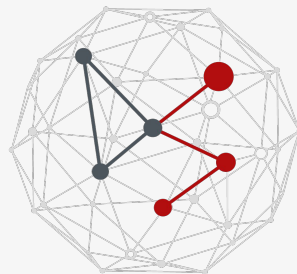


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UNIVERSITÀ  
DEGLI STUDI  
DI PADOVA

  DIPARTIMENTO  
**MATEMATICA**



**DATA SCIENCE**  
UNIVERSITY OF PADOVA

# COMPARATIVE MODELLING

Master of Science in Data Science

**Damiano Piovesan**



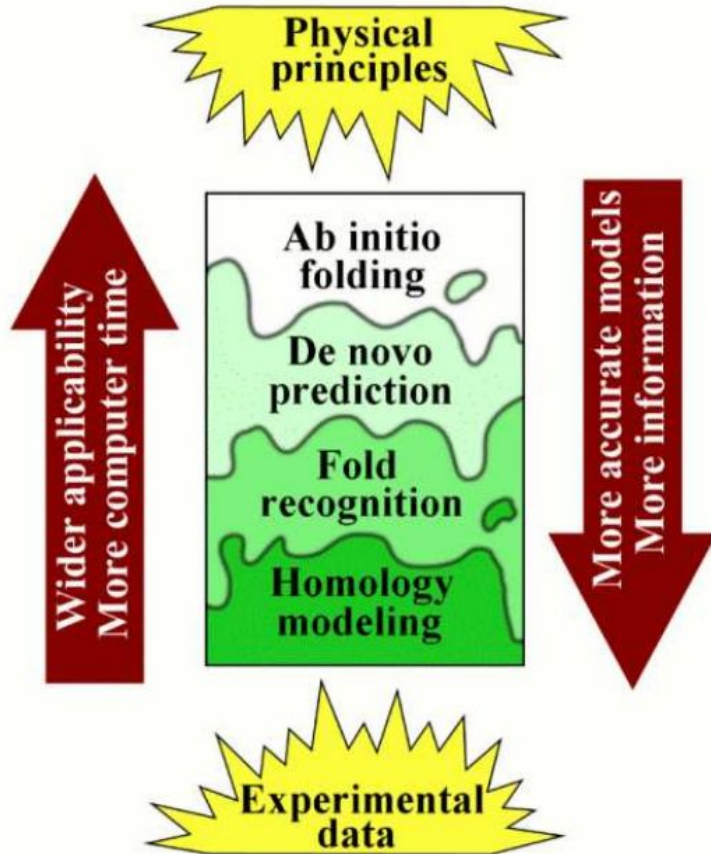
# Motivation

The **protein structure** allows a molecular and mechanistic understanding of the **protein function**

- Identification of **active sites** and the positions of **key residues**
- Prediction of protein-protein and protein-ligand **interactions**, which are mostly determined by steric (shape) and chemical (e.g. charge) **complementarity**
- Filling the **sequence / structure gap**. Million sequences are known, while the PDB contain only ca. 200K structures

However

- Many proteins have the sequence not similar enough to build an *in silico* model by homology
- Many folds are not represented in the PDB



- **De novo prediction / Ab initio**

- Secondary structure prediction; conformation of short fragments (Rosetta); molecular dynamics; Monte Carlo; quantum mechanics (unfeasible)

- Tough computation

- **Fold recognition**

- Try to fit with known folds

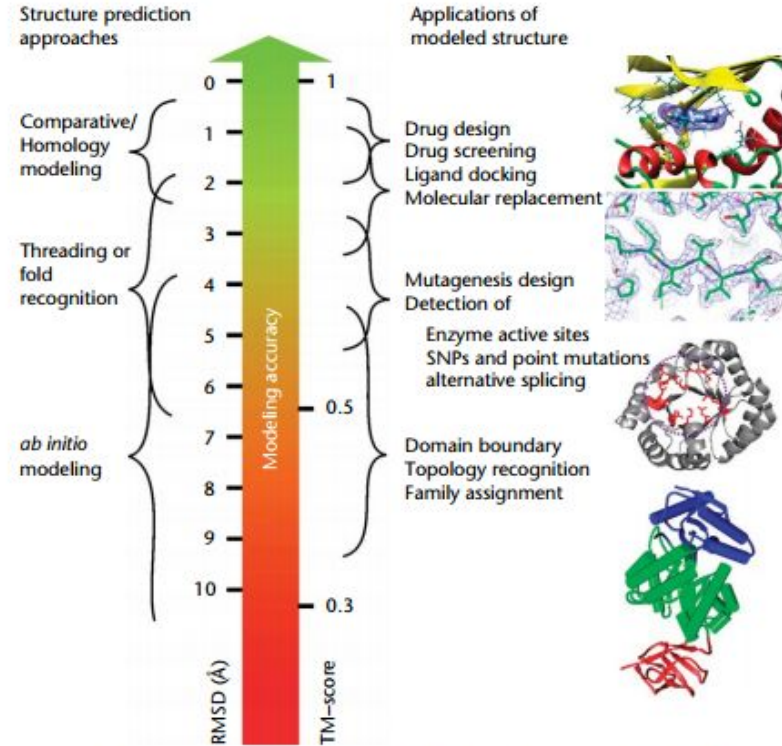
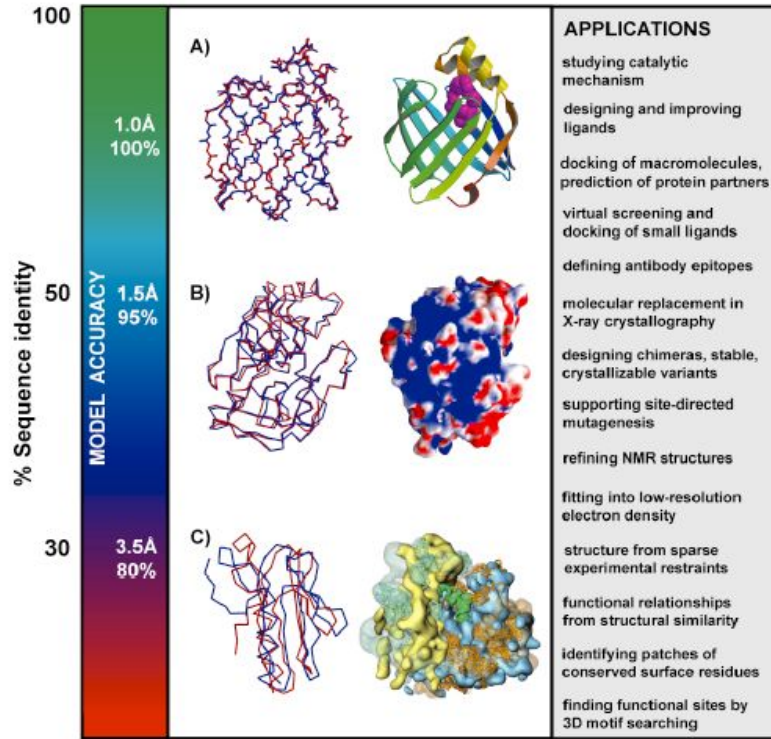
- The fold space is not completely known (50% success)

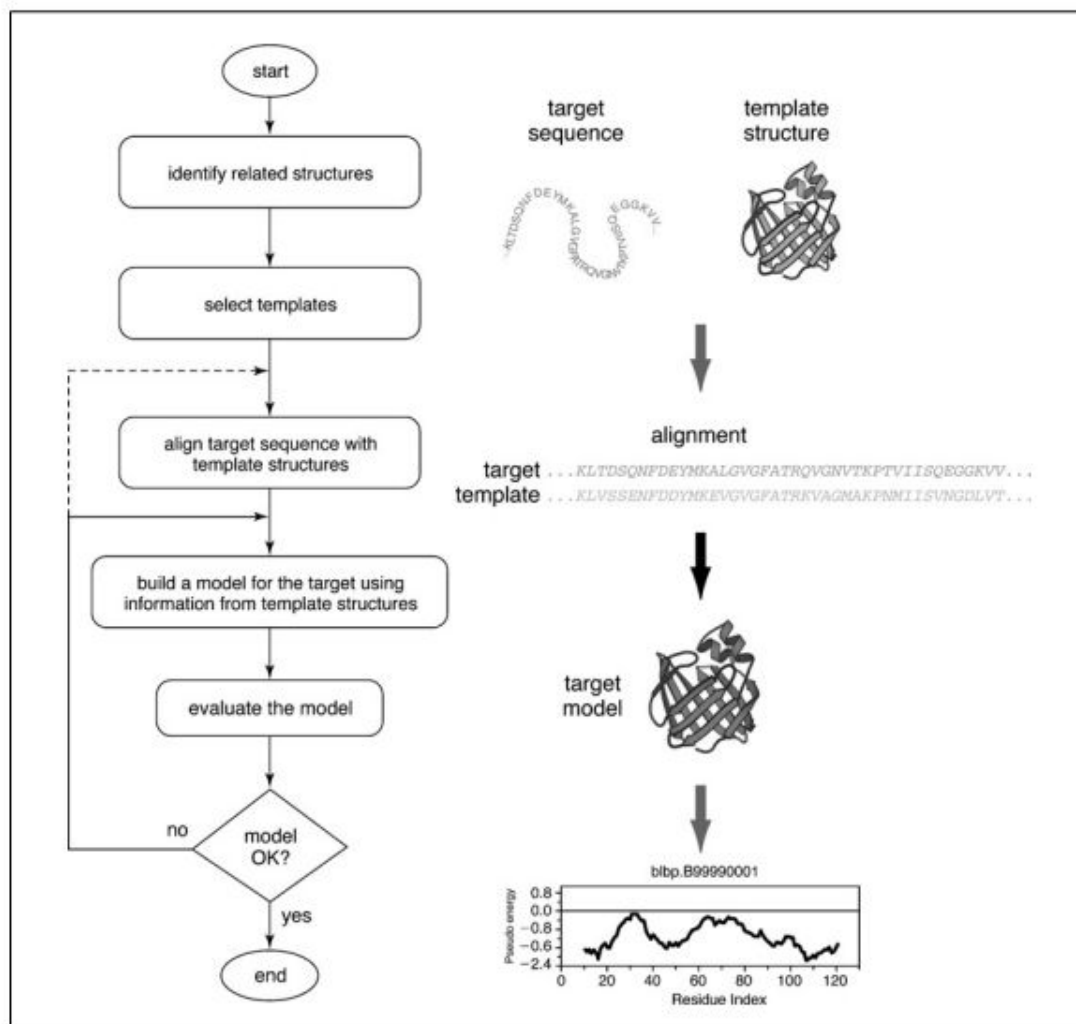
- **Homology modelling**

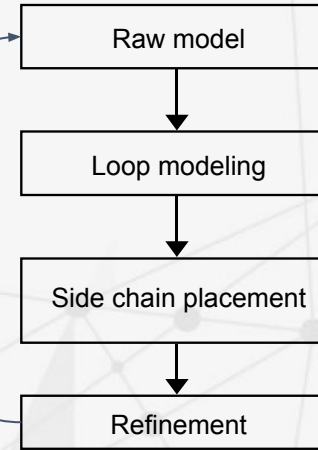
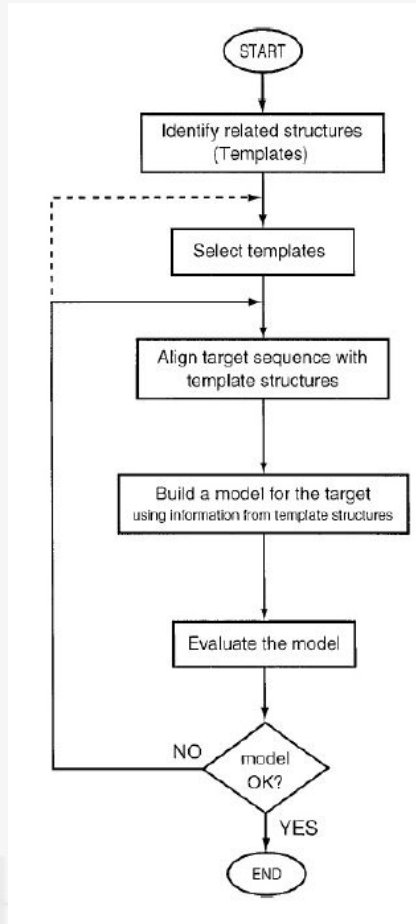
- Similar sequences have similar structures (+50% sequence identity)

- 40% of genes are not homologous to known structures



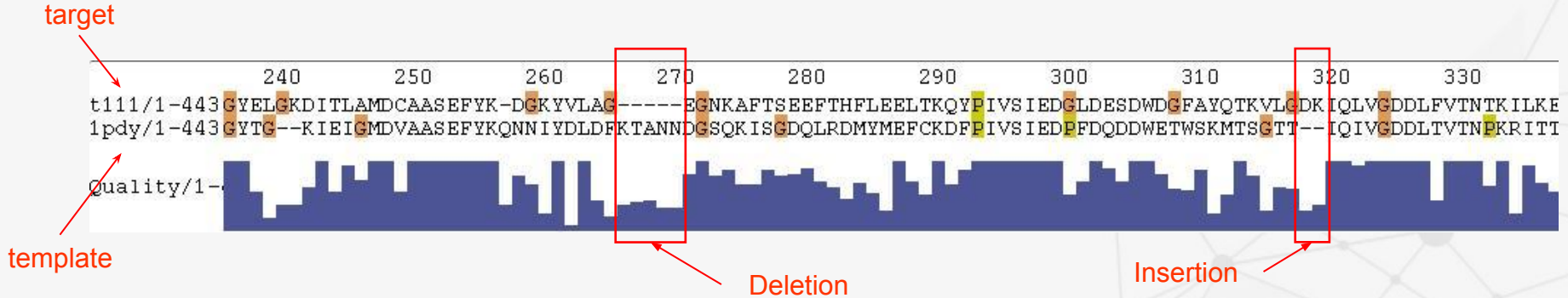








# Alignment



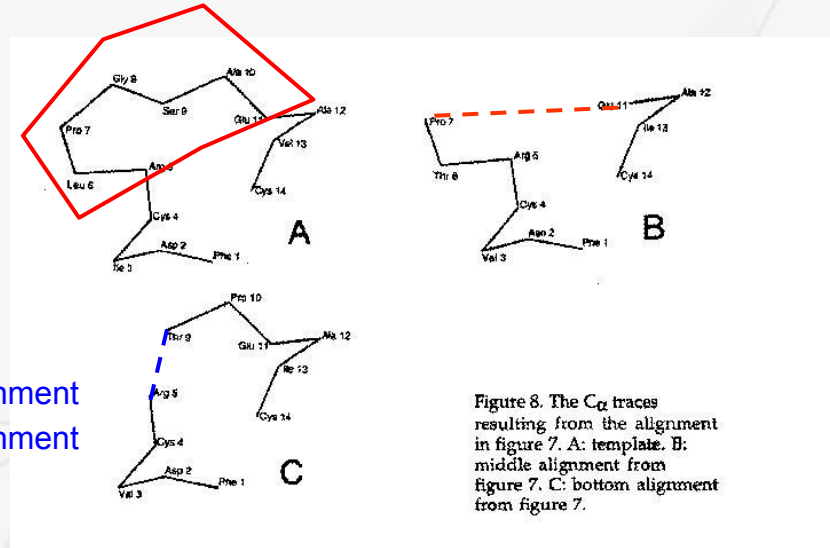
- **Database search** - Find homologous sequences with known structure. Generally a heuristic **PSI-BLAST / BLAST**
- Assign **equivalent positions** between target and template. Determine insertion and deletion. Optimal alignment with **Smith-Waterman** (local) or **Needleman-Wunsch** (global) algorithms



# Improve the sequence alignment

- Errors in the alignment cannot be corrected in the following steps!
- Often the best sequence alignment is non optimal for the structure

How you model this?



Worse sequence alignment  
Better structure alignment

	1	2	3	4	5	6	7	8	9	10	11	12	13	14	
A	PHE	ASP	ILE	CYS	ARG	LEU	PRO	GLY	SER	ALA	GLU	ALA	VAL	CYS	(green in fig 8)
B	PHE	ASN	VAL	CYS	ARG	THR	PRO	---	---	---	GLU	ALA	ILE	CYS	(red in fig 8)
C	PHE	ASN	VAL	CYS	ARG	---	---	---	THR	PRO	GLU	ALA	ILE	CYS	(blue in fig 8)

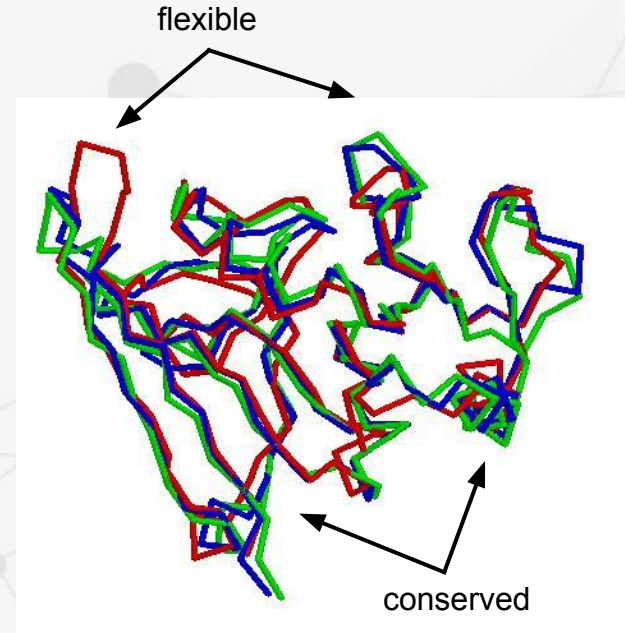
Figure 7. Example of sequence alignment in an area where a deletion needs to be modelled.



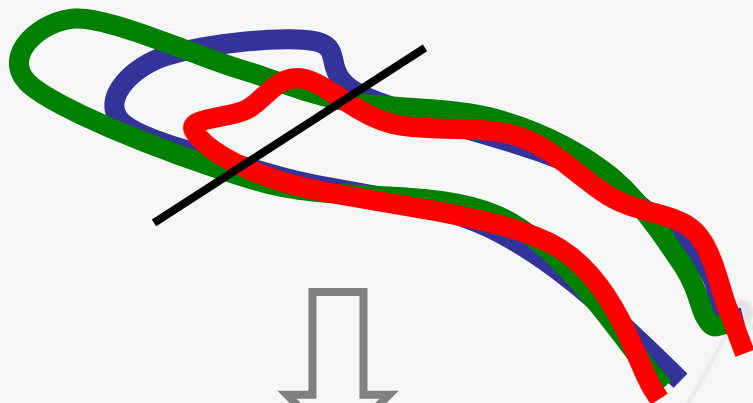


# Building the raw model

- 3D coordinates of the **template** residues can be directly used
- The **variable regions** of the structures (generally loops) and in particular position near indels have to be **predicted**
- Two principal methods are used for the construction
  - Fragment-based
  - Restraint-based



# Fragment-based building



Idea → Copy “useful” coordinates of fragments

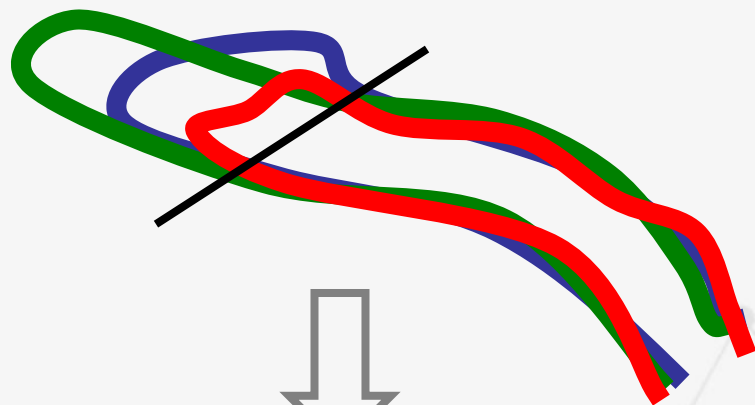
- Build a “sharp” set
- Keeps the geometry, eg the active site

Software

- 3D-JIGSAW (Bates et al.)
- COMPOSER (Blundell et al.)
- HOMER (Tosatto et al.)



# Restraint-based building



Idea → Use the template to derive restrictions at the atomic positions. Optimize the structure based on the restrictions

- “Spread” errors on the whole structure, but minimize it globally
- Does not ensure the local geometry, eg. in the active site

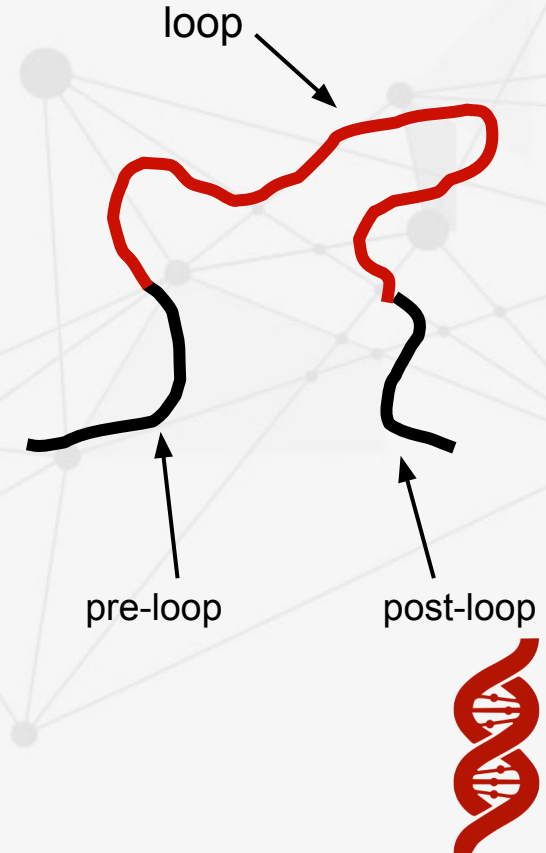
Software

- MODELLER (Šali et al.)



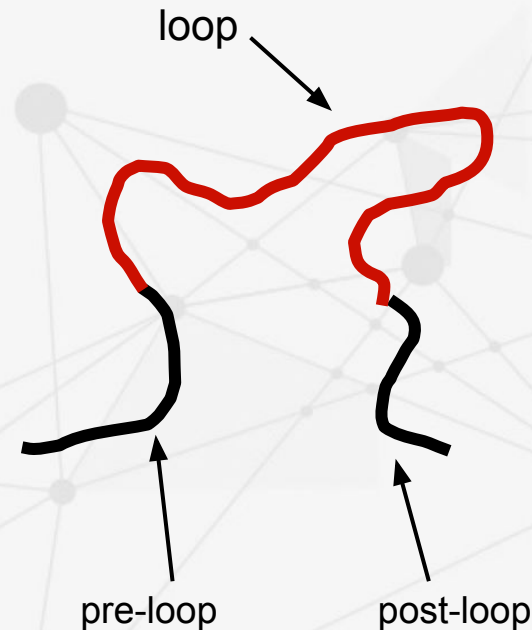
# Loop modeling

- Entire fragments of backbone can be missing in the raw-model
  - Not conserved in the protein family
  - Insertion
  - Deletion
- Problem description
  - Identify the conformation of the fragment (loop,  $k$  residues) that can connect the pre- to the post-loop
  - $\Phi$  and  $\psi$  are the only free parameters



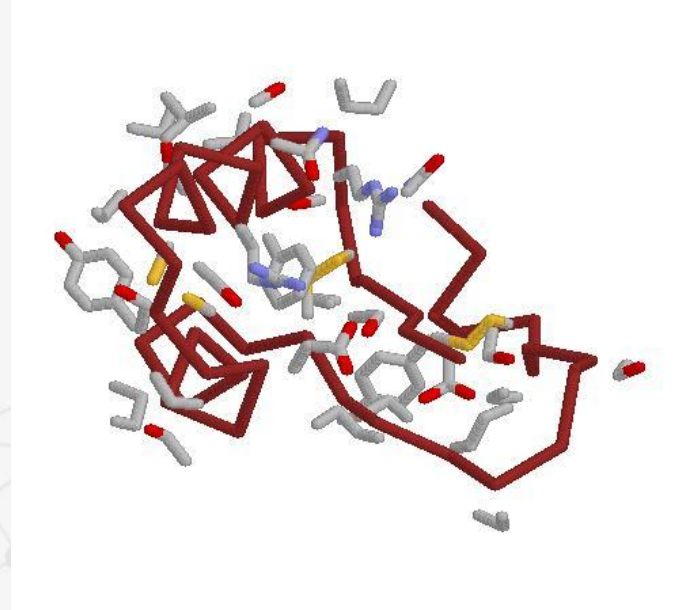
# Loop modeling

- Database methods
  - Extract loop fragments from PDB
  - Choose the fragment that fits better, based on geometric constraints
  - Not all possible conformations are available in PDB
- Ab initio methods
  - Identify best conformations based on the geometric constraints (torsion angles)
  - Select the “best” fragment
  - Problem: computing time



# Side chains

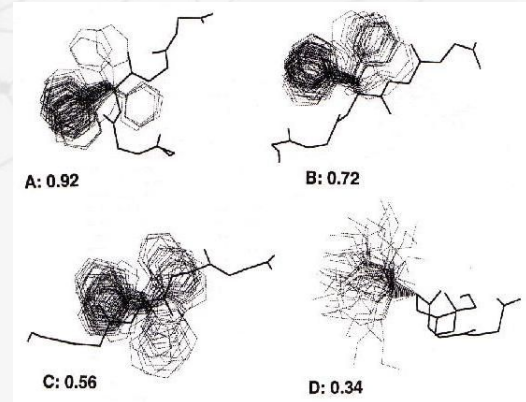
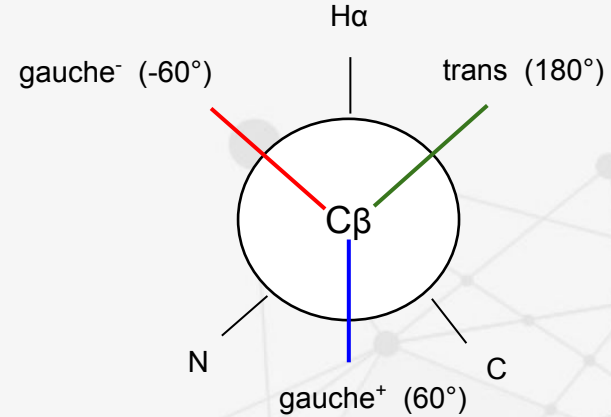
- **Amino acid differences** are not managed when applying the coordinates of the template to the sequence of the target (dimension and position of the side chains)
- Assuming **50% sequences identity**, half side-chains are replaced
- The **RMSD** change is relatively low, but the conformation of important residues (eg. active site) may change
- Effective methods exist to solve this problem, eg. **SCWRL**



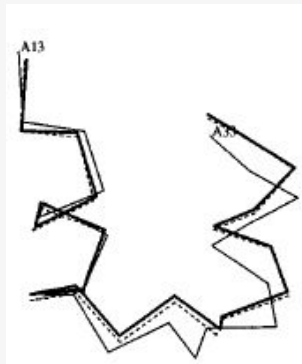


# Side chains

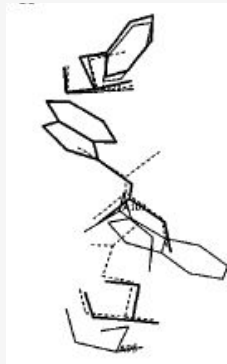
- Rotamers
  - 3 preferred positions for each torsion angles  $\chi$
- The propensity of a rotamer depends on the backbone torsion angles ( $\phi$ ,  $\psi$ ) and the type of amino acid
- Interdependence, domino effect
- Where possible, it is better to maintain the conformation of the template side chains



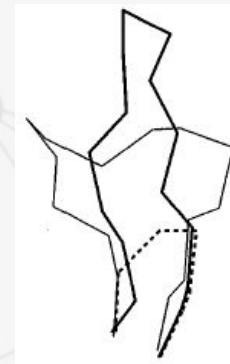
# Typical errors



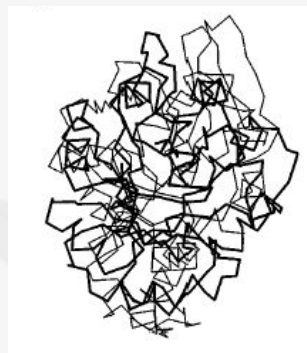
Shift



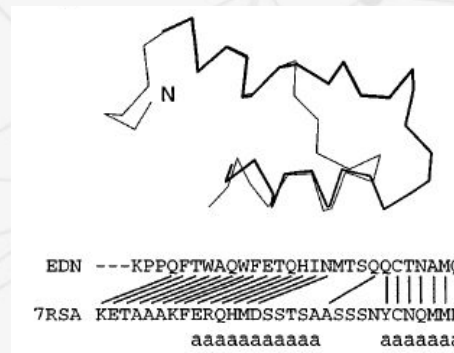
Side chains



Loops

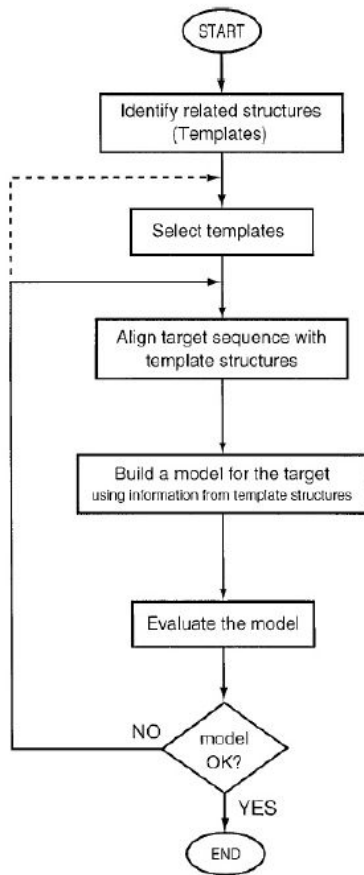


Wrong template



Wrong alignment



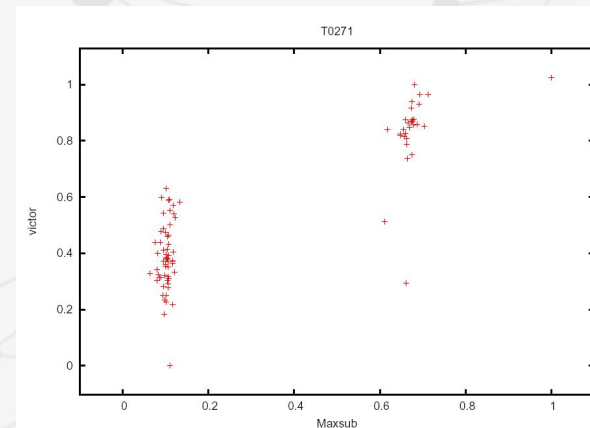


- How to solve these “problems” without worsening the model?
- Models from **alternative alignments** considering **energy profiles** can improve
  - It is necessary to know which part of the alignment to change
- **CASP** suggested the “*Don’t touch it*” philosophy for a long time
  - It is better to avoid local modifications of the structure
  - Changed over the last few years



# Model assessment

- Quality parameters
  - Steric hindrance and **clashes**
  - Deviation from the **geometry** of standard parameters
  - Frequency profiles or energy (**statistical potentials**)
- Software
  - *PROCHECK*
  - *VERIFY-3D*
  - *FRST*
  - *QMEAN*





## Welcome to SWISS-MODEL

SWISS-MODEL is a fully automated protein structure homology-modelling server, accessible via the ExPASy web server, or from the program DeepView (Swiss Pdb-Viewer). The purpose of this server is to make protein modelling accessible to all life science researchers worldwide.

[Start Modelling](#)

### Protein Structure Bioinformatics Group

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**BIOZENTRUM**  
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Molecular Life Sciences


When you publish or report results using SWISS-MODEL, please cite the relevant publications:

- Biasini, M., Bienert, S., Waterhouse, A., Arnold, K., Studer, G., Schmidt, T., Kiefer, F., Cassarino, T.G., Bertoni, M., Bordoli, L., Schwede, T. SWISS-MODEL: modelling protein tertiary and quaternary structure using evolutionary information. *Nucleic Acids Res.* 42, W252-W258 (2014). [doi>](#)
- Bienert, S., Waterhouse, A., de Beer, T.A., Tauriello, G., Studer, G., Bordoli, L., Schwede, T. The SWISS-MODEL Repository - new features and functionality. *Nucleic Acids Res.* 45, D313-D319 (2017). [doi>](#)
- Guex, N., Peitsch, M.C., Schwede, T. Automated comparative protein structure modeling with SWISS-MODEL and Swiss-PdbViewer: A historical perspective. *Electrophoresis* 30, S162-S173 (2009). [doi>](#)
- Benkert, P., Biasini, M., Schwede, T. Toward the estimation of the absolute quality of individual protein structure models. *Bioinformatics* 27, 343-350 (2011). [doi>](#)
- Bertoni, M., Kiefer, F., Biasini, M., Bordoli, L., Schwede, T. Modeling protein quaternary structure of homo- and hetero-oligomers beyond binary interactions by homology. *Scientific Reports* 7 (2017). [doi>](#)

