

# Phylogenetics

## Beyond topology

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## Today's objectives

Understand the goal of phylogenetic comparative methods.

Models of trait evolution:

- Brownian motion
- Ornstein-Uhlenbeck

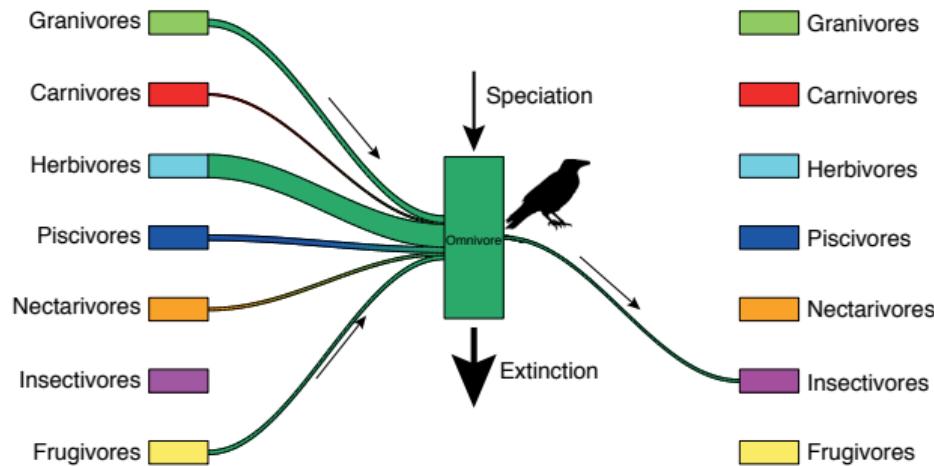
Onto the next adventure

**Phylogenetic inference** → estimating relationships between taxa

**Phylogenetic comparative methods (PCMs)** → to test hypotheses about characteristics or traits that drive the diversification of species

# Phylogenetic comparative methods

**Aim:** to test hypotheses about characteristics or traits that drive the diversification of species.



- How did traits evolve through time? • What factors or traits drive speciation & extinction? • Why are some lineages more specious? • When and why do we see shifts in diversification? • What are the features of extinct ancestors?

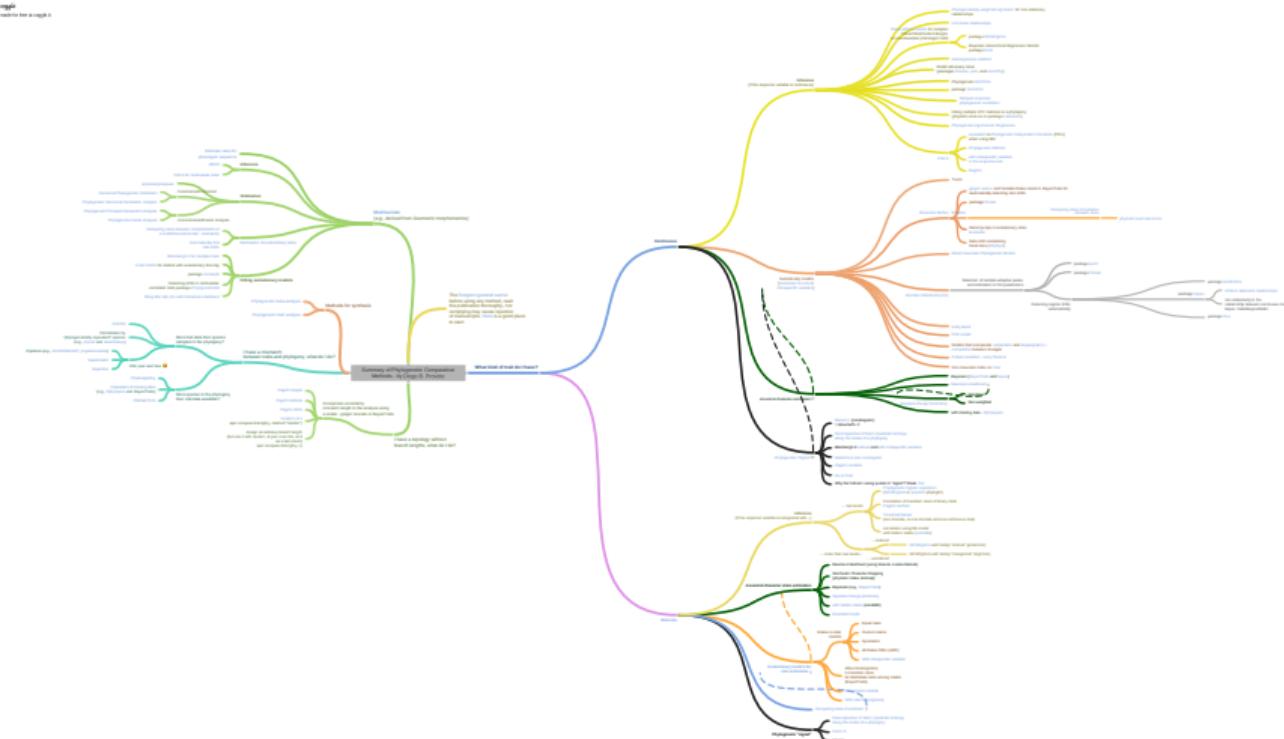
Cornwell & Nakagawa (2017) *Phylogenetic comparative methods*.

## Phylogenetic comparative methods

There are two types of traits: discrete (e.g. biogeographic area) and continuous (e.g. body size).

Next we'll focus on continuous trait models.

# There are now a huge number of PCMs



Explore Diogo Provete's [infographic](#) to learn more.

# Where do phylogenetic comparative methods fit in?

## A. Phylogenetic data collection

<i>Inia</i>	0 Ma
[A T G C]	[0 1 0 1]
<i>Tursiops</i>	5.3 - 0 Ma
[A T G C]	[0 1 0 1]

<i>Isthminia</i> †	5 - 2 Ma
[■ ■ ■ ■]	[0 1 0 1]

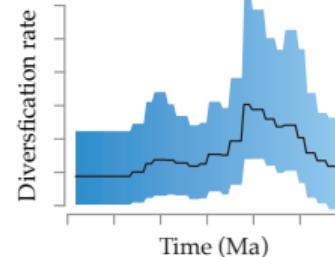
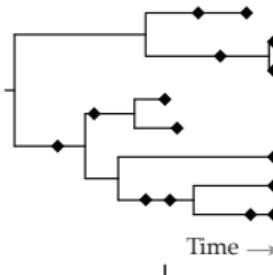
  

<i>Eurhinodelphis</i> †	12 - 9 Ma
[■ ■ ■ ■]	[■ ■ ■ ■]

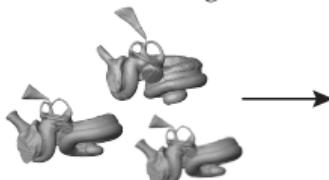
## B. Analysis in BEAST2



## C. Output part 1



## D. CT scanning



## E. Trait data collection

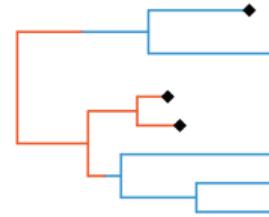
$$\begin{pmatrix} X_{11} & X_{21} & X_{31} & \dots & X_{N1} \\ X_{12} & X_{22} & X_{32} & \dots & X_{N2} \\ \dots \\ X_{1k} & X_{2k} & X_{3k} & \dots & X_{Nk} \end{pmatrix}$$

matrix of N taxa & k traits

## F. Analysis in PCMBASE



## G. Output part 2



Example project workflow combining phylogenetic inference and PCMs. It is becoming increasingly possible to do both simultaneously (and this might even be preferable).

# Comparative methods

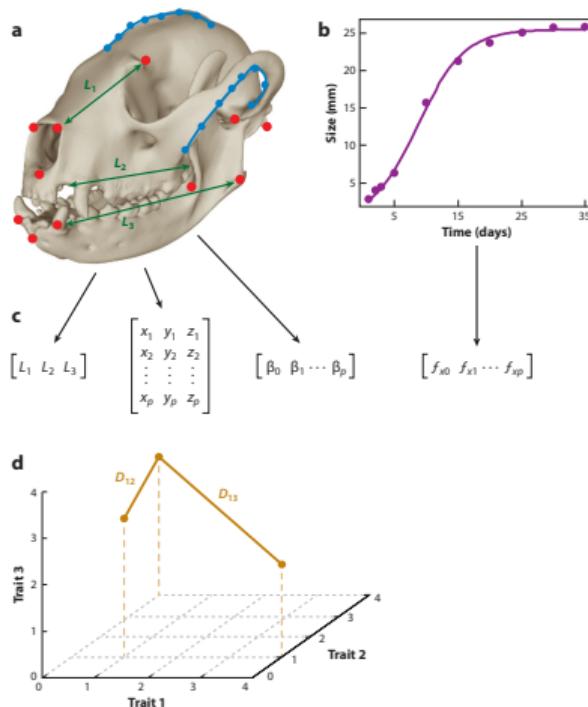


Photo Matt Artz Unsplash

Adams & Collyer (2019) *Annual Reviews Eco Evo Sys.* Phylogenetic comparative methods and the evolution of multivariate phenotypes

## Phylogenetic non-independence

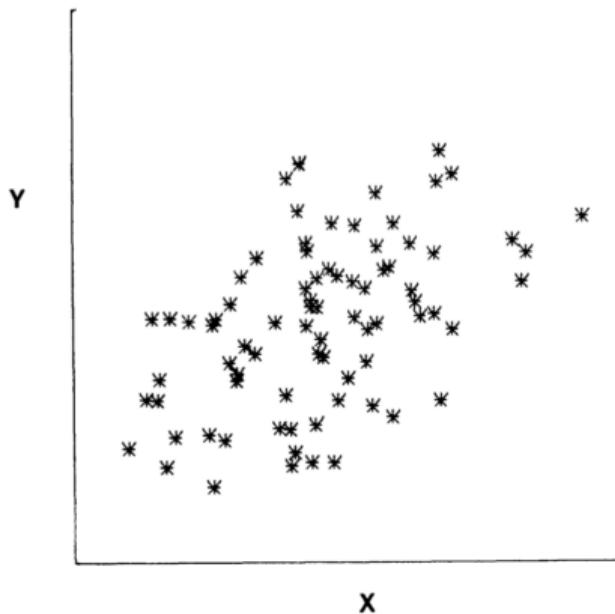


FIG. 6.—A typical data set that might be generated for the phylogeny in fig. 5 using the model of independent Brownian motion (normal increments) in each character.

Felsenstein (1985) *American Naturalist*

## Phylogenetic non-independence

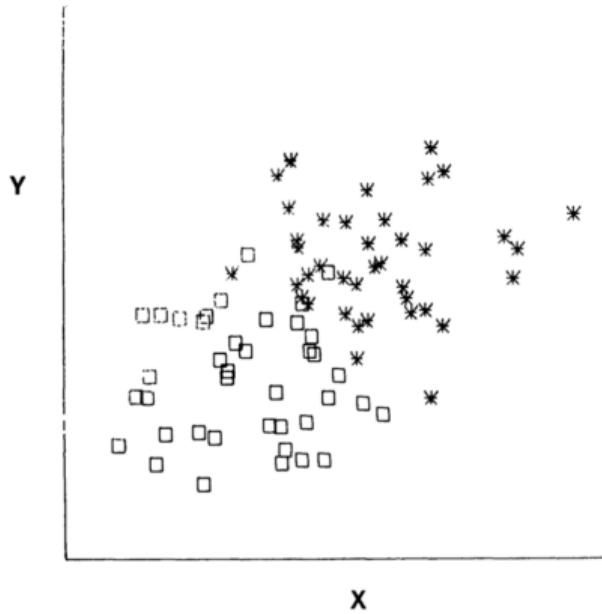
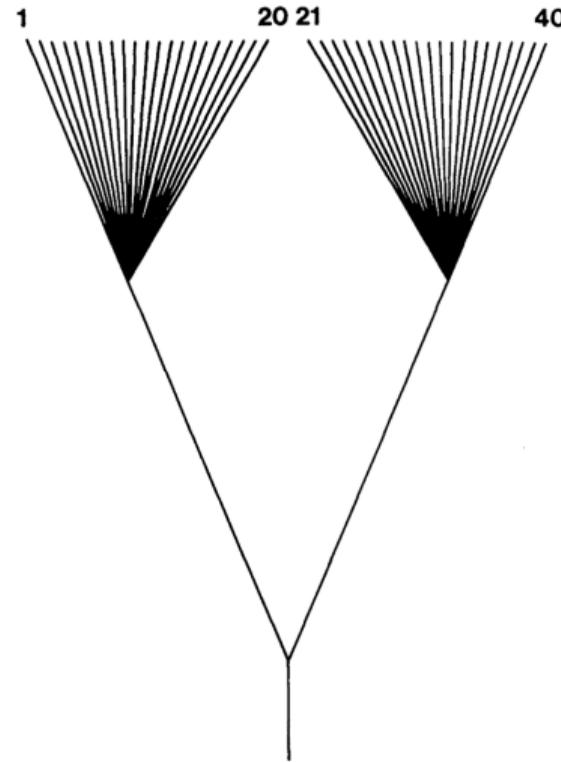


FIG. 7.—The same data set, with the points distinguished to show the members of the 2 monophyletic taxa. It can immediately be seen that the apparently significant relationship of fig. 6 is illusory.

Felsenstein (1985) *American Naturalist*

## Phylogenetic non-independence



Felsenstein (1985) *American Naturalist*

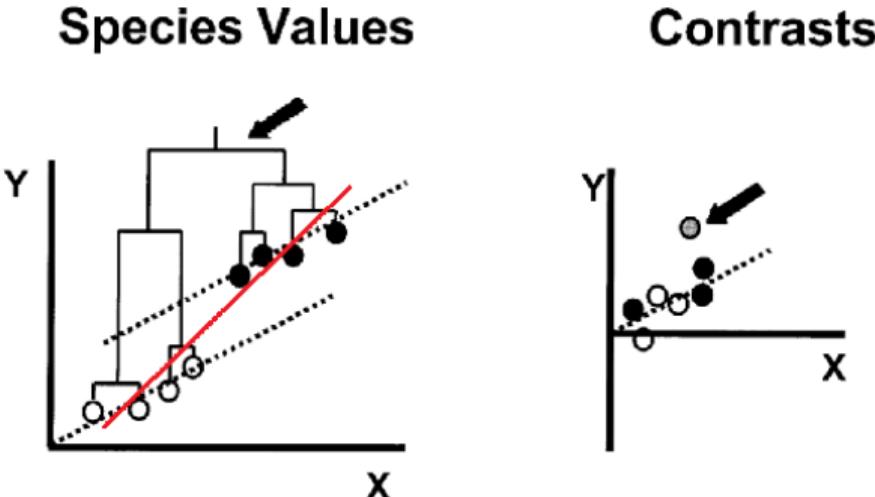
## Taking phylogenetic non-independence into account

**The problem:** species in the same clade are more similar because of their shared evolutionary history.

Regression analysis assumes individual data points are statistically independent – this assumption is violated for species data.

Phylogenetic independent contrasts (PIC) and phylogenetic generalised least squares (PGLS) are two early approaches that take phylogenetic non-independent into account in the study of trait evolution.

## Taking phylogenetic non-independence into account



The red line would be the slope of a standard least squares regression.

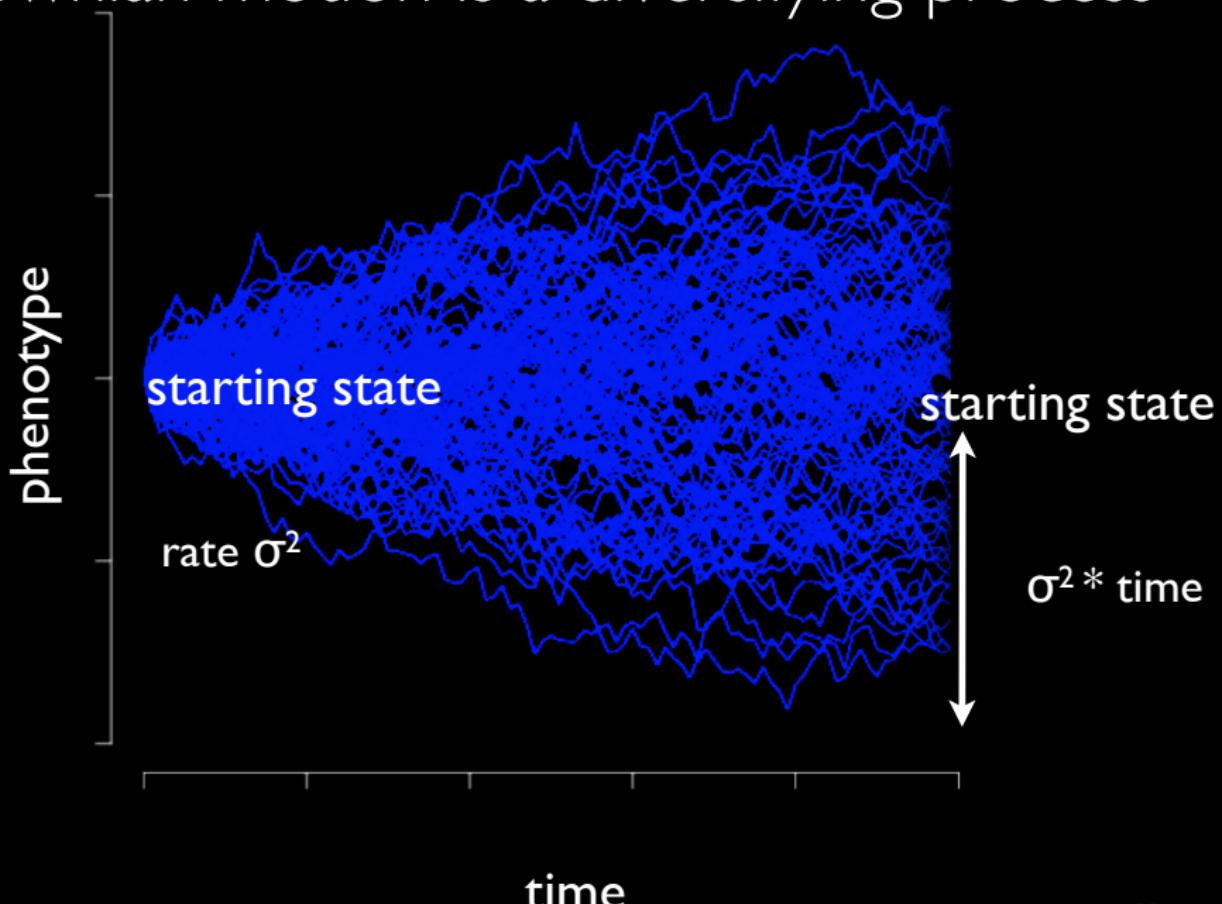
Image source: Laura Soul's [PCM tutorial](#). Adapted from Nunn and Barton (2001).

# Model's of continuous trait evolution applied to mammals

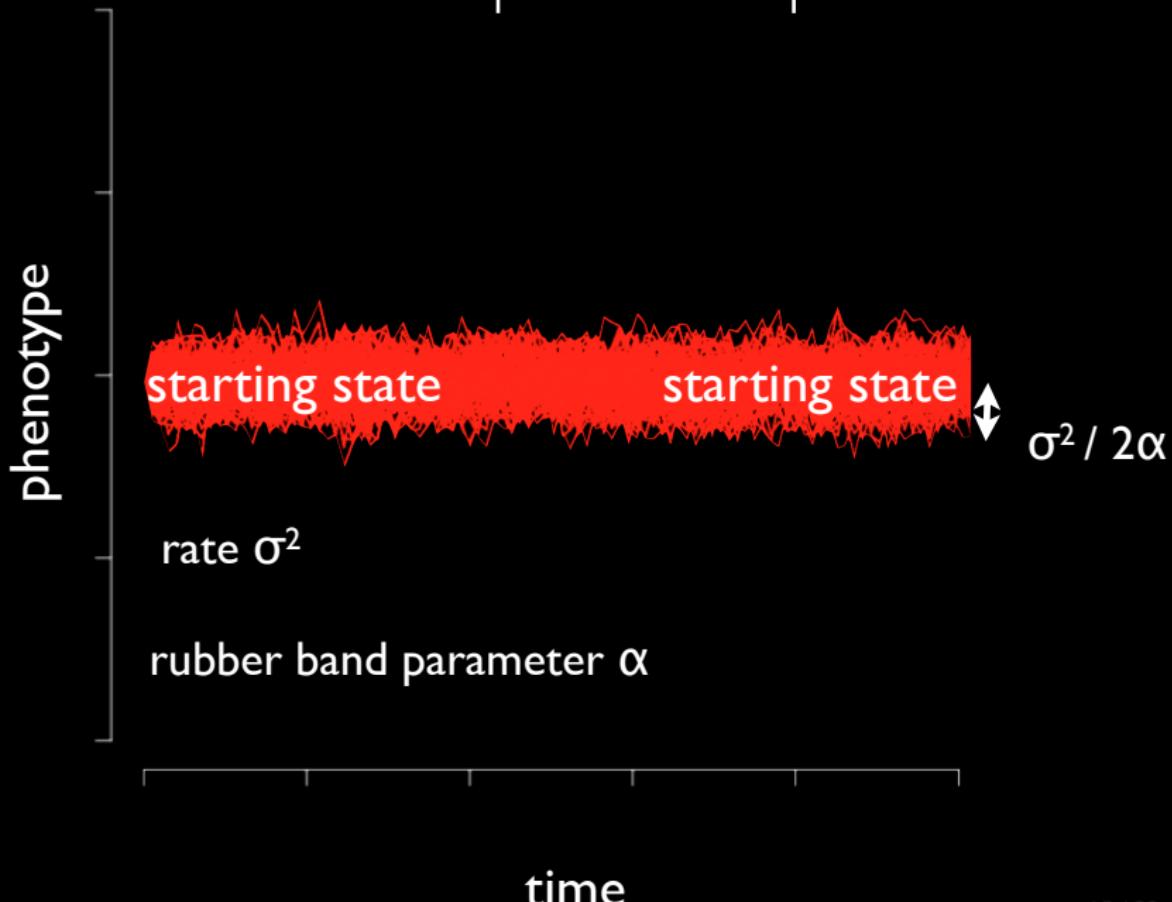
Slides borrowed from Graham Slater's phylosemiar.

See also Slater (2013). *Methods Evolution & Ecology*.

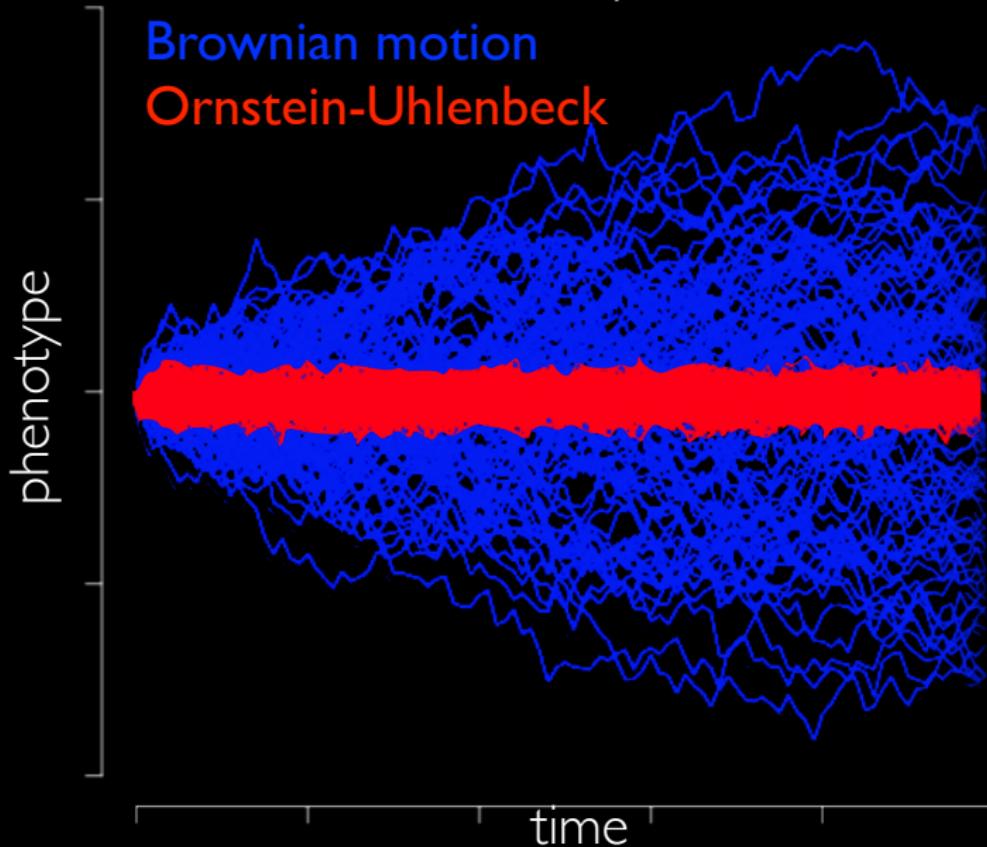
# Brownian motion is a diversifying process



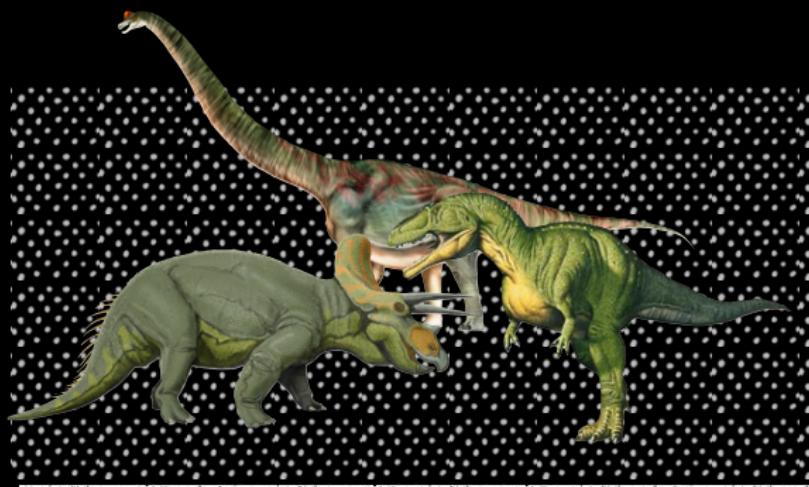
# OU is an equilibrium process



BM and OU simulated at the same rate give very different disparities

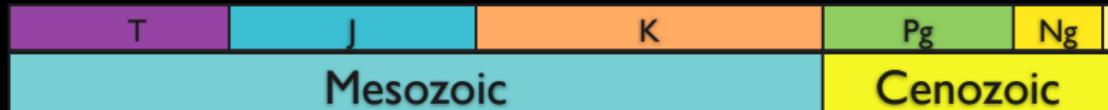
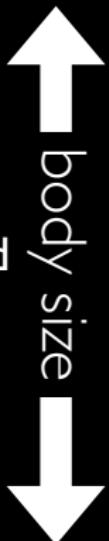


# variation in mode



evolution  
unconstrained

evolution constrained



# 3 paleo-motivated models for mammalian body size evolution

K-Pg Shift

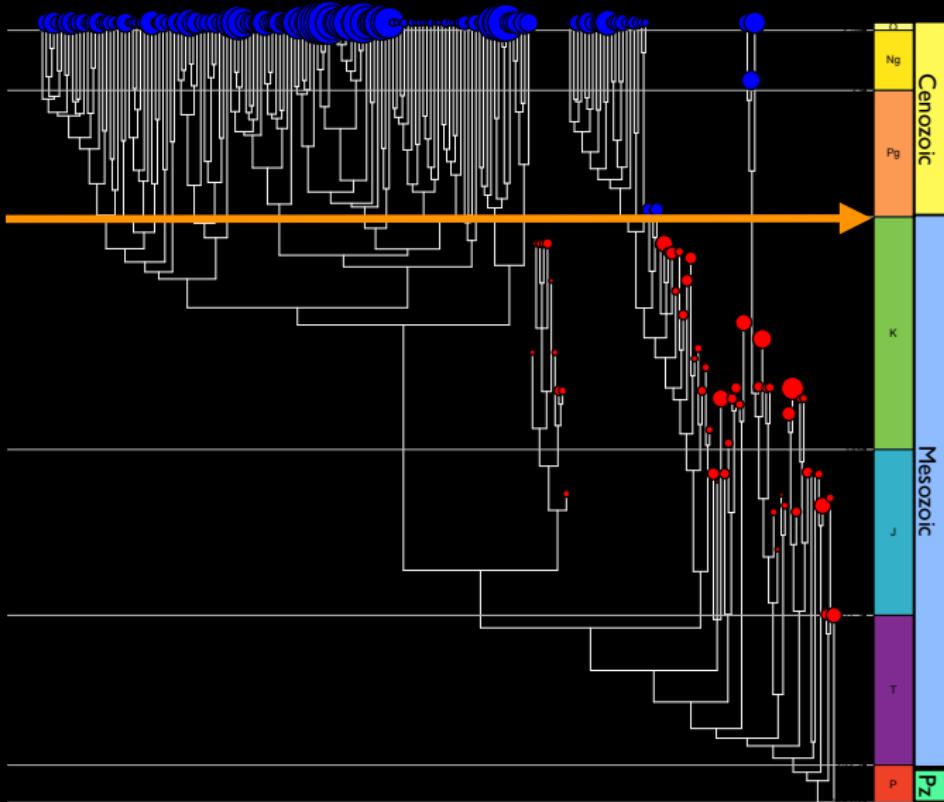


ecological release

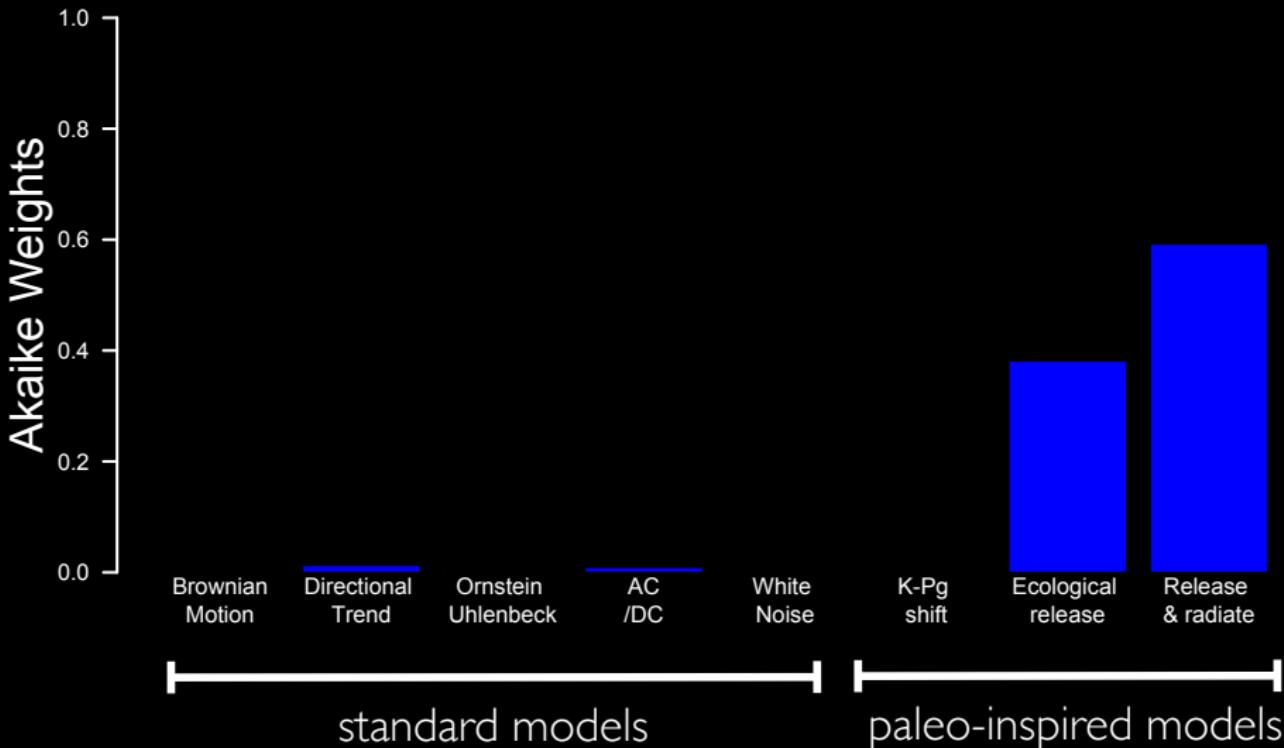


release and radiate



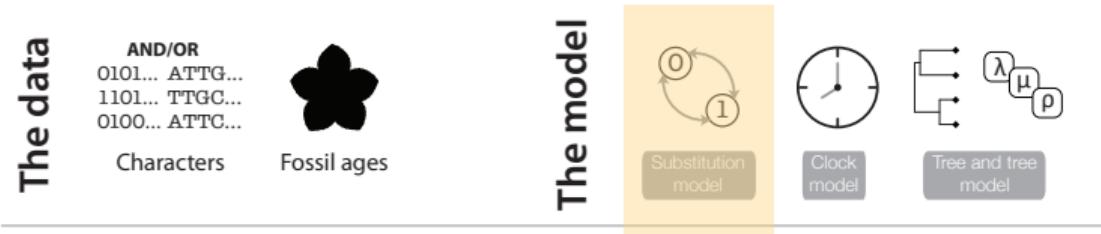


release & radiate fits best but ecological release is almost as good



# Continuous trait models in Bayesian inference

# Bayesian phylogenetic dating



posterior

$$P(\boxed{\text{Tree} \ \lambda \ \mu \ \rho \ \text{Clock} \ \text{Flower}} \mid \text{0101...} \atop \text{1101...} \atop \text{0100...} \text{ Flower}) =$$

probability of the character data given everything else\*      probability of the timetree given the timetree model      priors on model parameters

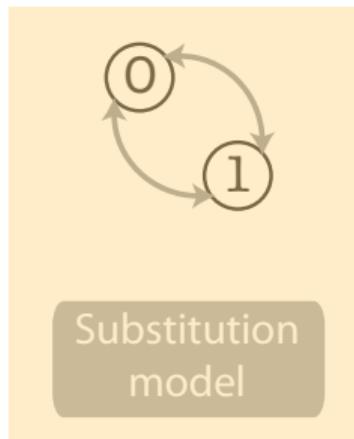
$$P(\text{0101...} \mid \boxed{\text{Tree} \ \lambda \ \mu \ \rho \ \text{Clock}}) P(\boxed{\text{Tree}} \mid \text{Flower}) P(\lambda \ \mu \ \rho) P(\text{Clock})$$

marginal probability of the data

\*the timetree, the parameters and the tripartite model

## Recap: Bayesian phylogenetic dating requires three model components

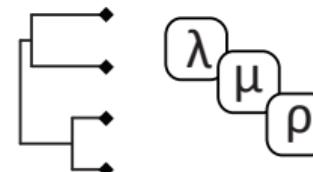
- The **substitution model** ← describes how sites evolve over time.
- The **clock model** ← describes how evolutionary rates vary across the tree.
- The **tree model** ← describes how trees grow over time. Temporal evidence is included here.



Substitution  
model

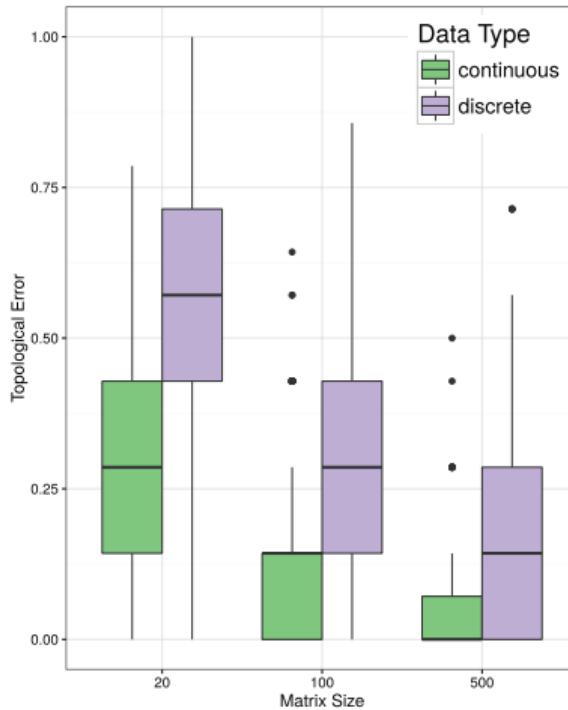


Clock  
model



Tree and tree  
model

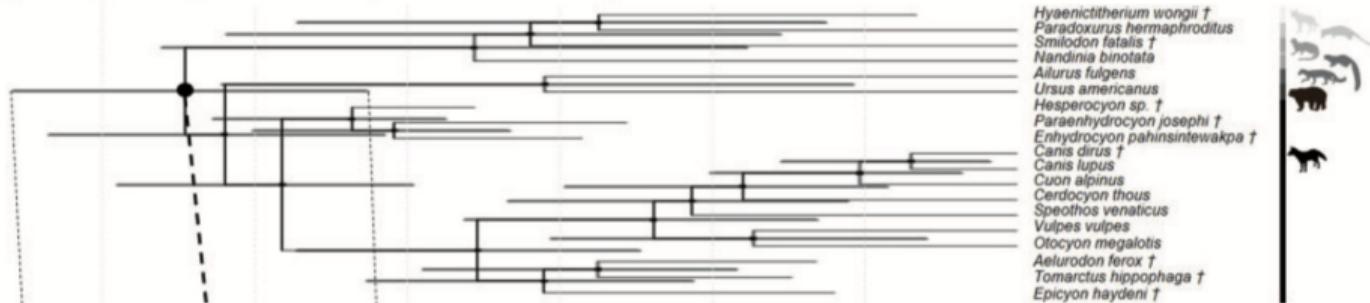
# Continuous trait models can be used to build trees



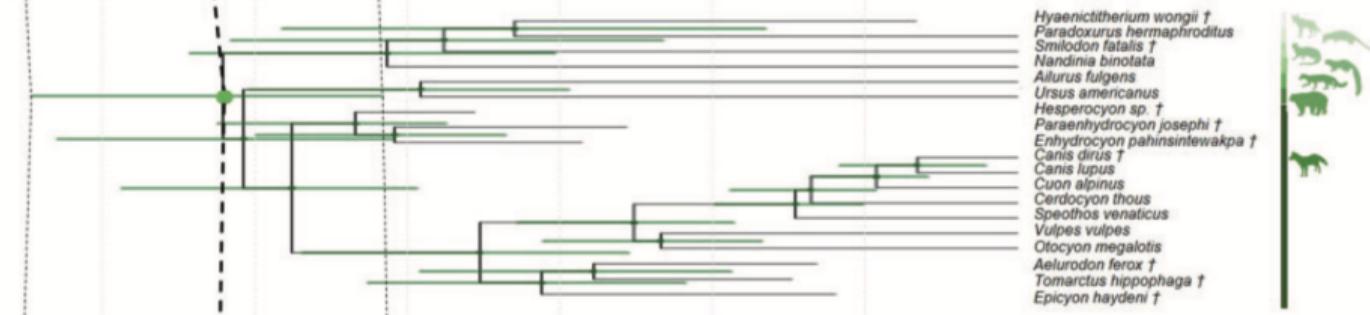
Parins-Fukuchi (2018) *Sys Bio*. Use of continuous traits can improve morphological phylogenetics.

# Continuous trait models can be used to build dated trees

## A) Morphological data ( $R = R^*$ , $c = 1$ )



## B) Morphological data ( $R = R^*$ , $c = 1$ ) + Molecular data

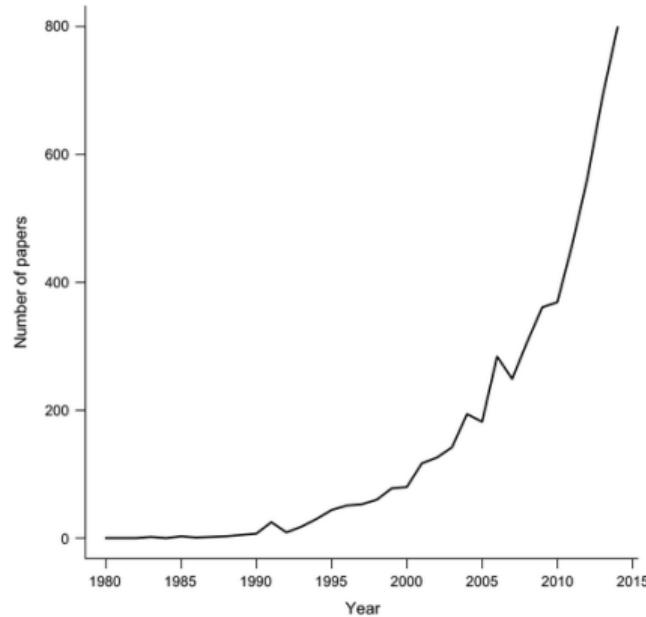


Álvarez et al. (2019) *Sys Bio.* Bayesian estimation of divergence times using correlated quantitative characters.

## Summary of what we can do with continuous trait models

1. Test hypotheses about evolutionary process
2. Build trees
3. Date trees

# The "dark side" of PCMs



Tree uncertainty, trait uncertainty, model uncertainty.

Limitations of methods are rarely addressed or well documented.

Cooper et al. (2016) *MEE*. Shedding light on the "dark side" of phylogenetic comparative methods.  
See also Cornwell & Nakagawa (2017) *Current Biology*. Phylogenetic comparative methods primer.

## The "dark side" of PCMs

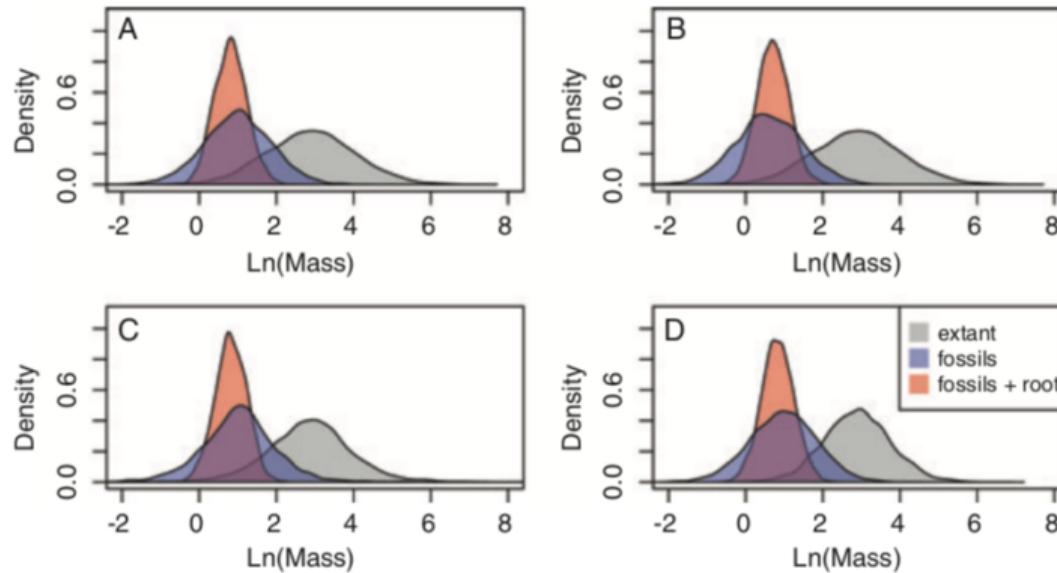
Make sure the method is appropriate for your question and data.

Try to understand the method assumption and limitations.

Don't over-interpret your results.

Taking phylogenetic history into account often gives you more information than ignoring it.

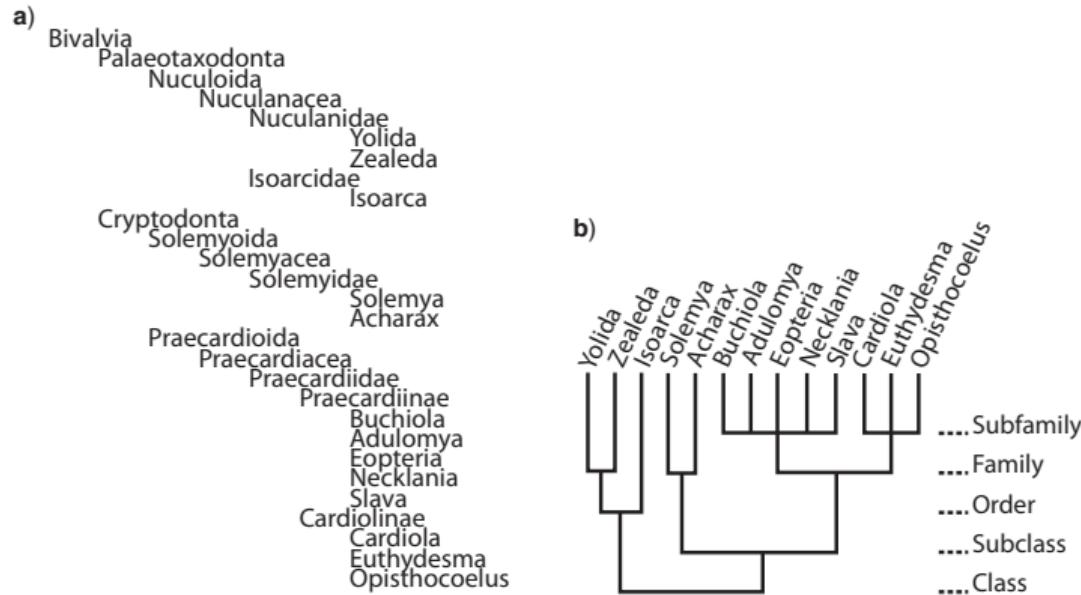
# The bright side of PCMs?



Simulations show that incorporating fossil information improves our ability to distinguish among models of quantitative trait evolution using comparative data.

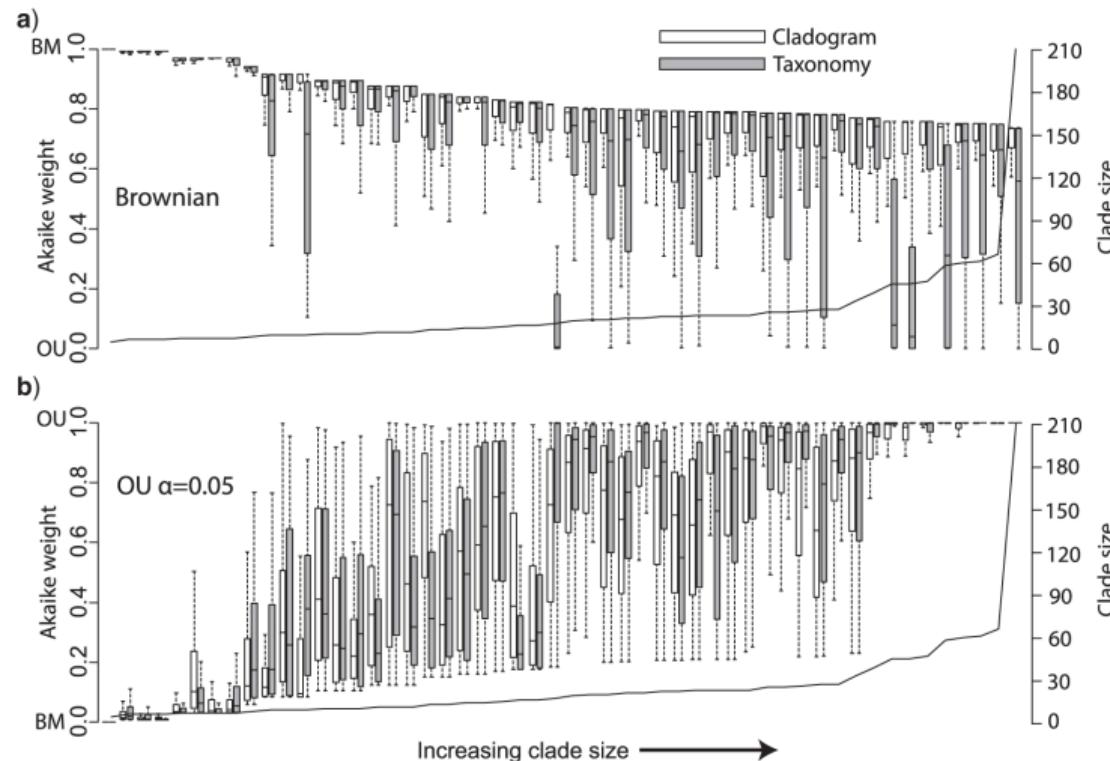
Slater et al. (2012). *Evolution*.

# The bright side of PCMs?



Soul & Friedman (2015). *Sys Bio.* Taxonomy and phylogeny can yield comparable results in comparative palaeontological analyses.

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Soul & Friedman (2015). *Sys Bio.* Taxonomy and phylogeny can yield comparable results in comparative palaeontological analyses.

## Take homes

Species as data points are not independent.

PCMs provide a statistical approach for studying the evolution of traits in a phylogenetic framework (among other things).

PCMs also have a **dark side** — they are statistical methods after all!

## Further reading

The PCM community it very R centric!

Laura Soul & David Wright have an excellent introductory [tutorial](#) for doing PCMs using R.

Luke Harmon has a brilliant [online book](#) all about PCMs.

Check out Graham Slater's [phylosemiar](#) to learn more about fossils and PCMs.