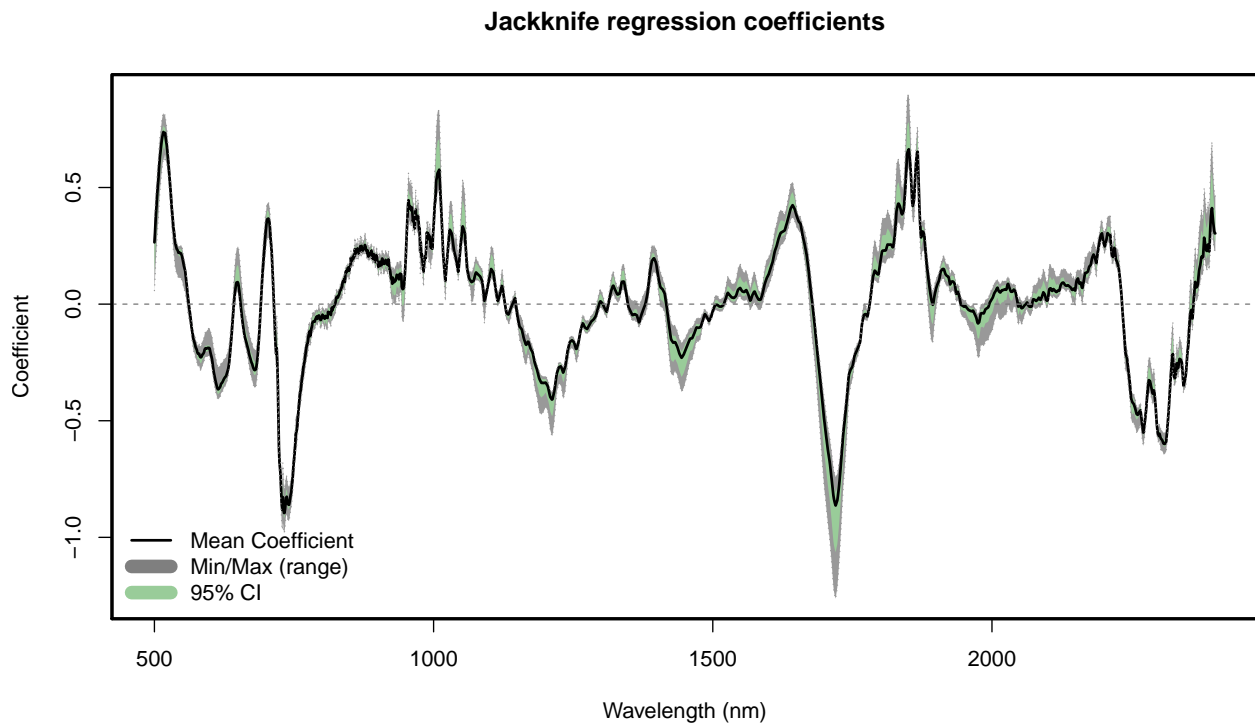
##          Plant_Species Species_Code Plot Narea_mg_mm2 Narea_g_m2 PLSR_Predicted

```

## 184	Jacobaea vulgaris	Jacvul	WC2	0.008756996	0.8756996	0.9462916
## 185	Potentilla reptans	Potrep	WC2	0.010313464	1.0313464	1.5386676
## 186	Rubus caesius	Rubcae	WC2	0.007968454	0.7968454	0.8790482
## 187	Urtica dioica	Urtdio	WC2	0.012737560	1.2737560	1.1241560
## 188	Ammophila arenaria	Ammare	WC3	0.028072806	2.8072806	2.4527108
## 189	Jacobaea vulgaris	Jacvul	WC3	0.010251687	1.0251687	1.1553688
##	PLSR_Residuals	LCI	UCI	LPI	UPI	
## 184	0.07059201	0.9154961	0.9532972	0.4623162	1.430267	
## 185	0.50732119	1.4875834	1.5528063	1.0540777	2.023258	
## 186	0.08220284	0.8472007	0.9329303	0.3936085	1.364488	
## 187	-0.14959995	1.1075928	1.1743800	0.6395189	1.608793	
## 188	-0.35456980	2.4248448	2.5638131	1.9651833	2.940238	
## 189	0.13020008	1.1262731	1.1615642	0.6713762	1.639361	

### Jackknife coefficient plot

```
spectratrait::f.plot.coef(Z = t(Jackknife_coef), vv = vv,
  plot_label="Jackknife regression coefficients",position = 'bottomleft')
abline(h=0,lty=2,col="grey50")
box(lwd=2.2)
```



```
dev.copy(png,file.path(outdir,paste0(inVar,'_Jackknife_Regression_Coefficients.png')),
  height=2100, width=3800, res=340)
```

```
## quartz_off_screen
## 3
```

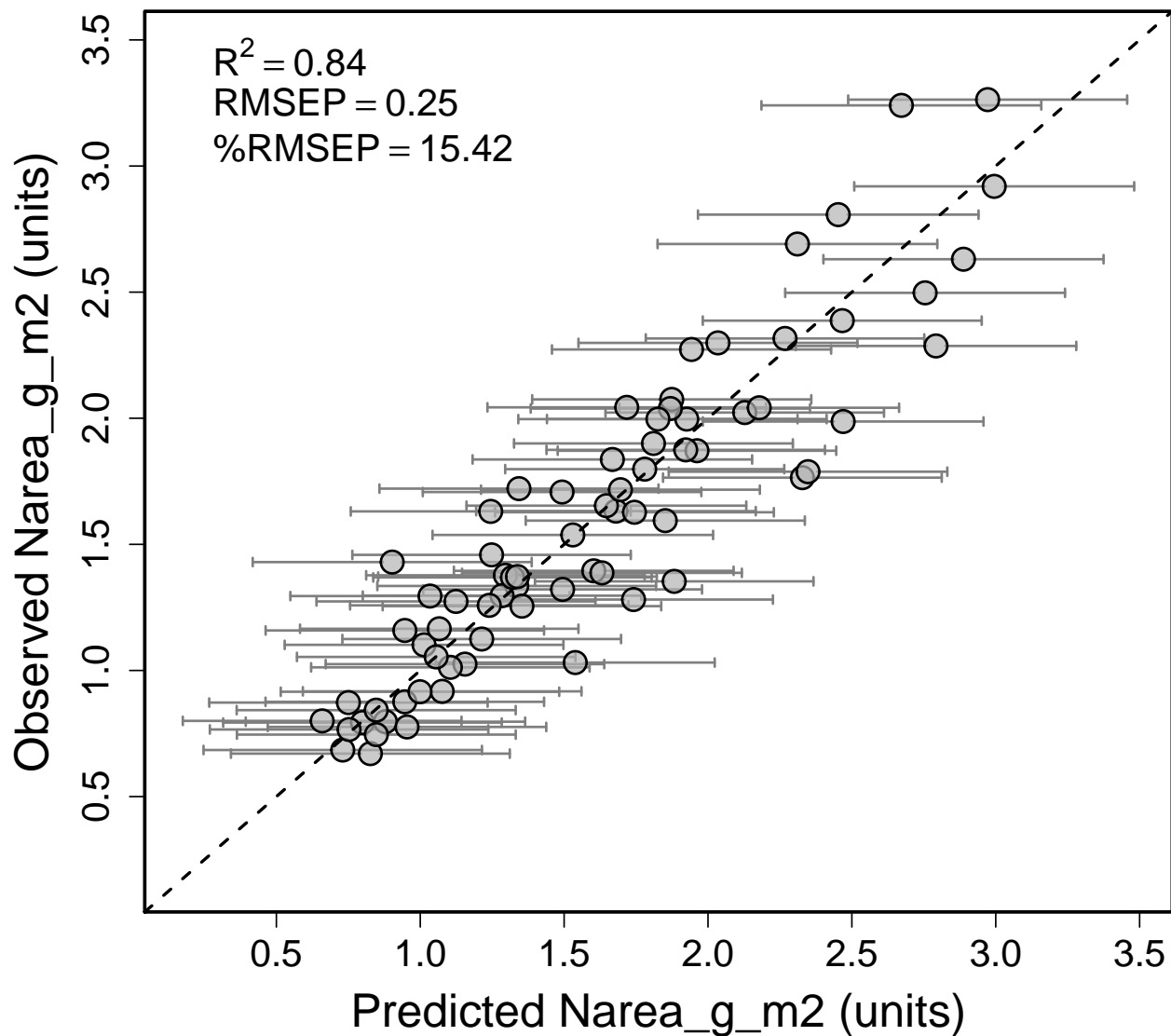
```
dev.off();
```

```
## pdf
## 2
```



## Jackknife validation plot

```
RMSEP <- sqrt(mean(val.plsr.output$PLSR_Residuals^2))
pecr_RMSEP <- RMSEP/mean(val.plsr.output[,inVar])*100
r2 <- round(pls::R2(plsr.out, newdata = val.plsr.data)$val[nComps+1],2)
expr <- vector("expression", 3)
expr[[1]] <- bquote(R^2==.(r2))
expr[[2]] <- bquote(RMSEP==.(round(RMSEP,2)))
expr[[3]] <- bquote("%RMSEP"==.(round(pecr_RMSEP,2)))
rng_vals <- c(min(val.plsr.output$LPI), max(val.plsr.output$UPI))
par(mfrow=c(1,1), mar=c(4.2,5.3,1,0.4), oma=c(0, 0.1, 0, 0.2))
plotrix::plotCI(val.plsr.output$PLSR_Predicted, val.plsr.output[,inVar],
  li=val.plsr.output$LPI, ui=val.plsr.output$UPI, gap=0.009, sfrac=0.004,
  lwd=1.6, xlim=c(rng_vals[1], rng_vals[2]), ylim=c(rng_vals[1], rng_vals[2]),
  err="x", pch=21, col="black", pt.bg=scales::alpha("grey70",0.7), scol="grey50",
  cex=2, xlab=paste0("Predicted ", paste(inVar), " (units)"),
  ylab=paste0("Observed ", paste(inVar), " (units)"),
  cex.axis=1.5, cex.lab=1.8)
abline(0,1,lty=2,lw=2)
legend("topleft", legend=expr, bty="n", cex=1.5)
box(lwd=2.2)
```



```
dev.copy(png,file.path(outdir,paste0(inVar,"_PLSR_Validation_Scatterplot.png")),
         height=2800, width=3200, res=340)
```

```
## quartz_off_screen
##           3
```

```
dev.off();
```

```
## pdf
##    2
```

#### Output jackknife results

```
out.jk.coefs <- data.frame(Iteration=seq(1,length(Jackknife_intercept),1),
                           Intercept=Jackknife_intercept,t(Jackknife_coef))
head(out.jk.coefs)[1:6]
```

```
##      Iteration  Intercept Wave_500 Wave_501 Wave_502 Wave_503
## Seg 1         1 -0.001089661 0.3156927 0.3524556 0.3947195 0.4329382
```

```
## Seg 2      2  0.082969588 0.2989509 0.3382983 0.3835509 0.4239103
## Seg 3      3  0.114879574 0.2716867 0.3122469 0.3574386 0.3982935
## Seg 4      4  0.178884696 0.2099486 0.2520760 0.3018899 0.3452178
## Seg 5      5  0.126339690 0.2898707 0.3311239 0.3762377 0.4163999
## Seg 6      6 -0.085381533 0.2805890 0.3195387 0.3625074 0.4023830

write.csv(out.jk.coefs,file=file.path(outdir,
                                     paste0(inVar,
                                              '_Jackkife_PLSR_Coefficients.csv')),
          row.names=FALSE)
```

## Create core PLSR outputs

```
print(paste("Output directory: ", outdir))

## [1] "Output directory:  /var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjQ/T//Rtmpi5zkMP"

# Observed versus predicted
write.csv(cal.plsr.output,file=file.path(outdir,
                                         paste0(inVar,'_Observed_PLSR_CV_Pred_',
                                                  nComps,'comp.csv')),
          row.names=FALSE)

# Validation data
write.csv(val.plsr.output,file=file.path(outdir,
                                         paste0(inVar,'_Validation_PLSR_Pred_',
                                                  nComps,'comp.csv')),
          row.names=FALSE)

# Model coefficients
coefs <- coef(plsr.out,ncomp=nComps,intercept=TRUE)
write.csv(coefs,file=file.path(outdir,
                               paste0(inVar,'_PLSR_Coefficients_',
                                       nComps,'comp.csv')),
          row.names=TRUE)

# PLSR VIP
write.csv(vips,file=file.path(outdir,
                              paste0(inVar,'_PLSR_VIPs_',
                                      nComps,'comp.csv')))
```

## Confirm files were written to temp space

```
print("**** PLSR output files: ")

## [1] "**** PLSR output files: "

print(list.files(outdir)[grep(pattern = inVar, list.files(outdir))])

## [1] "Narea_g_m2_Cal_PLSR_Dataset.csv"
## [2] "Narea_g_m2_Cal_Val_Histograms.png"
## [3] "Narea_g_m2_Cal_Val_Scatterplots.png"
## [4] "Narea_g_m2_Cal_Val_Spectra.png"
```

```
## [5] "Narea_g_m2_Coefficient_VIP_plot.png"
## [6] "Narea_g_m2_Jackknife_PLSR_Coefficients.csv"
## [7] "Narea_g_m2_Jackknife_Regression_Coefficients.png"
## [8] "Narea_g_m2_Observed_PLSR_CV_Pred_10comp.csv"
## [9] "Narea_g_m2_PLSR_Coefficients_10comp.csv"
## [10] "Narea_g_m2_PLSR_Component_Selection.png"
## [11] "Narea_g_m2_PLSR_Validation_Scatterplot.png"
## [12] "Narea_g_m2_PLSR_VIPs_10comp.csv"
## [13] "Narea_g_m2_Val_PLSR_Dataset.csv"
## [14] "Narea_g_m2_Validation_PLSR_Pred_10comp.csv"
## [15] "Narea_g_m2_Validation_RMSEP_R2_by_Component.png"
```