A basic PLSR example using leaf-level specrta and leaf mass per area (LMA) data from several CONUS NEON sites

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Overview

his is an R Markdown Notebook to illustrate how to conduct a basic model fit. This example shows you how to retrieve a dataset from the EcoSIS spectral database, choose the "optimal" number of plsr components, and fit a plsr model for leaf-mass area

When you click the **Knit** button in Rstudio a document will be generated that includes both content as well as the output of any embedded R code chunks within the document.

Getting Started

Installation

```
list.of.packages <- c("readr","httr","pls","dplyr","reshape2") # packages needed for script
# check for dependencies and install if needed
new.packages <- list.of.packages[!(list.of.packages %in% installed.packages()[,"Package"])]
if(length(new.packages)) install.packages(new.packages)</pre>
```

Load libraries

```
# load libraries needed for script
library(pls)
library(readr)
library(dplyr)
library(reshape2)
library(ggplot2)
library(dplyr)
```

Prepare helpers

```
# define function to grab PLSR model from GitHub
#devtools::source_gist("gist.github.com/christophergandrud/4466237")
source_GitHubData <-function(url, sep = ",", header = TRUE) {
    require(httr)
    request <- GET(url)
    stop_for_status(request)
    handle <- textConnection(content(request, as = 'text'))
    on.exit(close(handle))
    read.table(handle, sep = sep, header = header)
}</pre>
```

Setup temporary folder

```
print(paste0("Output temporary directory: ",outdir))
[1] "Output temporary directory: /var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjq/T//RtmpkzZB8m"
setwd(outdir) # set working directory
getwd() # check wd
```

[1] "/private/var/folders/xp/h3k9vf3n2jx181ts786_yjrn9c2gjq/T/RtmpkzZB8m"

Grab data from EcoSIS

```
print("**** Downloading Ecosis data ****")

[1] "**** Downloading Ecosis data ****"

ecosis_id <- "5617da17-c925-49fb-b395-45a51291bd2d" # NEON dataset

ecosis_file <- sprintf(
    "https://ecosis.org/api/package/%s/export?metadata=true",
    ecosis_id
)

message("Downloading data...")
dat_raw <- read_csv(ecosis_file)
message("Download complete!")
head(dat_raw)</pre>
```

A tibble: 6 x 2,162

```
Affiliation Common Name Domain Functional_type LMA Latin Genus 1 University~ black walnut D02 broadleaf 72.9 Juglans 2 University~ black walnut D02 broadleaf 72.9 Juglans 3 University~ black walnut D02 broadleaf 60.8 Juglans 4 University~ black walnut D02 broadleaf 60.8 Juglans 5 University~ black walnut D02 broadleaf 85.9 Juglans
```

```
6 University~ black walnut D02 broadleaf 85.9 Juglans
# ... with 2,156 more variables: Latin Species, PI, # Project, Sample_ID, USDA Symbol, 350, #
351, 352, 353, 354, 355, # 356, 357, 358, 359, 360, # 361, 362, 363, 364, 365, # 366, 367,
368, 369, 370, #371, 372, 373, 374, 375, #376, 377, 378, 379, 380, #381, 382, 383, 384,
385, # 386, 387, 388, 389, 390, # 391, 392, 393, 394, 395, # 396, 397, 398, 399, 400, # 401,
402, 403, 404, 405, \# 406, 407, 408, 409, 410, \# 411, 412, 413, 414, 415, \# 416, 417, 418,
419, 420, # 421, 422, 423, 424, 425, # 426, 427, 428, 429, 430, # 431, 432, 433, 434, 435, #
436 \;,\, 437 \;,\, 438 \;,\, 439 \;,\, 440 \;,\, \# \; 441 \;,\, 442 \;,\, 443 \;,\, 444 \;,\, \dots
names(dat raw)[1:40]
[1] "Affiliation" "Common Name" "Domain" "Functional_type" [5] "LMA" "Latin Genus" "Latin Species"
"PI"
[9] "Project" "Sample_ID" "USDA Symbol" "350"
[13] "351" "352" "353" "354"
[17] "355" "356" "357" "358"
[21] "359" "360" "361" "362"
[25] "363" "364" "365" "366"
[29] "367" "368" "369" "370"
[33] "371" "372" "373" "374"
[37] "375" "376" "377" "378"
```

Create PLSR dataset

```
Start.wave <- 500
End.wave <- 2400
wv <- seq(Start.wave,End.wave,1)

spectra <- data.frame(dat_raw[,names(dat_raw) %in% wv])
names(spectra) <- c(paste0("Wave_",wv))
head(spectra)[1:6,1:10]</pre>
```

```
sample_info <- dat_raw[,names(dat_raw) %notin% seq(350,2500,1)]
head(sample_info)</pre>
```

A tibble: 6×11

```
Affiliation Common Name Domain Functional_type LMA Latin Genus

1 University~ black walnut D02 broadleaf 72.9 Juglans

2 University~ black walnut D02 broadleaf 60.8 Juglans

3 University~ black walnut D02 broadleaf 60.8 Juglans

4 University~ black walnut D02 broadleaf 60.8 Juglans

5 University~ black walnut D02 broadleaf 85.9 Juglans

6 University~ black walnut D02 broadleaf 85.9 Juglans

# ... with 5 more variables: Latin Species , PI , Project , # Sample_ID , USDA Symbol
```

```
sample_info2 <- sample_info %>%
    select(Domain,Functional_type,Sample_ID,USDA_Species_Code=`USDA Symbol`,LMA_gDW_m2=LMA)
head(sample_info2)
```

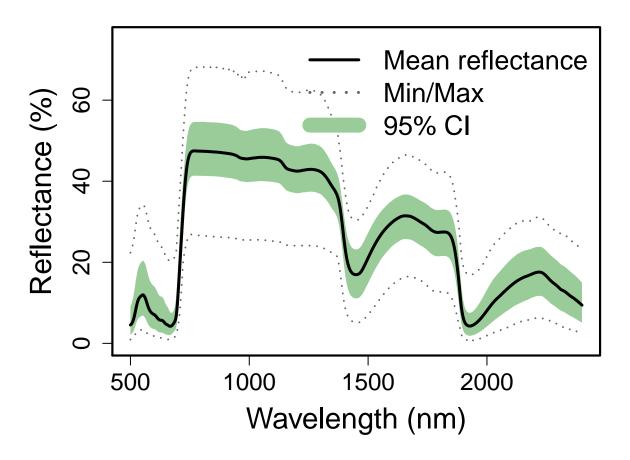
A tibble: 6 x 5

Domain Functional_type Sample_ID USDA_Species_Code LMA_gDW_m2 1 D02 broadleaf P0001 JUNI 72.9 2 D02 broadleaf L0001 JUNI 72.9 3 D02 broadleaf P0002 JUNI 60.8 4 D02 broadleaf L0002 JUNI 60.8 5 D02 broadleaf P0003 JUNI 85.9 6 D02 broadleaf L0003 JUNI 85.9

```
plsr_data <- data.frame(sample_info2,Spectra=I(as.matrix(spectra)))
inVar <- "LMA_gDW_m2"</pre>
```

Plot the spectra

```
cexaxis <- 1.5
cexlab <- 1.8
ylim <- 75
mean_spec <- colMeans(spectra[,which(names(spectra) %in% paste0("Wave_",wv))])</pre>
spectra_quantiles <- apply(spectra[,which(names(spectra) %in% paste0("Wave_",wv))],</pre>
                           2,quantile,na.rm=T,probs=c(0,0.025,0.05,0.5,0.95,0.975,1))
print("**** Plotting specrtal data ****")
[1] "**** Plotting specrtal data ****"
par(mfrow=c(1,1), mar=c(4.5,5.7,0.3,0.4), oma=c(0.3,0.9,0.3,0.1)) # B, L, T, R
plot(wv,mean_spec,ylim=c(0,ylim),cex=0.00001, col="white",xlab="Wavelength (nm)",
     ylab="Reflectance (%)",cex.axis=cexaxis, cex.lab=cexlab)
polygon(c(wv ,rev(wv)),c(spectra_quantiles[5,]*100, rev(spectra_quantiles[3,]*100)),
        col="#99CC99",border=NA)
lines(wv,mean_spec*100,lwd=3, lty=1, col="black")
lines(wv,spectra quantiles[1,]*100,lwd=1.85, lty=3, col="grey40")
lines(wv,spectra quantiles[7,]*100,lwd=1.85, lty=3, col="grey40")
legend("topright",legend=c("Mean reflectance","Min/Max", "95% CI"),lty=c(1,3,1),
       lwd=c(3,3,15),col=c("black","grey40","#99CC99"),bty="n", cex=1.7)
box(1wd=2.2)
```

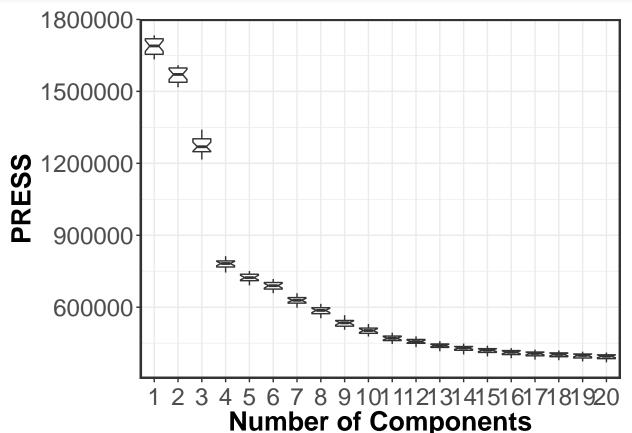


Run Jackknife test to find number of components - simple example

```
dims <- dim(plsr_data)</pre>
nComps <- 20
iterations <- 20
seg <- 5
prop <- 0.70
jk.out <- matrix(data=NA, nrow=iterations, ncol=nComps)</pre>
pls.options(parallel = parallel::detectCores()-1) # Use mclapply
print("*** Running jacknife permutation test. Please hang tight, this can take awhile ***")
[1] "*** Running jacknife permutation test. Please hang tight, this can take awhile ***"
start.time <- Sys.time()</pre>
for (i in 1:iterations) {
  rows <- sample(1:nrow(plsr_data),floor(prop*nrow(plsr_data)))</pre>
  sub.data <- plsr_data[rows,]</pre>
  plsr.out <- plsr(as.formula(paste(inVar,"~","Spectra")), scale=FALSE, center=TRUE, ncomp=nComps,
                    validation="CV", segments = seg, segment.type="interleaved", trace=FALSE, data=sub.d
  resPRESS <- as.vector(plsr.out$validation$PRESS)</pre>
  jk.out[i,seq(plsr.out$validation$ncomp)]=resPRESS
}
end.time <- Sys.time()
end.time - start.time
```

Time difference of 1.753883 mins

PRESS plot



Calculate optimal number of components

```
if (j > nComps) {
    break
}

results <- data.frame(seq(2,nComps,1),results)
names(results) <- c("Component", "P.value")
results</pre>
```

 $\begin{array}{c} \text{Component P.value 1 2 0 2 3 0 3 4 0 4 5 0 5 6 5.3e-07 6 7 0 7 8 0 8 9 0 9 10 1.3e-06 10 11 3e-08 11 12 0.0026535 12 13 3.129e-05 13 14 0.00918502 14 15 0.02547395 15 16 0.03095163 16 17 0.12853979 17 18 0.35387817 18 19 0.21422969 19 20 0.41859186 \\ \end{array}$

```
# *** based on t.test - optimal components are 16 ***
# NOTE: Becuase the jacknife test above depends on random selection
# the optimal components may change slightly between differe runs of this script
# This is expected given different permutatoins and uncertainty in the data
```

Final PLSR model fit

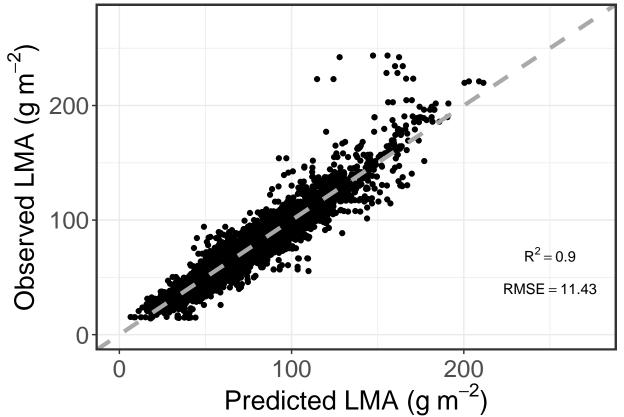
```
# Simple final model validated with cross-validation. Segmented cross-validation used
# given the very large sample size. For models with fewer observations (e.g. <100)
# LOO or leave-one-out cross validation is recommended
#nComps <- 14
first <- min(which(as.numeric(as.character(results$P.value)) > 0.05))
nComps <- results$Component[first]</pre>
print(paste0("*** Optimal number of components based on t.test: ", nComps))
[1] "*** Optimal number of components based on t.test: 17"
segs <- 30
pls.options(parallel = NULL)
plsr.out <- plsr(as.formula(paste(inVar,"~","Spectra")),scale=FALSE,ncomp=nComps,validation="CV",
                 segments=segs, segment.type="interleaved",trace=TRUE,data=plsr_data)
Segment: 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30
fit <- plsr.out$fitted.values[,1,nComps]</pre>
plot_data <- data.frame(plsr_data[, which(names(plsr_data) %notin% "Spectra")], Fitted=fit)</pre>
plot data <- plot data %>%
  mutate(Residuals = Fitted-LMA_gDW_m2)
head(plot_data)
```

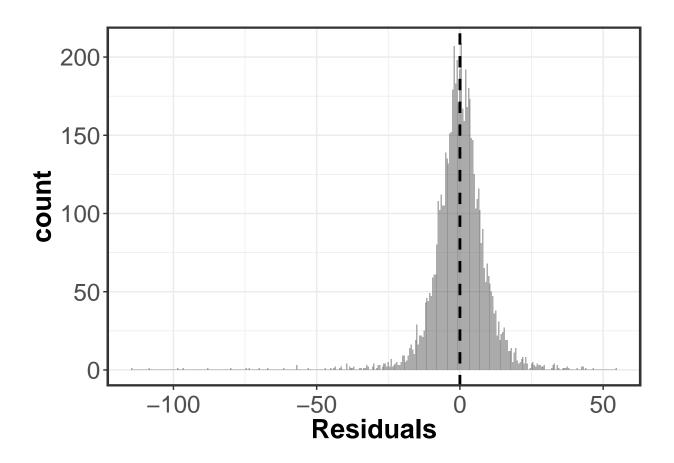
Domain Functional_type Sample_ID USDA_Species_Code LMA_gDW_m2 Fitted 1 D02 broadleaf P0001 JUNI 72.87 74.66709 2 D02 broadleaf L0001 JUNI 72.87 80.85354 3 D02 broadleaf P0002 JUNI 60.77 63.70678 4 D02 broadleaf L0002 JUNI 60.77 53.18834 5 D02 broadleaf P0003 JUNI 85.92 83.66721 6 D02 broadleaf L0003 JUNI 85.92 85.28082 Residuals 1 1.7970935 2 7.9835431 3 2.9367822 4 -7.5816605 5 -2.2527884 6 -0.6391764

Basic scatter plot of results

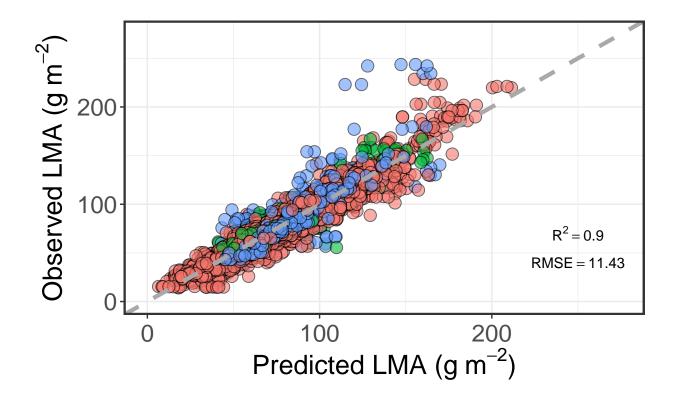
```
scatter_plot <- ggplot(plot_data, aes(x=Fitted, y=LMA_gDW_m2)) +
   theme_bw() + geom_point() + geom_abline(intercept = 0, slope = 1, color="dark grey",</pre>
```

```
linetype="dashed", size=1.5) + xlim(0, 275) + ylim(0, 275) +
labs(x=expression(paste("Predicted LMA (",g~m^{-2},")")),
    y=expression(paste("Observed LMA (",g~m^{-2},")"))) +
annotate("text", x=250, y=70, label = paste0("R^2 == ", round(pls::R2(plsr.out)[[1]][nComps],2)), par
annotate("text", x=250, y=40, label = paste0("RMSE == ", round(pls::RMSEP(plsr.out)[[1]][nComps],2)),
theme(axis.text=element_text(size=18), legend.position="none",
    axis.title=element_text(size=20, face="bold"),
    axis.text.x = element_text(angle = 0,vjust = 0.5),
    panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))
scatter_plot
```

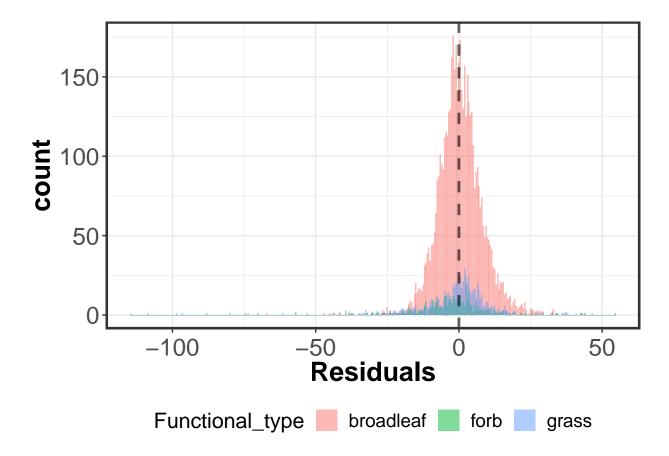




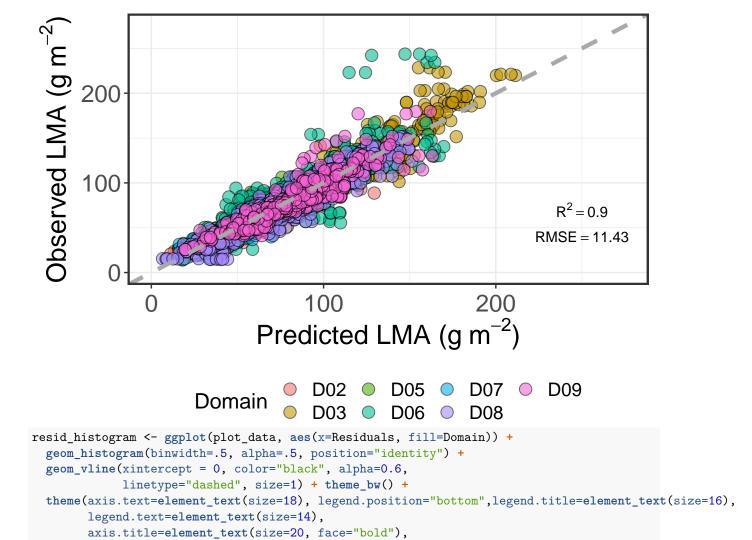
Scatter plot by Functional_type



Functional_type • broadleaf • forb • grass



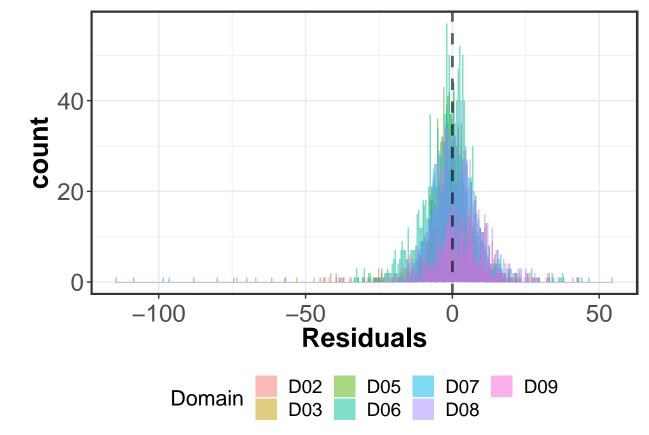
Scatter plot by Domain



axis.text.x = element_text(angle = 0, vjust = 0.5),

resid_histogram

panel.border = element_rect(linetype = "solid", fill = NA, size=1.5))



eof