

Phylogenetic dating

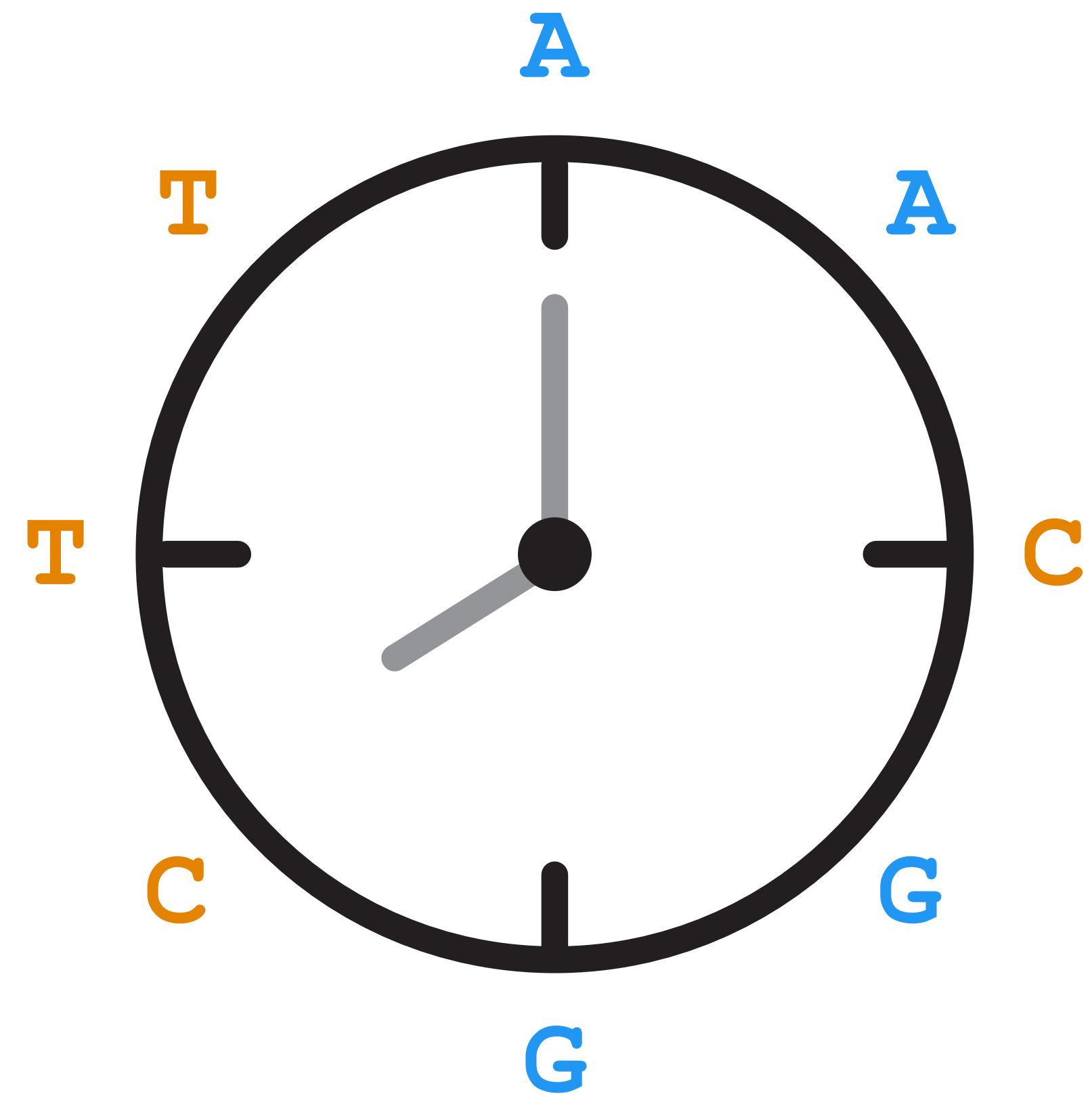
Analytical Paleobiology

Rachel Warnock
14.08.24



Today's objectives

- Recap
 - Bayesian inference
 - MCMC
- Intro to molecular dating



Recap

Bayes' theorem

$$\Pr(\text{model} \mid \text{data}) = \frac{\Pr(\text{data} \mid \text{model}) \Pr(\text{model})}{\Pr(\text{data})}$$

Bayes' theorem

Likelihood

The probability of the data given the model assumptions and parameter values

$$\Pr(\text{model} \mid \text{data}) = \frac{\Pr(\text{data} \mid \text{model}) \Pr(\text{model})}{\Pr(\text{data})}$$

Bayes' theorem

Priors

This represents our prior knowledge of the model parameters

$$\Pr(\text{model} \mid \text{data}) = \frac{\Pr(\text{data} \mid \text{model}) \Pr(\text{model})}{\Pr(\text{data})}$$

Bayes' theorem

$$\Pr(\text{model} \mid \text{data}) = \frac{\Pr(\text{data} \mid \text{model}) \Pr(\text{model})}{\Pr(\text{data})}$$

Marginal probability

The probability of the data, given all possible parameter values. Can be thought of as a normalising constant

Bayes' theorem

posterior

Reflects our combined knowledge based on the likelihood and the priors

$\Pr(\text{model} \mid \text{data}) =$

$$\frac{\Pr(\text{data} \mid \text{model}) \Pr(\text{model})}{\Pr(\text{data})}$$

Bayesian tree inference

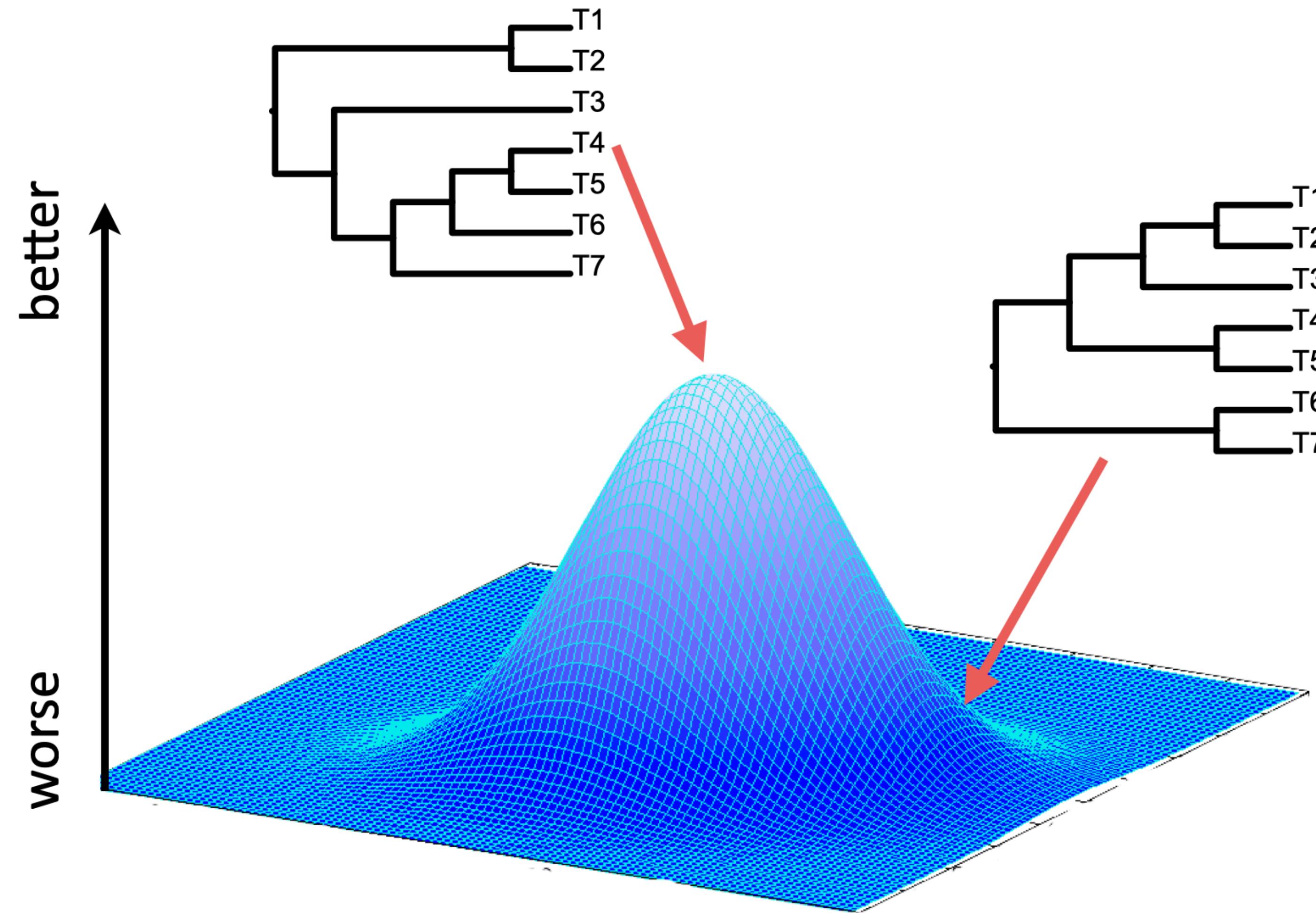
$$\text{posterior} \quad P(E \mid \text{0101...}, \text{1101...}, \text{0100...}) = \frac{\text{likelihood} \quad P(\text{0101...} \mid E) \quad P(E)}{\text{priors} \quad P(\text{0101...}, \text{1101...}, \text{0100...})}$$

Diagram illustrating the components of Bayesian tree inference:

- posterior**: $P(E \mid \text{0101...}, \text{1101...}, \text{0100...})$
- likelihood**: $P(\text{0101...} \mid E)$
- priors**: $P(E)$
- marginal probability**: $P(\text{0101...}, \text{1101...}, \text{0100...})$

The diagram shows a phylogenetic tree with two terminal nodes. The left node is labeled with '0' and the right node with '1'. Arrows indicate the direction of evolution from root to leaves. The likelihood term $P(\text{0101...} \mid E)$ corresponds to the probability of observing the sequence '0101...' at the first node given the tree E . The prior term $P(E)$ represents the probability of the tree structure itself. The marginal probability $P(\text{0101...}, \text{1101...}, \text{0100...})$ is the joint probability of all observed sequences.

How do we find the ‘best’ tree?



It depends how you measure ‘best’

Method	Criterion (tree score)
Maximum parsimony	Minimum number of changes
.....
Maximum likelihood	Likelihood score (probability), optimised over branch lengths and model parameters
.....
Bayesian inference	Posterior probability, integrating over branch lengths and model parameters

Both maximum likelihood and Bayesian inference are model-based approaches

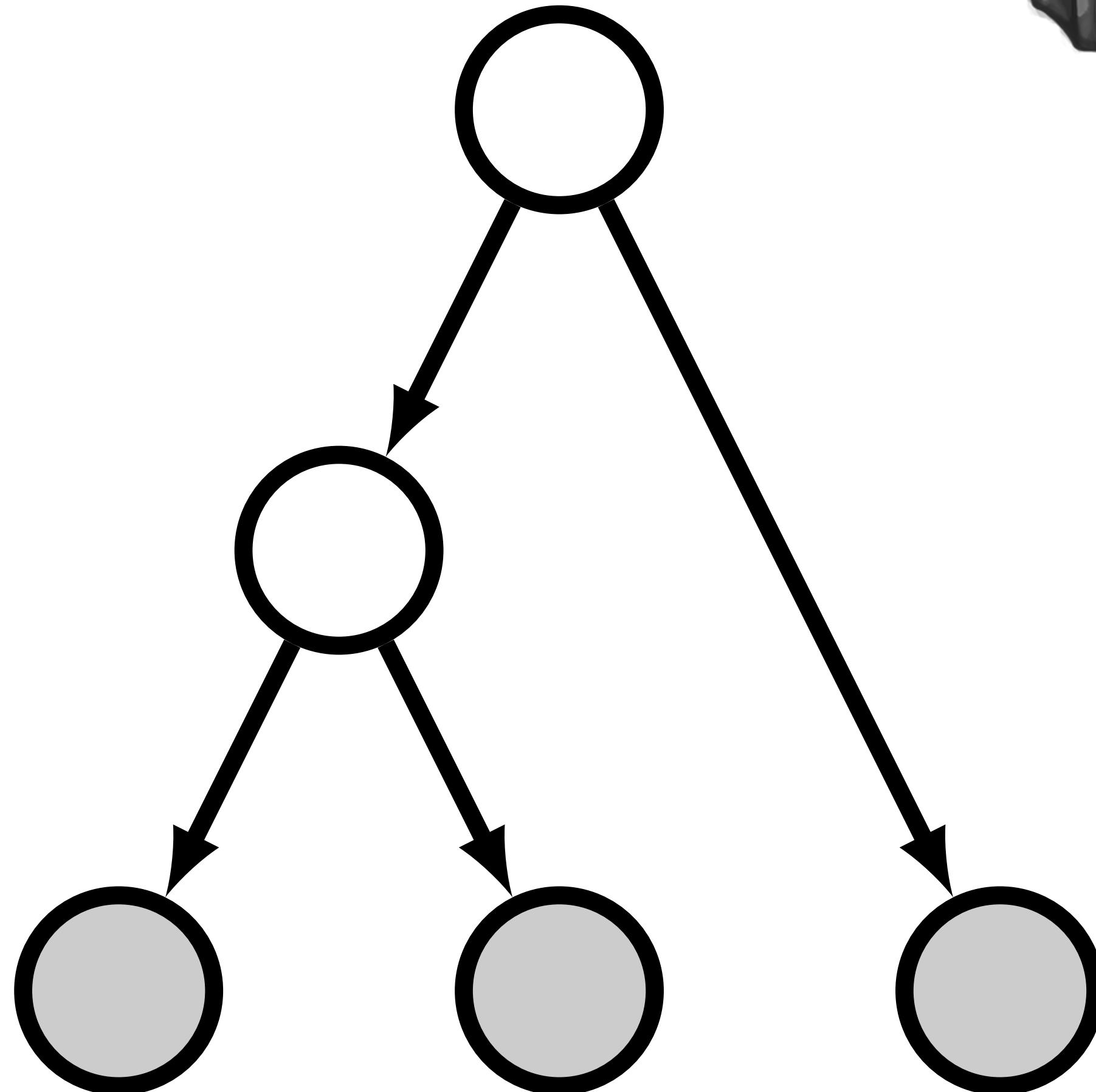
Note these are not the only approaches to tree-building but they are the most widely used

Graphical models

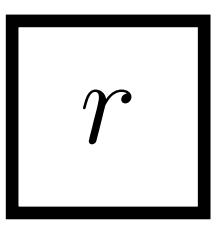


Provide tools for visually and computationally representing complex, parameter-rich models

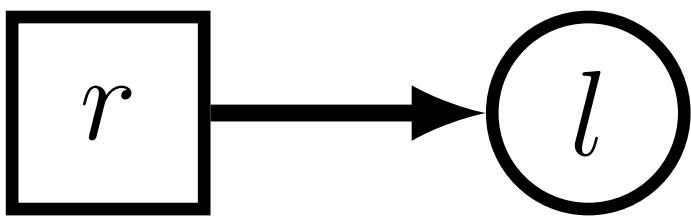
Depict the conditional dependence structure of parameters and other random variables



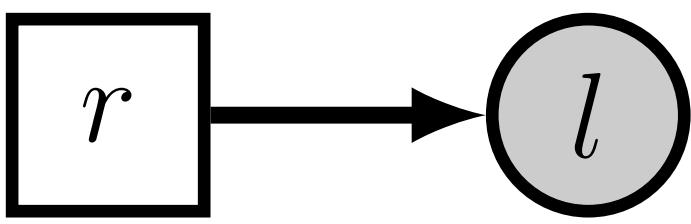
a)



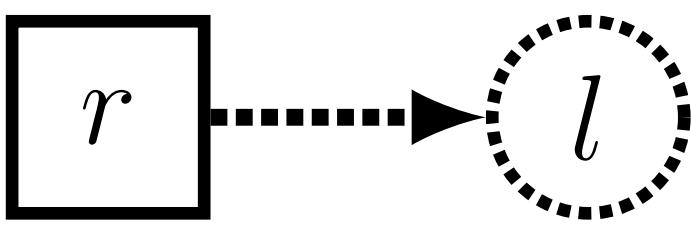
b)



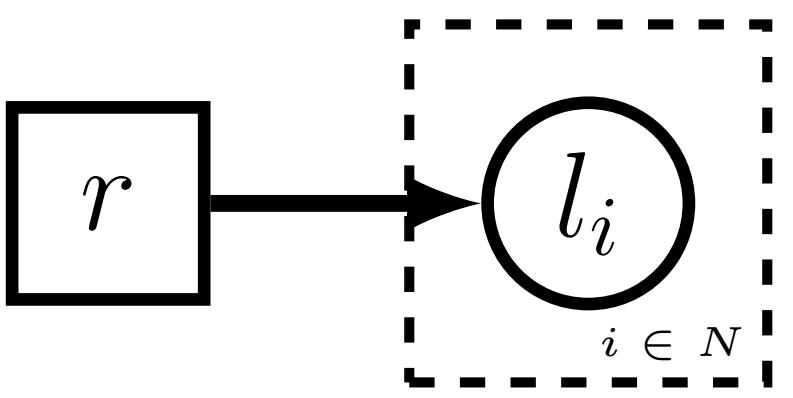
c)



d)



e)



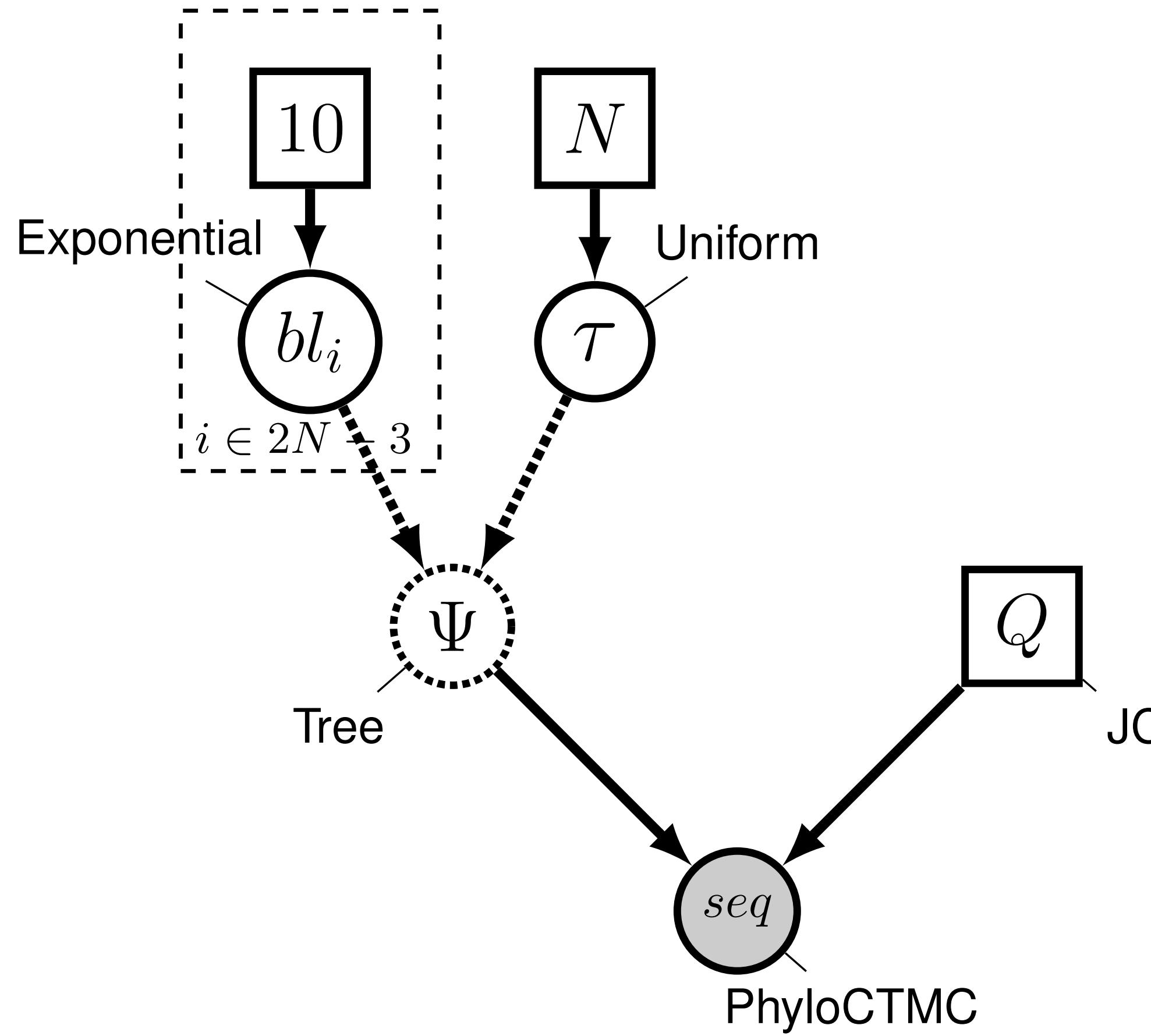
```
# constant node  
r <- 10
```

```
# stochastic node  
l ~ dnExp(r)
```

```
# stochastic node (observed)  
l.clamp(0.1)
```

```
# deterministic node  
l := exp(r)
```

```
# stochastic nodes (iid)  
for (i in 1:N) {  
  l[i] ~ dnExp(r)  
}
```



```

for (I in 1:n_branches) {
  bl[I] ~ dnExponential(10.0)
}
topology ~ dnUniformTopology(taxa)
psi := treeAssembly(topology, bl)

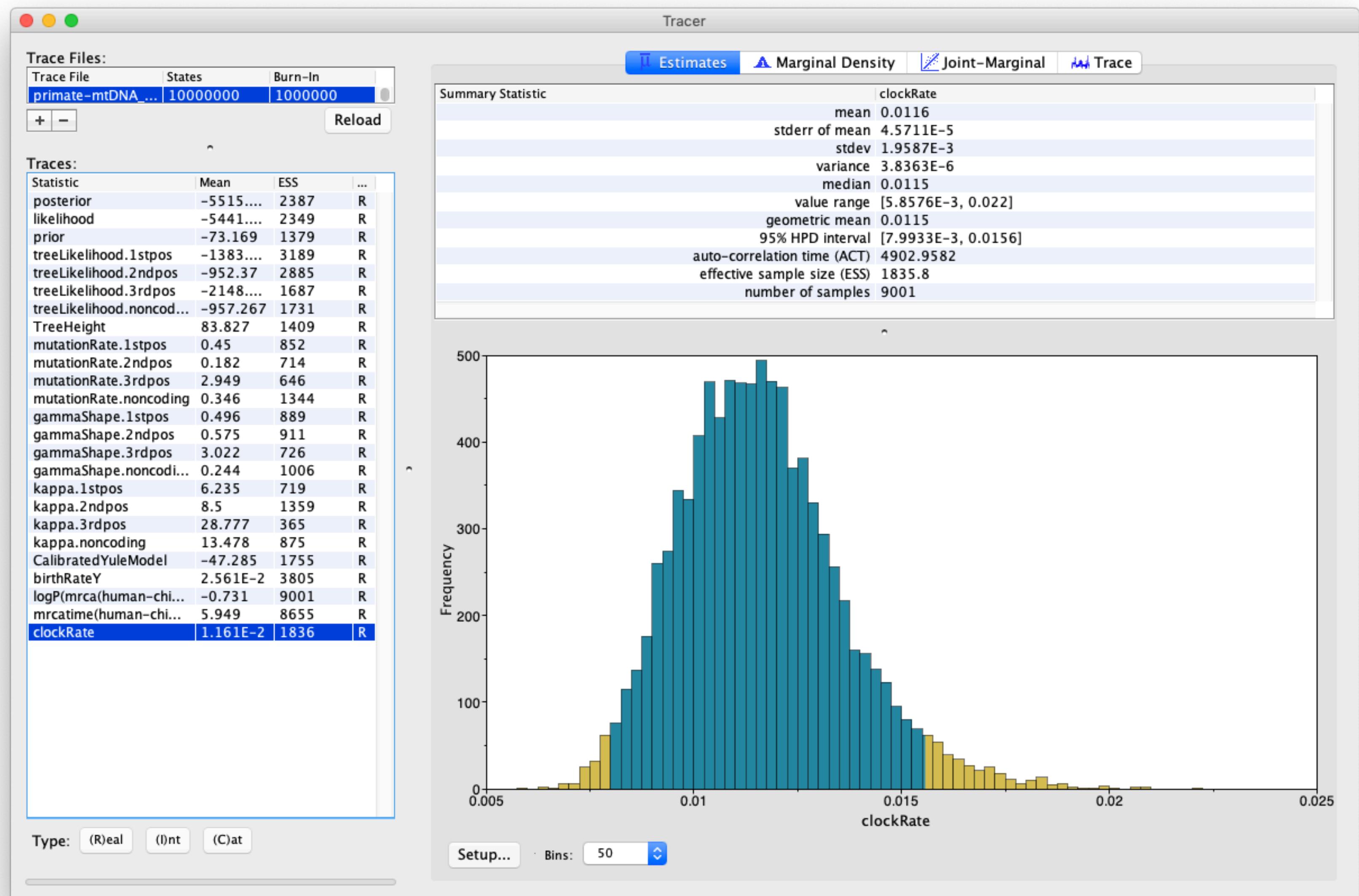
Q_morpho <- fnJC(2)

phyMorpho ~ dnPhyloCTMC( tree=psi,
siteRates=rates_morpho, Q=Q_morpho,
type="Standard", coding="variable" )
phyMorpho.clamp( data )

```

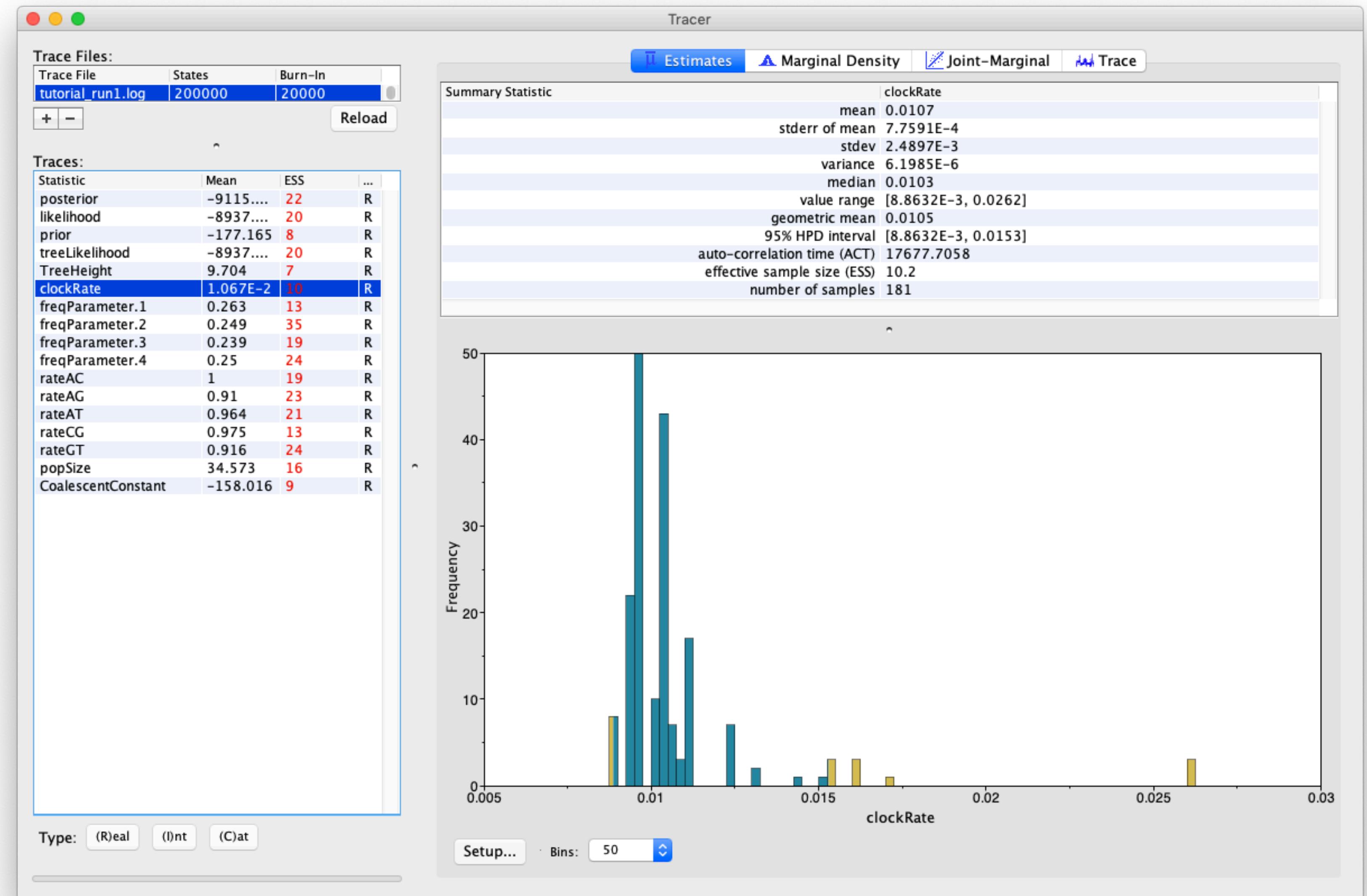
Summarising the posterior

Tracer is an amazing program for exploring MCMC output



Summarising the posterior

Tracer is an amazing program for exploring MCMC output

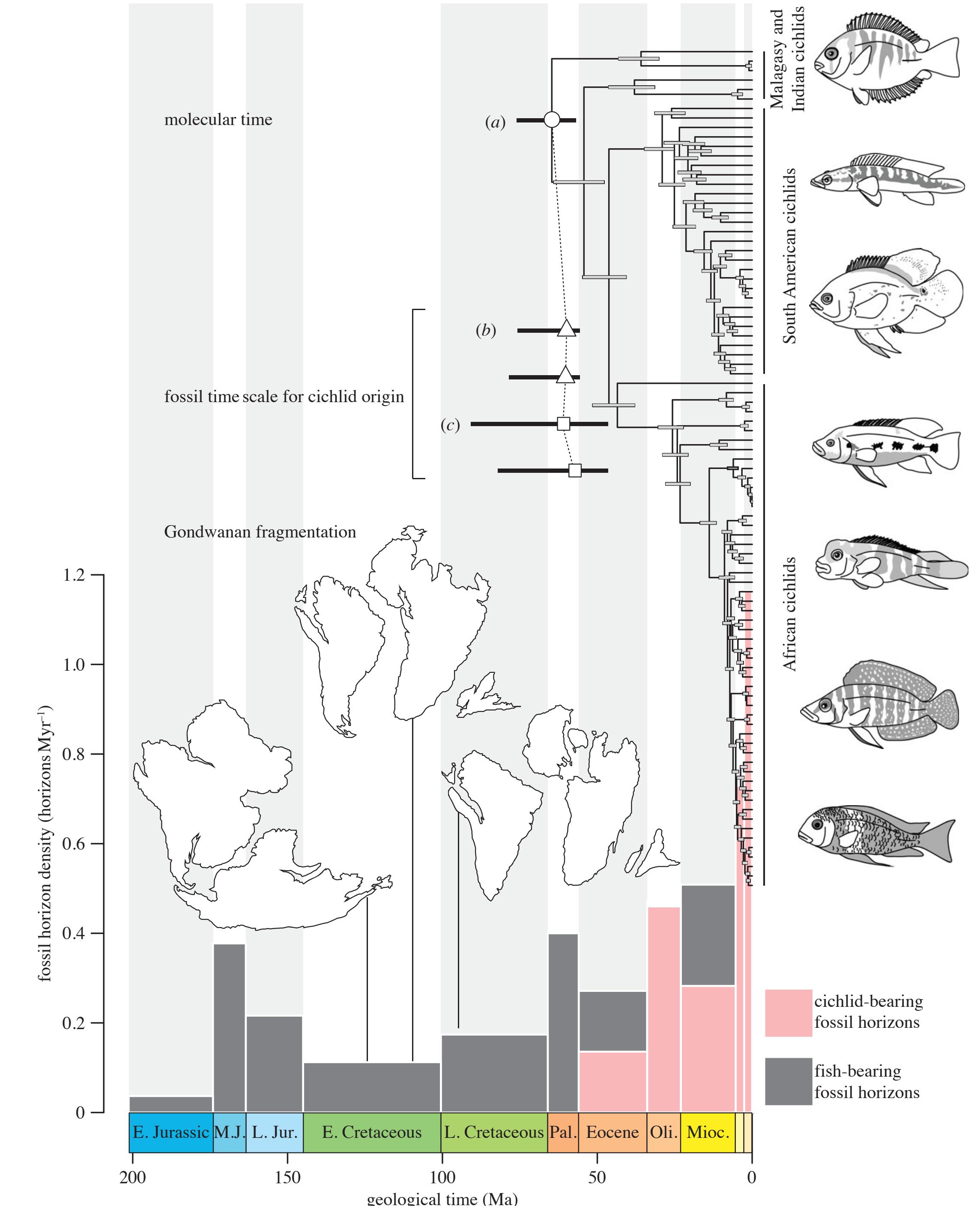


Introduction to molecular dating

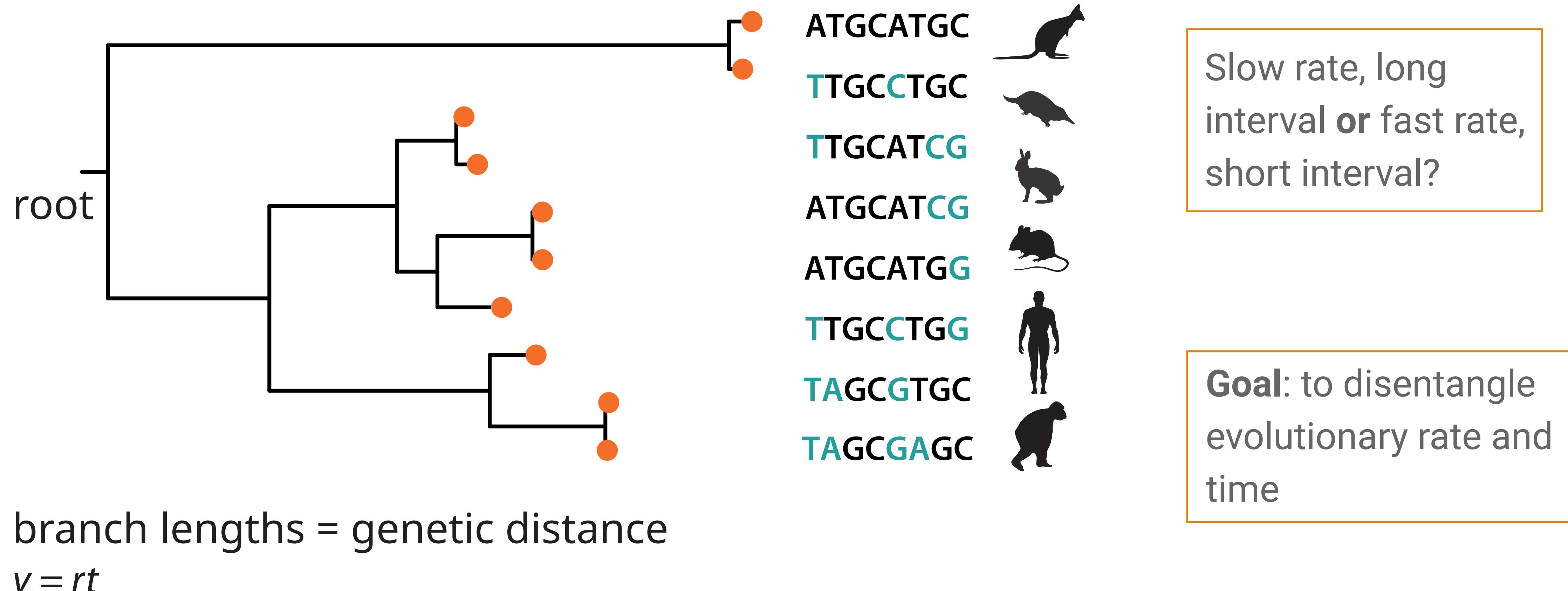
What can we learn from trees?

- Evolutionary relationships
- Timing of diversification events
- Geological context
- Rates of phenotypic evolution
- Diversification rates

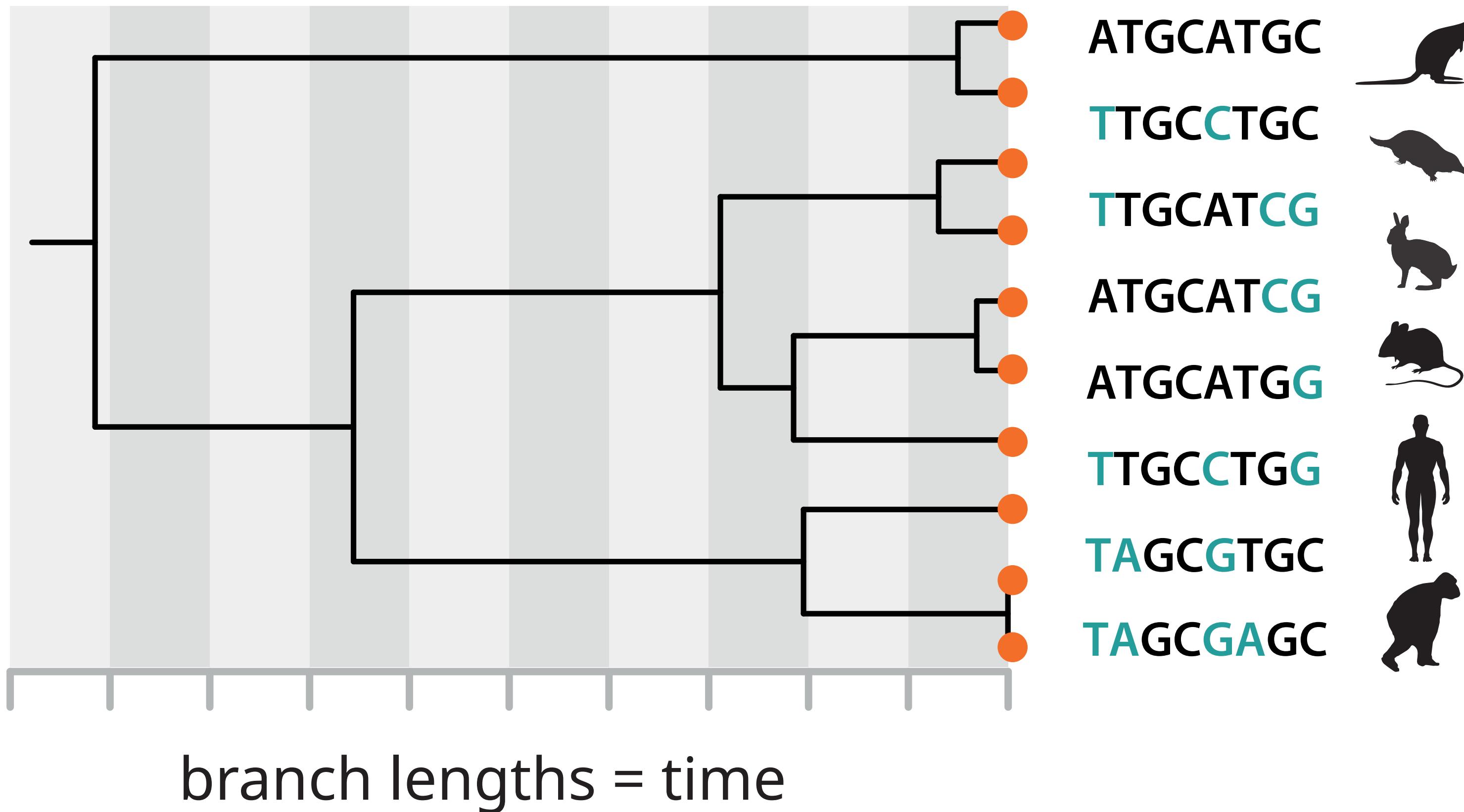
Image adapted from Friedmann et al. (2013)



Molecular (or morphological) characters are not independently informative about time

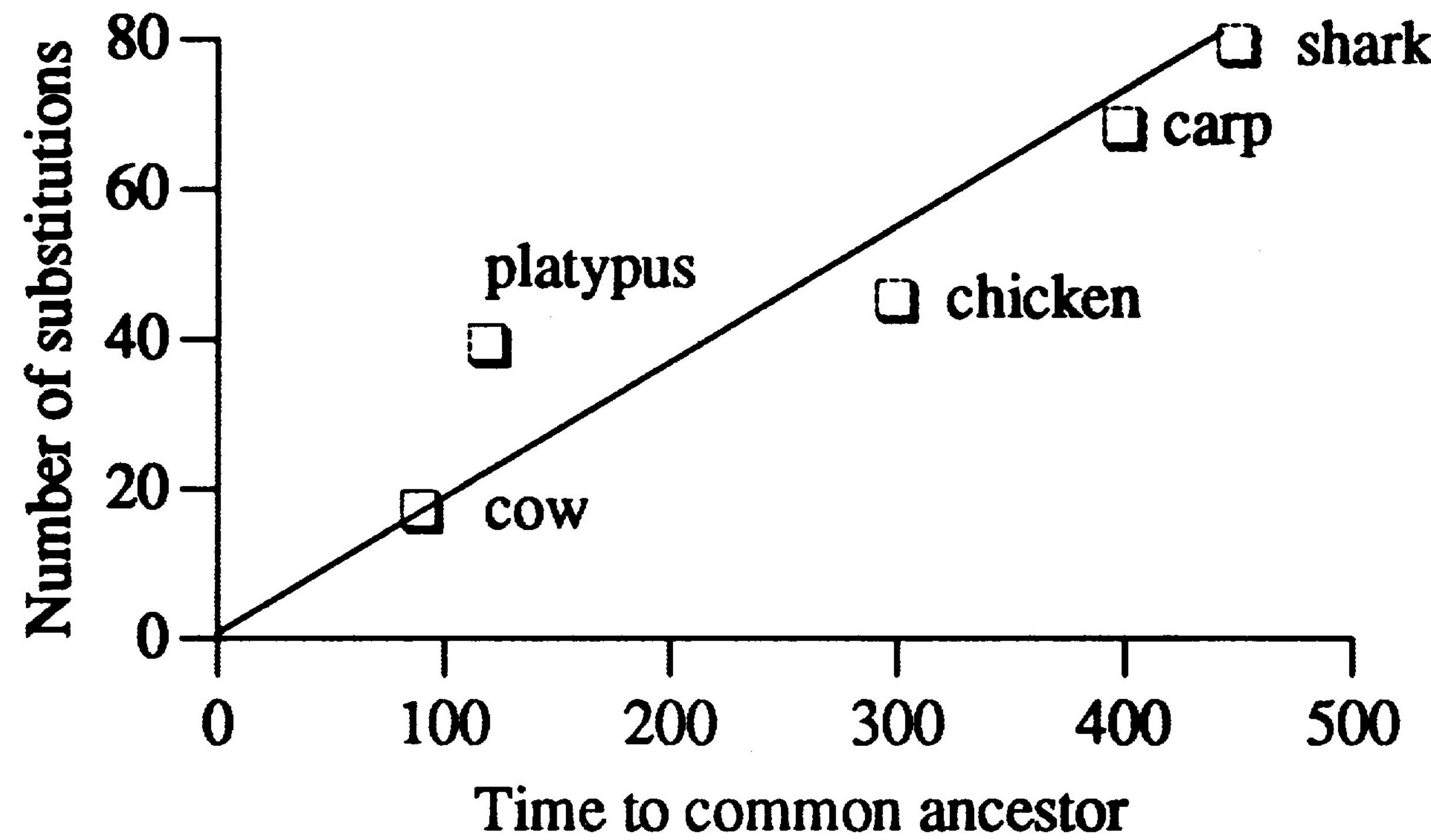


Molecular (or morphological) characters are not independently informative about time



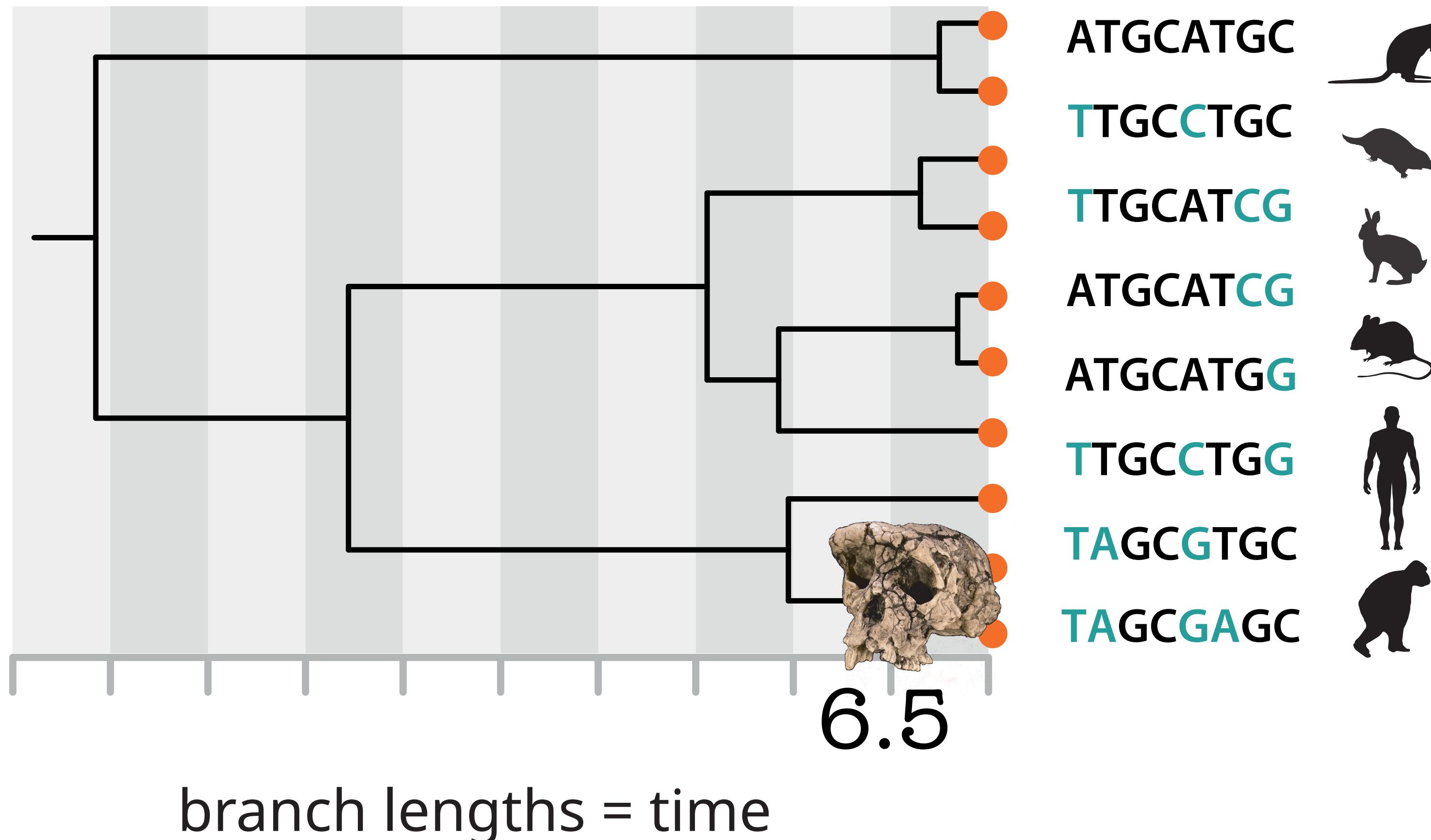
Goal: to disentangle
evolutionary rate and
time

The molecular clock hypothesis



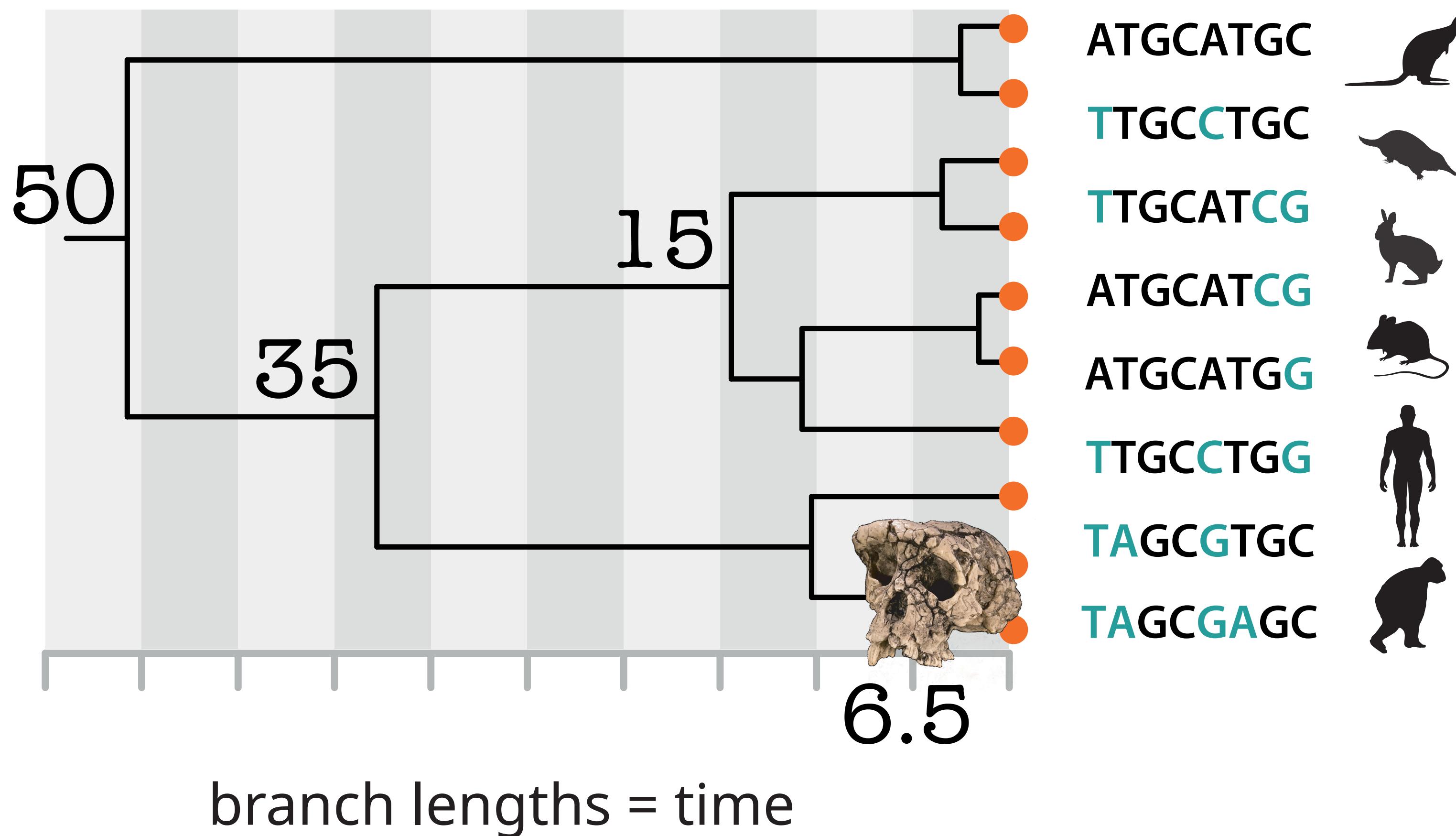
Molecules as documents of evolutionary history Zuckerkandl & Pauling ([1965](#))
A history of the molecular clock Morgan ([1998](#))

Calibrating the substitution rate



Temporal evidence of divergence for one species pair let's us **calibrate** the average rate of molecular evolution

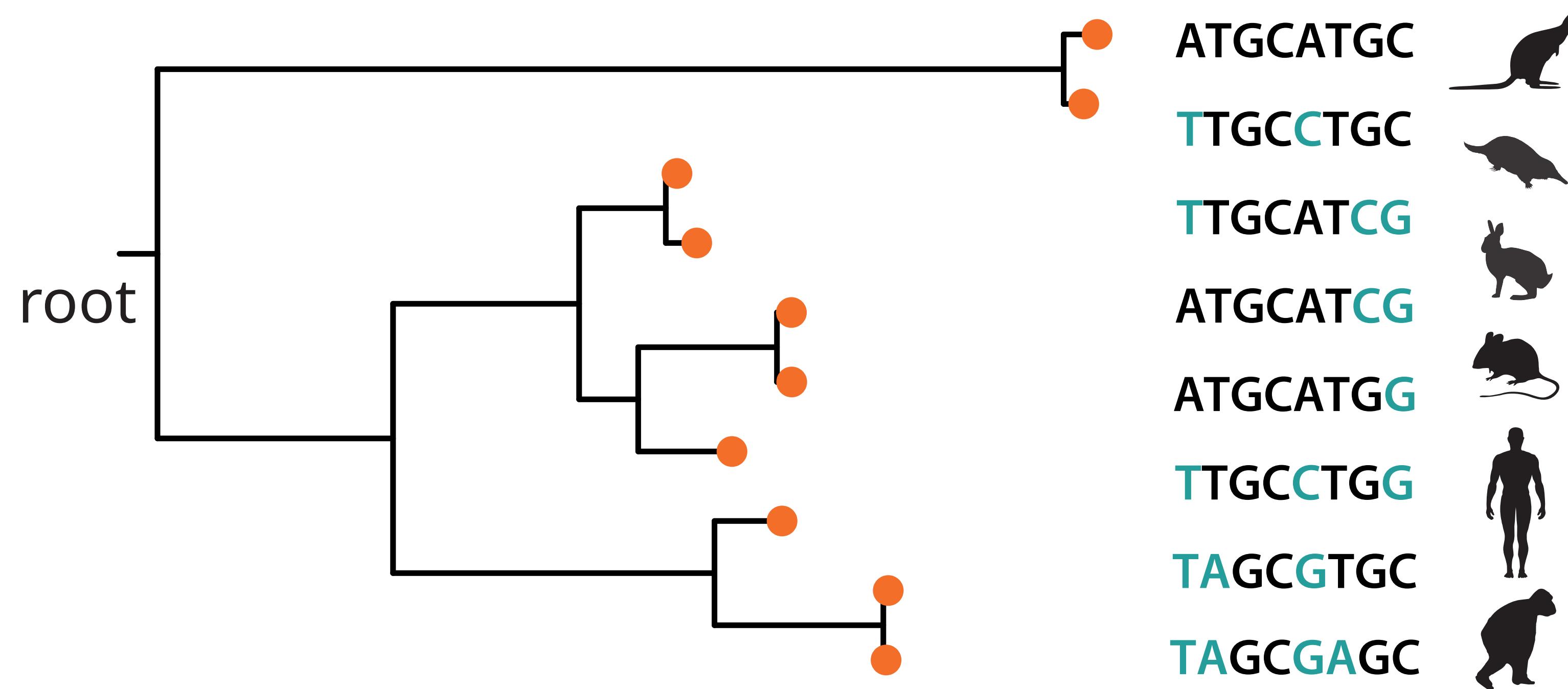
Calibrating the substitution rate



We can use this rate to extrapolate the divergence times for other species pairs

Molecular dating: challenges

Rate and time are not fully identifiable!

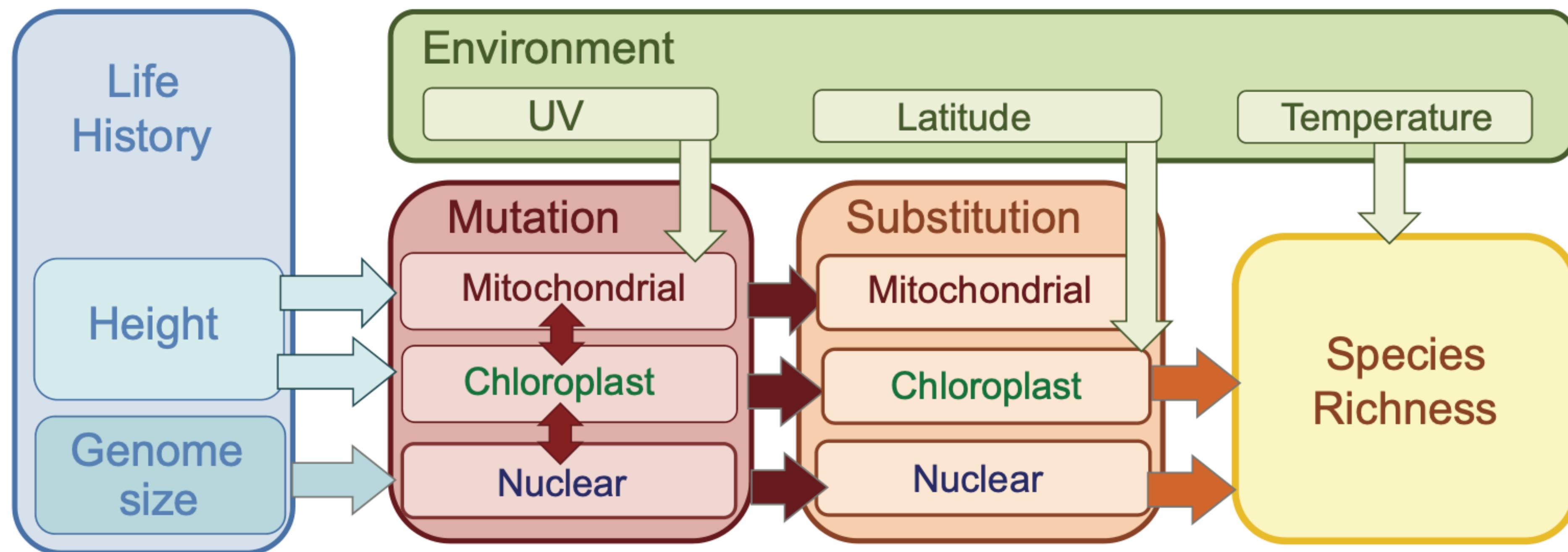


branch lengths = genetic distance

$$v = rt$$

Molecular dating: challenges

Many variables contribute to variation in the substitution rate



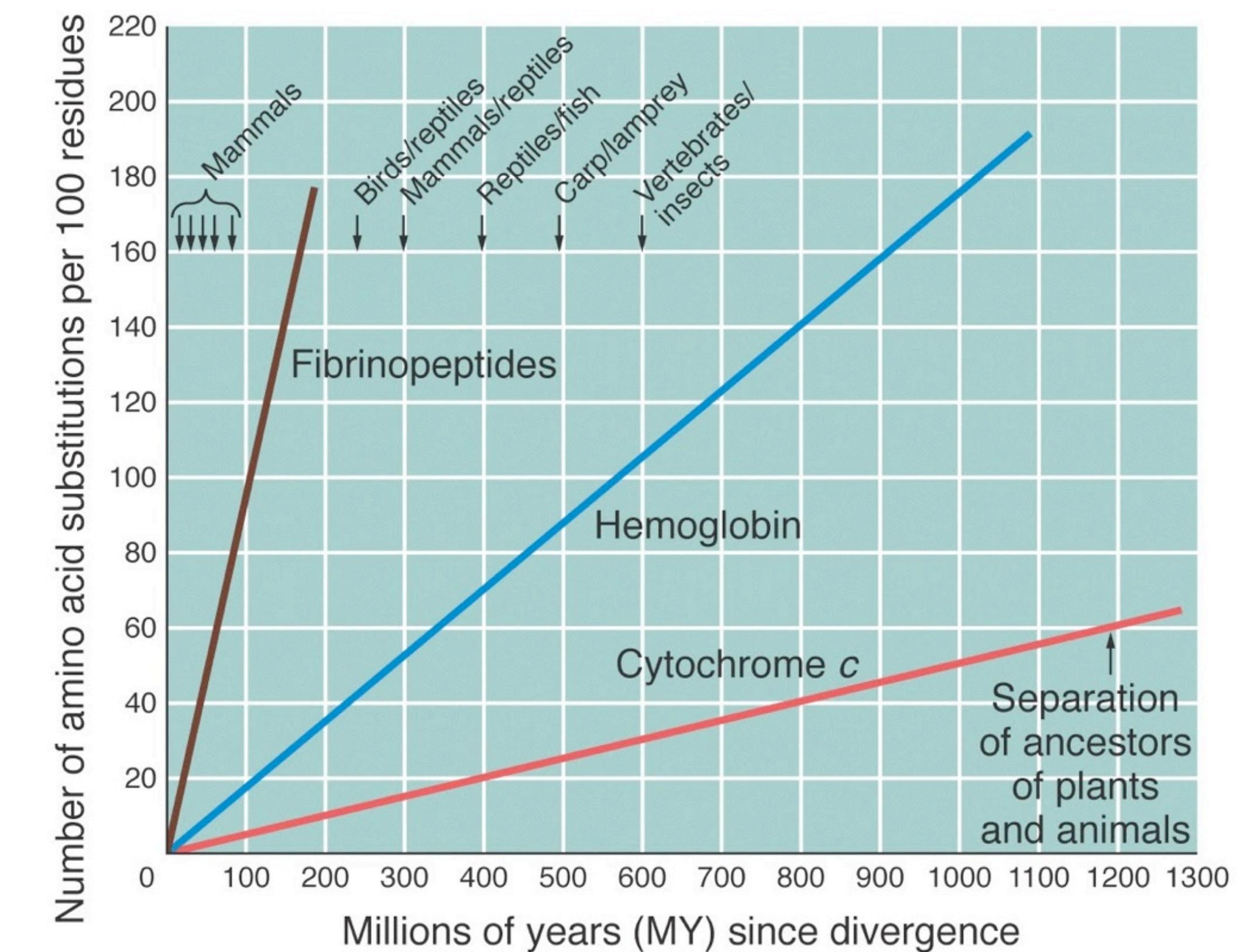
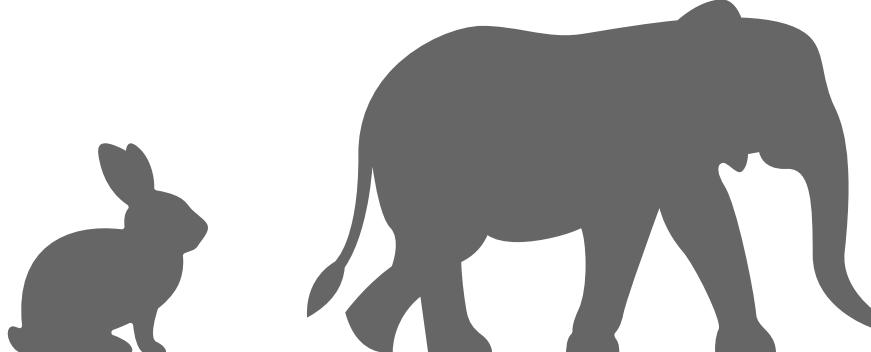
Molecular dating: challenges

Many variables contribute to **variation in the substitution rate**

The molecular clock is not constant

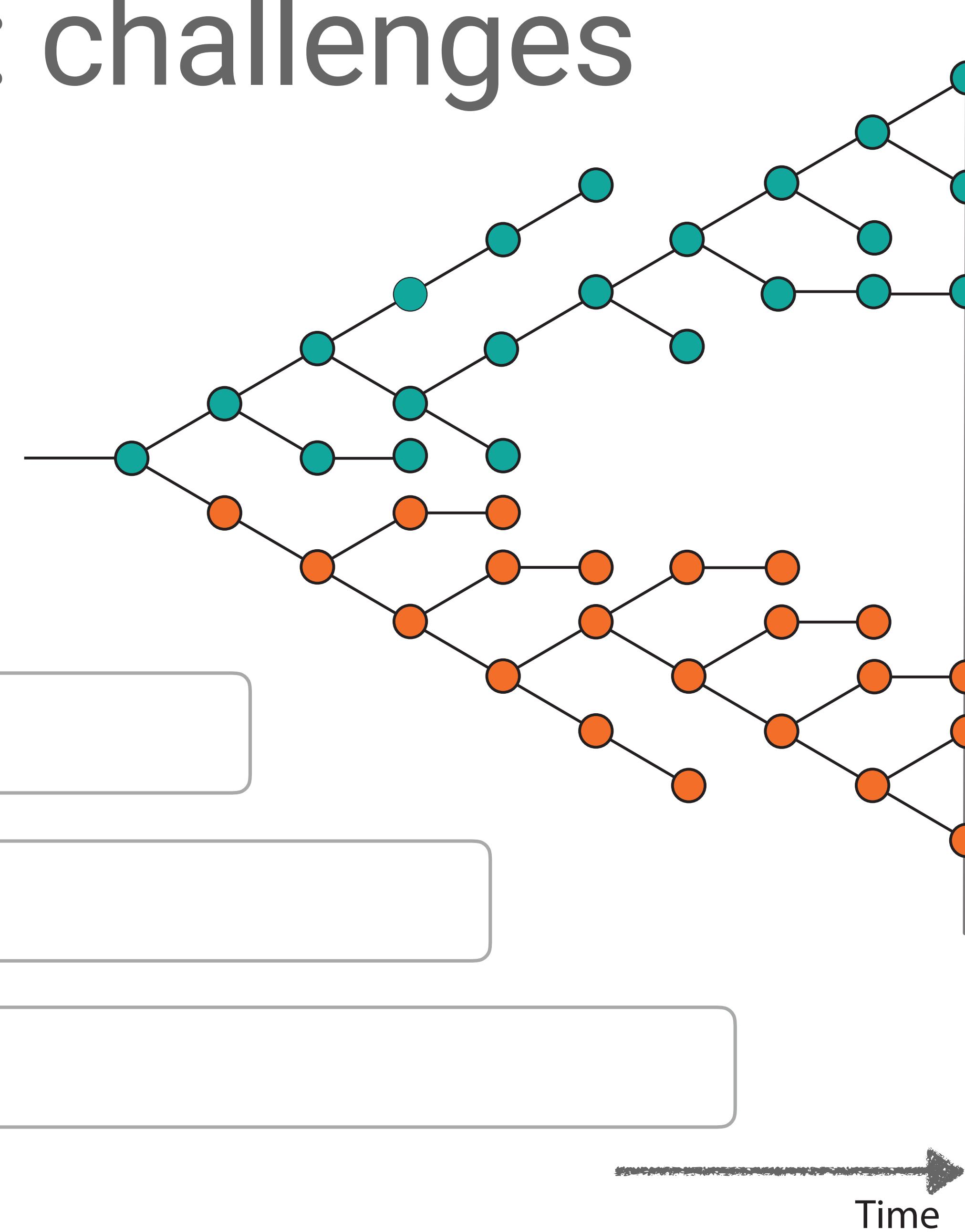
Rates vary across:

- taxa
- time
- genes
- sites within the same gene



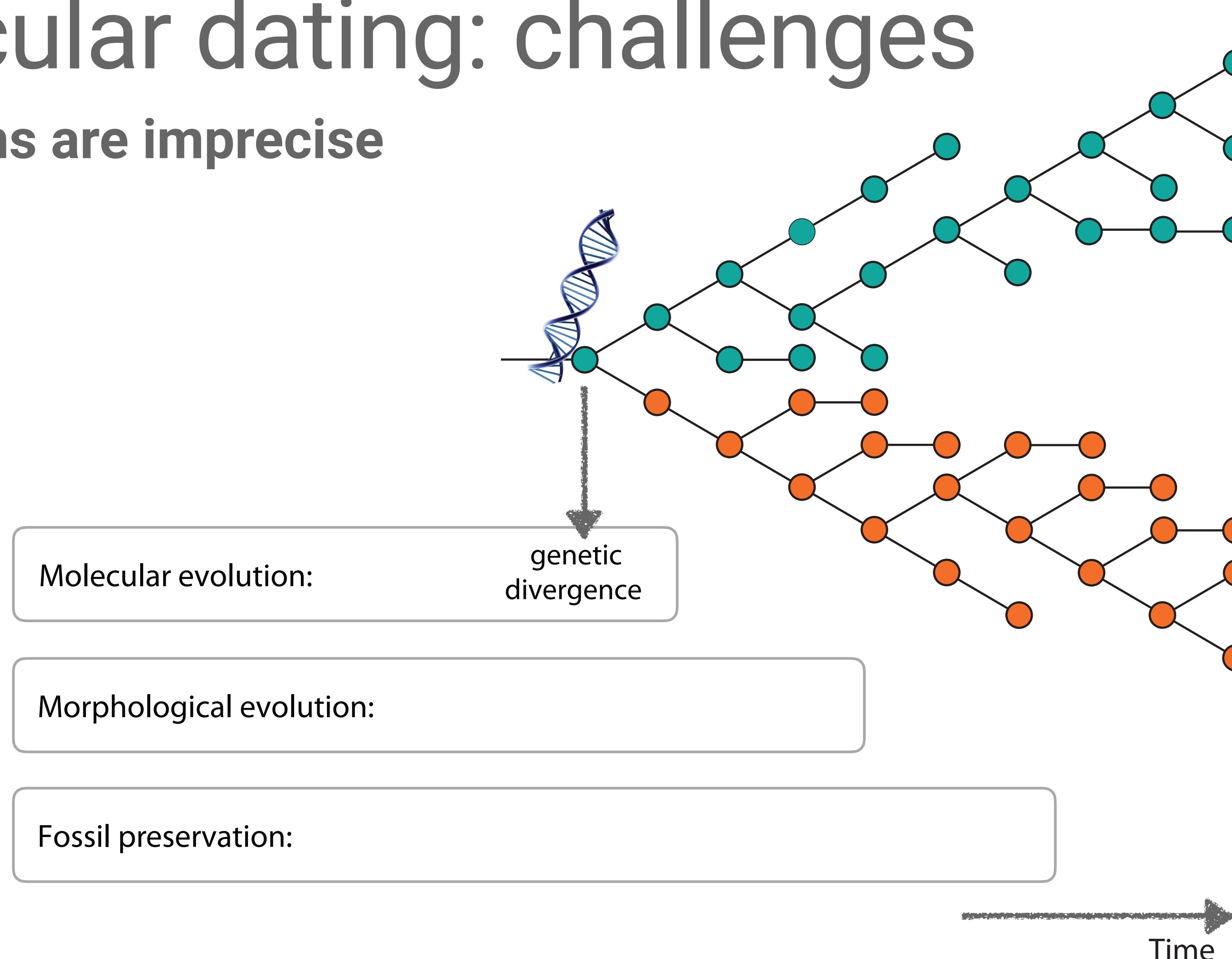
Molecular dating: challenges

Calibrations are imprecise



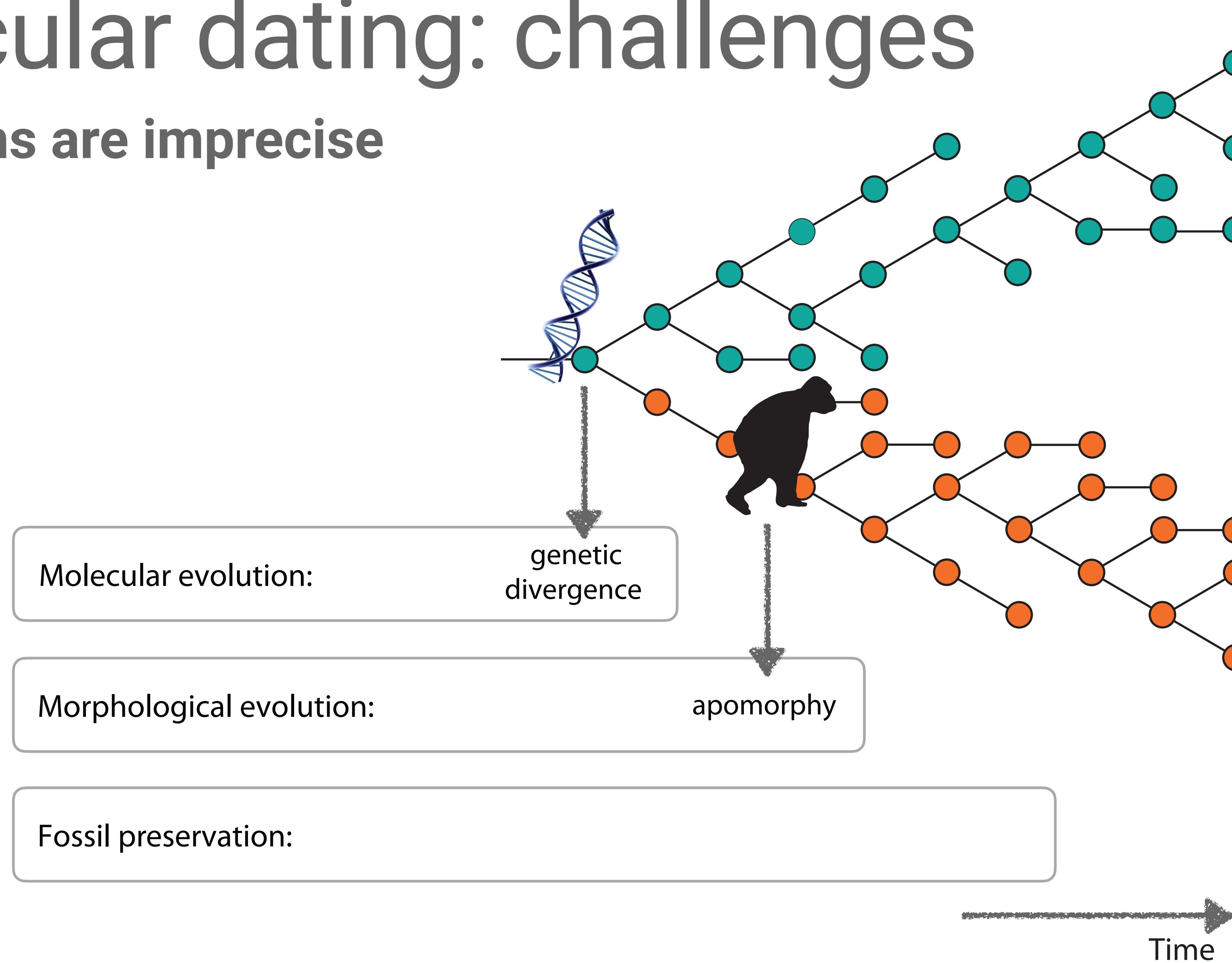
Molecular dating: challenges

Calibrations are imprecise



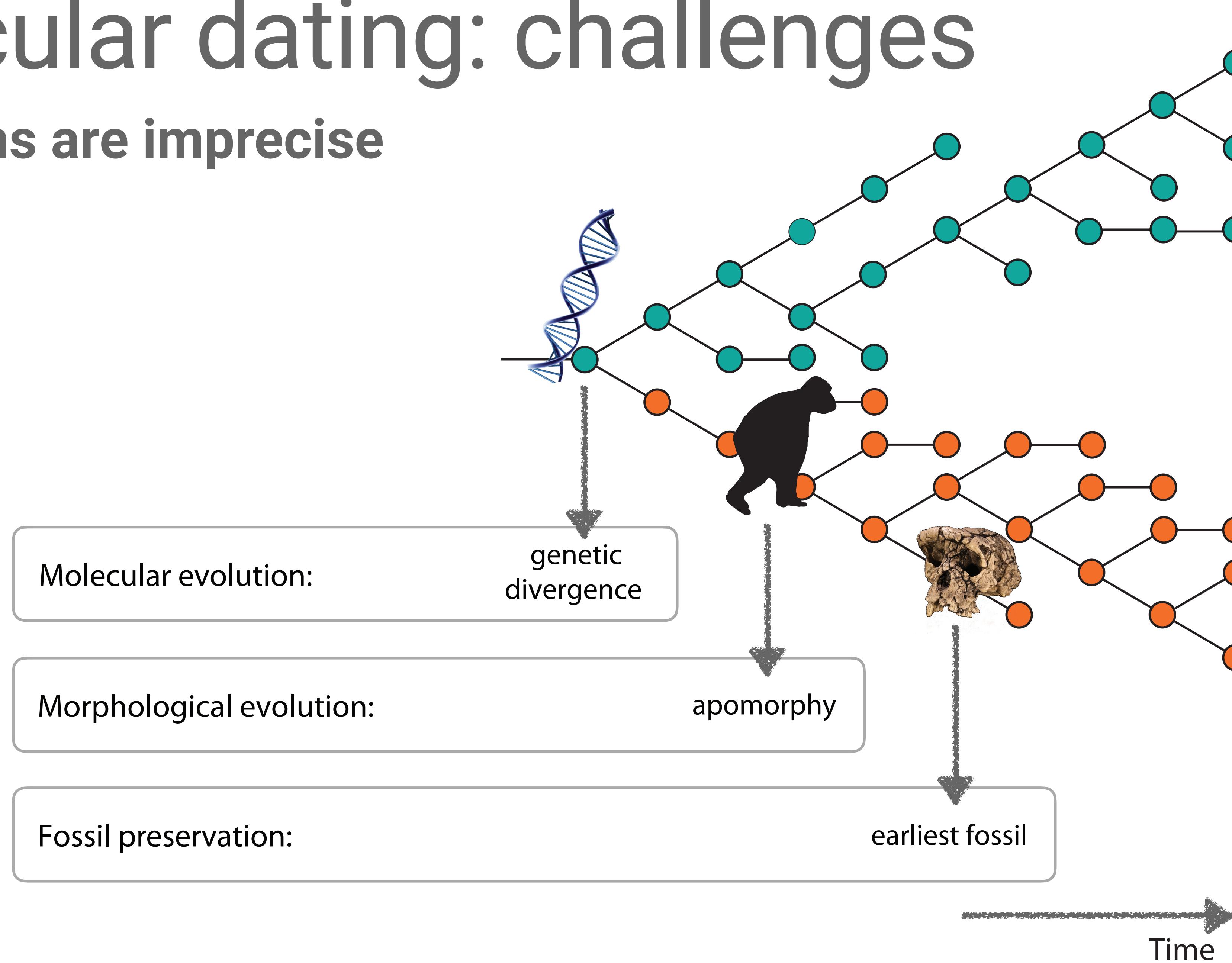
Molecular dating: challenges

Calibrations are imprecise



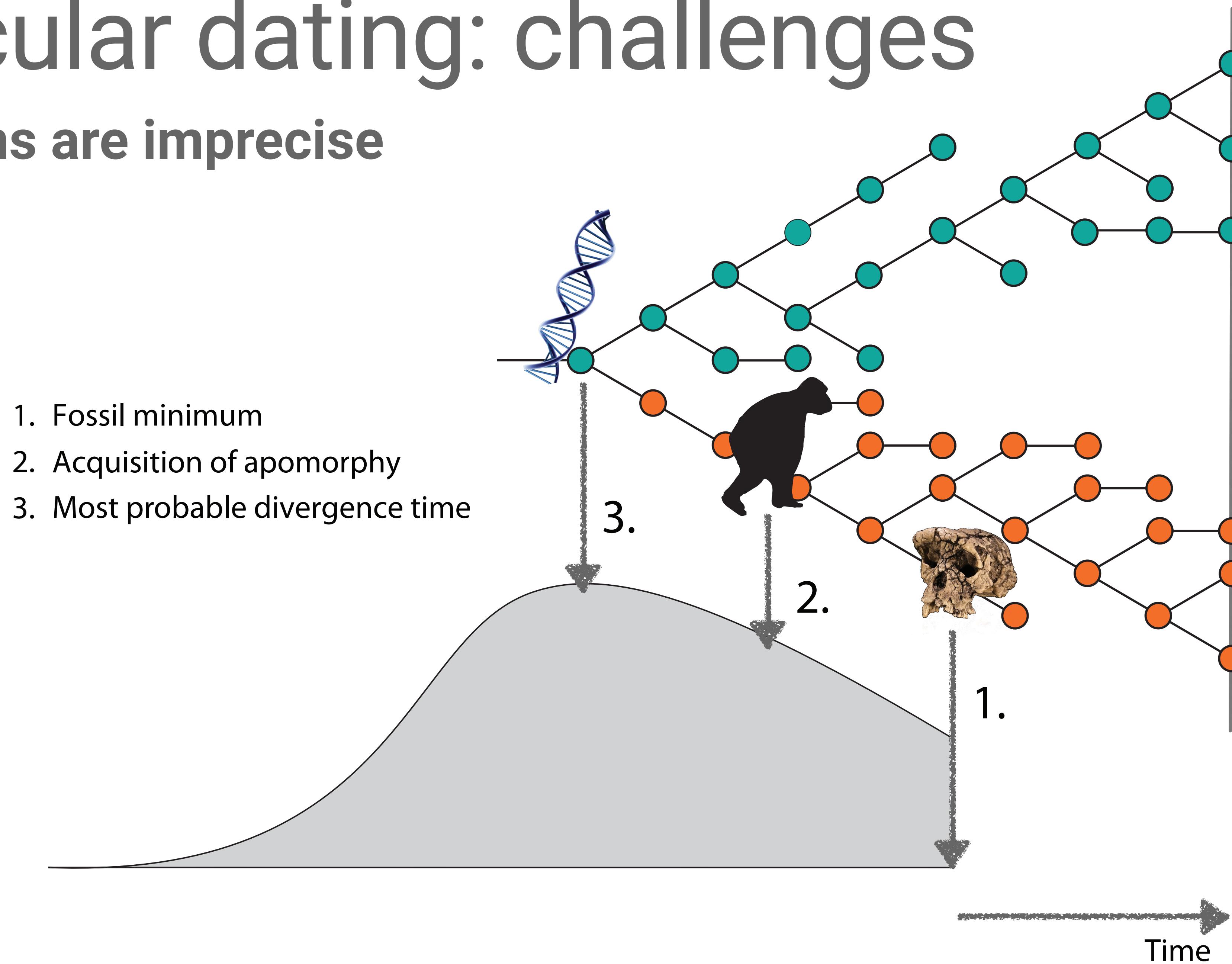
Molecular dating: challenges

Calibrations are imprecise



Molecular dating: challenges

Calibrations are imprecise



Molecular dating: challenges

Summary

1. Rate and time are not fully identifiable

2. The substitution rate varies

3. Calibrations are imprecise

→ we need a flexible statistical framework that deals well with uncertainty!

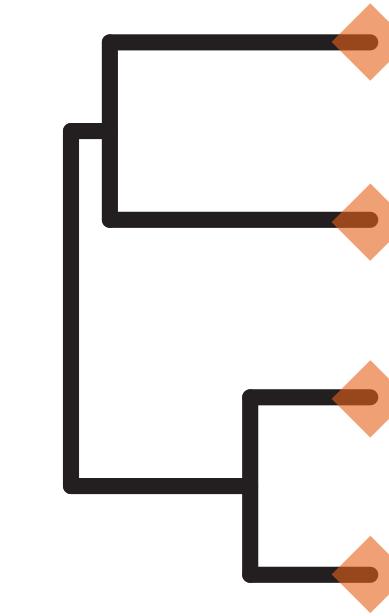
Bayesian divergence time estimation

Components used to infer trees

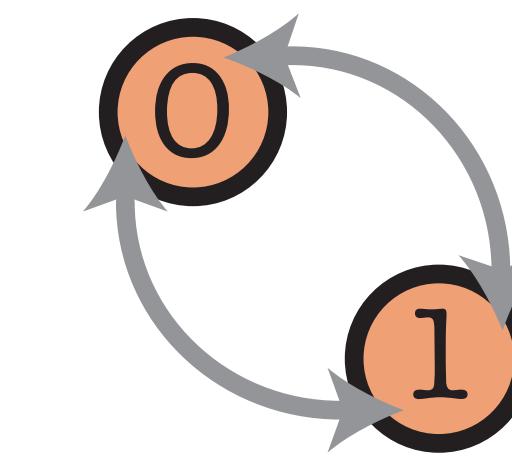
without considering time

0101...
1101...
0100...

data
sequences or
characters



tree
topology and
branch lengths



substitution
model

Bayesian tree inference

$$\text{posterior} \quad P(E \mid \text{0101...}, \text{1101...}, \text{0100...}) = \frac{\text{likelihood} \quad P(\text{0101...}, \text{1101...}, \text{0100...} \mid E, \text{prior})}{\text{priors} \quad P(E)}$$

marginal probability

We use a Bayesian framework

$$P(\text{ model } | \text{ data }) = \frac{P(\text{ data } | \text{ model }) P(\text{ model })}{P(\text{ data })}$$

likelihood

priors

posterior

marginal probability of the data

Bayesian divergence time estimation

The data

AND/OR

0101... ATTG...

1101... TTGC...

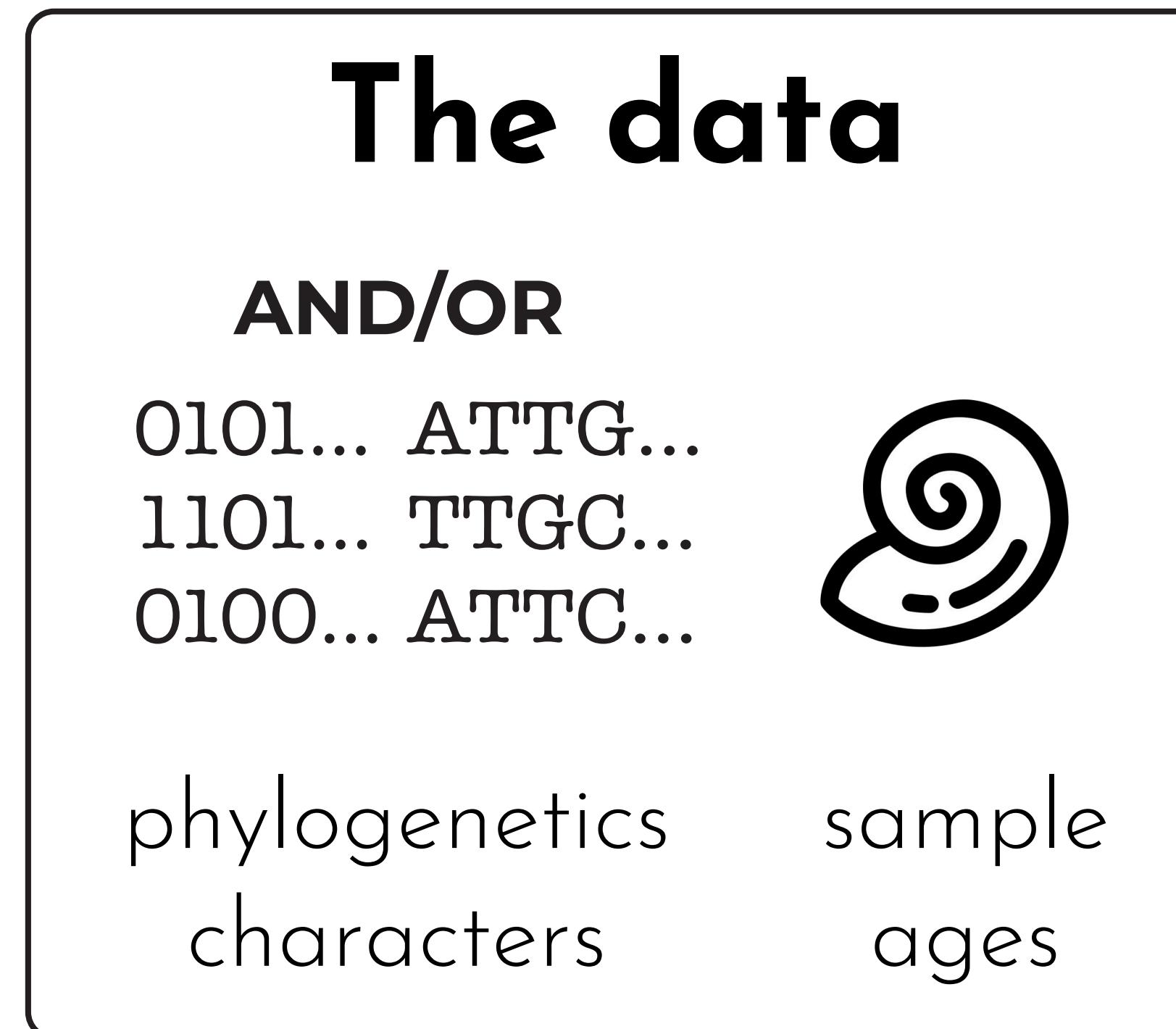
0100... ATTC...



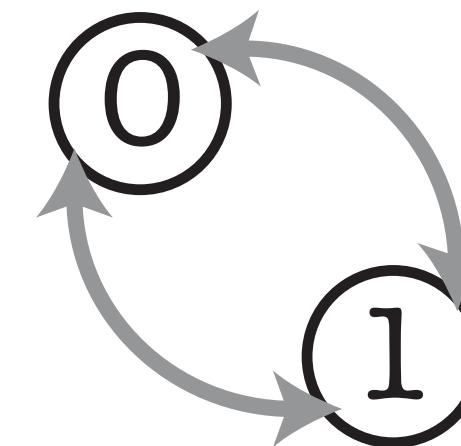
phylogenetics
characters

sample
ages

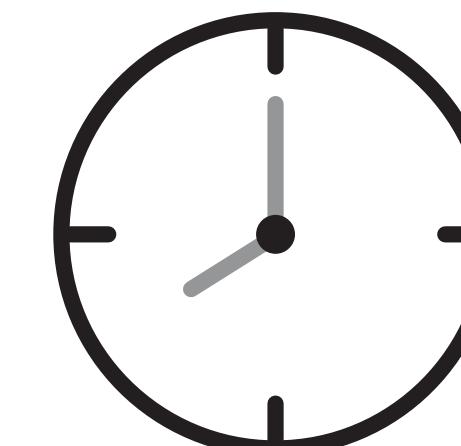
Bayesian divergence time estimation



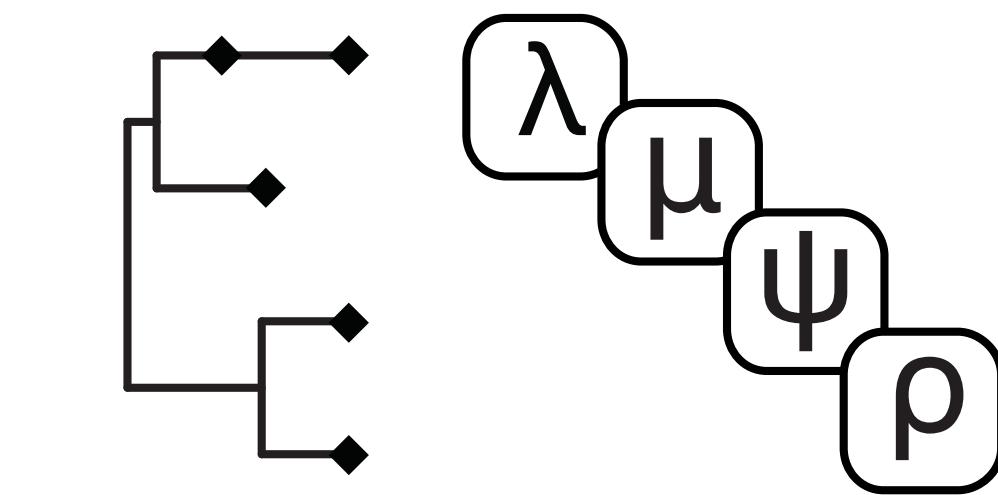
3 model components



substitution
model

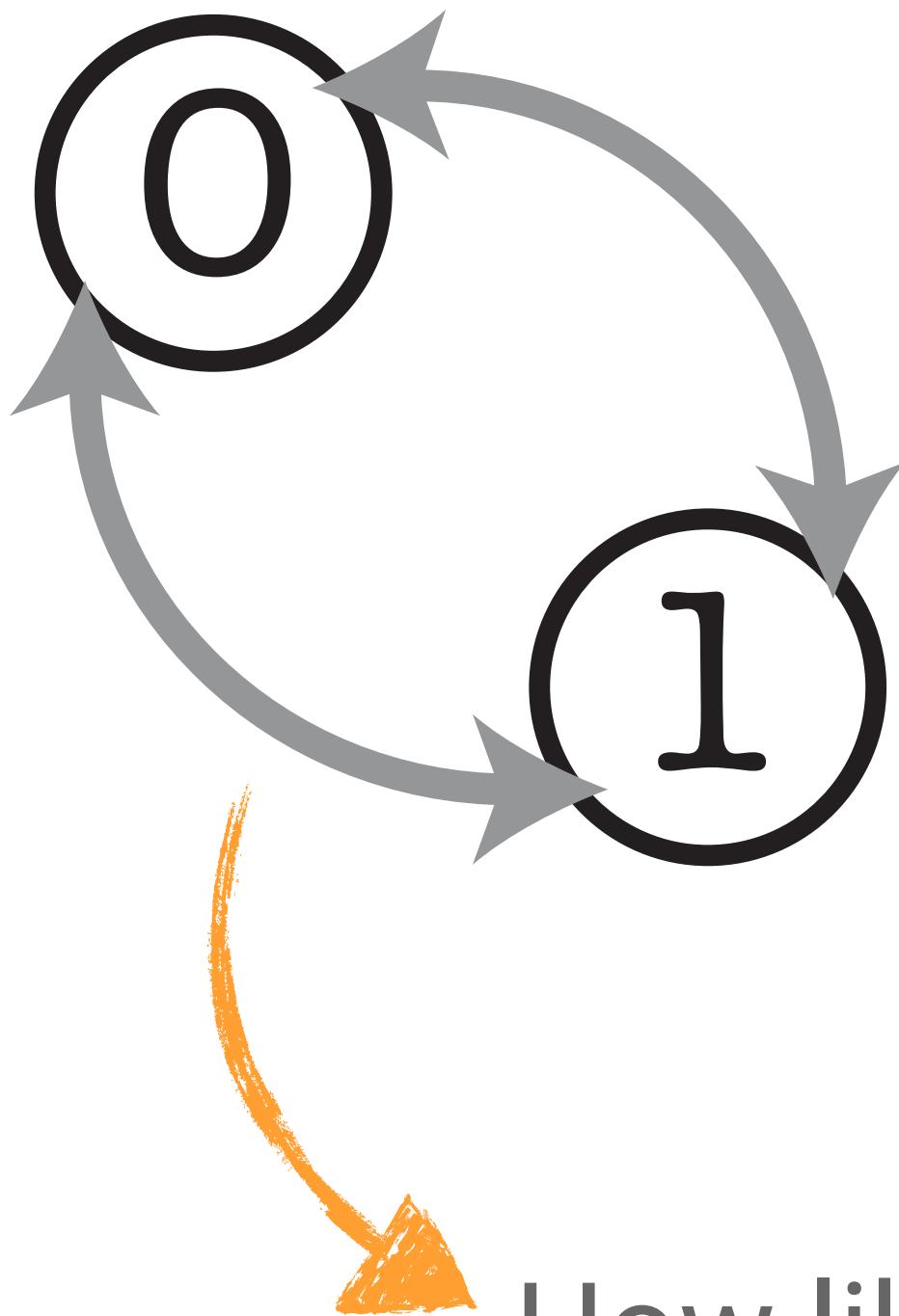


clock
model

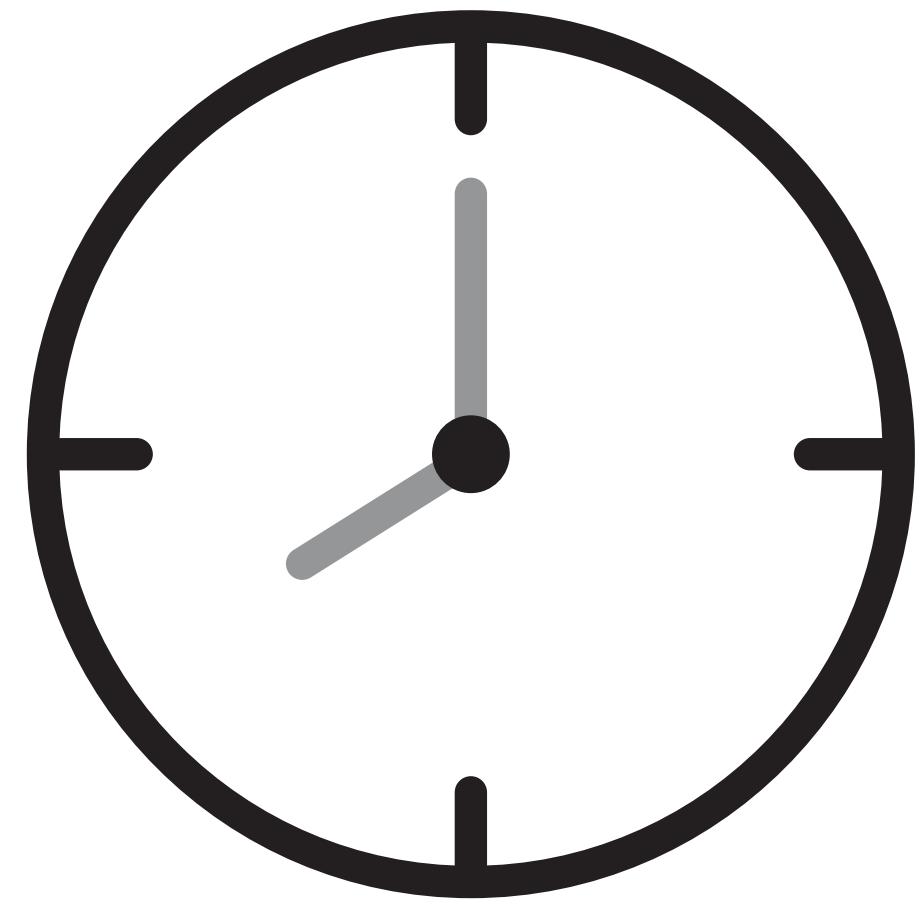


tree and
tree model

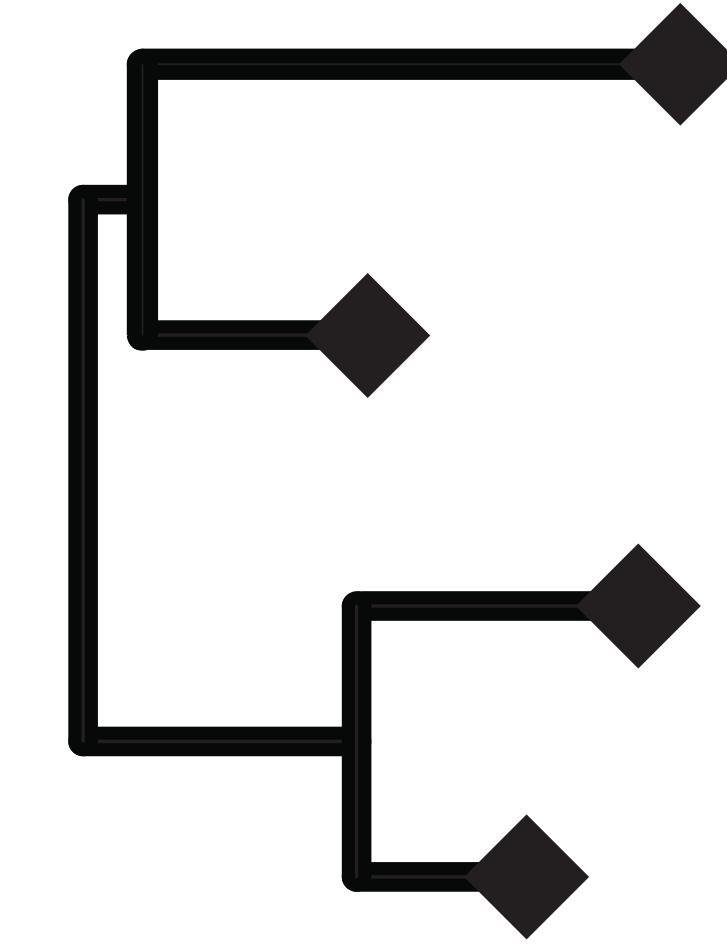
substitution model



clock model

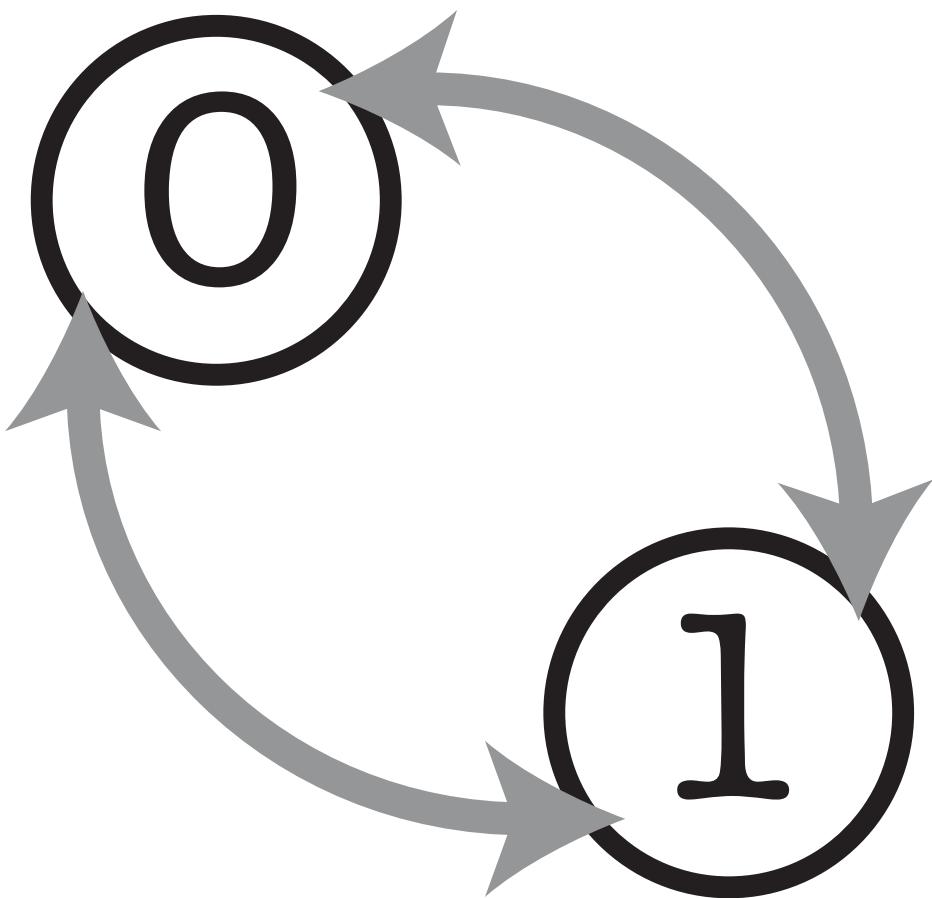


tree model

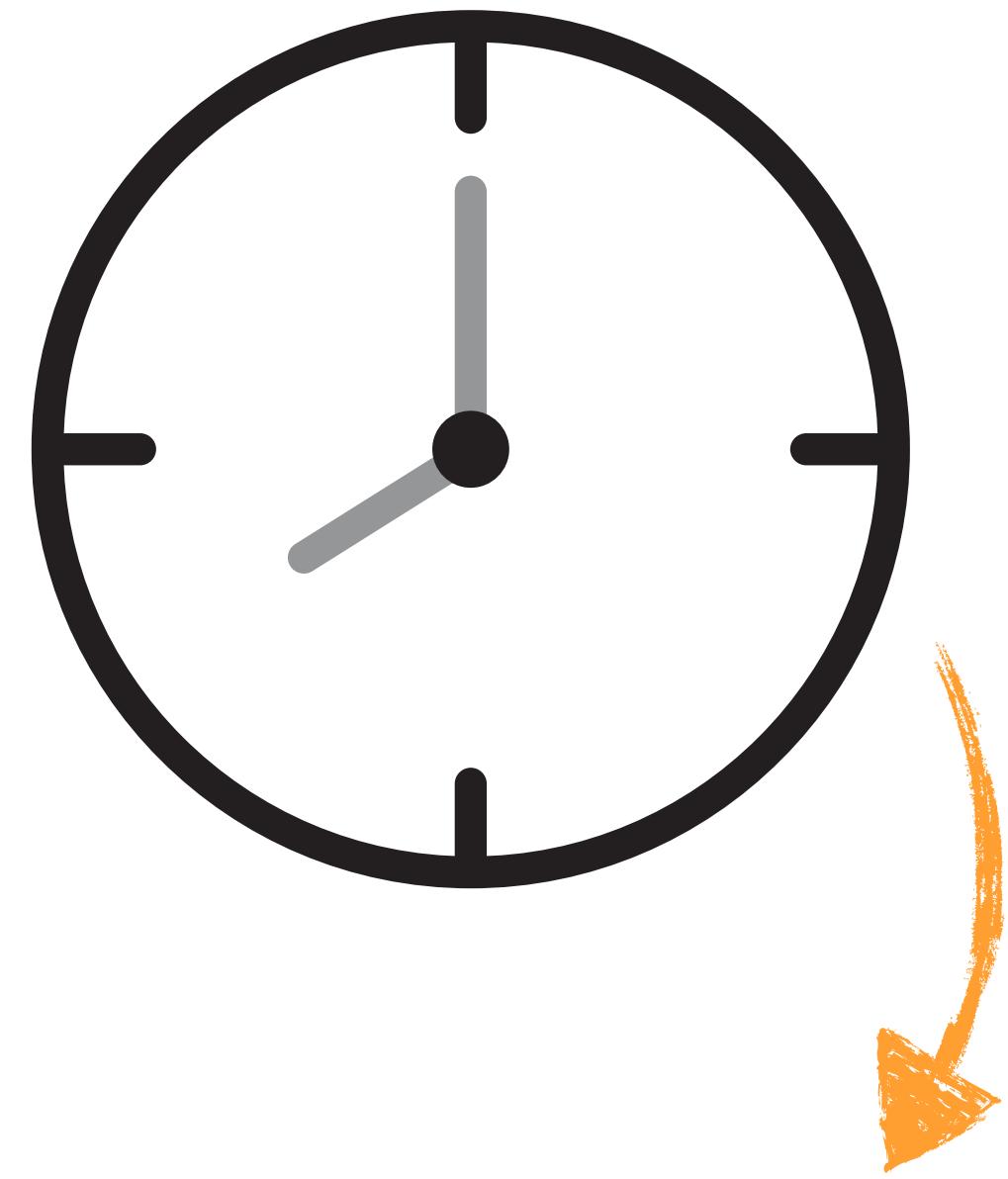


How likely are we to observe a change
between character states? e.g., A → T

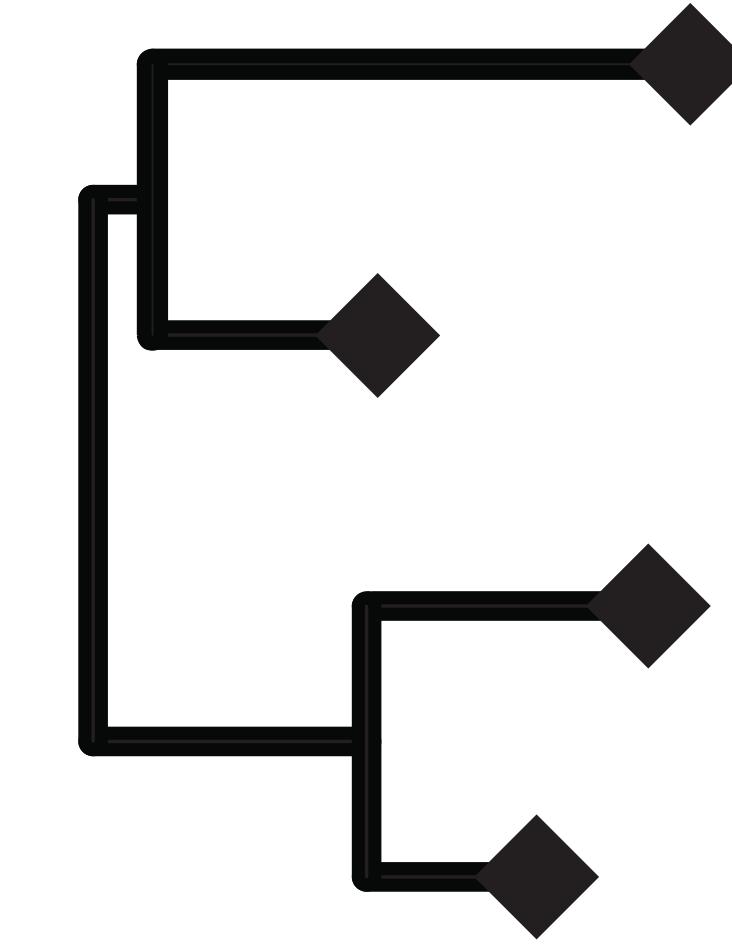
substitution model



clock model

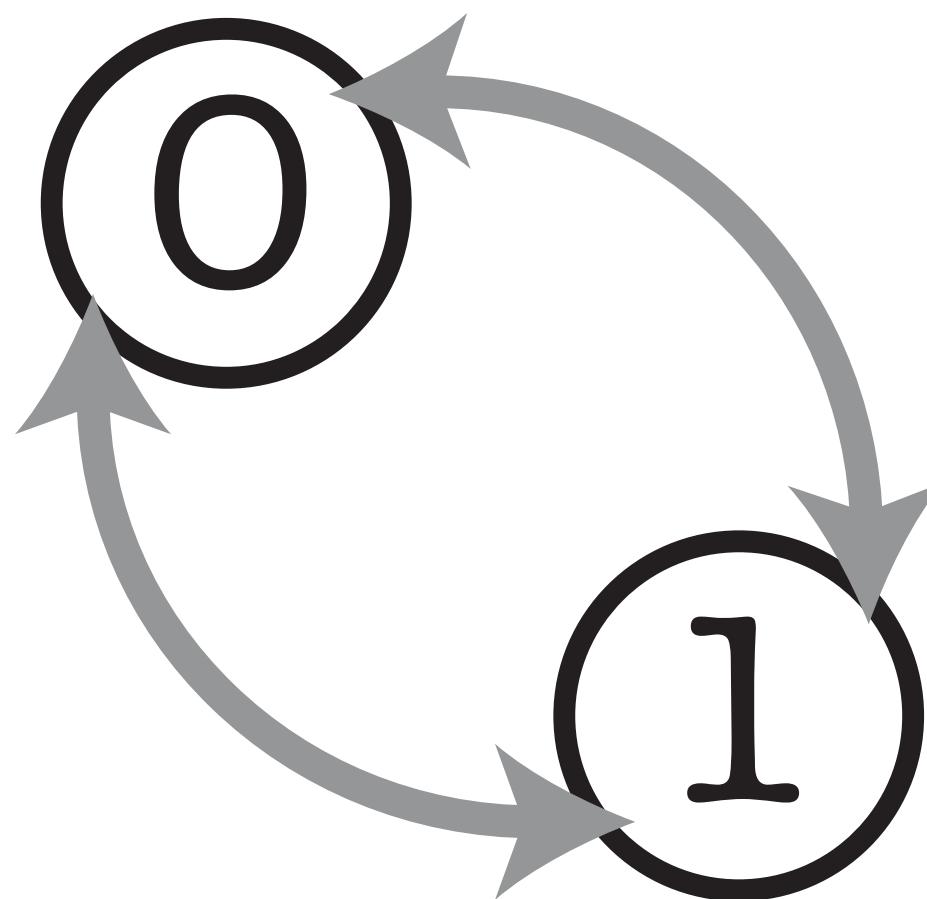


tree model

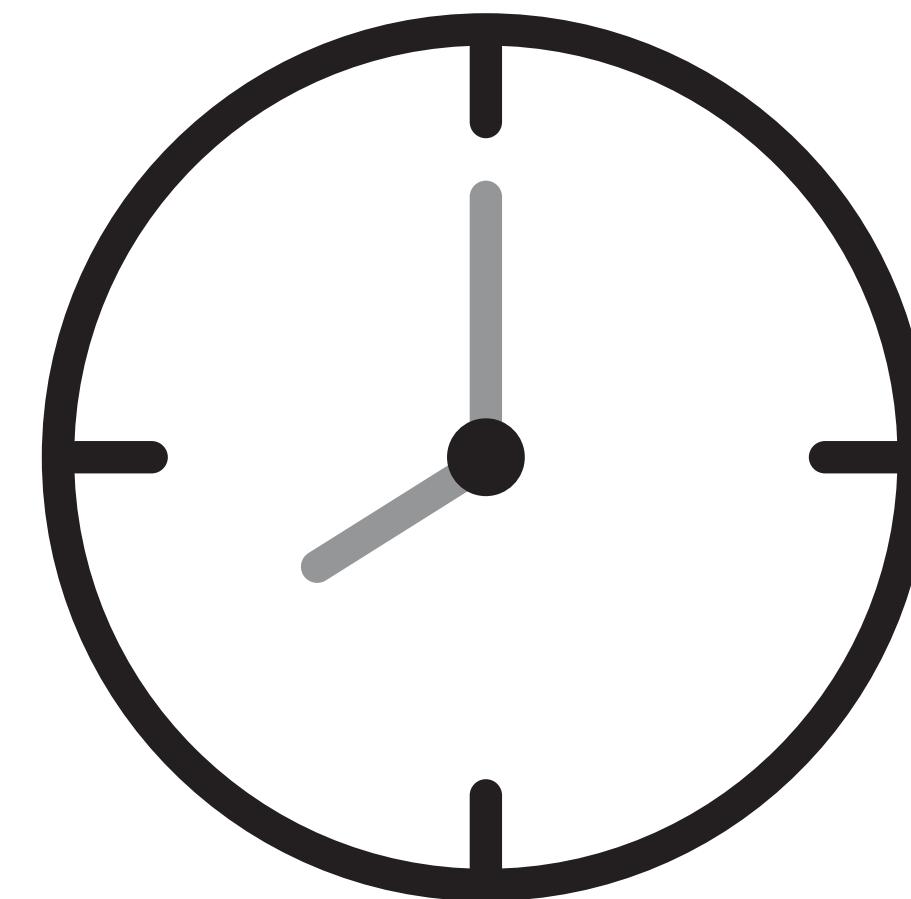


How have rates of evolution varied (or not)
across the tree?

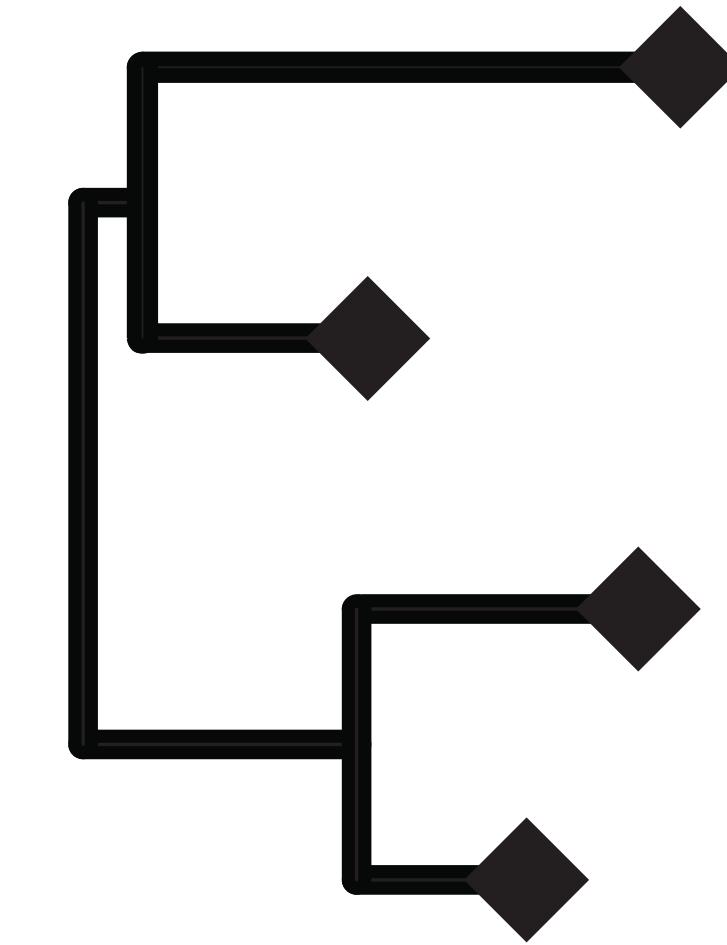
substitution model



clock model



tree model



How have species originated, gone extinct and been sampled through time?

Note: the tree model is often referred to as the **tree prior** even though the fossil sampling times are also data. See [May & Rothfels 2023](#)

Bayesian divergence time estimation

posterior

$$P(E \mid \lambda, \mu, \psi, p, O, t \mid 0101\dots, 1101\dots, 0100\dots, \text{snail}) =$$

likelihood

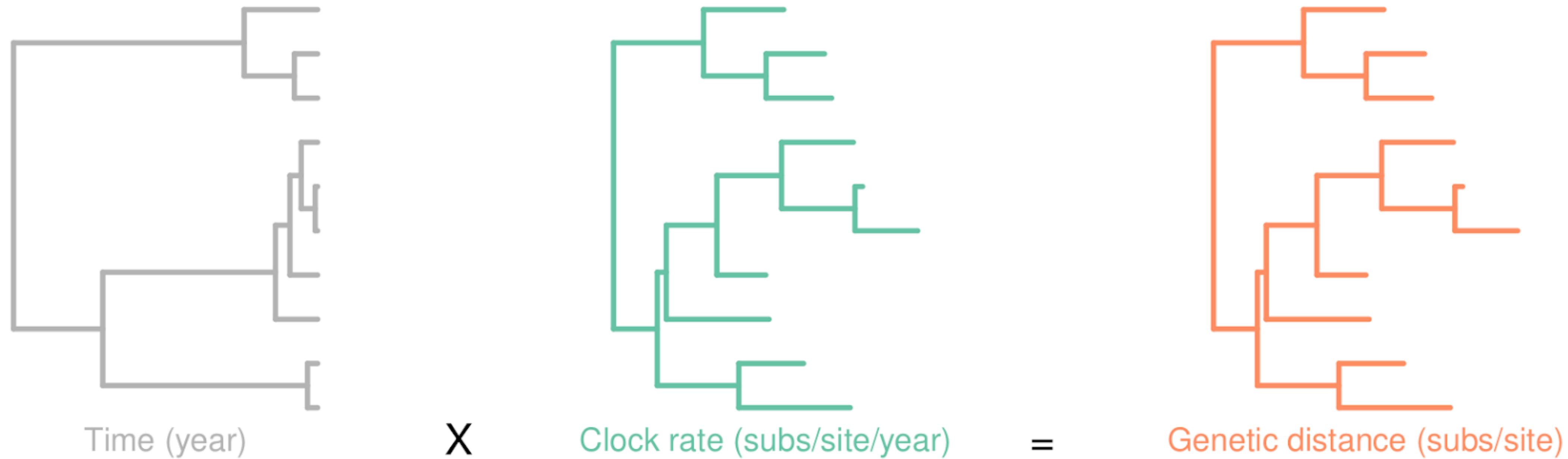
probability of the
time tree

priors

$$P(0101\dots, 1101\dots, 0100\dots \mid E) P(E \mid \lambda, \mu, \psi, p, O, t) = P(O \mid \lambda, \mu, \psi, p) P(O \mid t) P(t)$$

marginal pr of the data

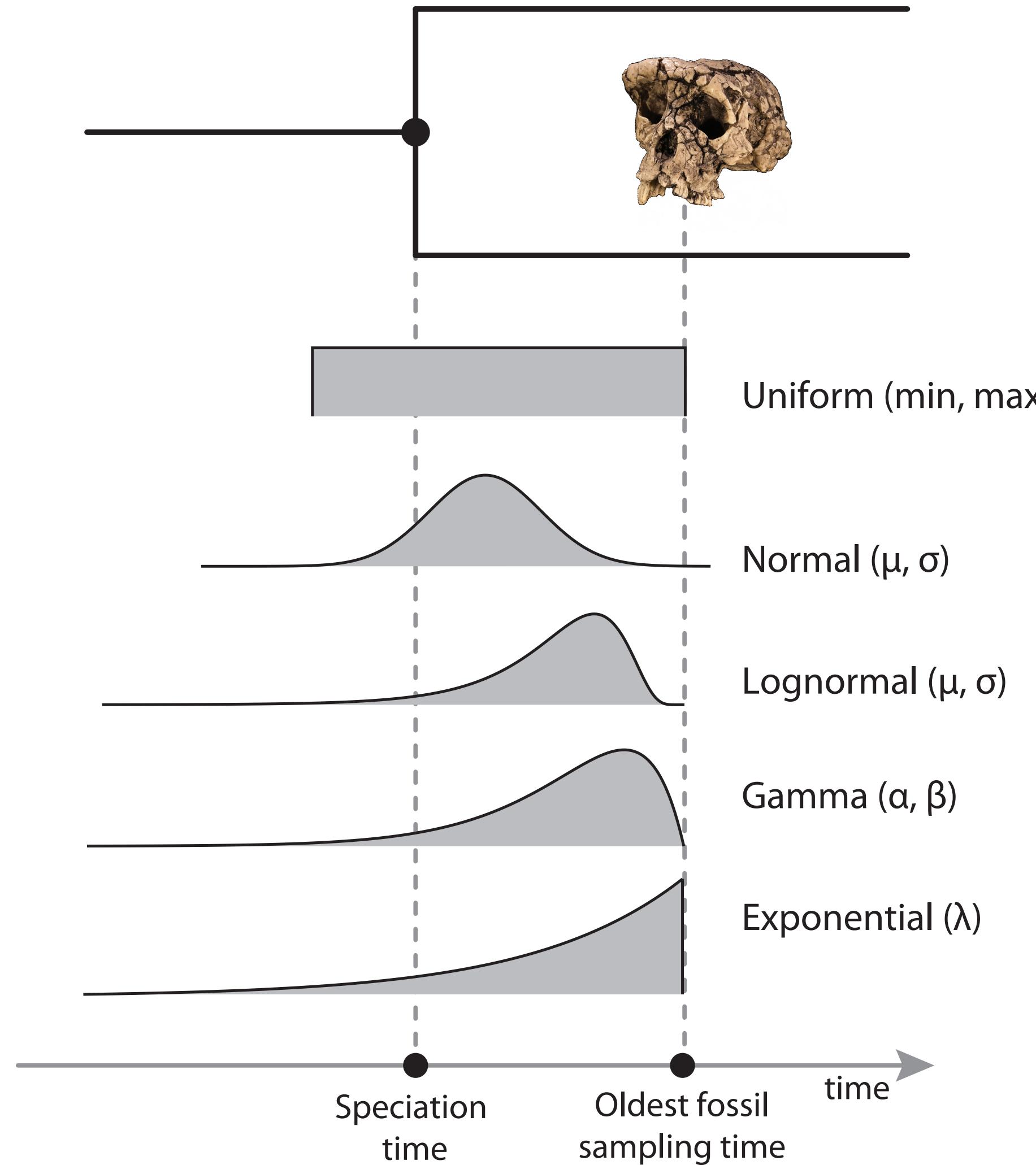
Calculating the likelihood



Based on the calibration times we can estimate the rate over time

Once we have the rate we can transform evolutionary rates in genetic distance

Node dating



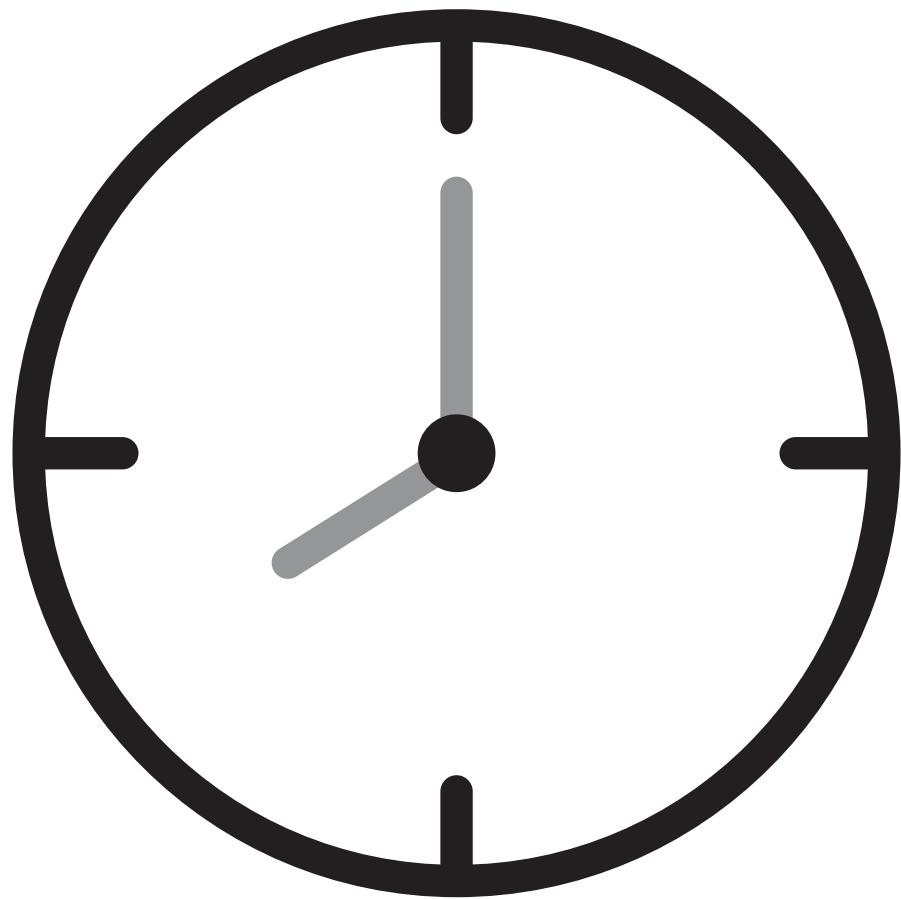
We can use a **calibration density** to constrain internal node ages

We typically use a **birth-death process** model to describe the tree generating process

Adapted from Heath 2012. Sys Bio

clock model

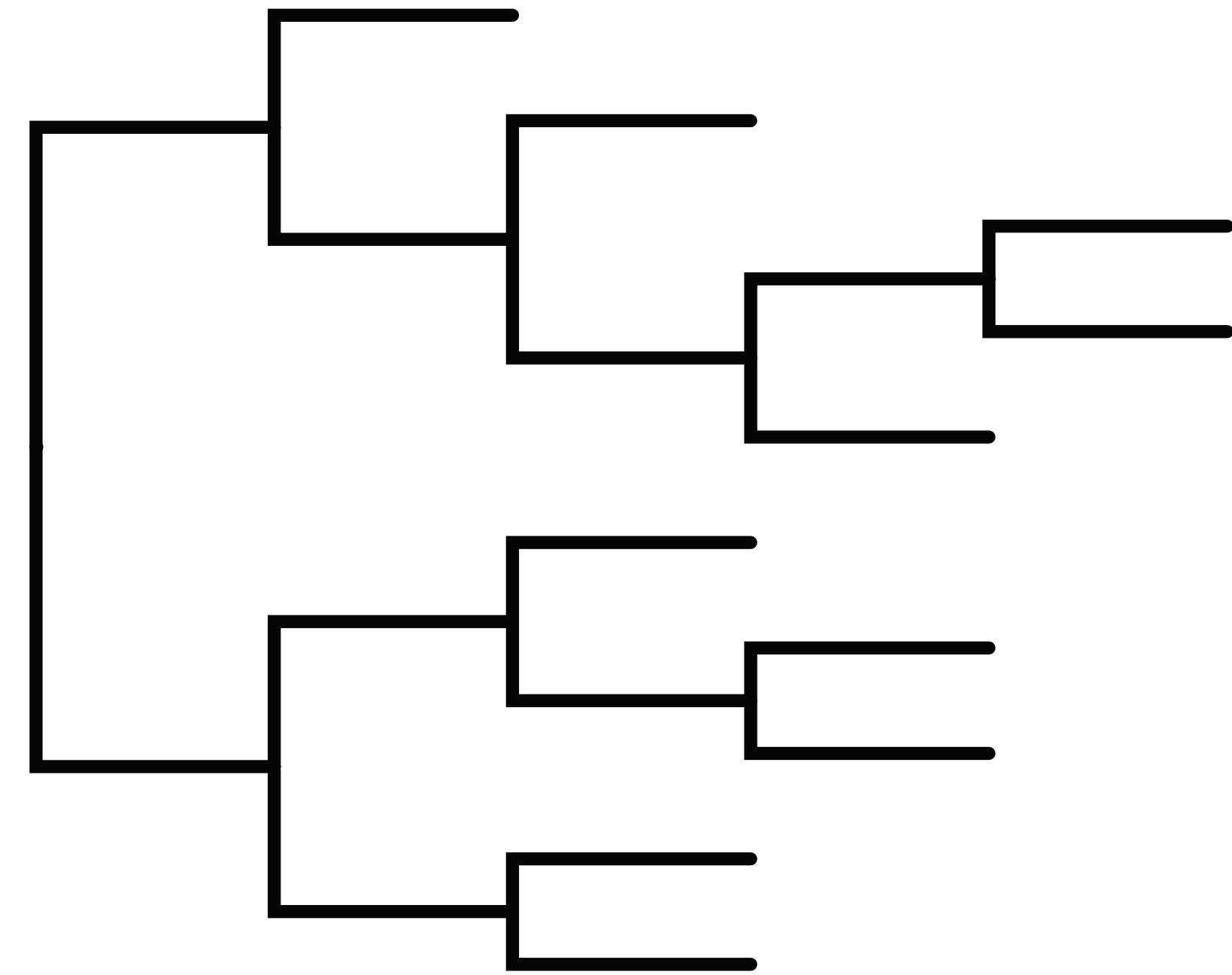
The clock model describes how evolutionary rates vary (or not) across the tree



The strict / constant molecular clock model

Assumptions

- The substitution rate is constant over time
 - All lineages share the same rate



branch length = substitution rate

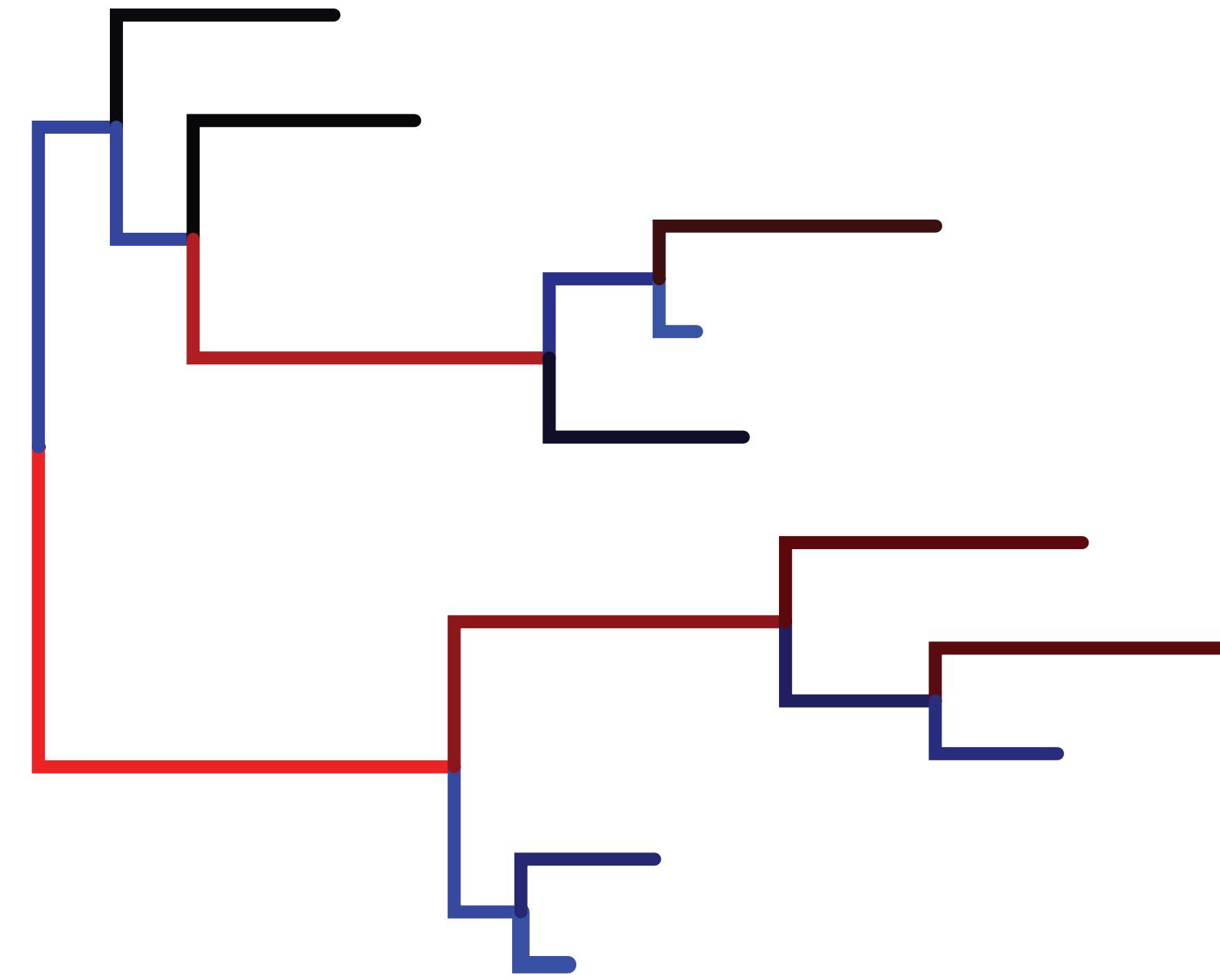


low high

Relaxed clock models

Assumptions

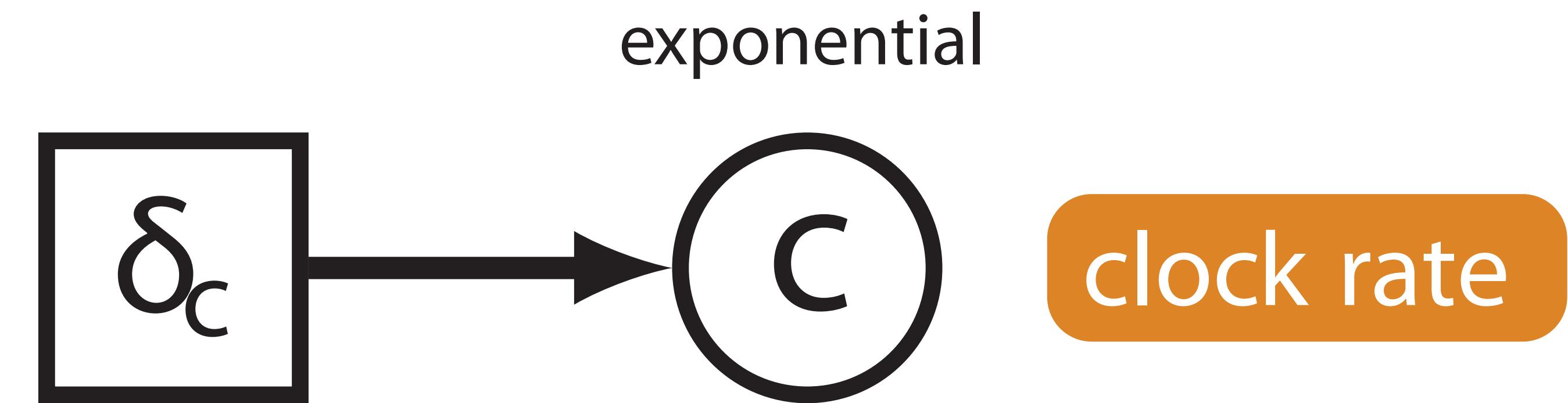
- Lineage-specific rates
- The rate assigned to each branch is drawn from some underlying distribution



branch length = substitution rate
low high

Graphical models: strict clock model

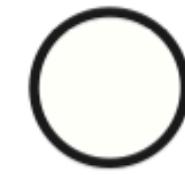
- a) Constant node
- b) Stochastic node
- c) Deterministic node
- d) Clamped node
(observed)
- e) Plate



Graphical models: relaxed clock model



a) Constant node



b) Stochastic node



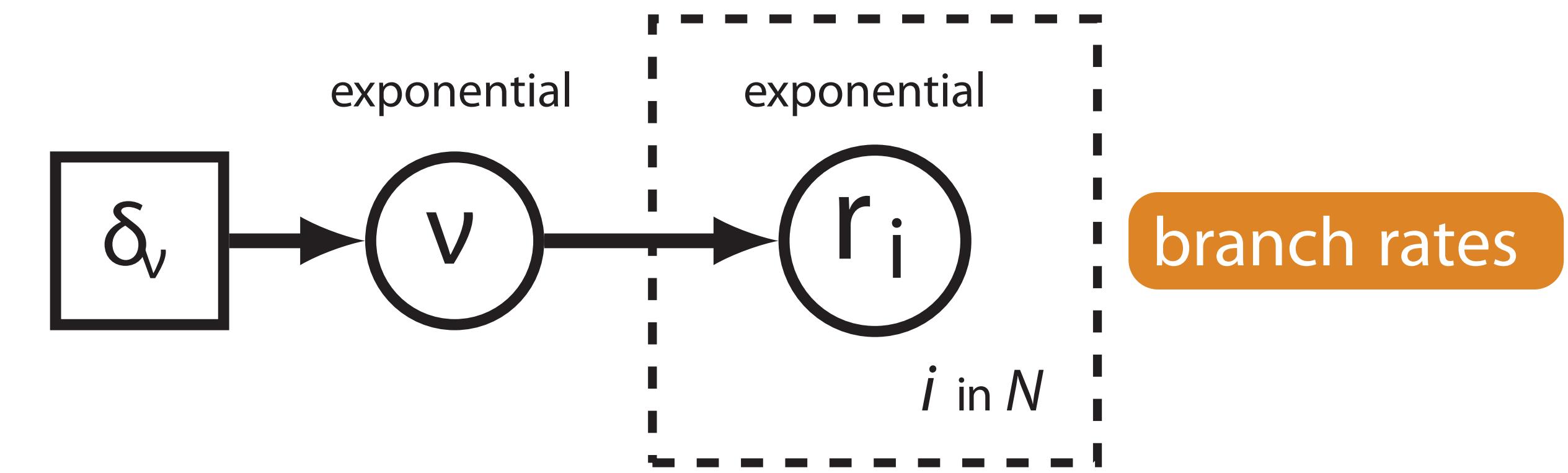
c) Deterministic node



d) Clamped node
(observed)



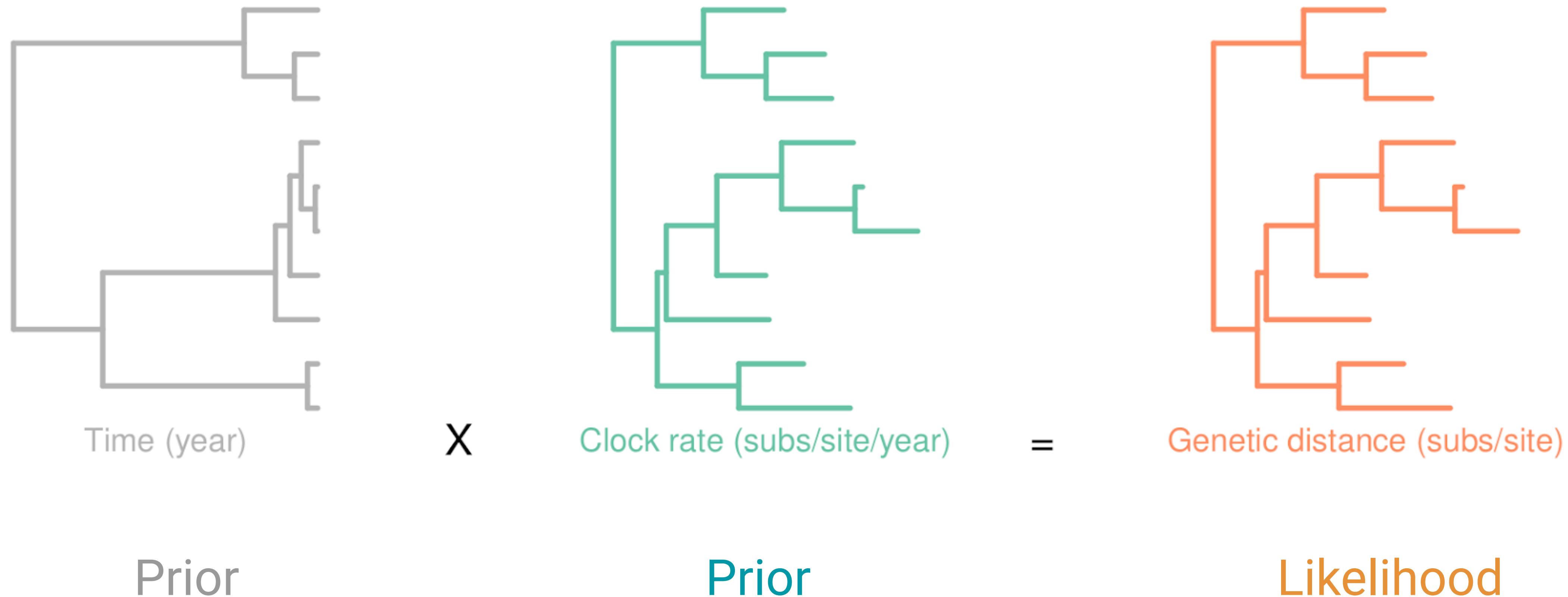
e) Plate



There are many different clock models

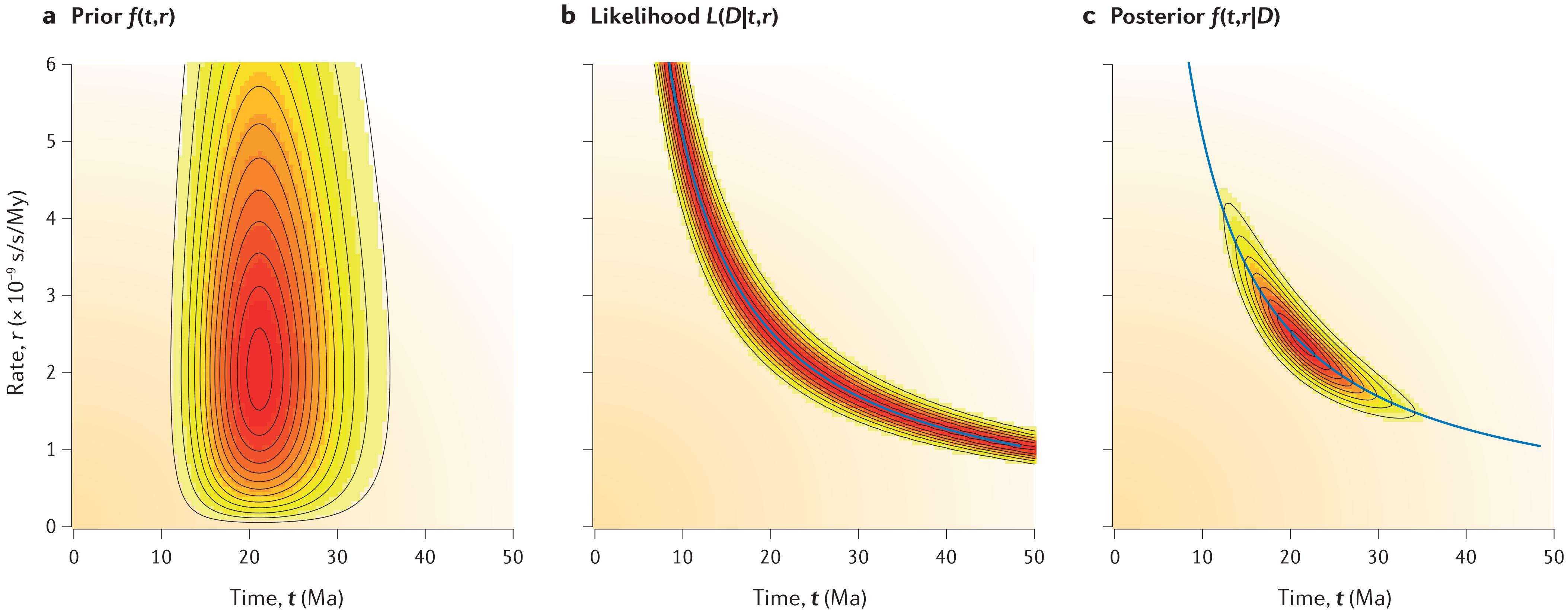
- Strict clock
- Uncorrelated or independent clock (= the favourite)
- Autocorrelated clock
- Local clocks
- Mixture models

Times and rates are not fully identifiable!



Slide adapted from Sebastian Duchene

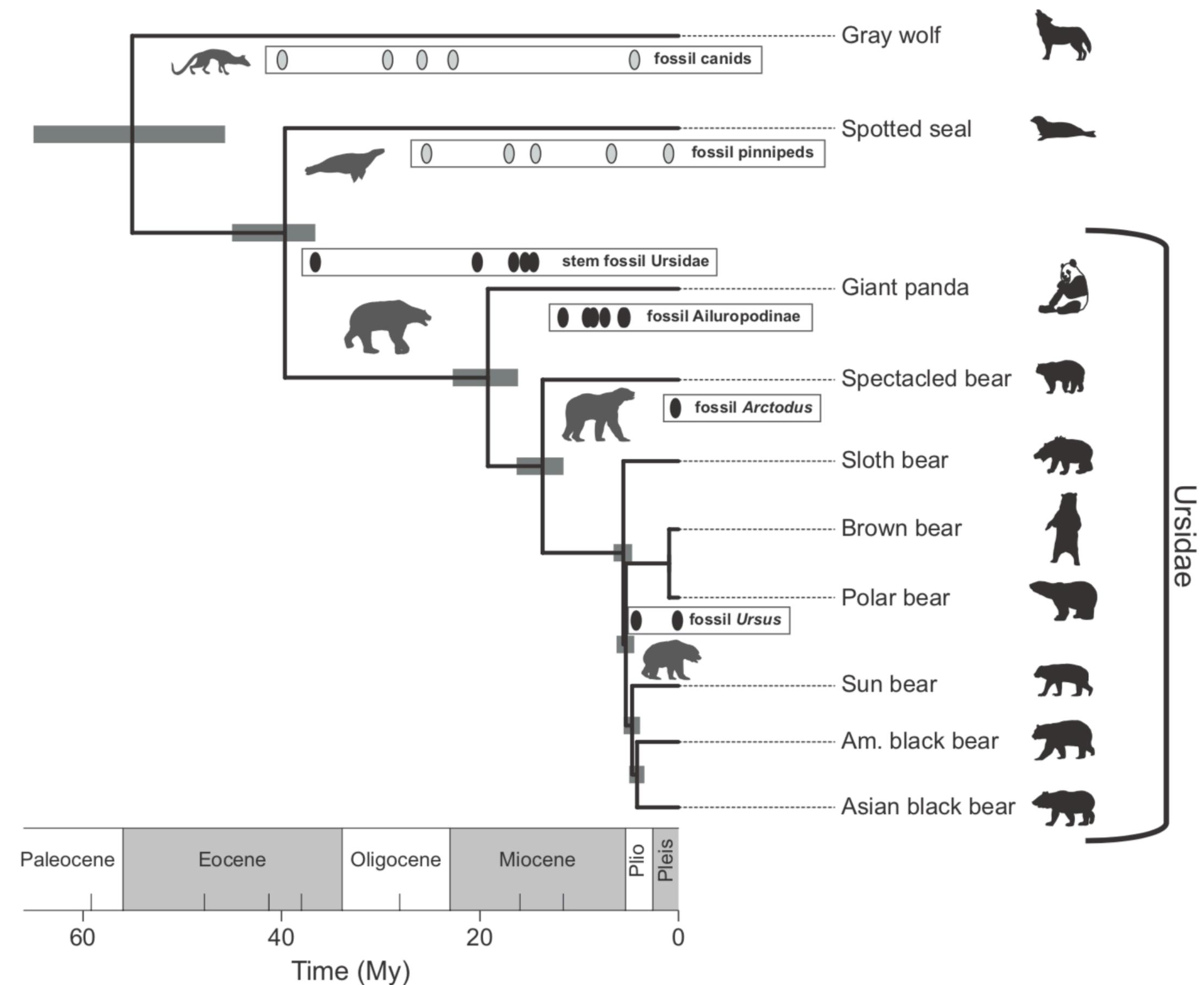
The priors will always influence the results



Exercise

Next objectives

- Recap
 - Tripartite framework
 - The fossilised birth-death process
 - Total-evidence dating
 - Phylodynamics



Bayesian divergence time estimation

Recap

We use a Bayesian framework

$$P(\text{ model } | \text{ data }) = \frac{P(\text{ data } | \text{ model }) P(\text{ model })}{P(\text{ data })}$$

likelihood

priors

posterior

marginal probability of the data

Bayesian divergence time estimation

The data

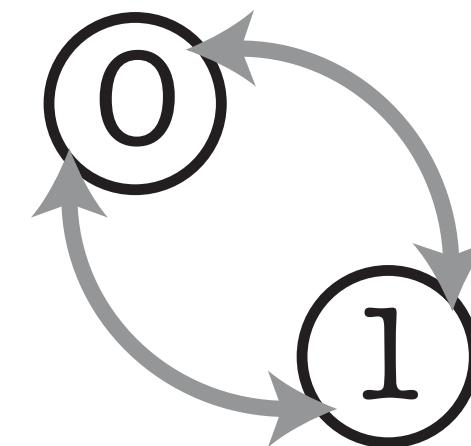
and / or
0101... ATTG...
1101... TTGC...
0100... ATTC...



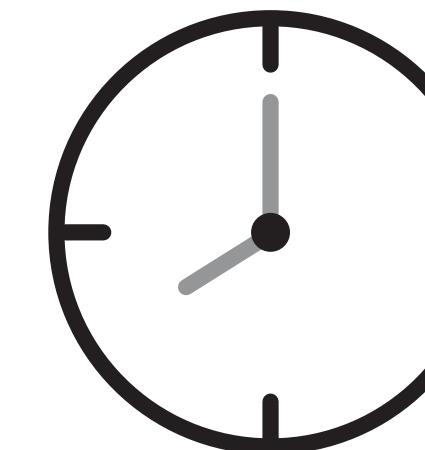
phylogenetics
characters

sample
ages

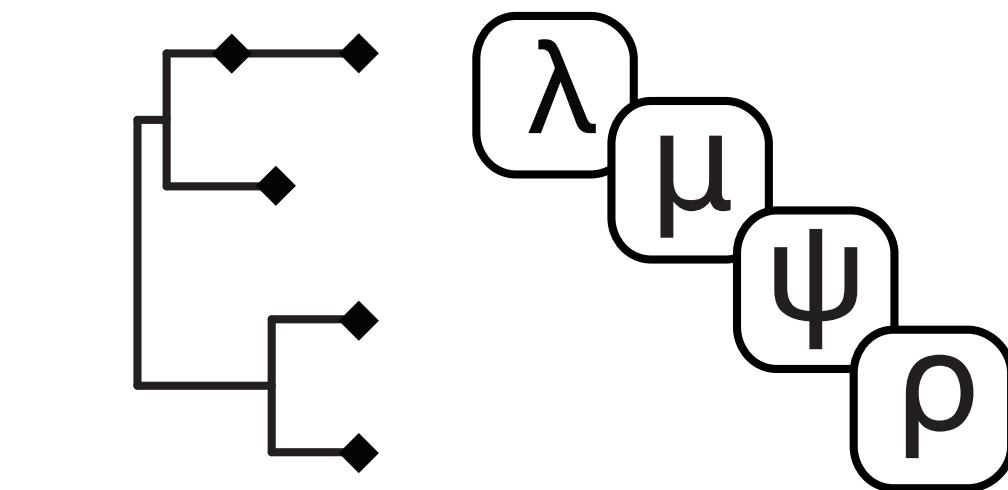
3 model components



substitution
model



clock
model

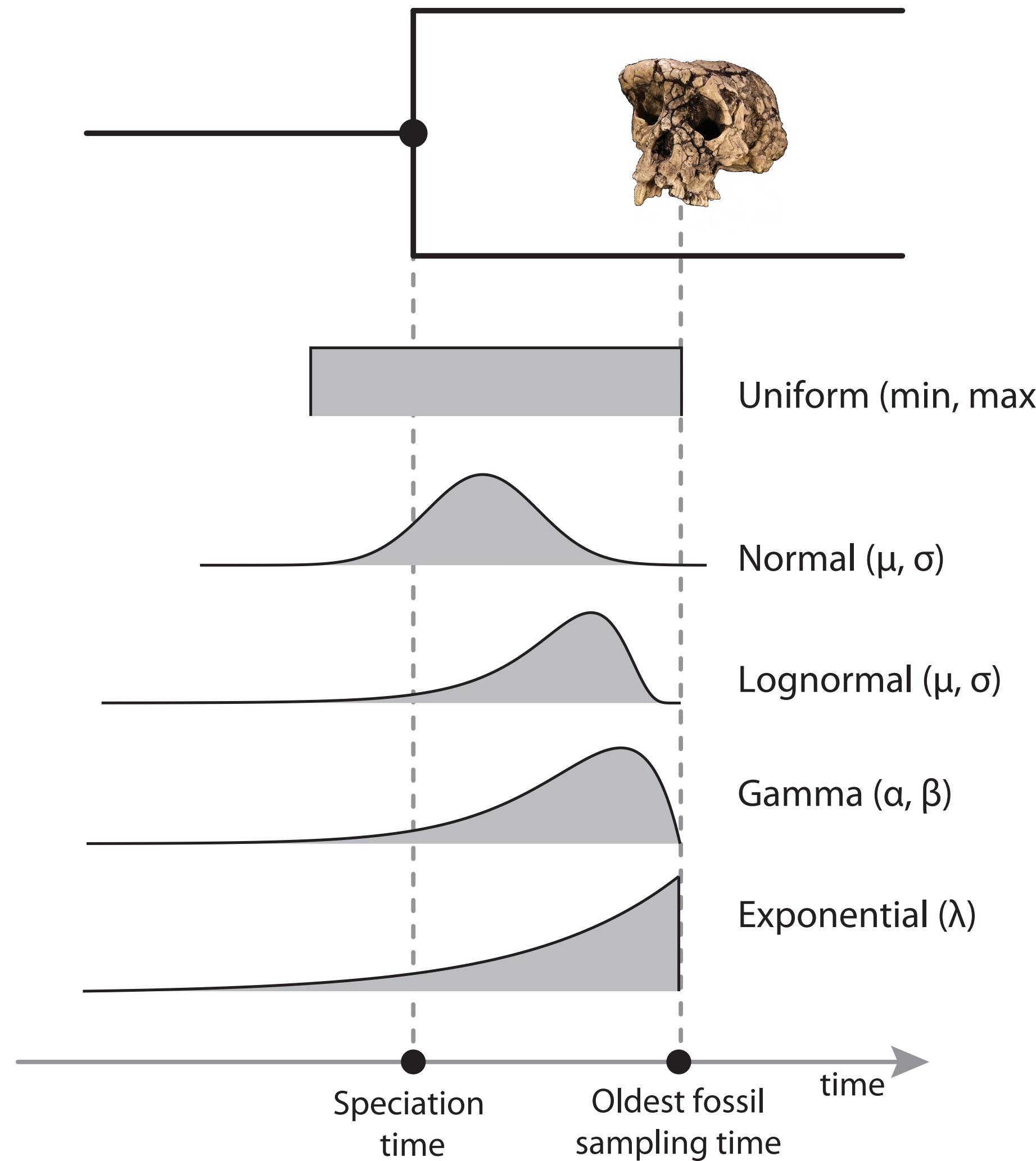


tree and tree
model

probability of the
time tree

$$P(E | \mathcal{G}^{\lambda, \mu, \psi, p})$$

Recap: Node dating



We can use a **calibration density** to constrain internal node ages

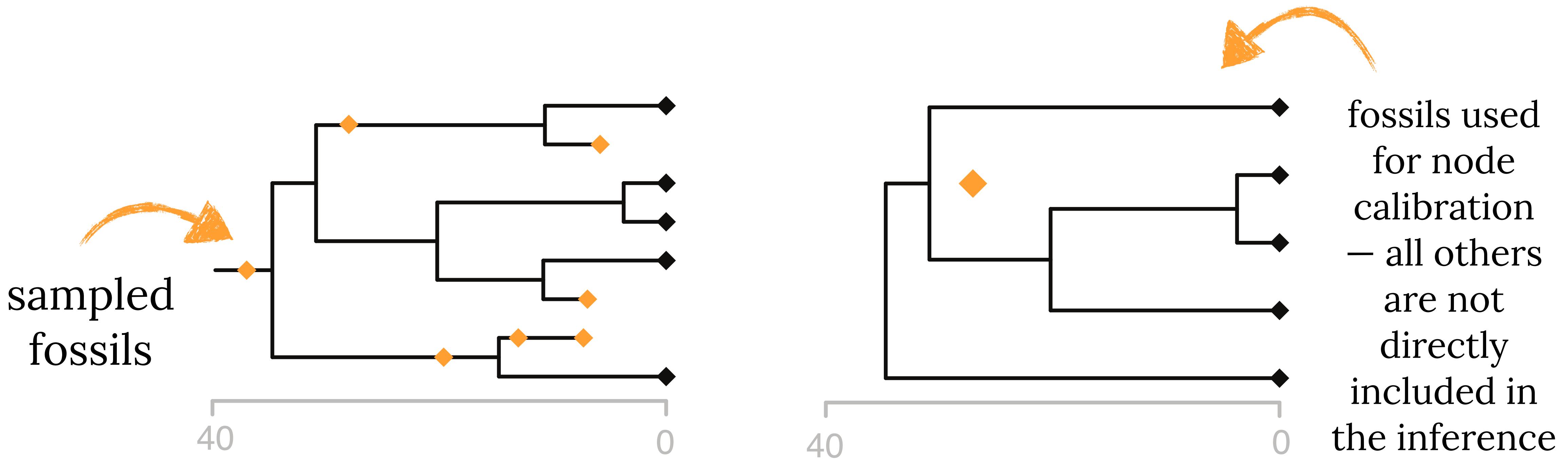
We typically use a **birth-death process** model to describe the tree generating process

Adapted from Heath (2012). Sys Bio

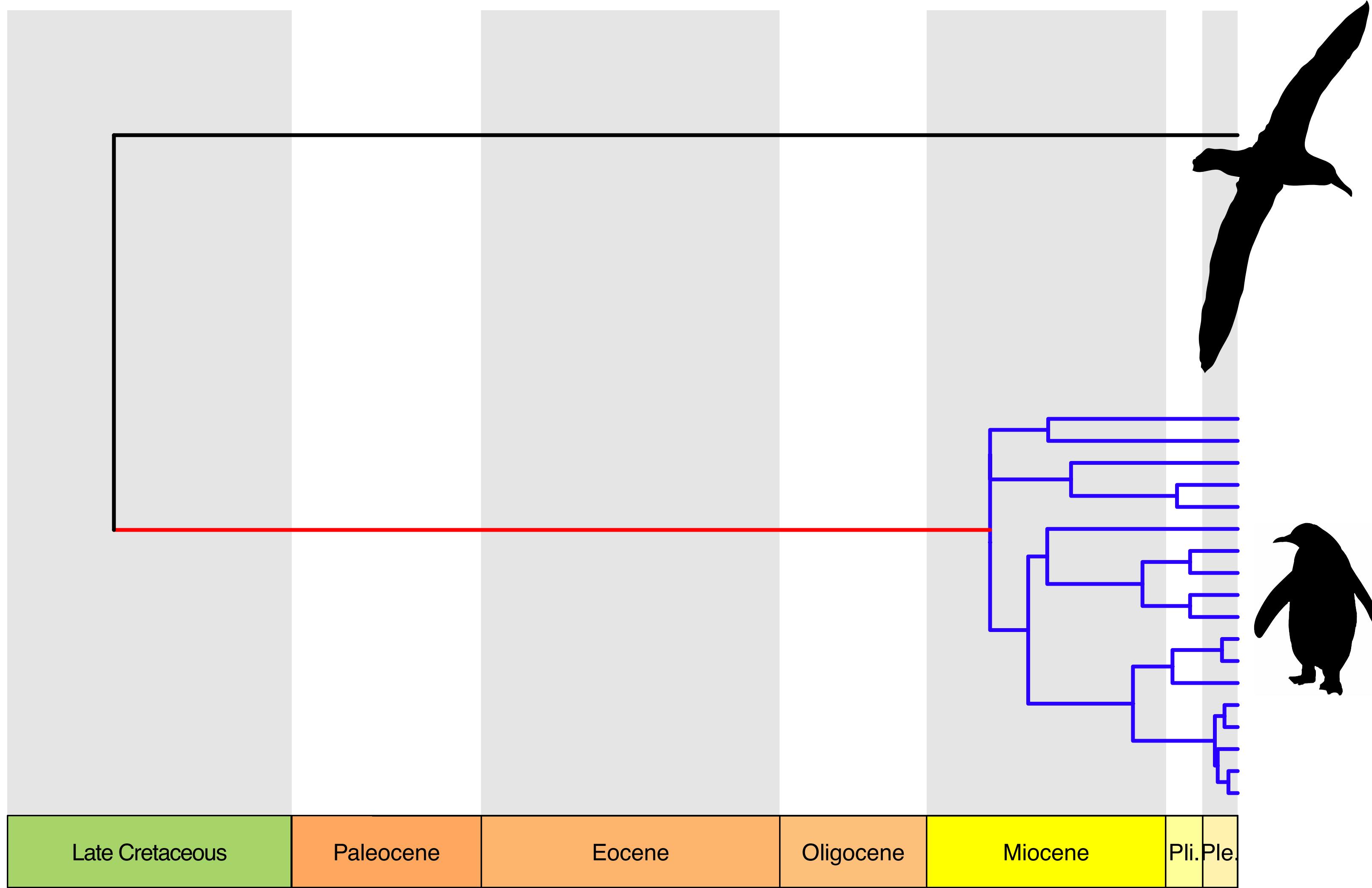
Node dating: potential issues

There are many!

A lot of information is excluded, since typically we assign one fossil per calibration node

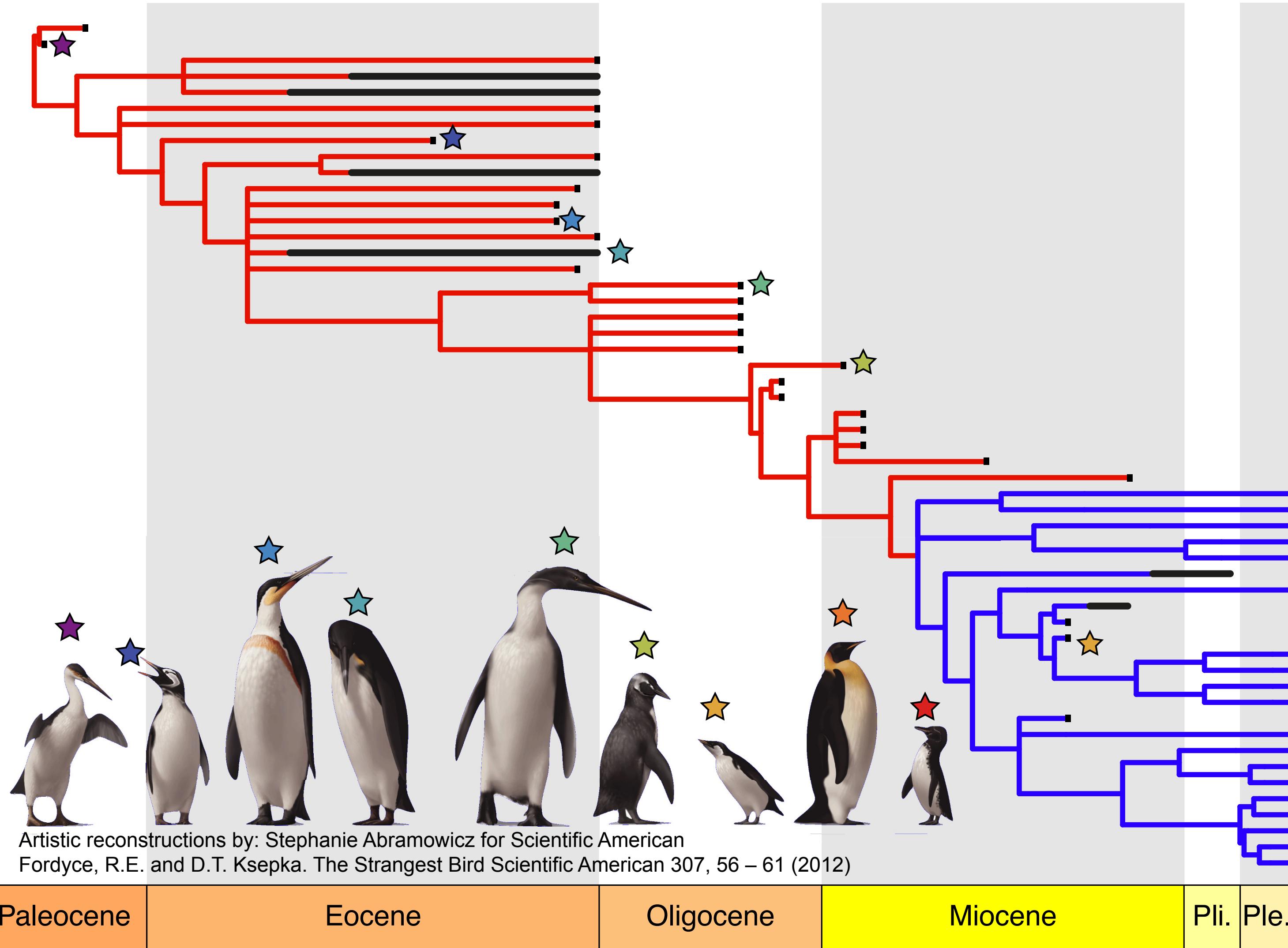


Example: living penguins



Nearest living relative is the group containing falcons - separated by ~60 Ma

Example: living penguins



But penguins
have a rich
fossil record!

Node dating: potential issues

The model doesn't describe the process that generated the fossil sampling times, meaning the model is **statistically incoherent**

The calibration priors are difficult to specify objectively and can have a massive impact on the divergence times. They can also interact with each other and / or the birth-death process prior in unintuitive ways

Some references on issues with specified vs effective priors

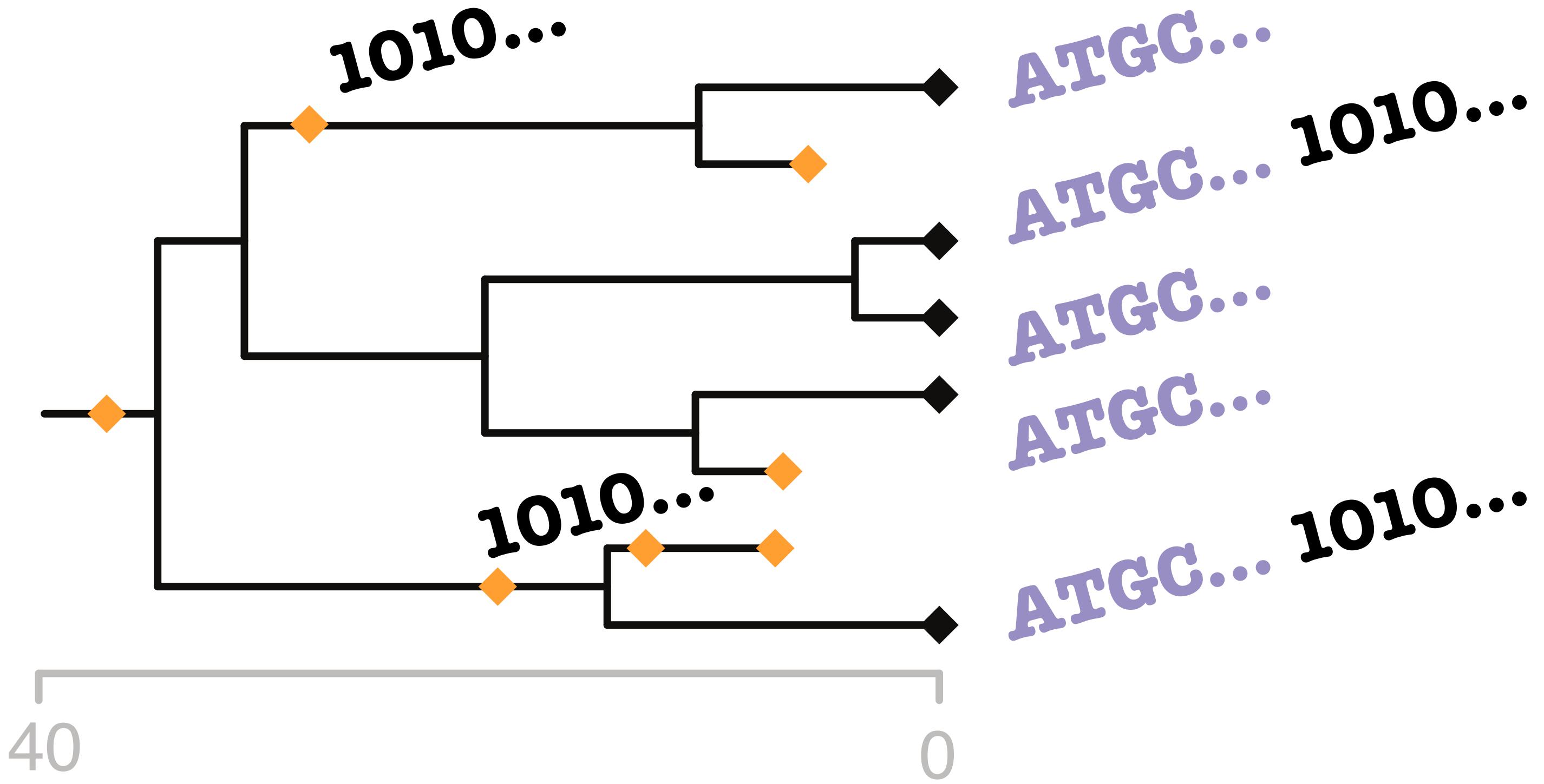
Yang and Rannala. [2006](#). MBE

Heled and Drummond. [2012](#). Sys Bio

Warnock et al. [2012, 2015](#)

Total-evidence dating

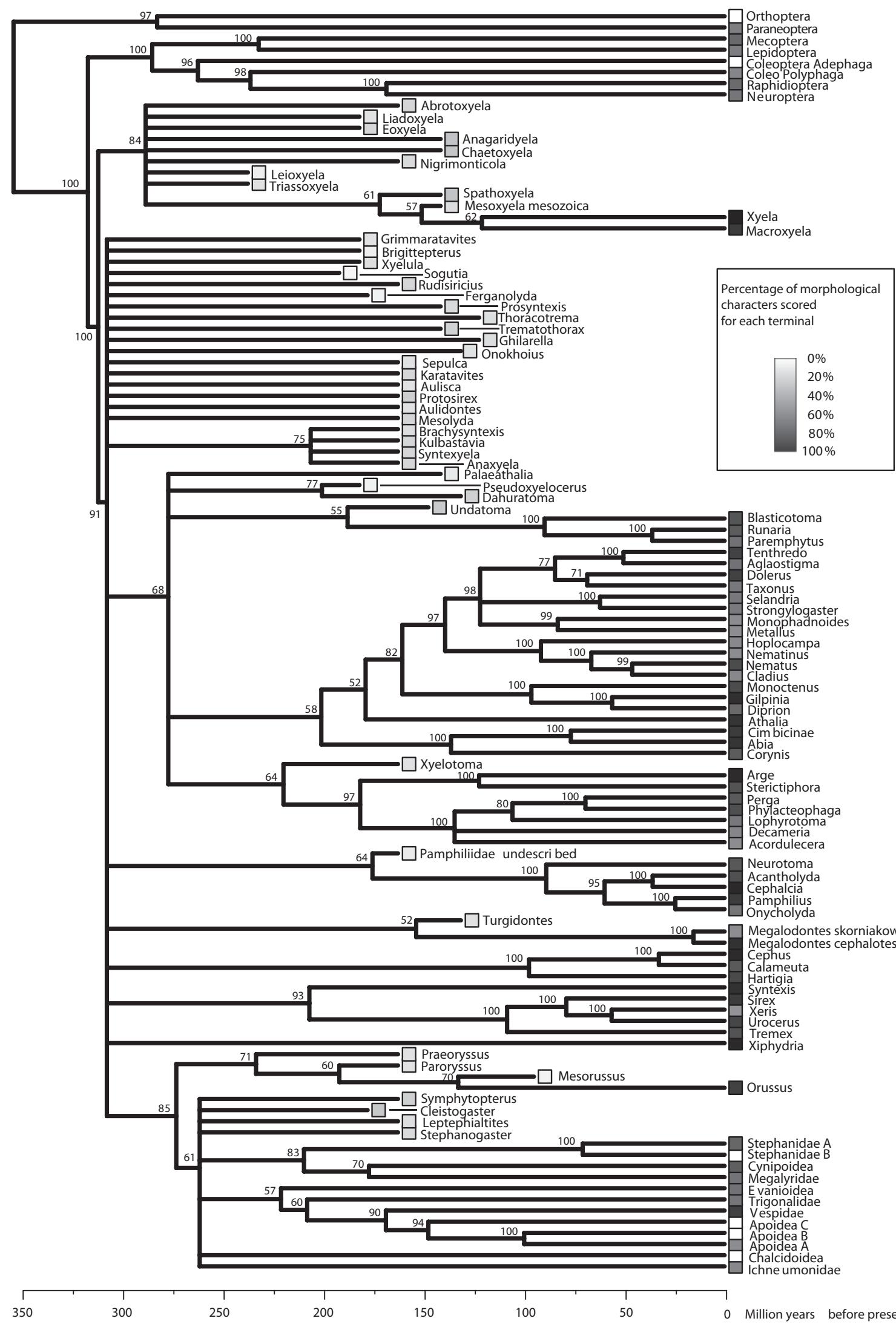
Tip-dating or “total-evidence” dating



We have DNA for living species. We have morphology for living *and* fossil species

Fossils can be positioned on the basis of morphology
→ accounts for uncertainty in fossil placement

The uniform tree prior

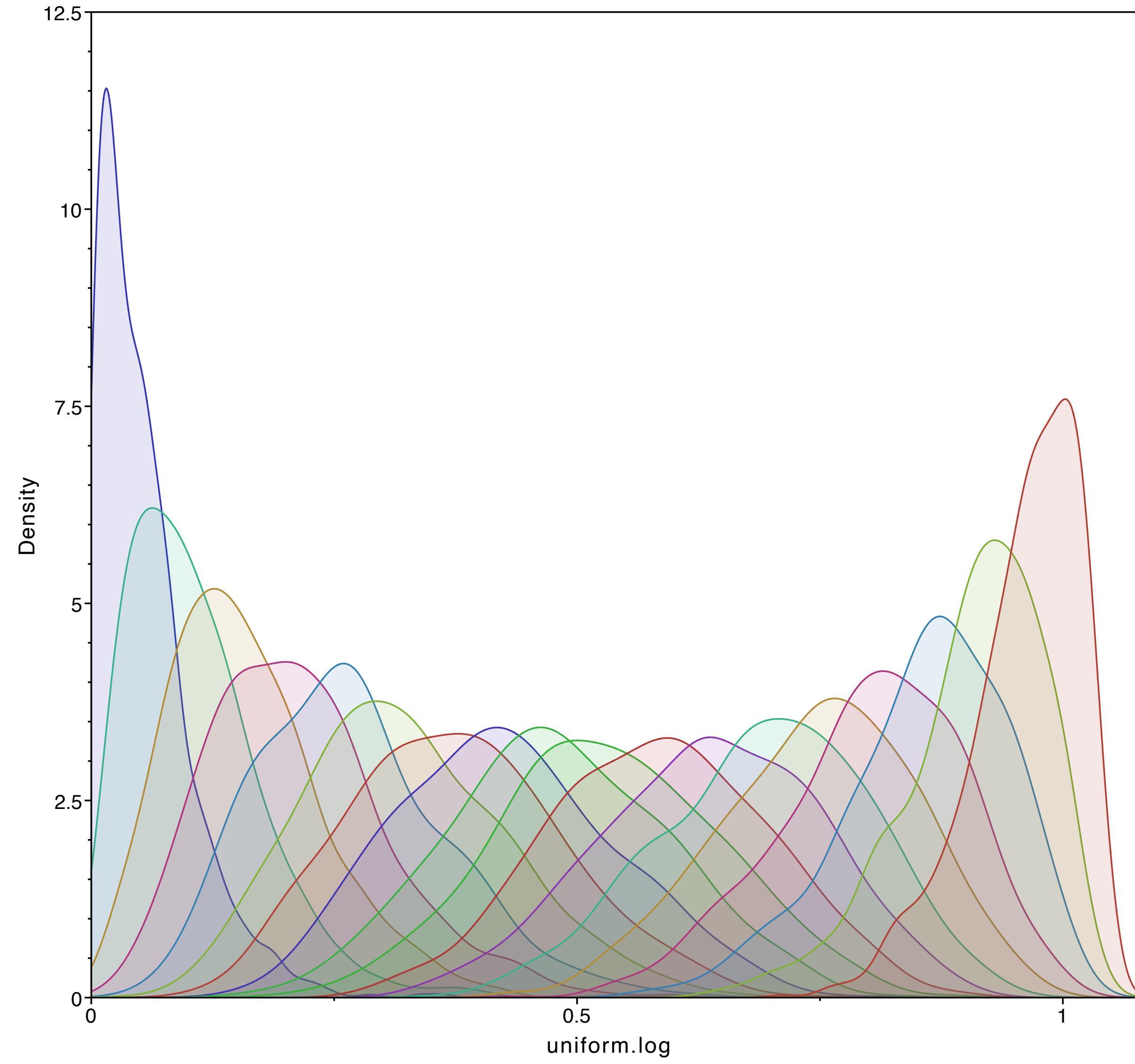


Dated tree of Hymenoptera



The uniform tree prior assumes all trees and branch lengths are equally likely within the bounds of the fossil ages (+ a max upper bound)

It does not explicitly account for the fossil sampling process



A uniform tree prior implies time till the next split is independent of how many lineages there are present

This is in contrast to birth-death processes, where more lineages mean a higher chance of observing a split in one of these lineages

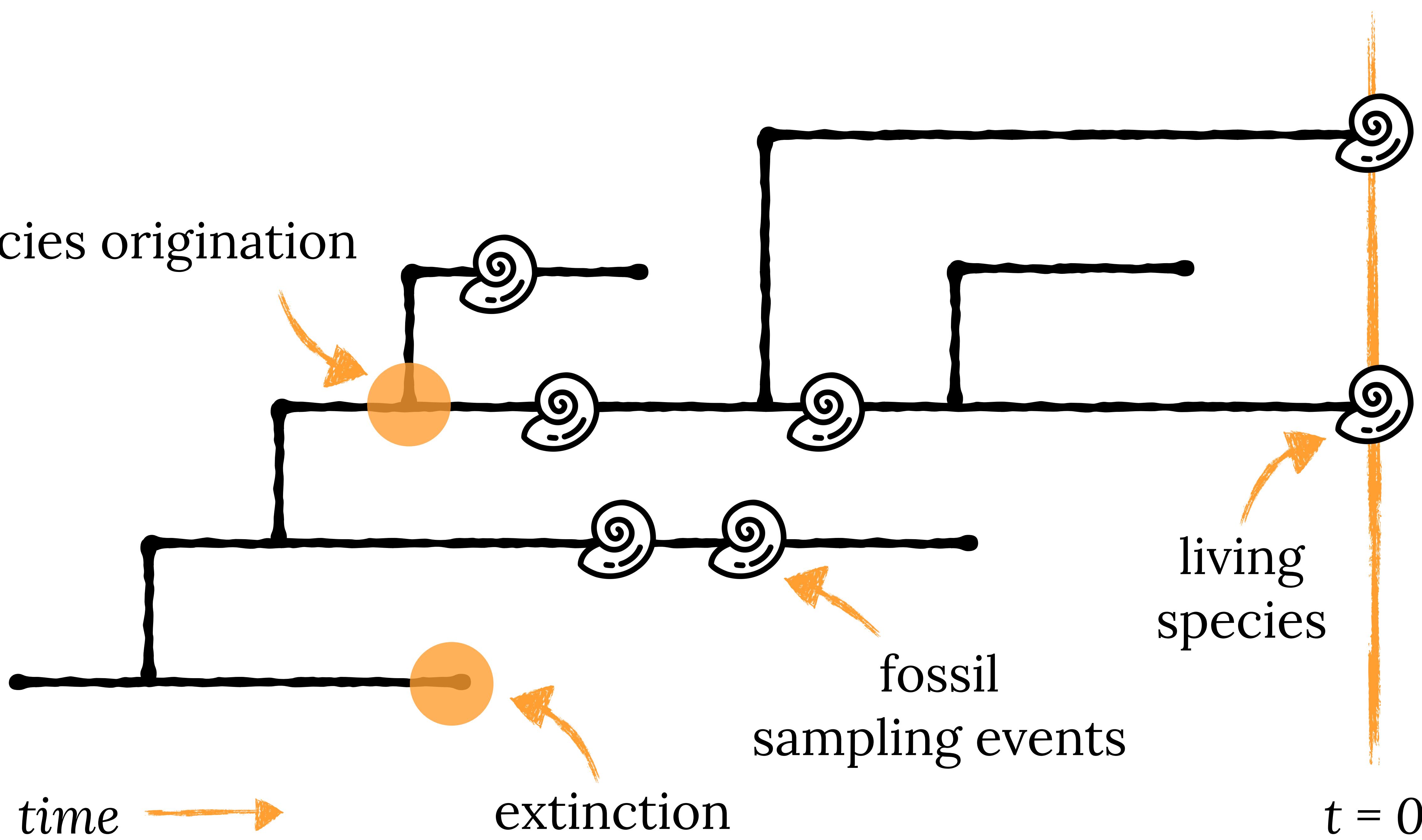
[Uniform tree priors - why not use them?](#)

Remco Bouckaert

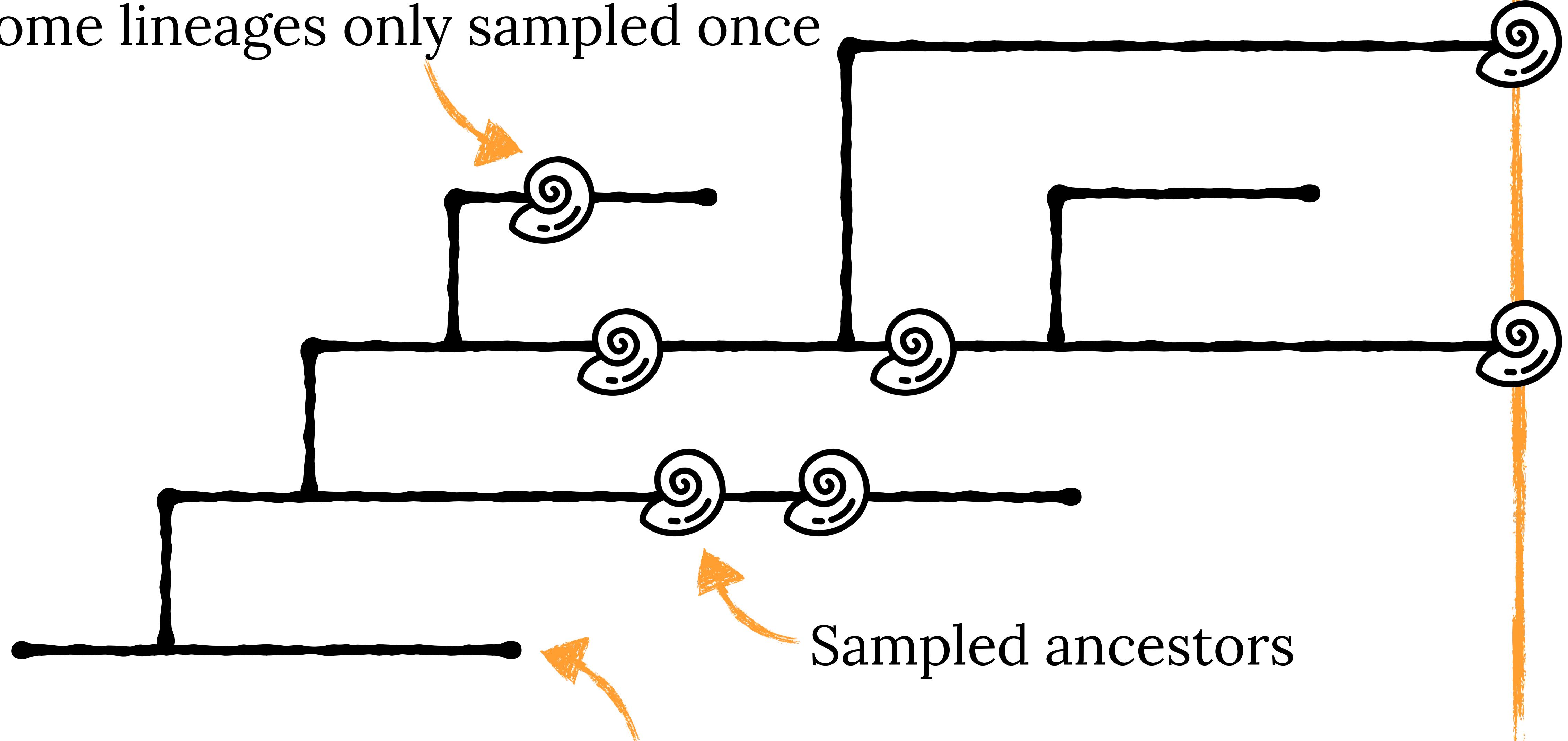
What does a generating prior for the fossil record look like?

The fossilised birth-death process

species origination



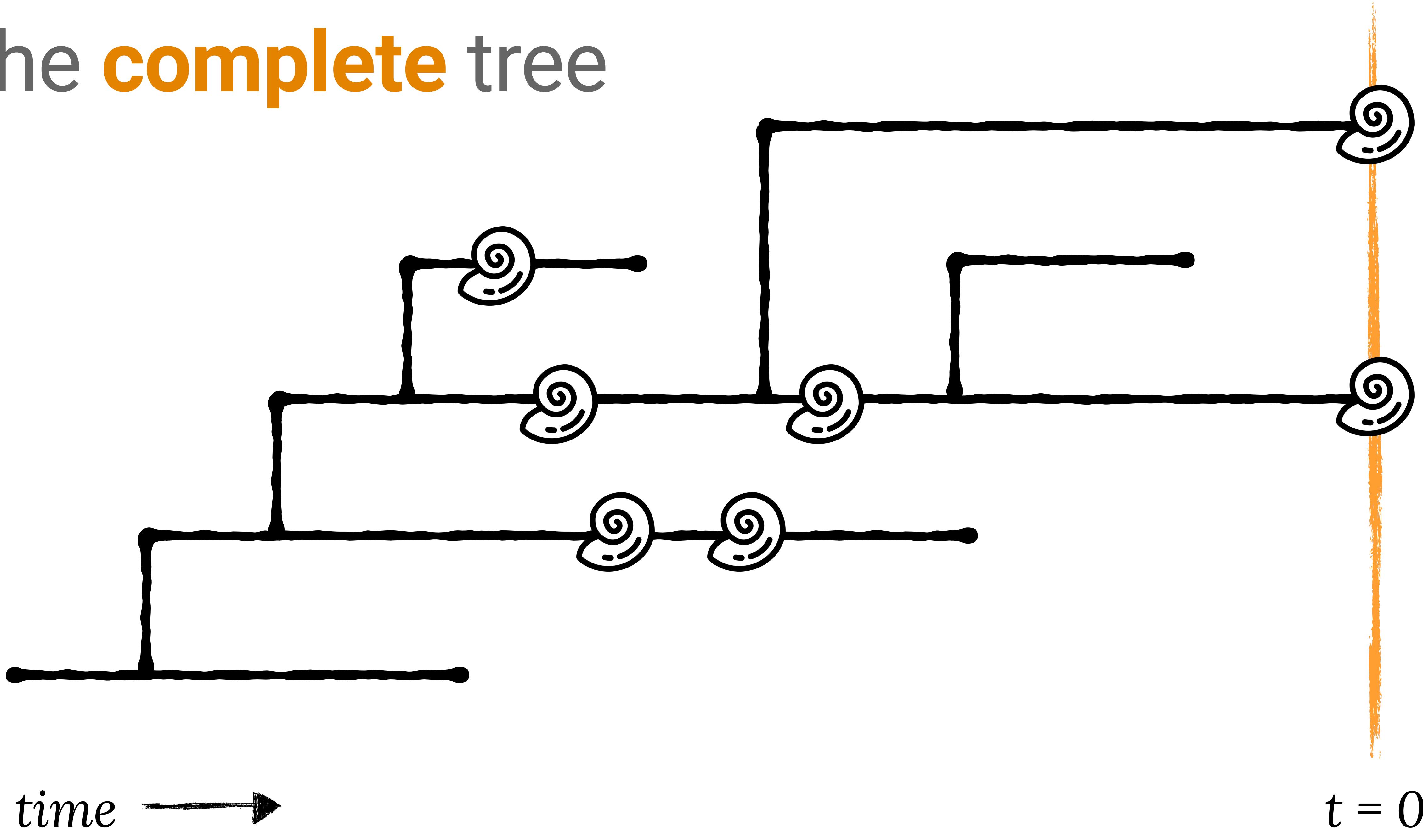
Some lineages only sampled once



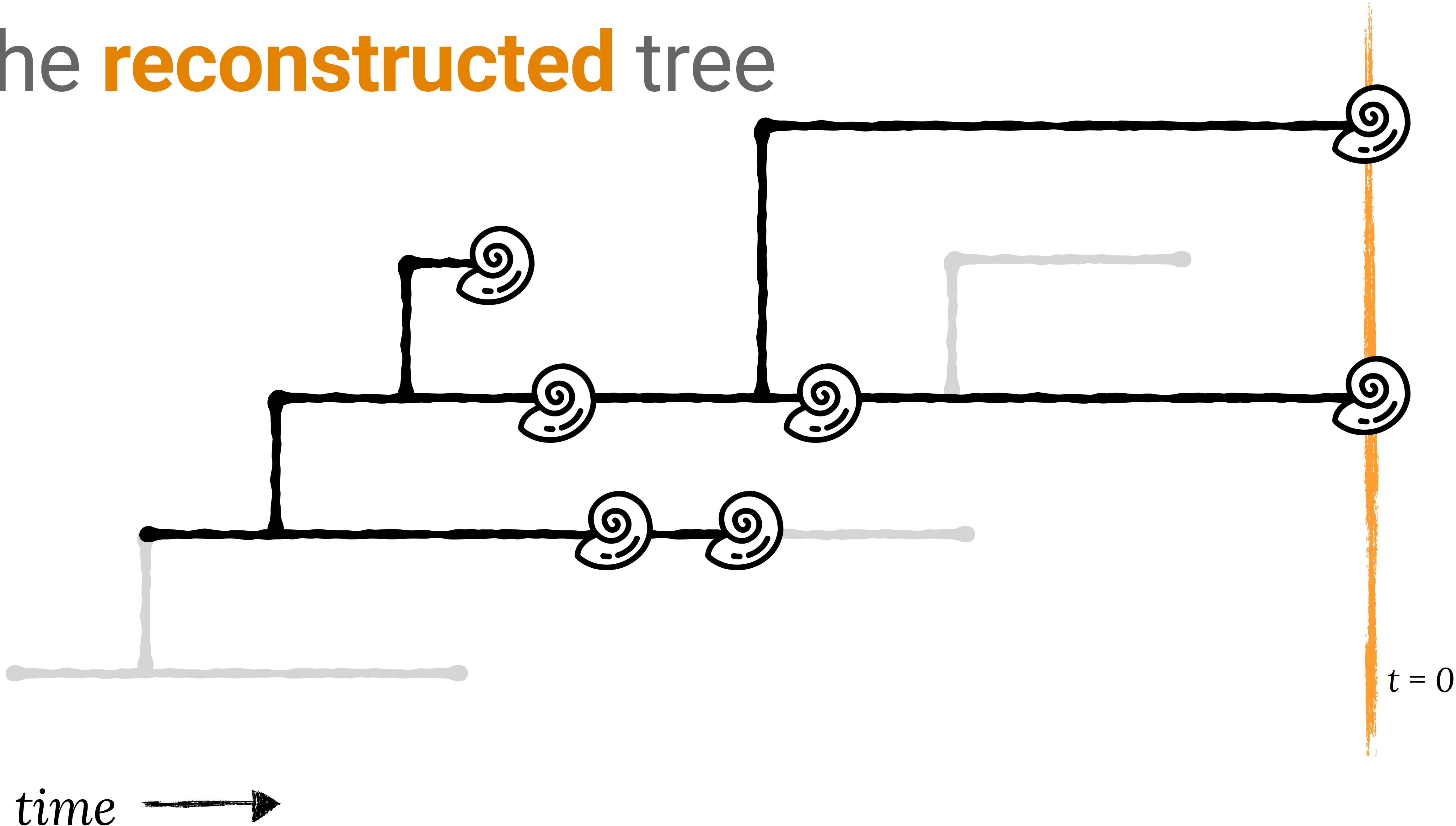
Some lineages go completely unsampled

$t = 0$

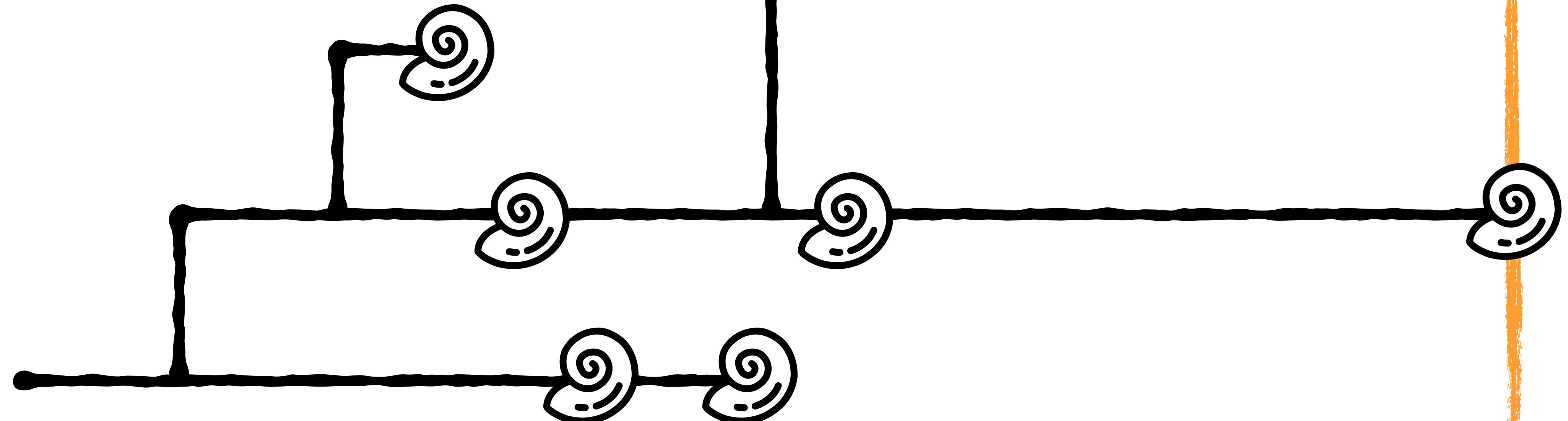
The complete tree



The reconstructed tree



The **fossilised birth-death (FBD) process** allows us to calculate the probability of observing the reconstructed tree

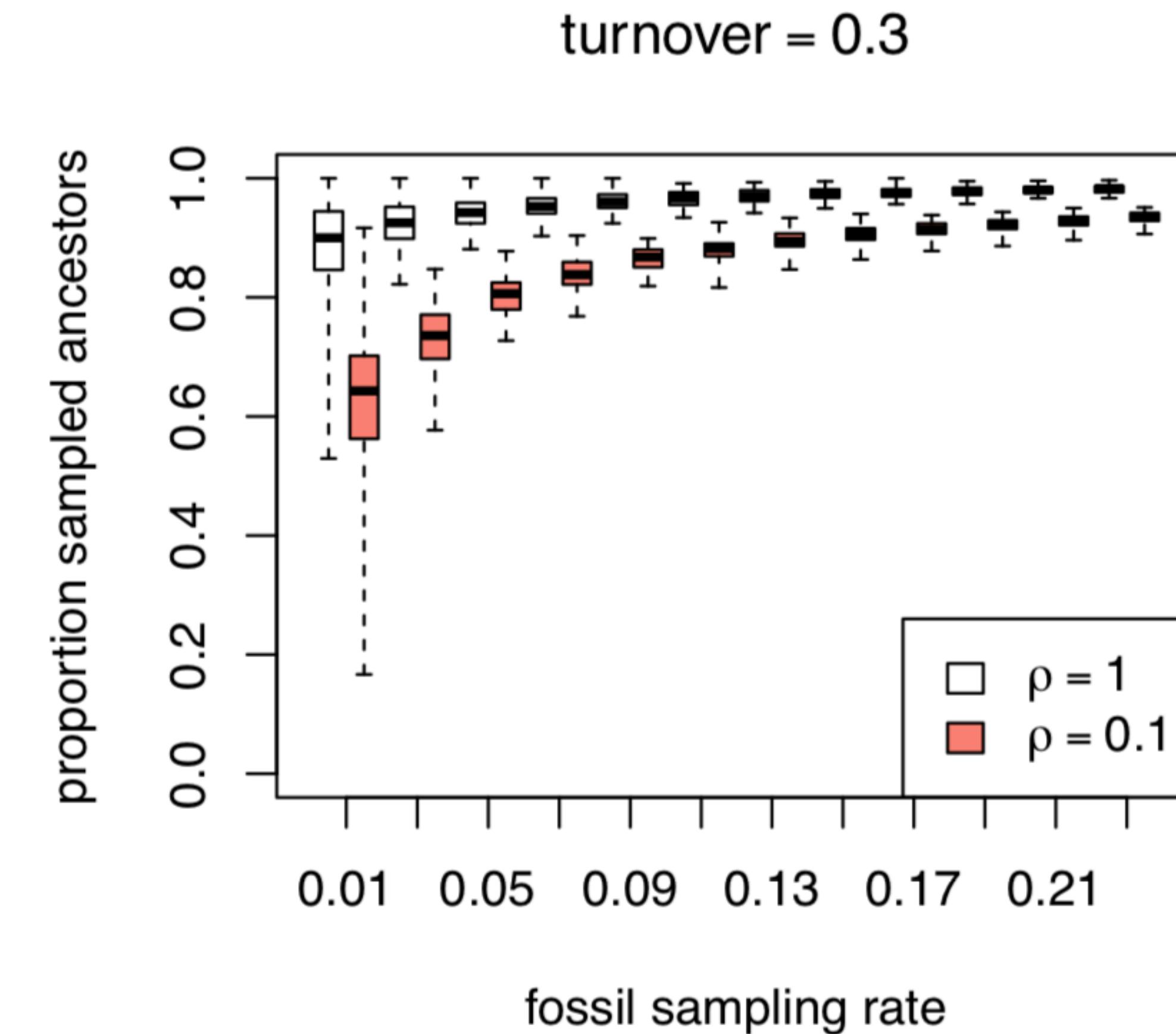
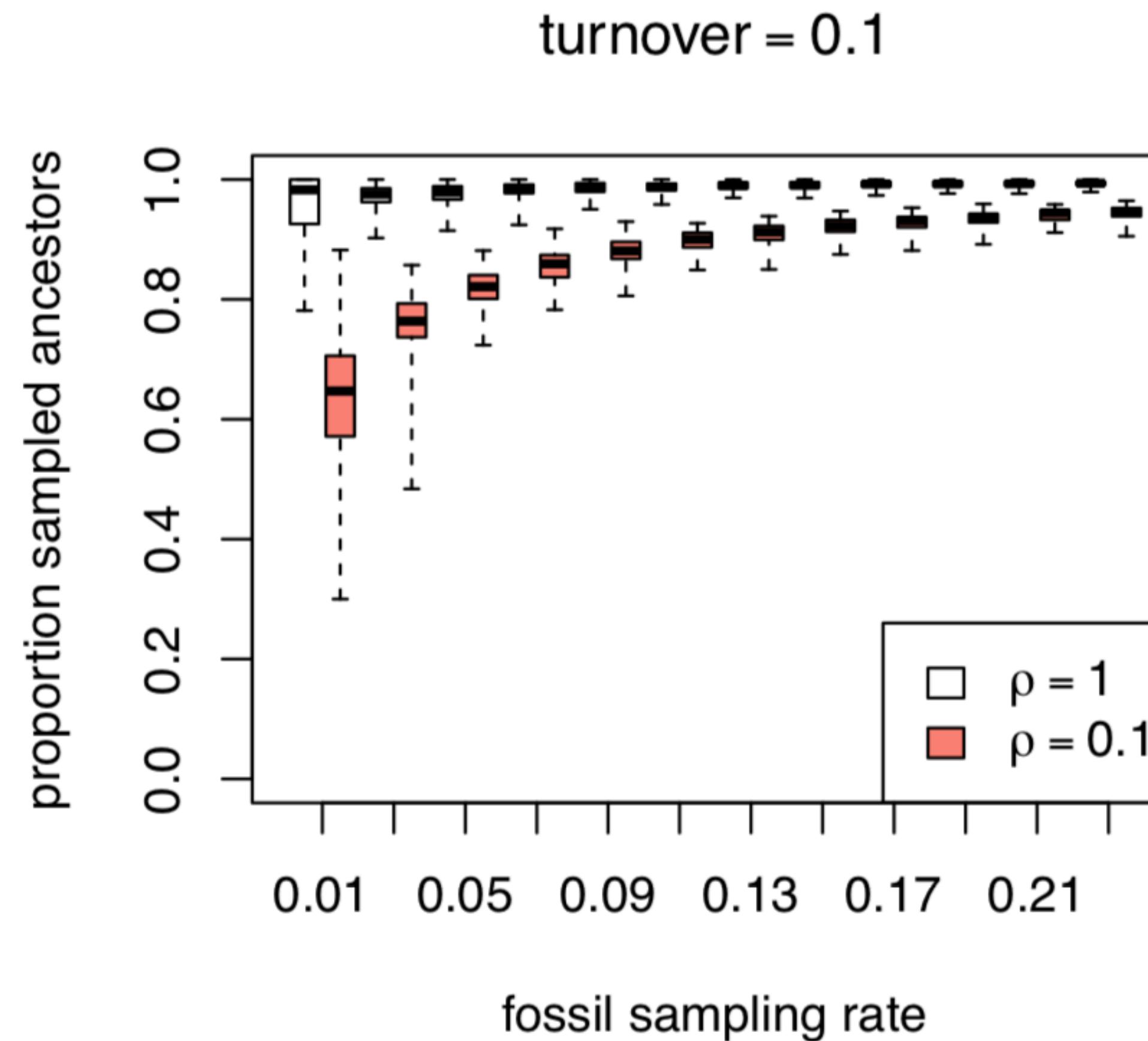


$$P(E | \text{snail}, \lambda, \mu, \psi, \rho)$$

Sampling-through-time in birth-death trees. Stadler. (2010)
First implemented: Heath et al. (2014) and Gavryushkina et al. (2014)

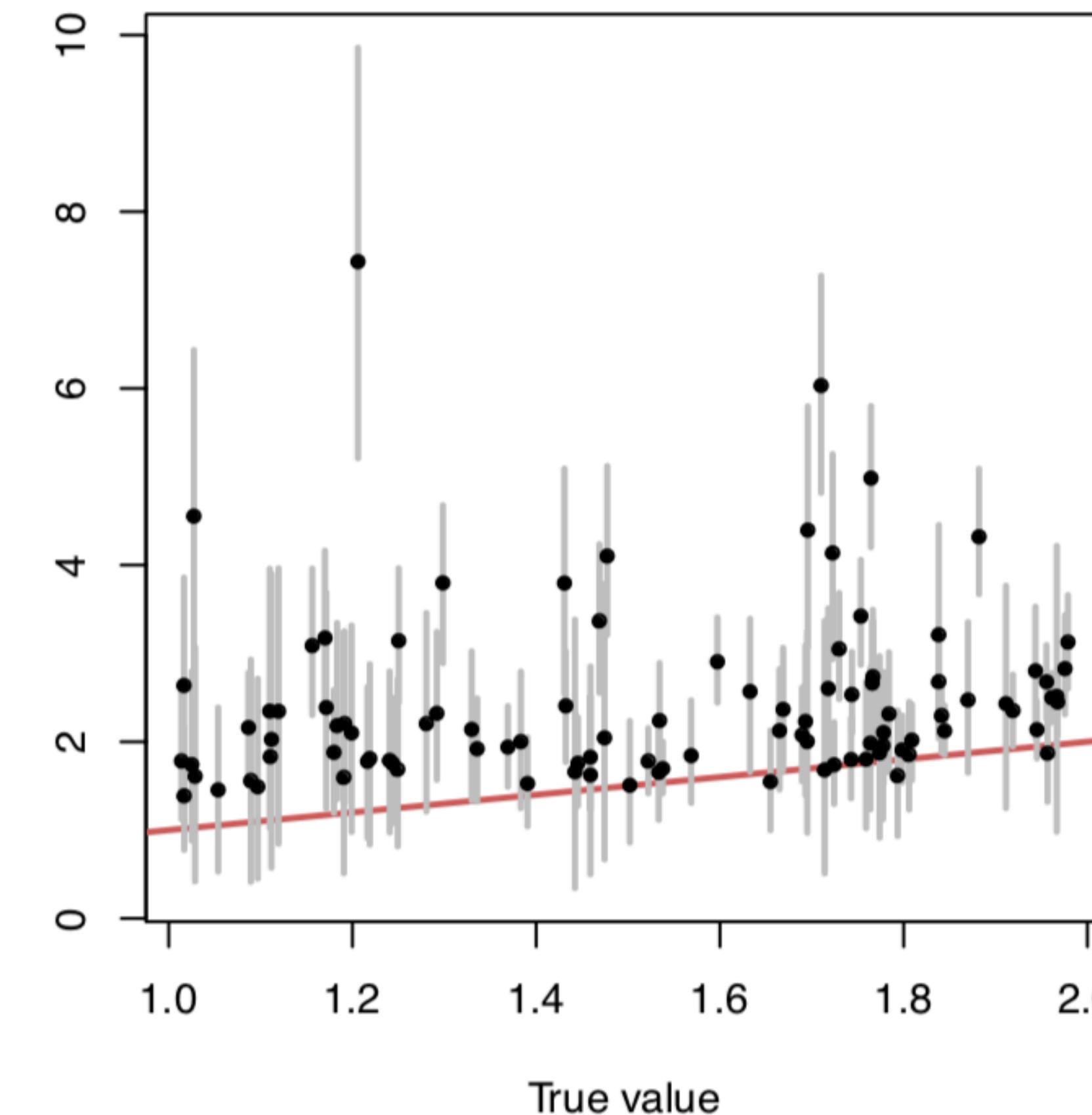
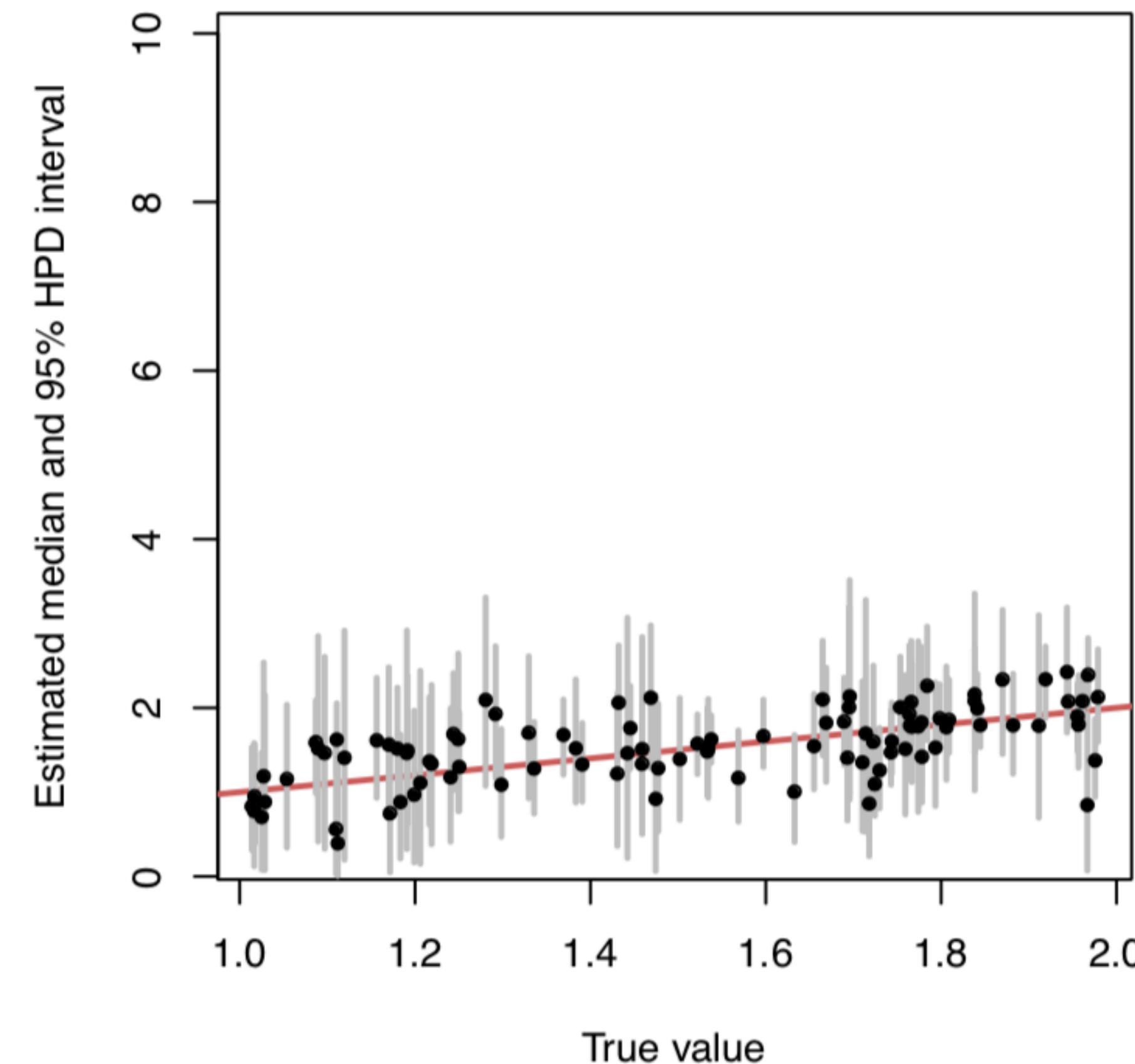
Sampled ancestors

The proportion increases with higher turnover (birth - death) or higher sampling

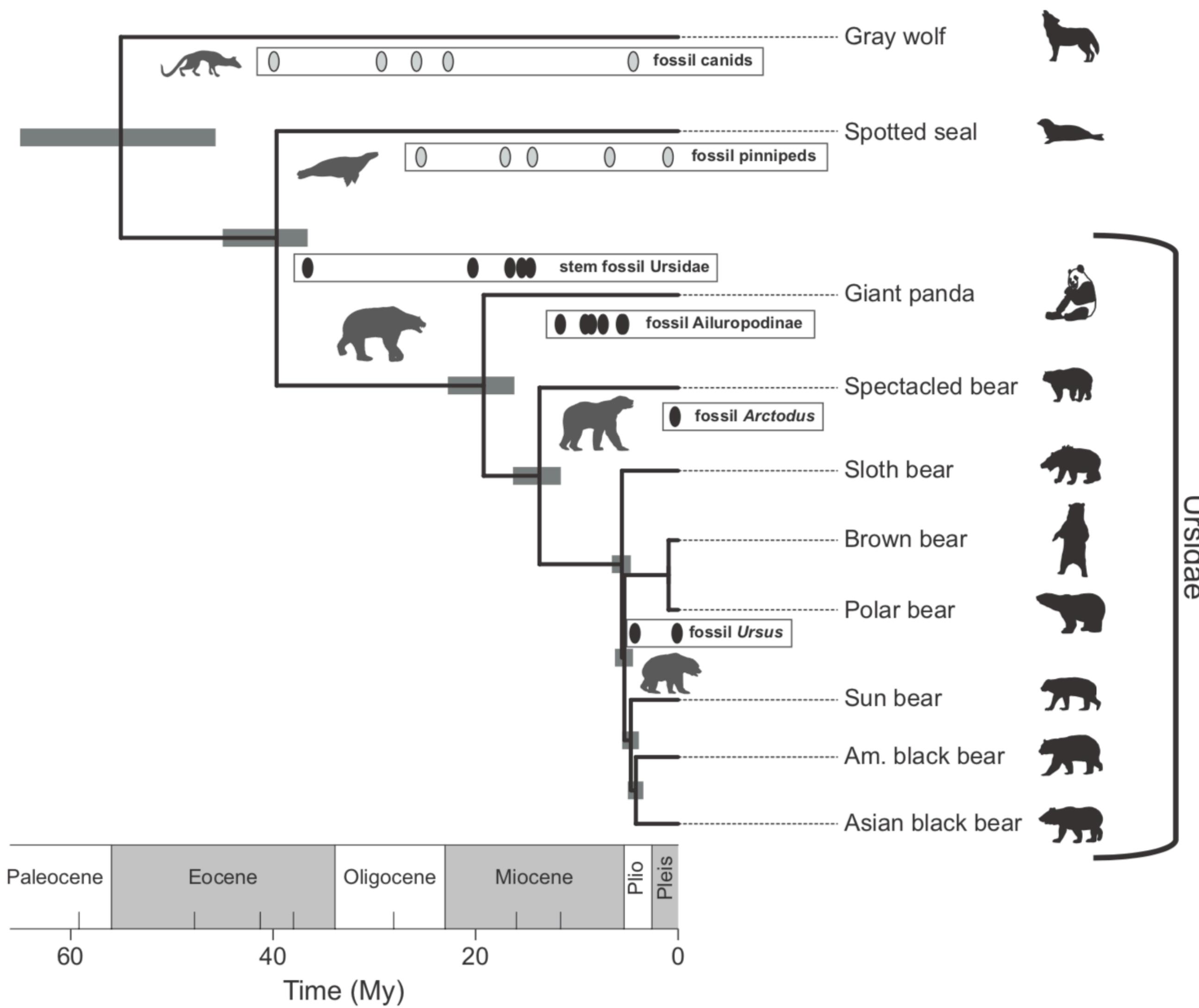


Sampled ancestors

Ignoring sampled ancestors can lead to inaccurate parameter estimates



Time calibrated tree of living and fossil bears



First application of the FBD model.

Fossils are incorporated via constraints, not character data. Their precise placement can not be inferred, but this uncertainty will be reflected in the posterior

Exercise

Fossils can be incorporated via **taxonomy** or **character data** (total-evidence)

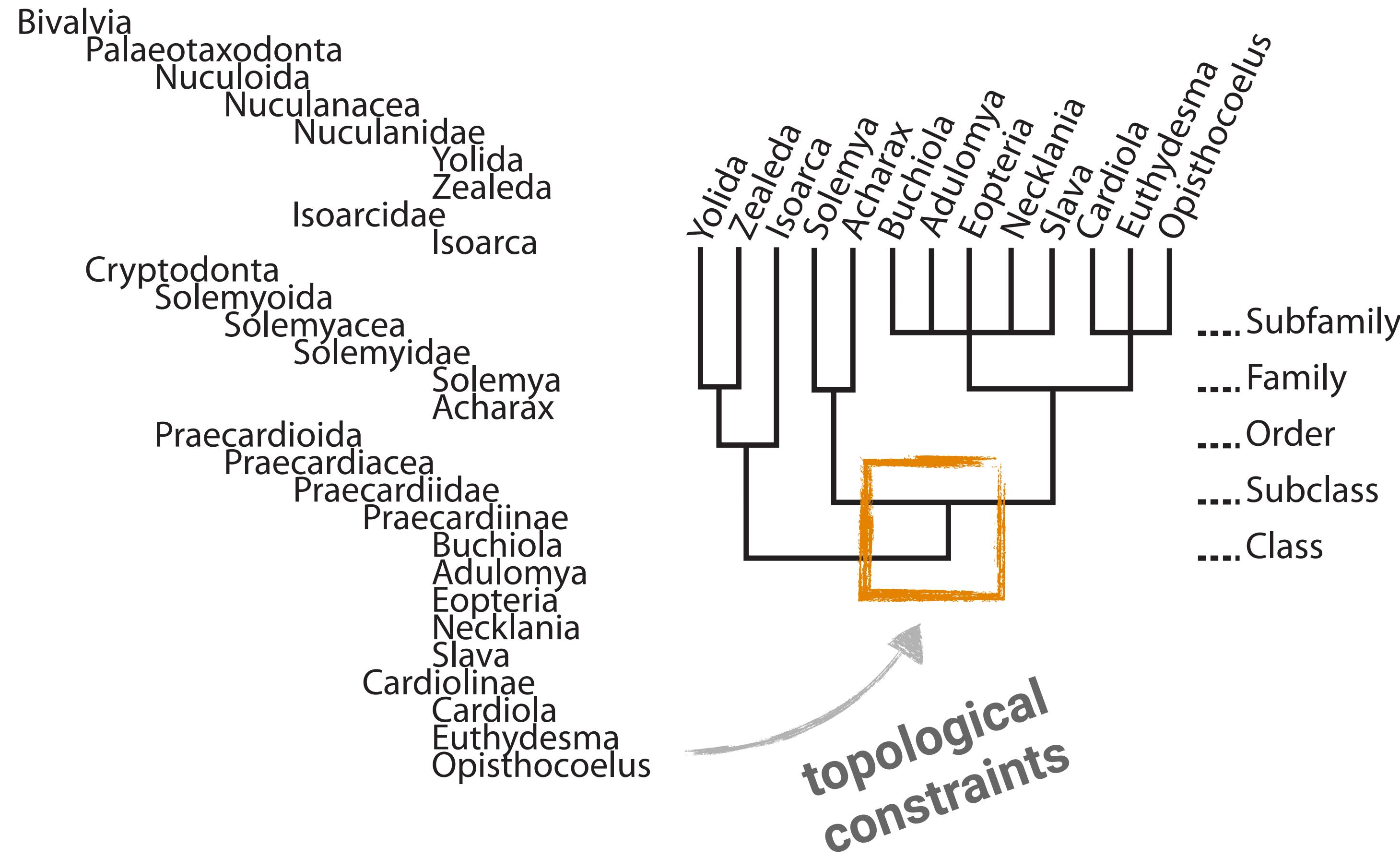
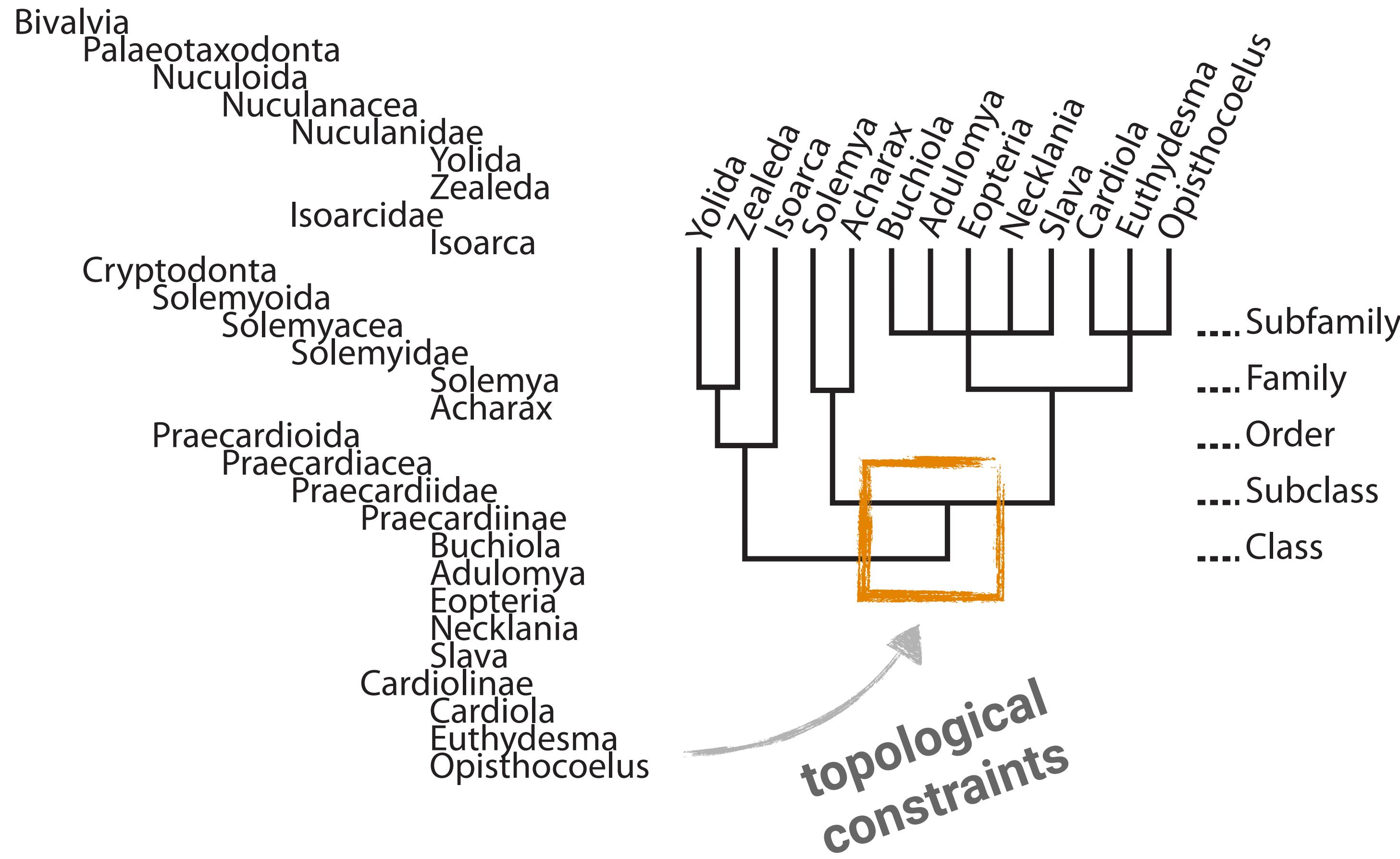


Image source Soul & Friedman (2015)

Fossils can be incorporated via **taxonomy** or **character data** (total-evidence)



ATAT...

TCACT...

?????...

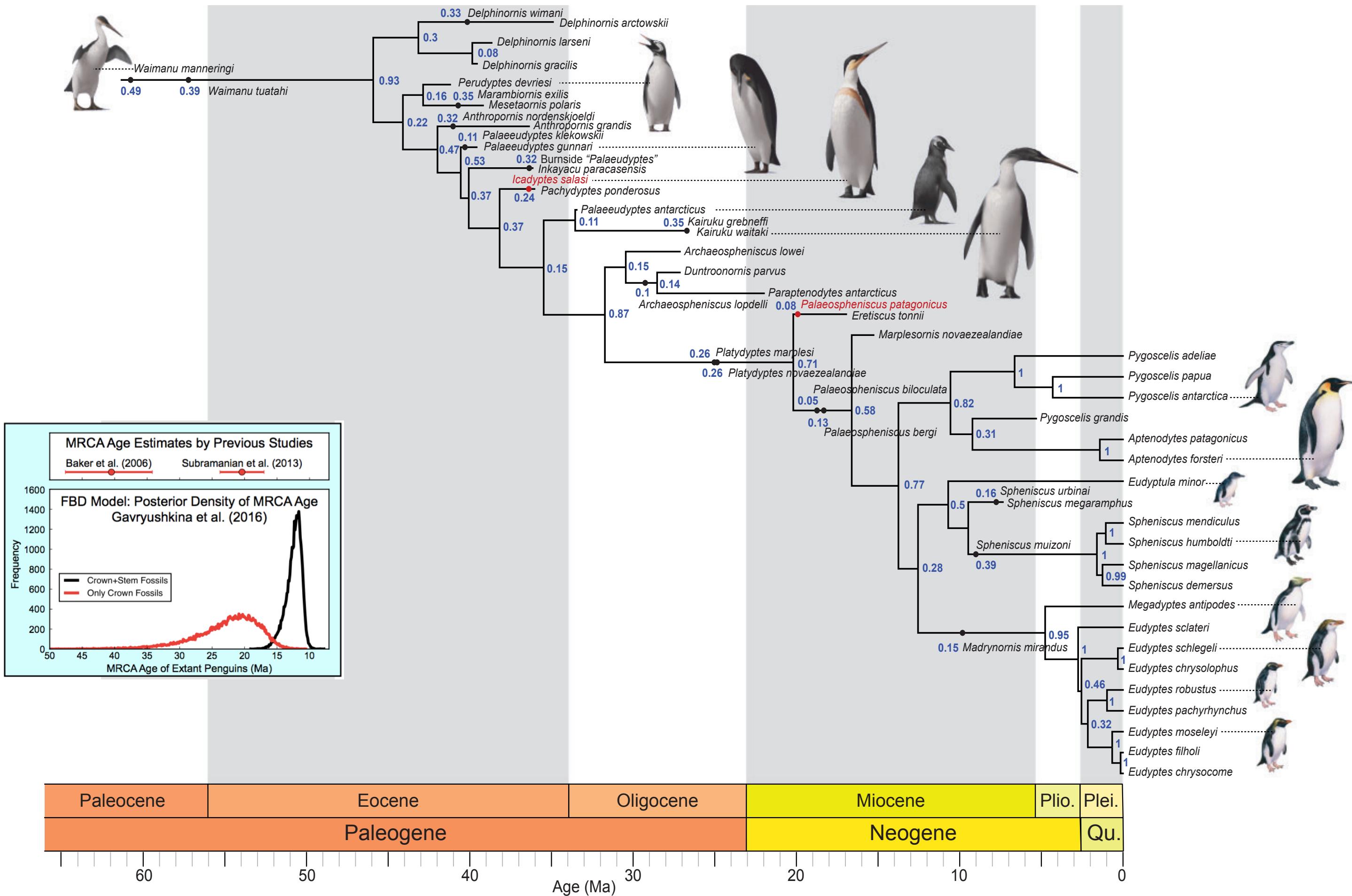
OR

1001...

1101...

0100...

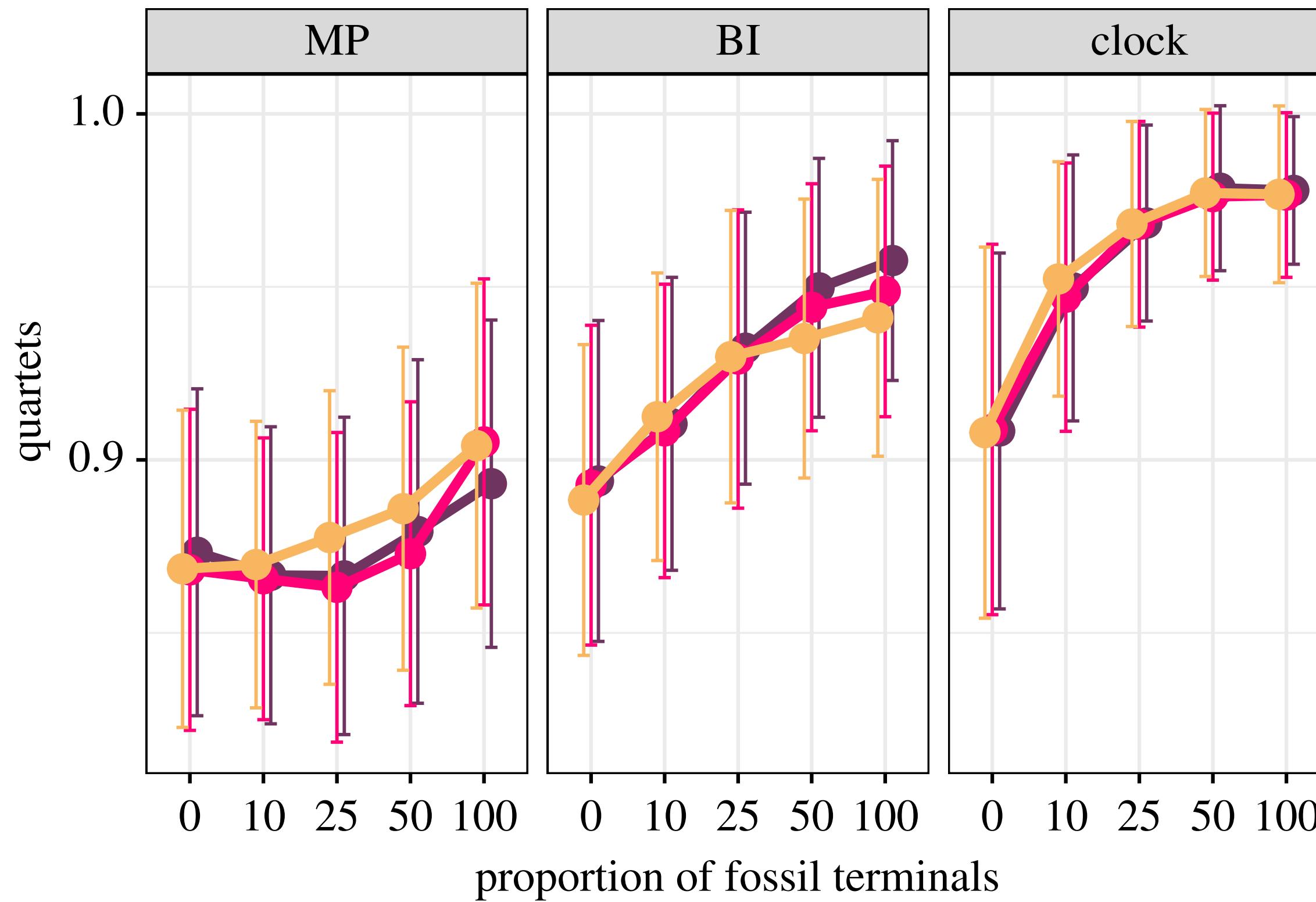
Time calibrated tree of living and fossil penguins



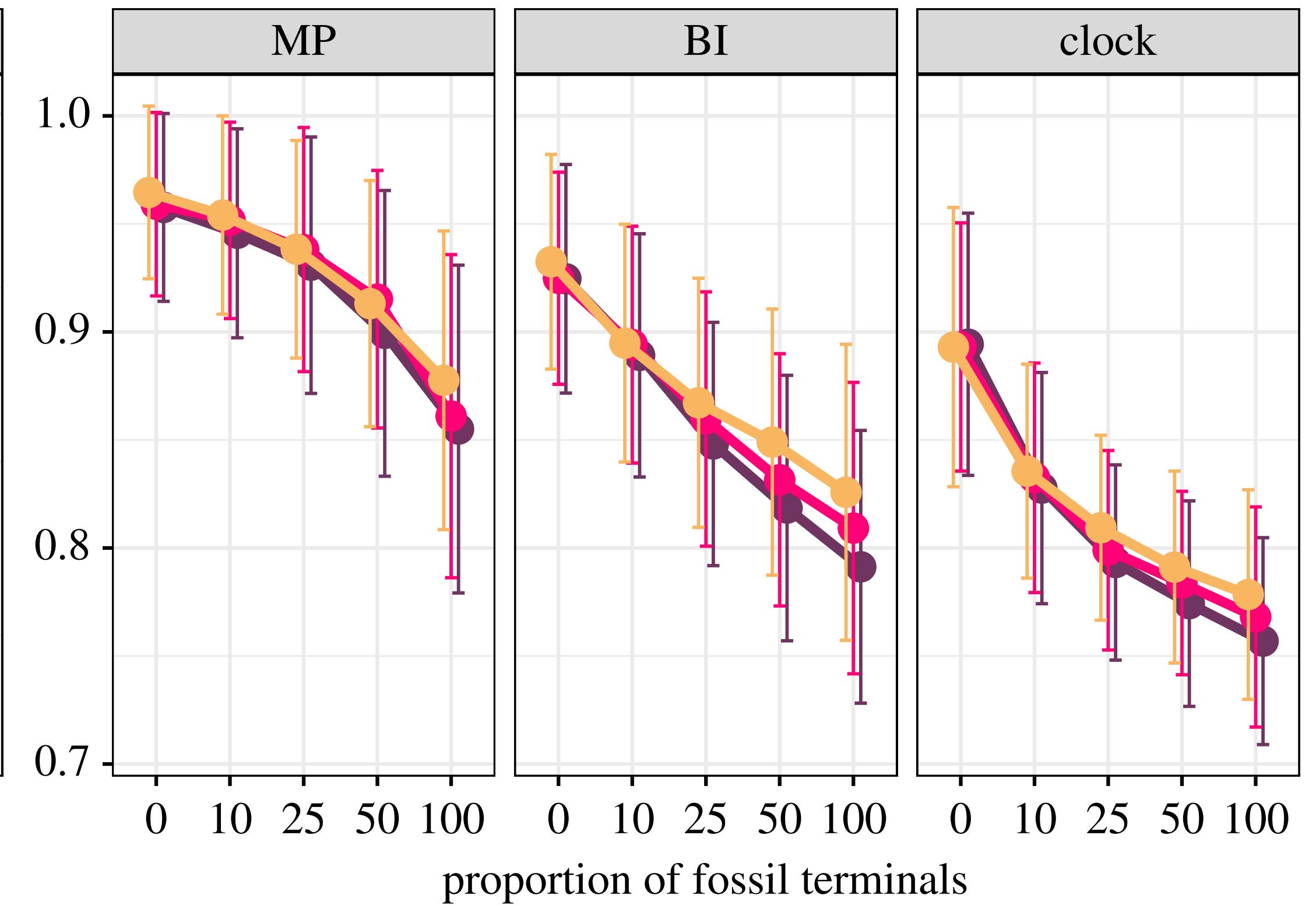
First application of total evidence dating using the FBD model

Fossils are incorporated using character data

accuracy



precision



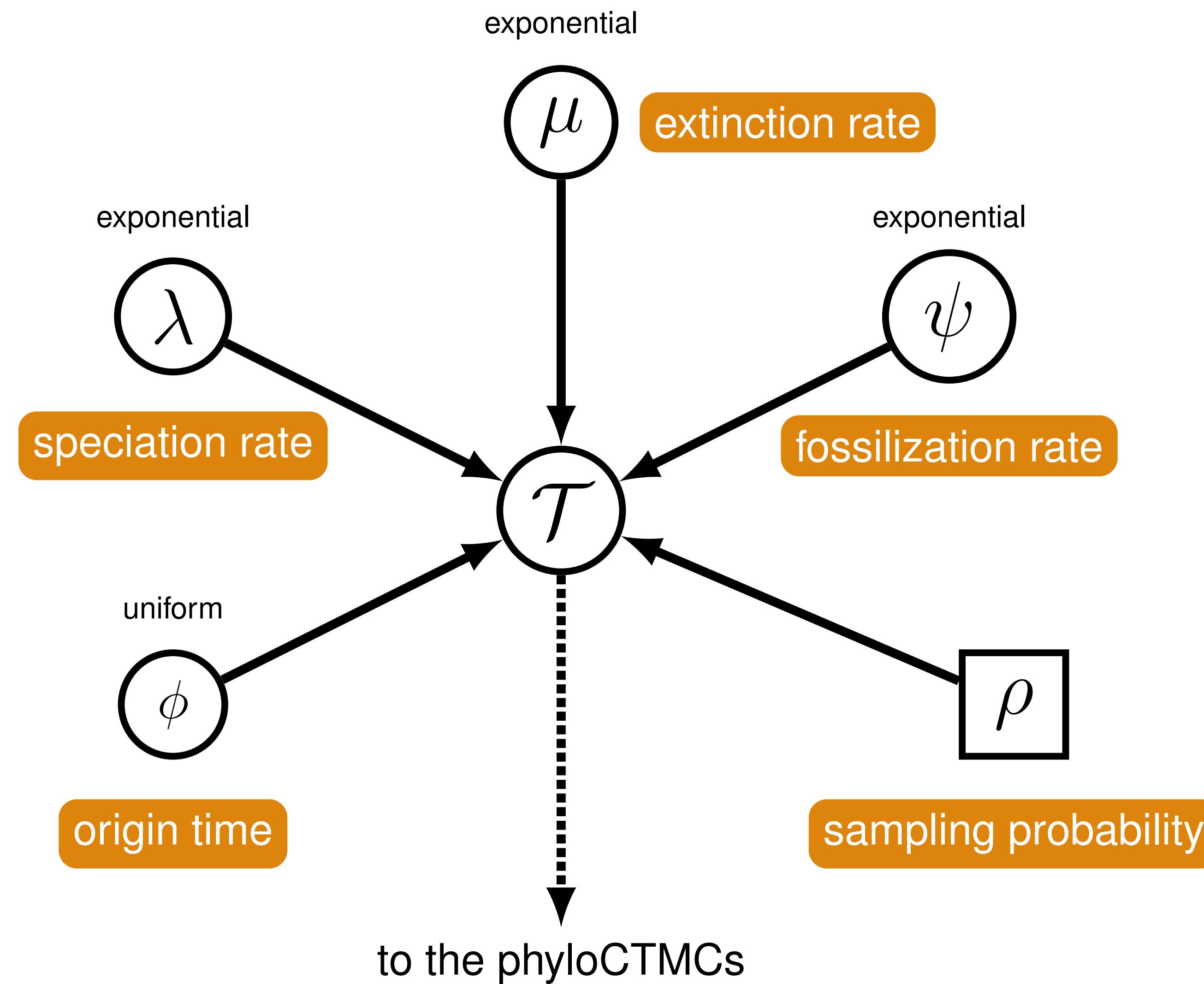
level of missing data ● none ● low ● high

Fossils improve phylogenetic analyses of morphological characters
Koch, Garwood, Parry. 2020. Proc B

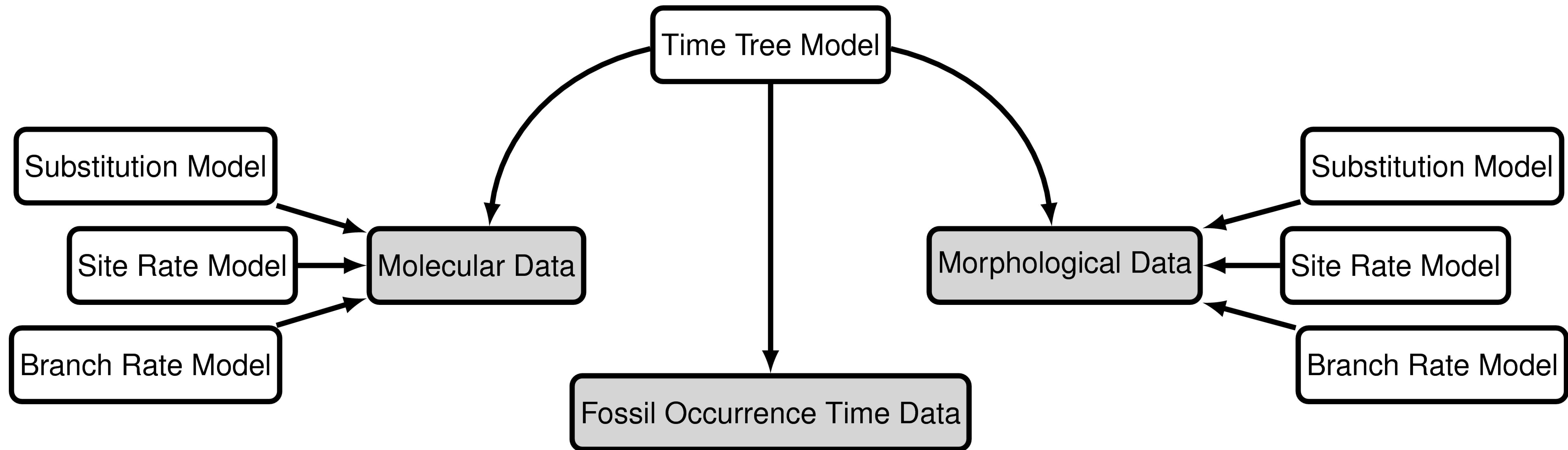
Some notes

- The topology of extant taxa is largely unaffected by how fossils are incorporated
- Fossils *and* age information help inform topology
- Divergence times are much more sensitive to errors in fossil placement and model misspecification
- Total-evidence dating is more robust to model misspecification

Graphical model representation of the FBDP



Bringing everything together

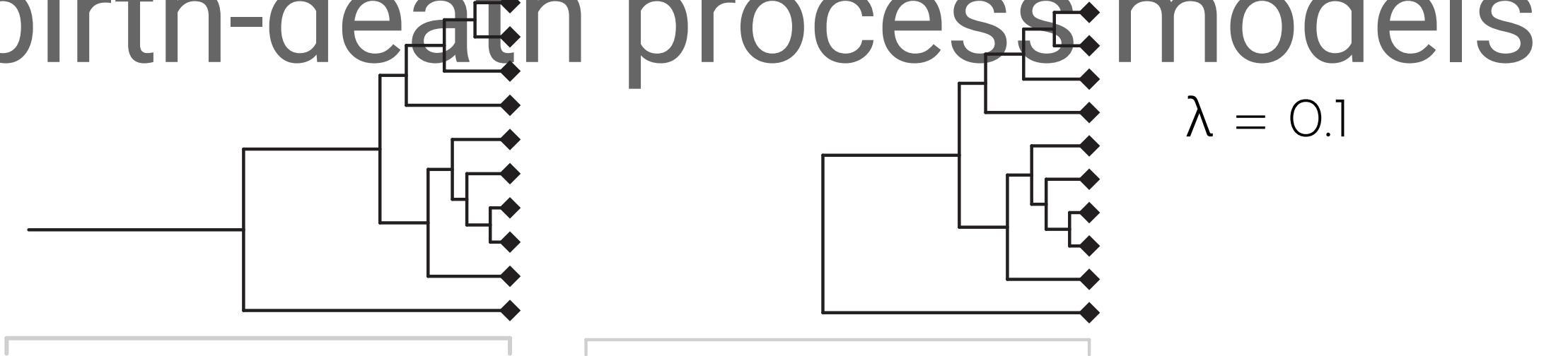


Relationship to (some) other birth-death process models

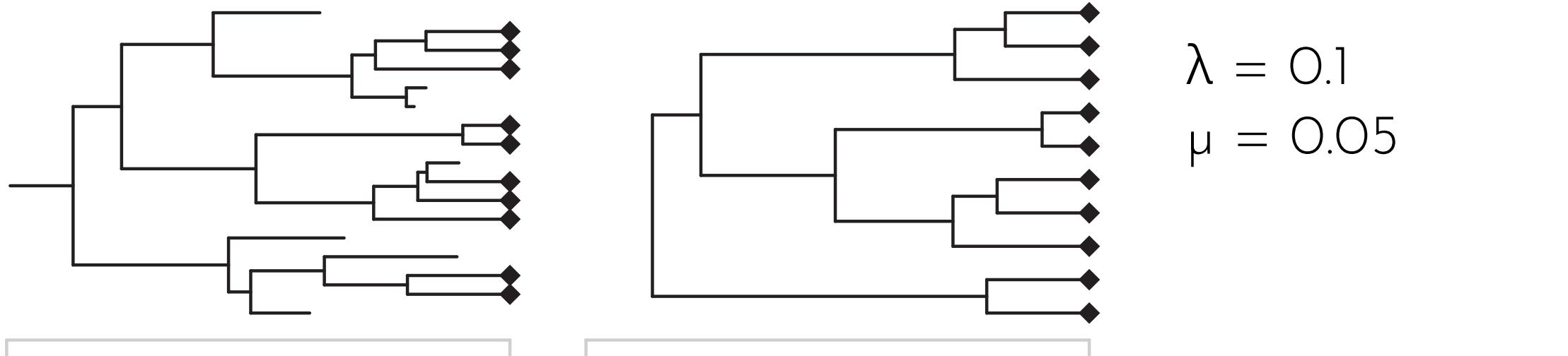
These models are special cases of the FBD process, with fossil sampling (ψ) = zero.

complete vs. reconstructed trees

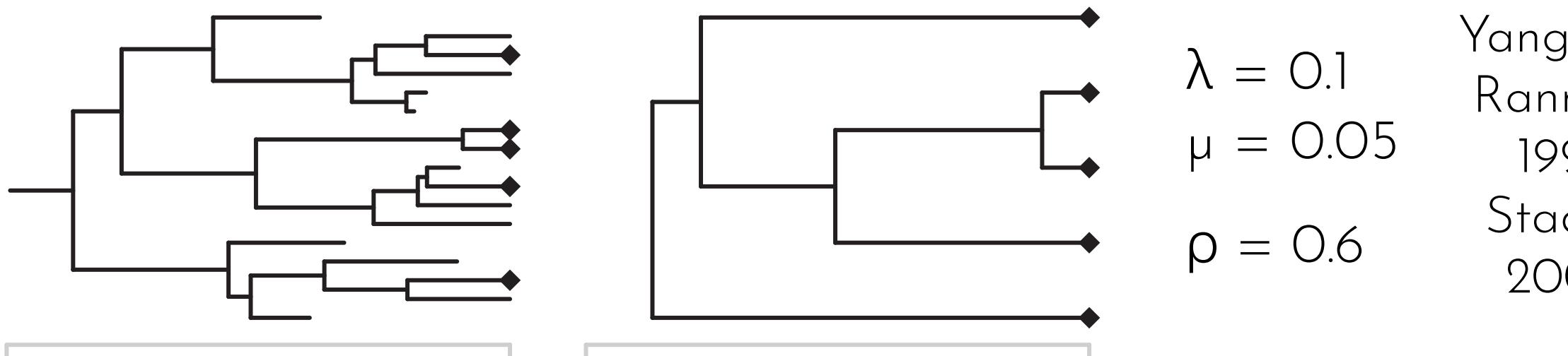
complete vs. reconstructed trees



$$\lambda = 0.1$$

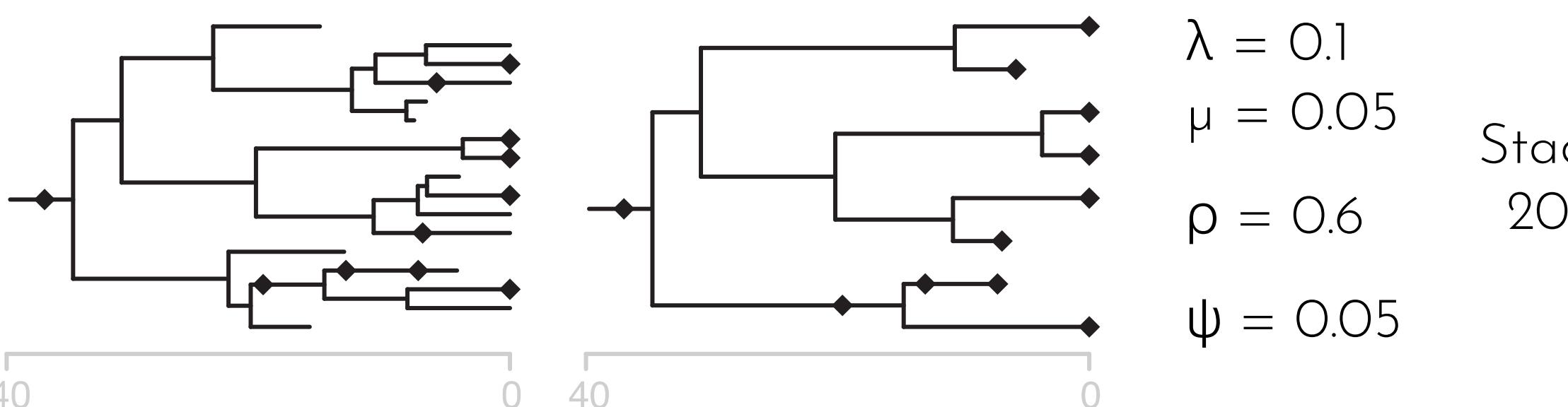


$$\begin{aligned} \lambda &= 0.1 \\ \mu &= 0.05 \end{aligned}$$



$$\begin{aligned} \lambda &= 0.1 \\ \mu &= 0.05 \\ \rho &= 0.6 \end{aligned}$$

Yang and
Rannala
1997
Stadler
2009



$$\begin{aligned} \lambda &= 0.1 \\ \mu &= 0.05 \\ \rho &= 0.6 \\ \psi &= 0.05 \end{aligned}$$

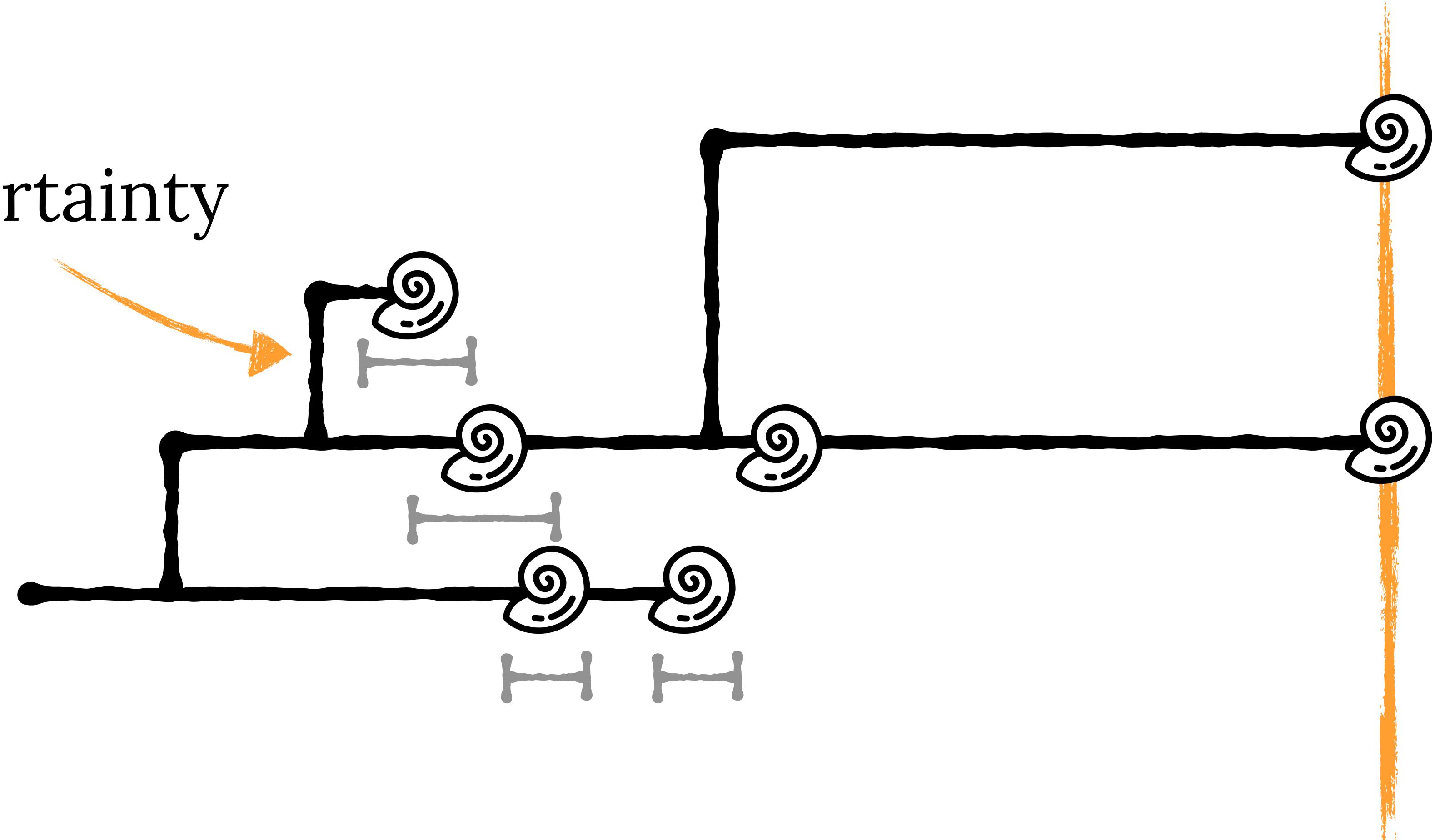
Stadler
2010

Stadler et al. 2012
See also: Stadler and Yang 2013

Sample age uncertainty



age uncertainty

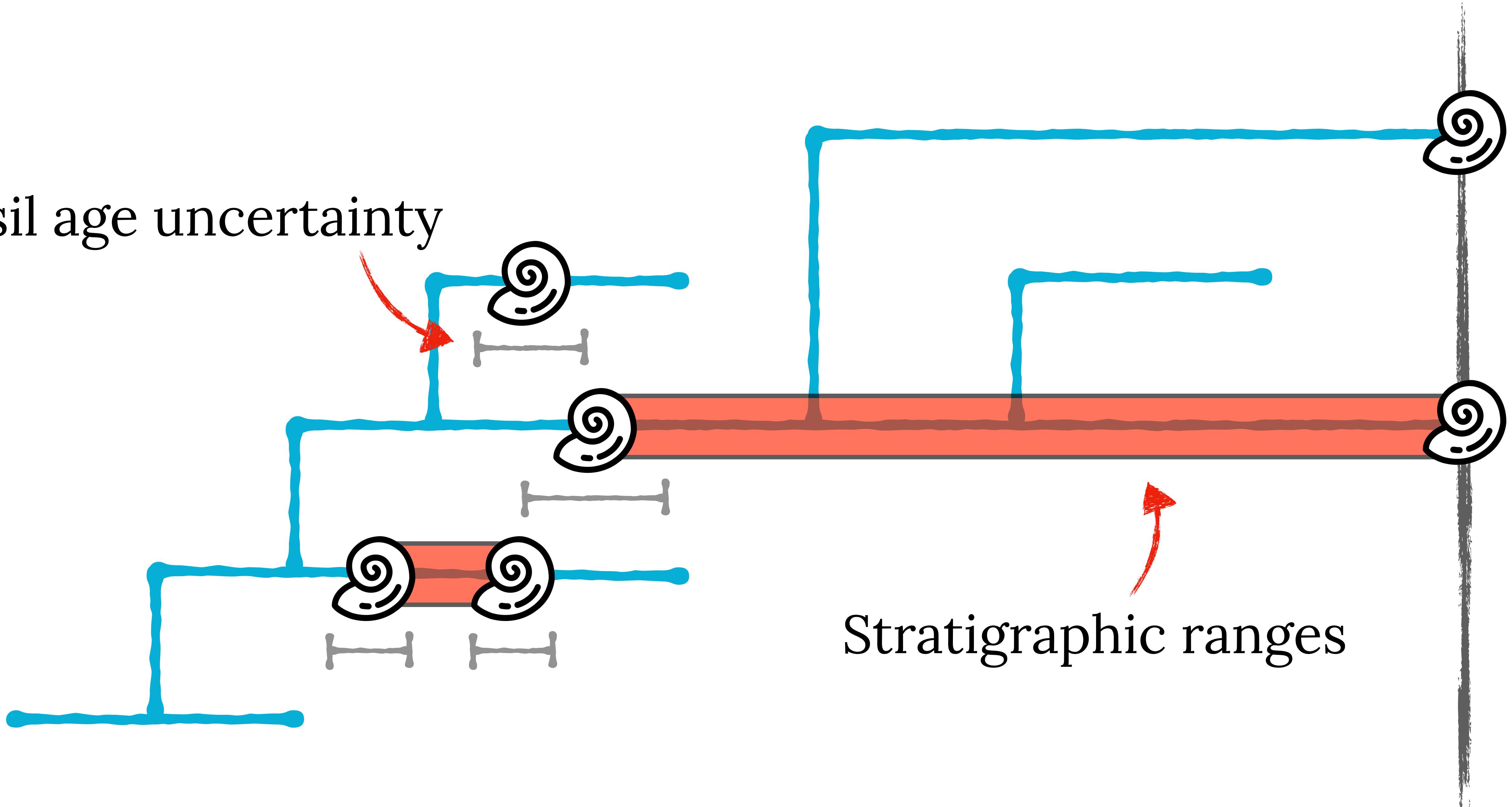


Ignoring Fossil Age Uncertainty Leads to Inaccurate Topology in Time Calibrated Tree Inference

Barido-Sottani et al. 2018, 2020

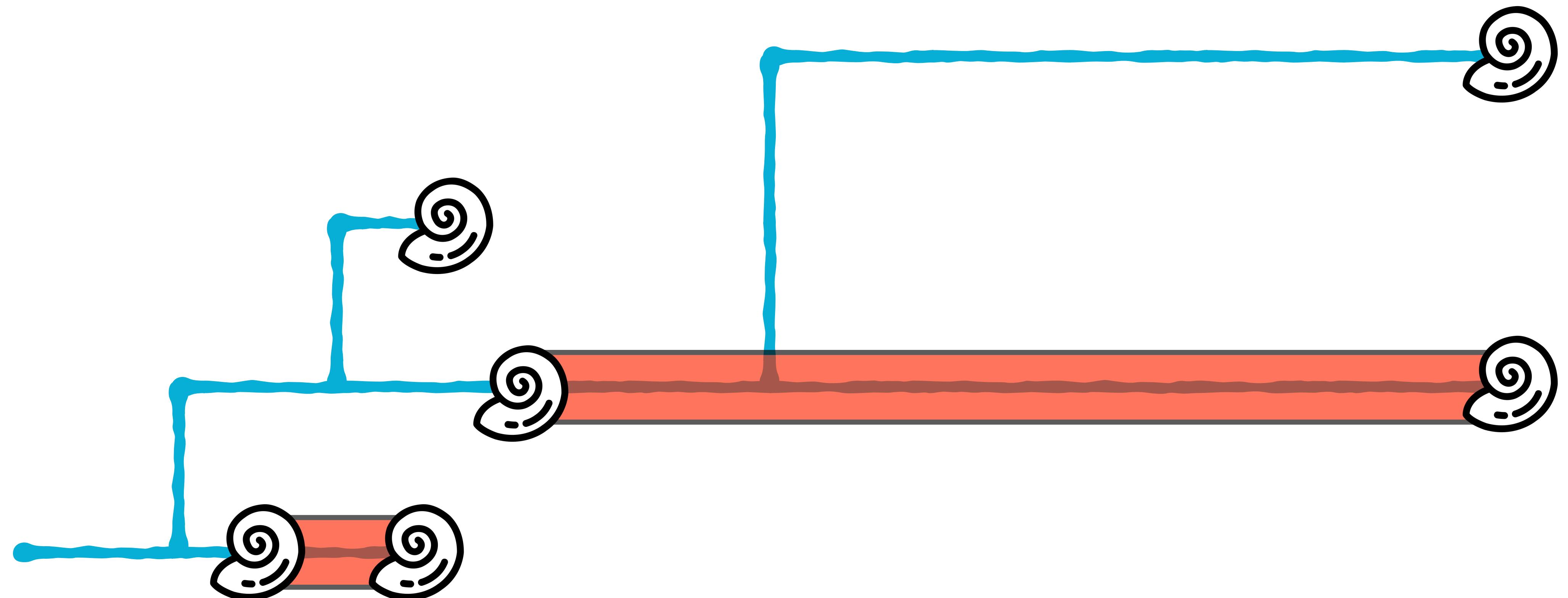
Putting the F in FBD analyses: tree constraints or morphological data? Barido-Sottani et al. 2023₈₈

fossil age uncertainty

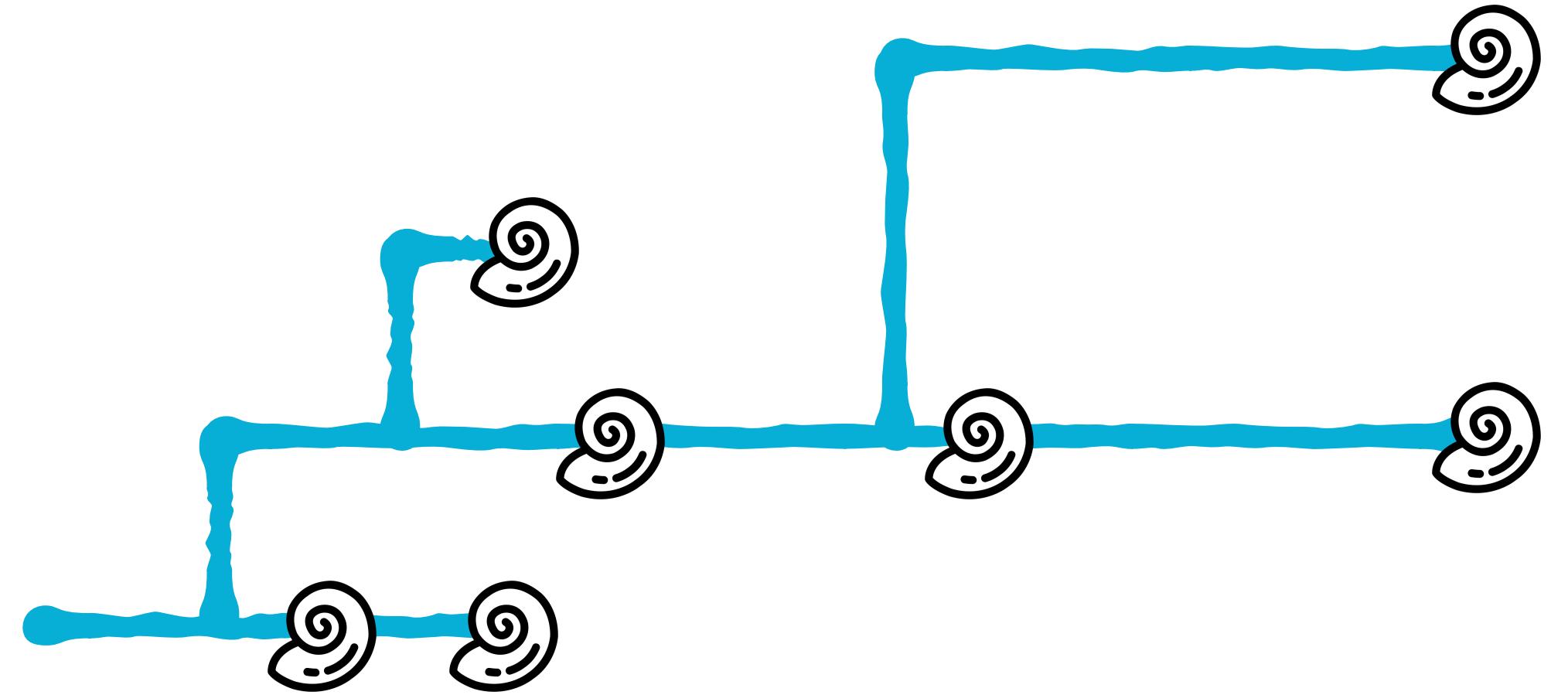


The inseparability of sampling and time and its influence on attempts to unify the molecular & fossil records
Hopkins et al. 2018. Paleobiology

The fossilised birth-death range process



*The fossilised birth-death model for the analysis of stratigraphic range data under different speciation modes
Stadler et al. 2018. JTB*

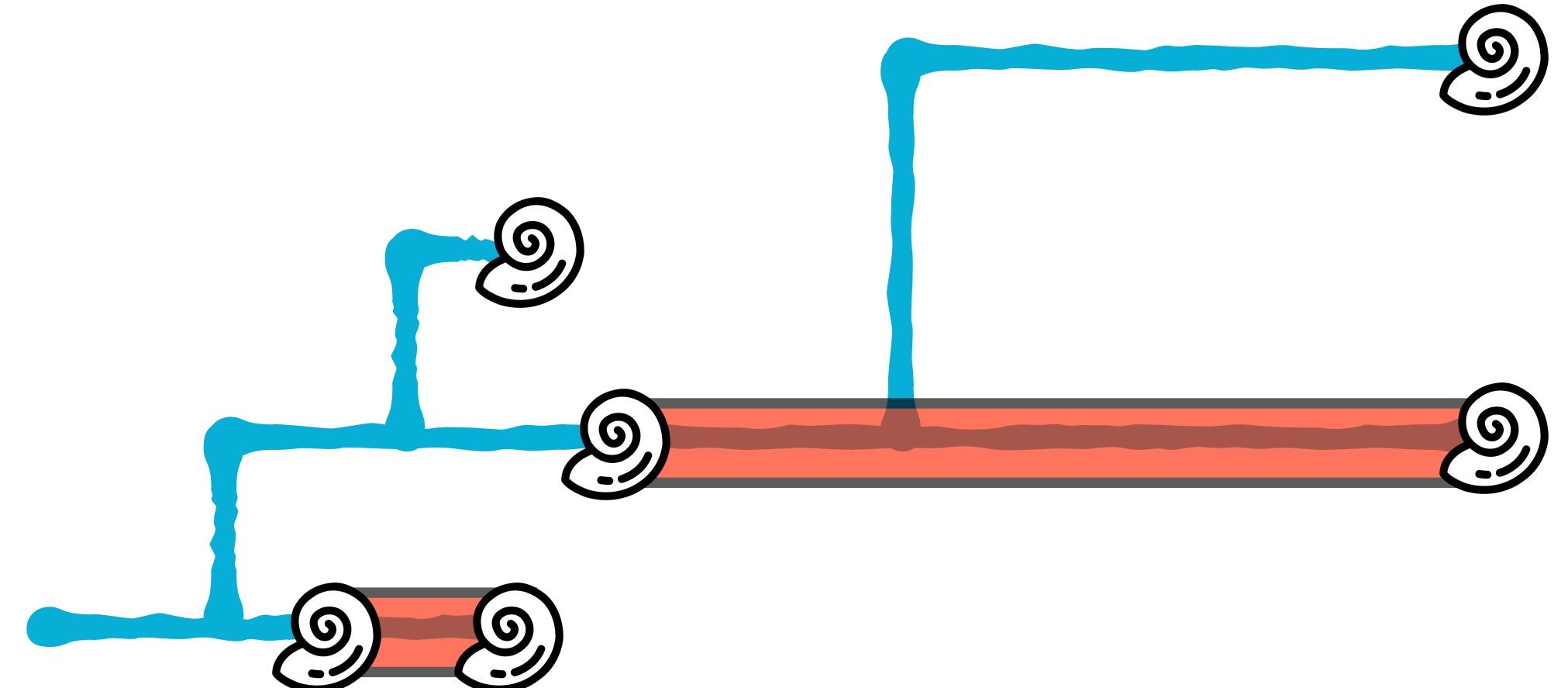


FBD process for analysis of specimen level data

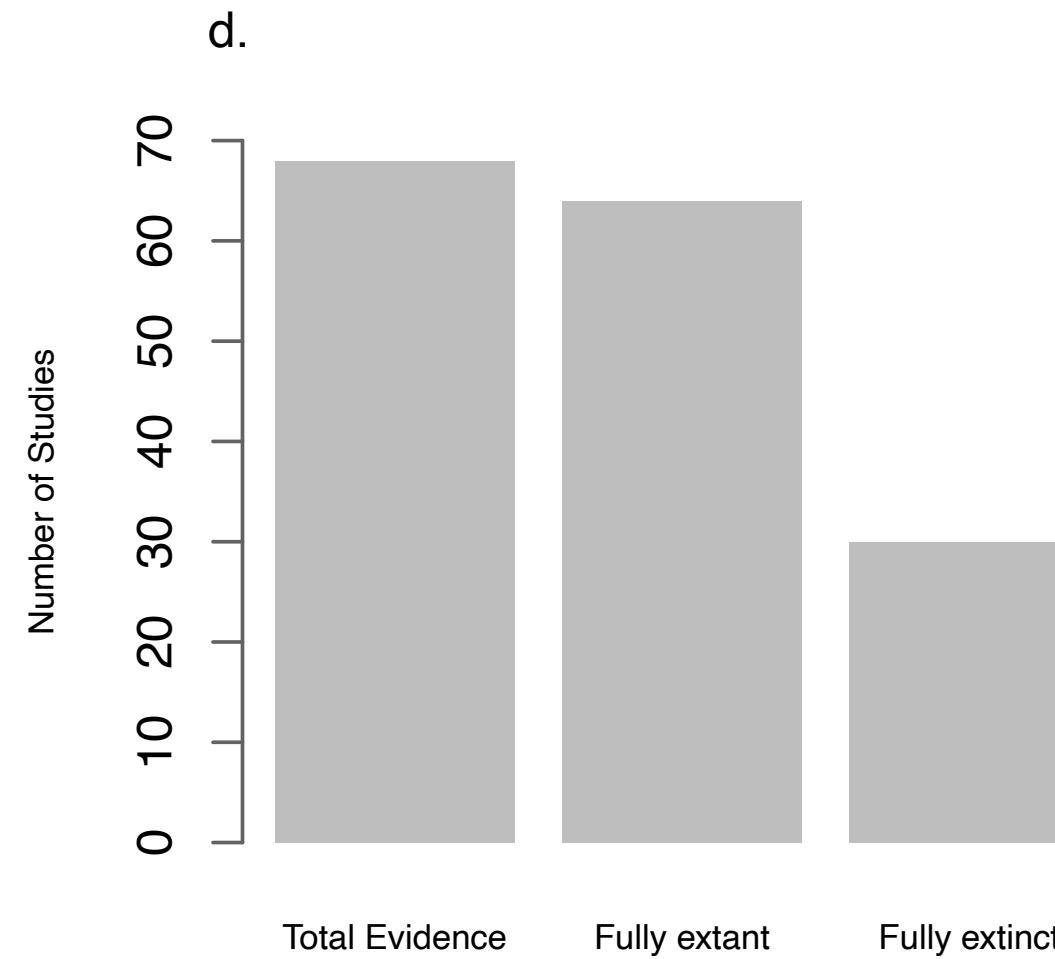
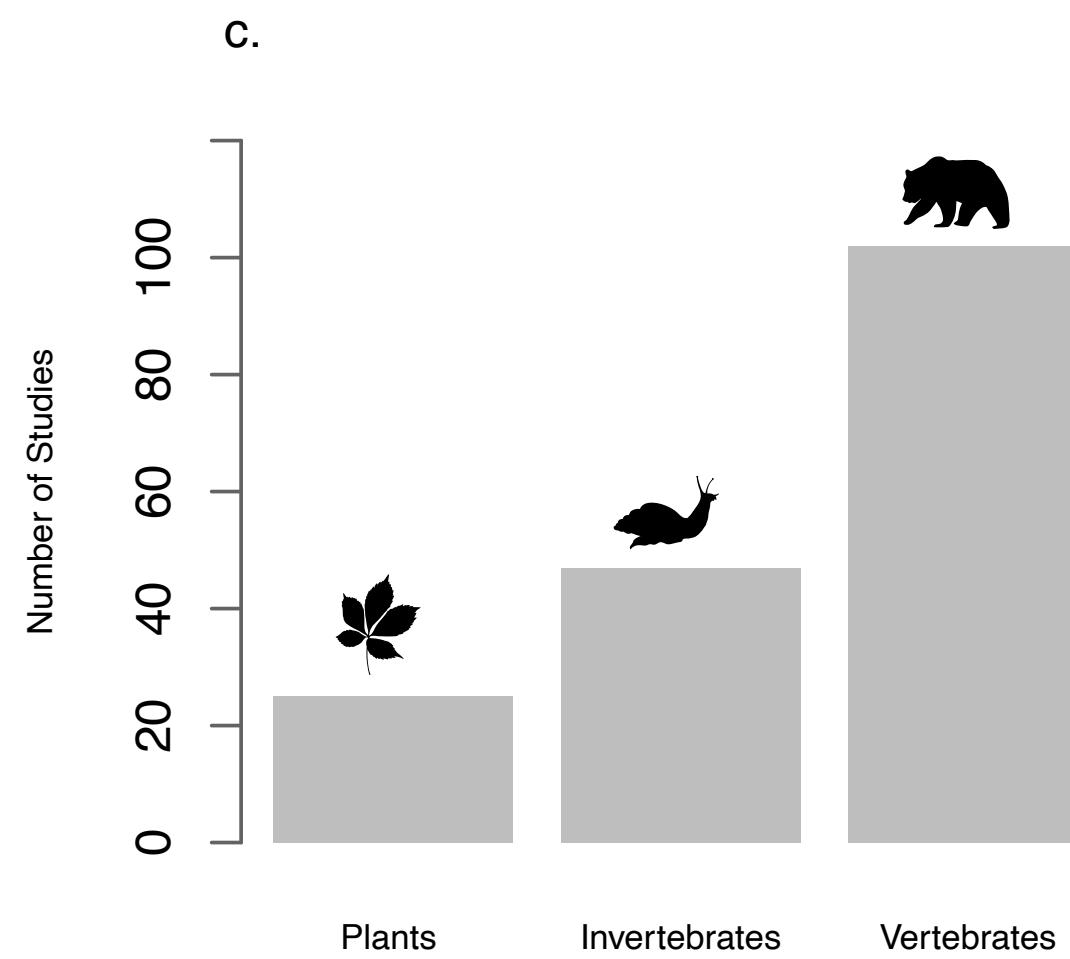
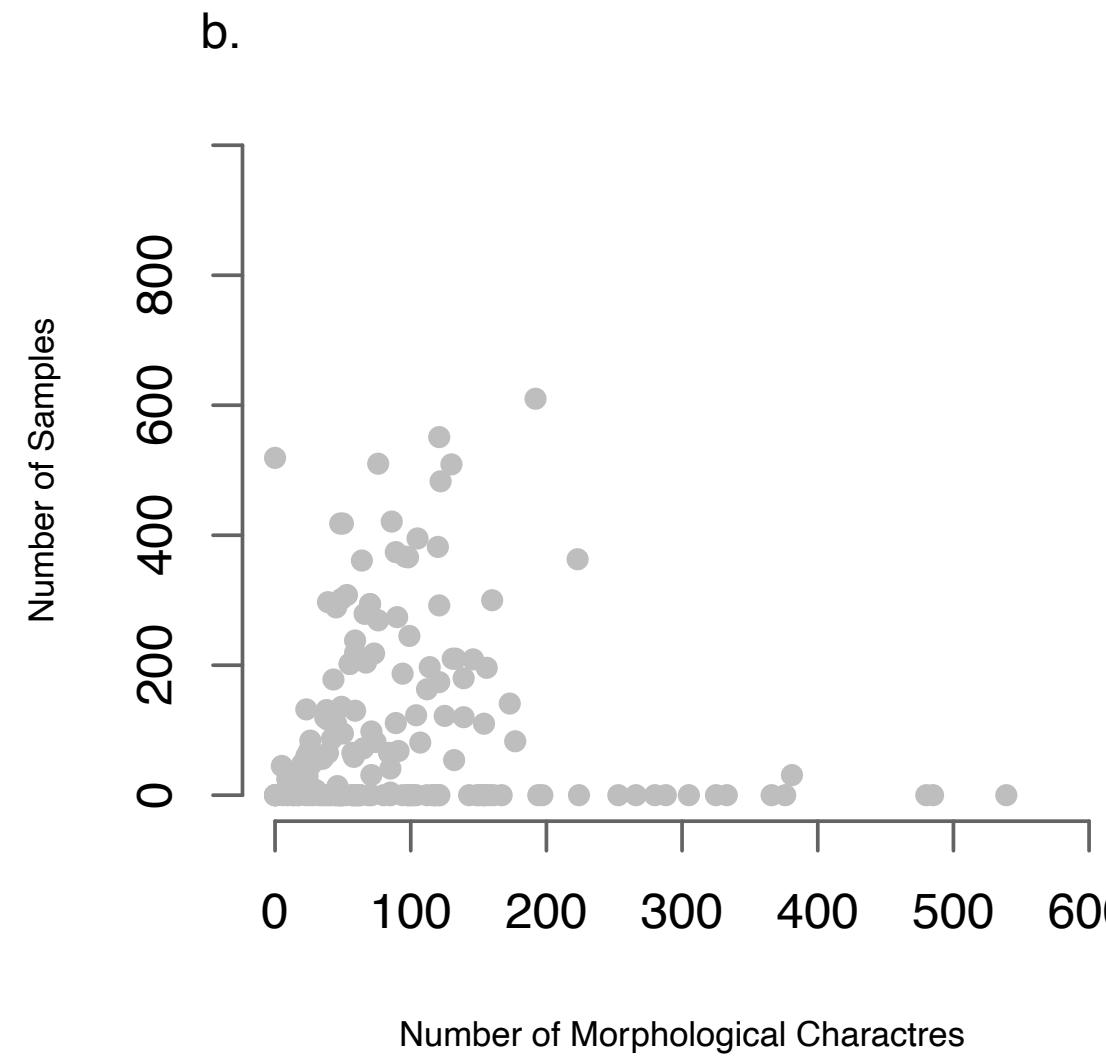
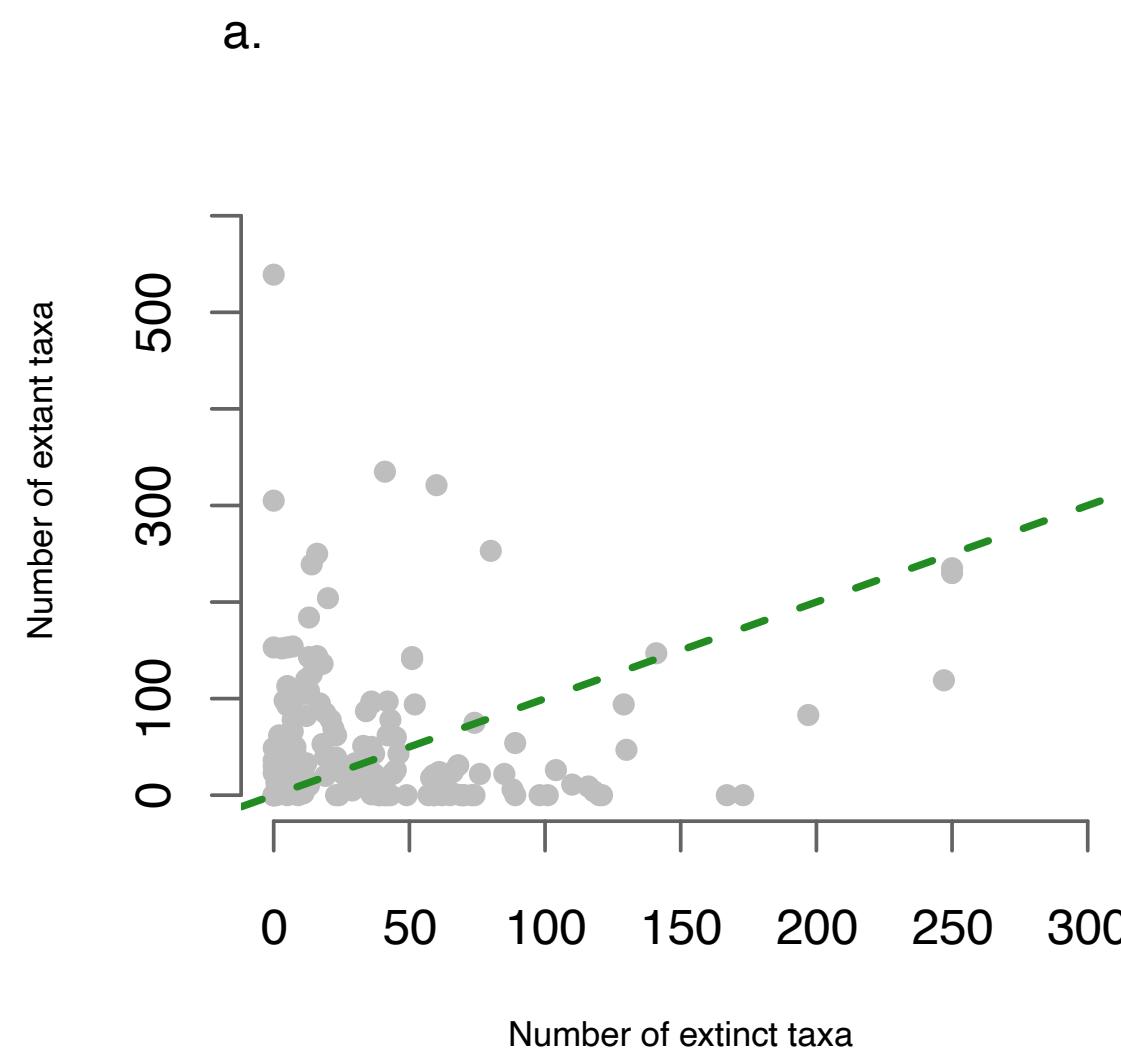
$$P(E | \text{spiral}^{\lambda, \mu, \psi, p})$$

The FBD range process for analysis of stratigraphic ranges

$$Pr(E | \text{spiral}^{\lambda, \mu, \psi})$$



When can you use the FBD process?



Used >170 times to date (mainly in macroevolution)

Can be applied to a wide range of scenarios

Upper limit is about 500 samples/tips

Not practical with large alignments

Computational cost comes from sampling tree space

Exercise

Phylogenetics

Diversification rate estimation

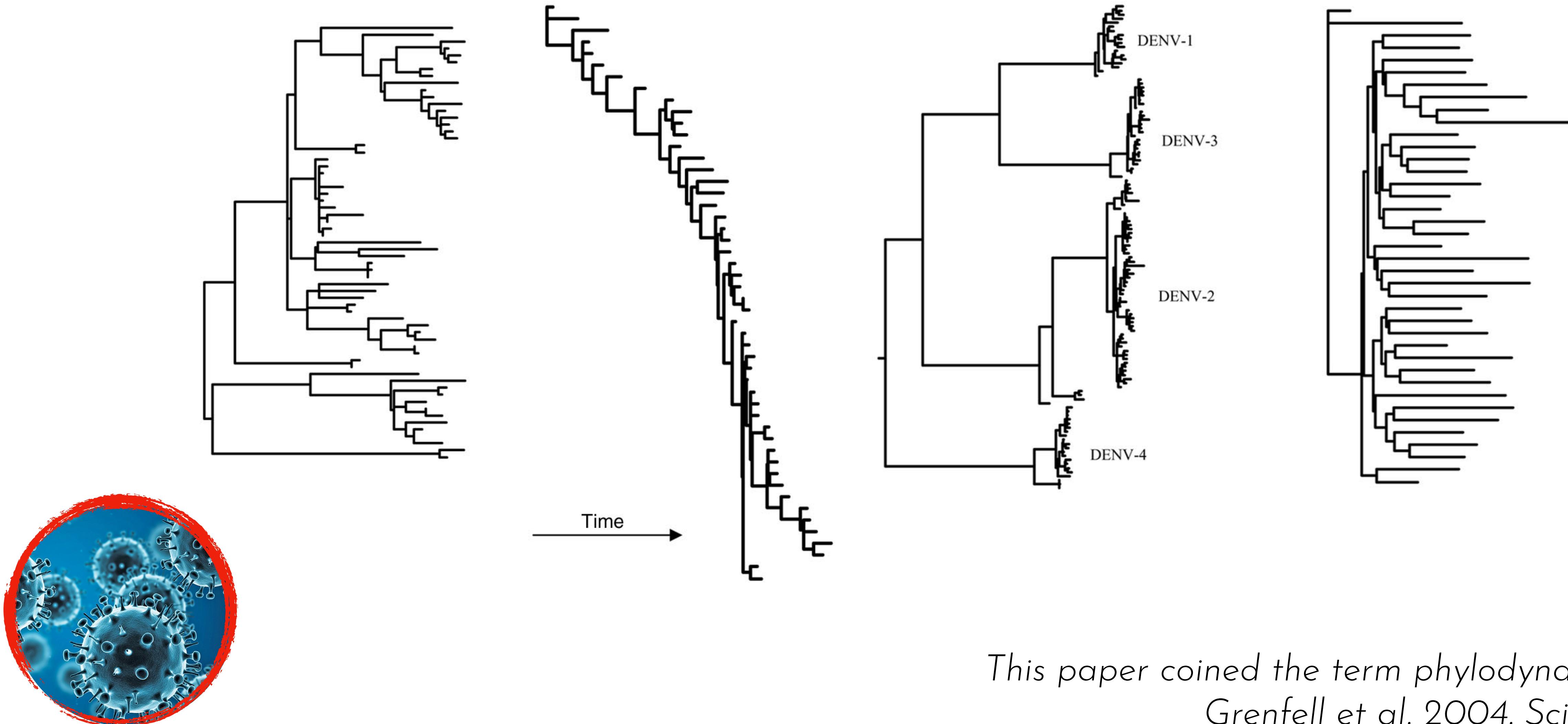
Bayesian divergence time estimation

$$P(E \mid \lambda, \mu, \psi, p, O, t \mid 0101\dots, 1101\dots, 0100\dots, \text{snail}) =$$

probability of the
time tree

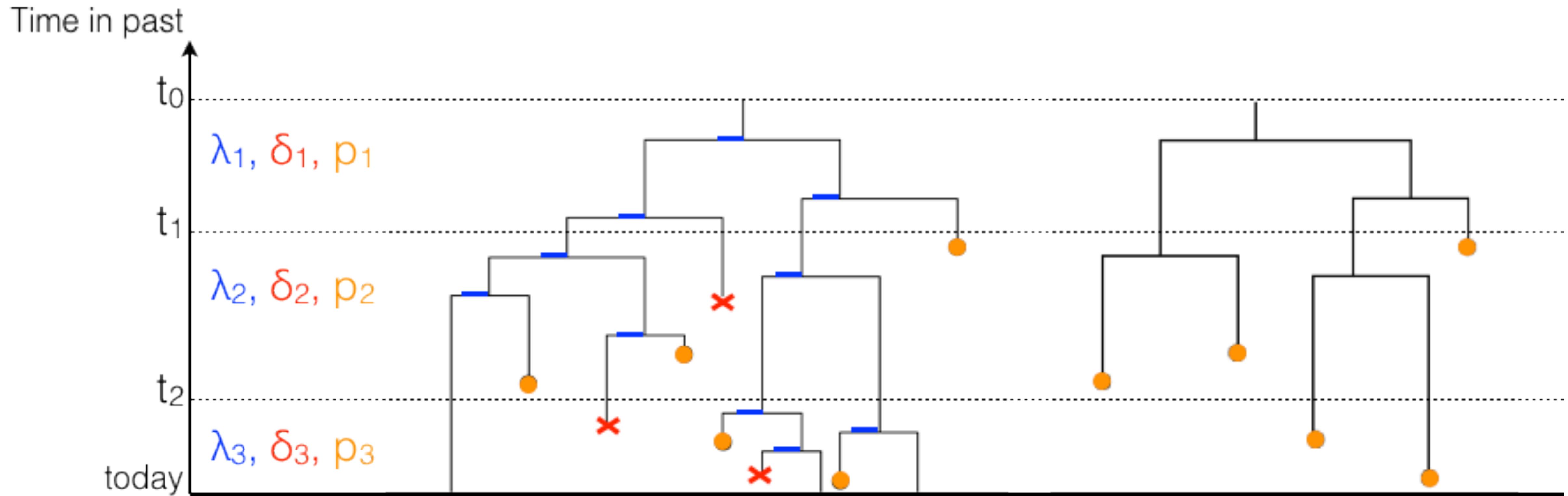
$$\frac{P(0101\dots, 1101\dots, 0100\dots \mid E) P(E \mid \lambda, \mu, \psi, p, O, t) P(\lambda, \mu, \psi, p) P(O) P(t)}{P(0101\dots, 1101\dots, 0100\dots, \text{snail})}$$

Tree shape is informative about underlying dynamics

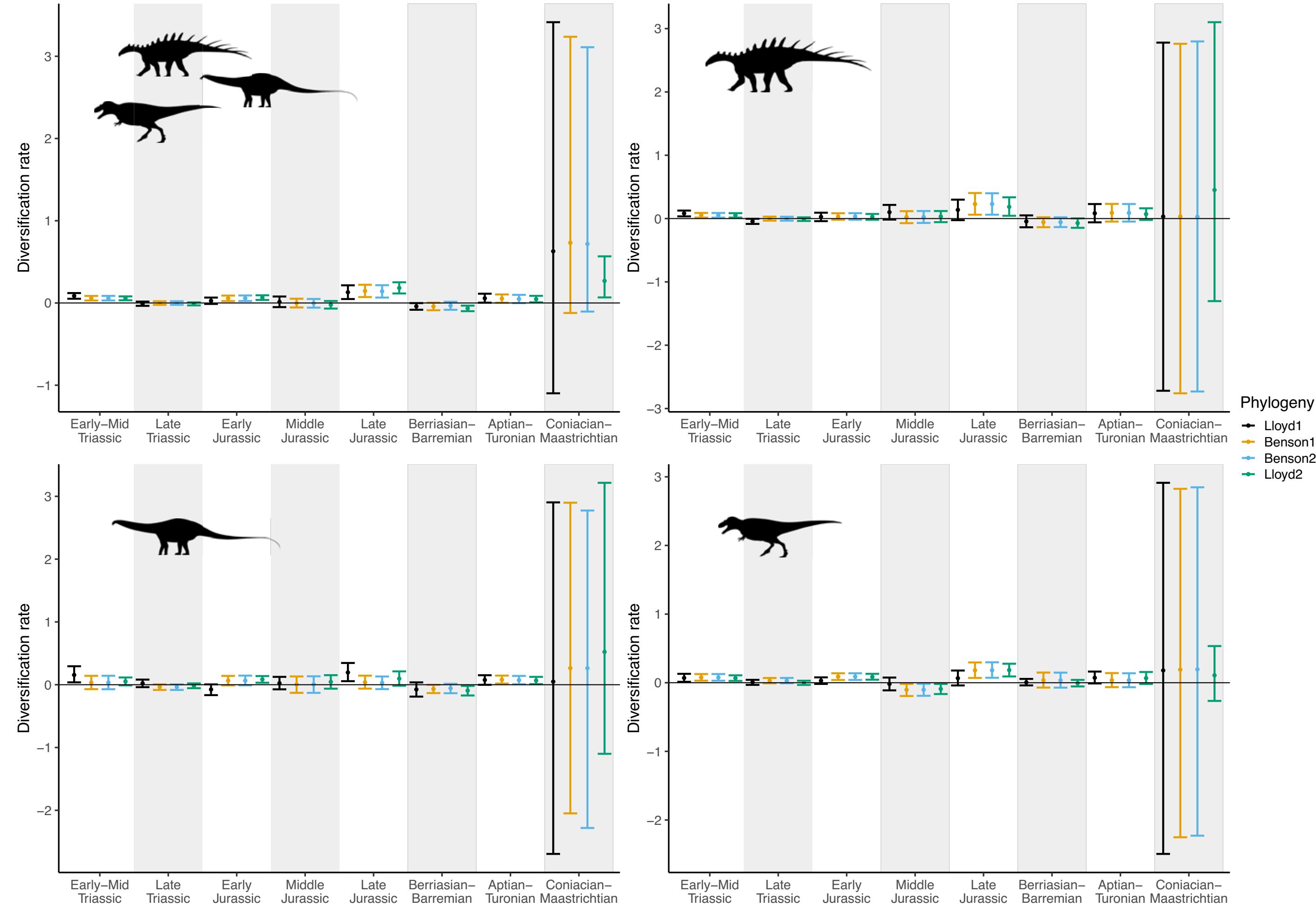


The skyline birth-death process

First used for tracking the spread of infectious diseases



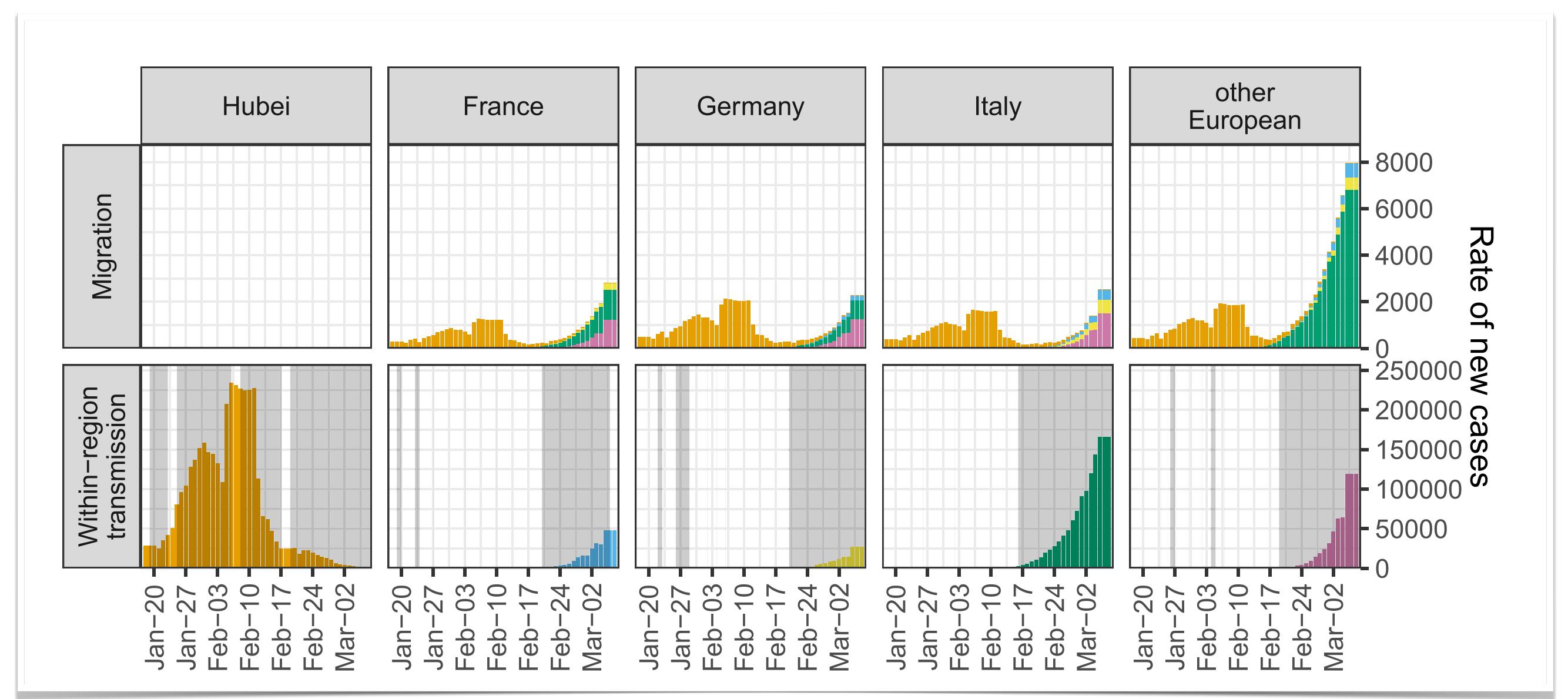
Macroevolutionary case study



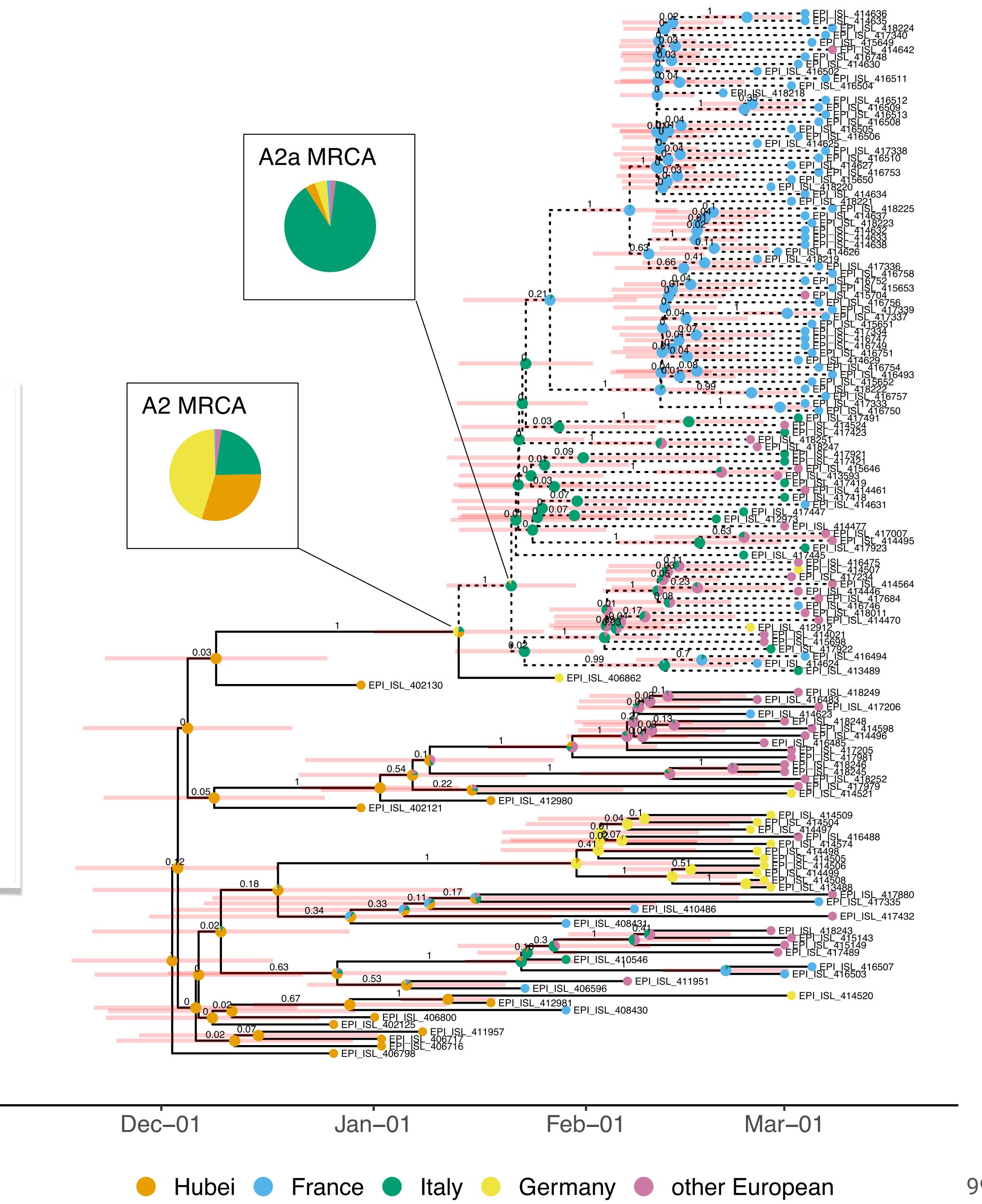
Phylogenies have been used to argue dinosaurs were incline prior to the KPg

FBD analyses suggest that we can not currently answer that question using phylogenies

Models that include migration



The origin and early spread of SARS-CoV-2 in Europe
Nadeau et al. 2021. PNAS



Bayesian divergence time estimation

The data

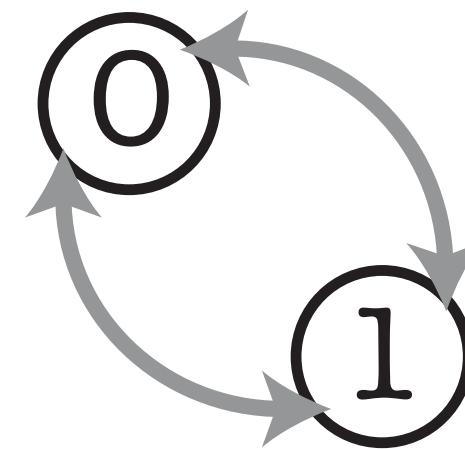
and / or
0101... ATTG...
1101... TTGC...
0100... ATTC...



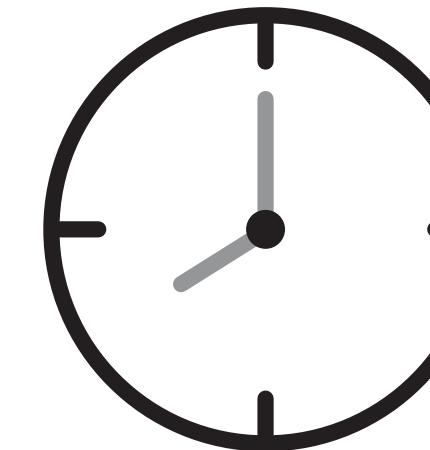
phylogenetics
characters

sample
ages

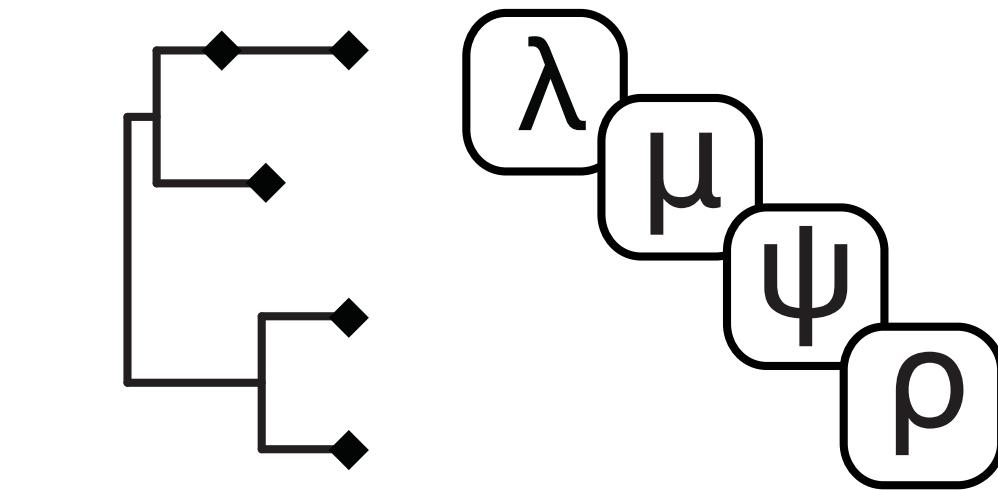
3 model components



substitution
model



clock
model



tree and tree
model

Using PCMs for dating

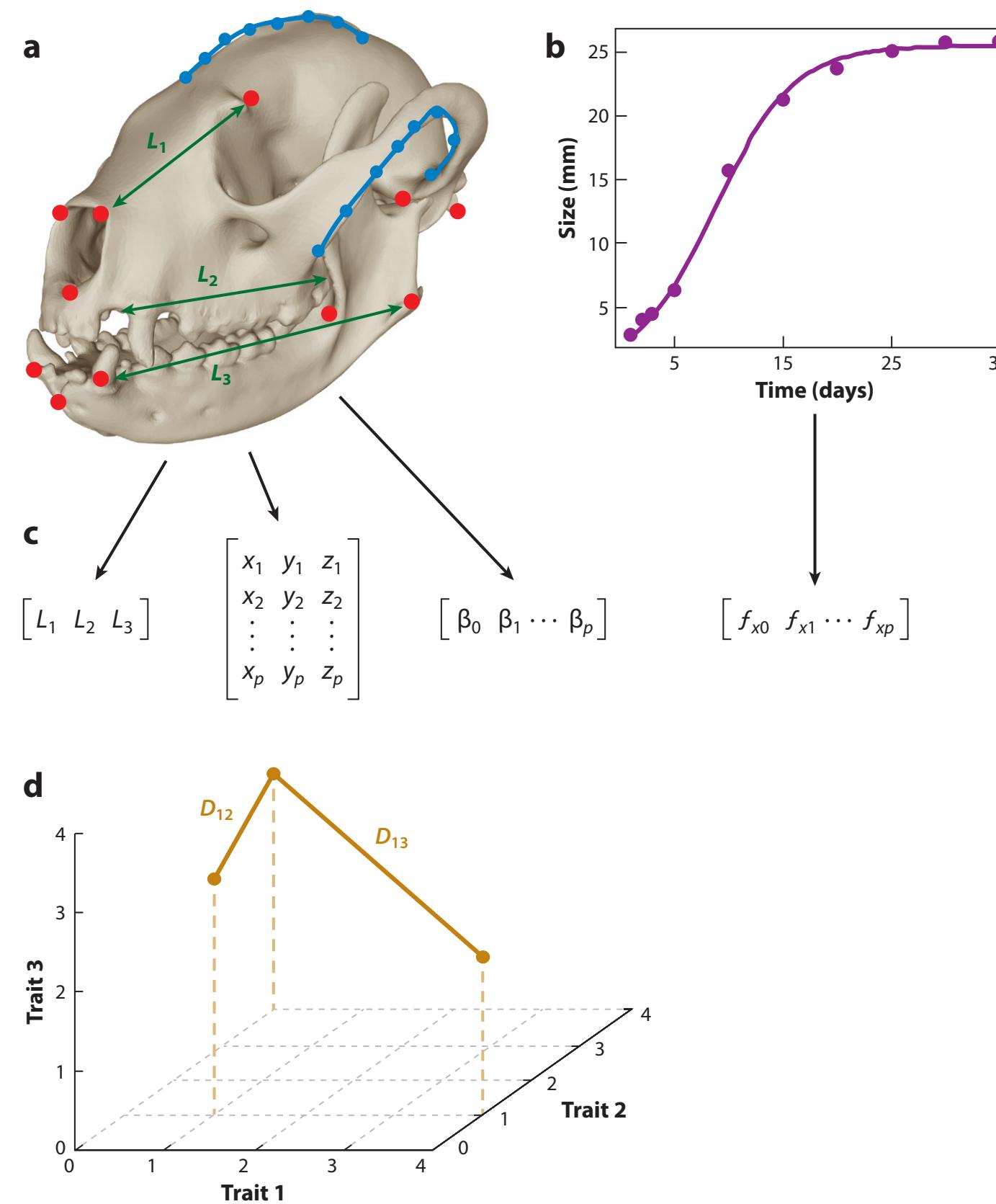
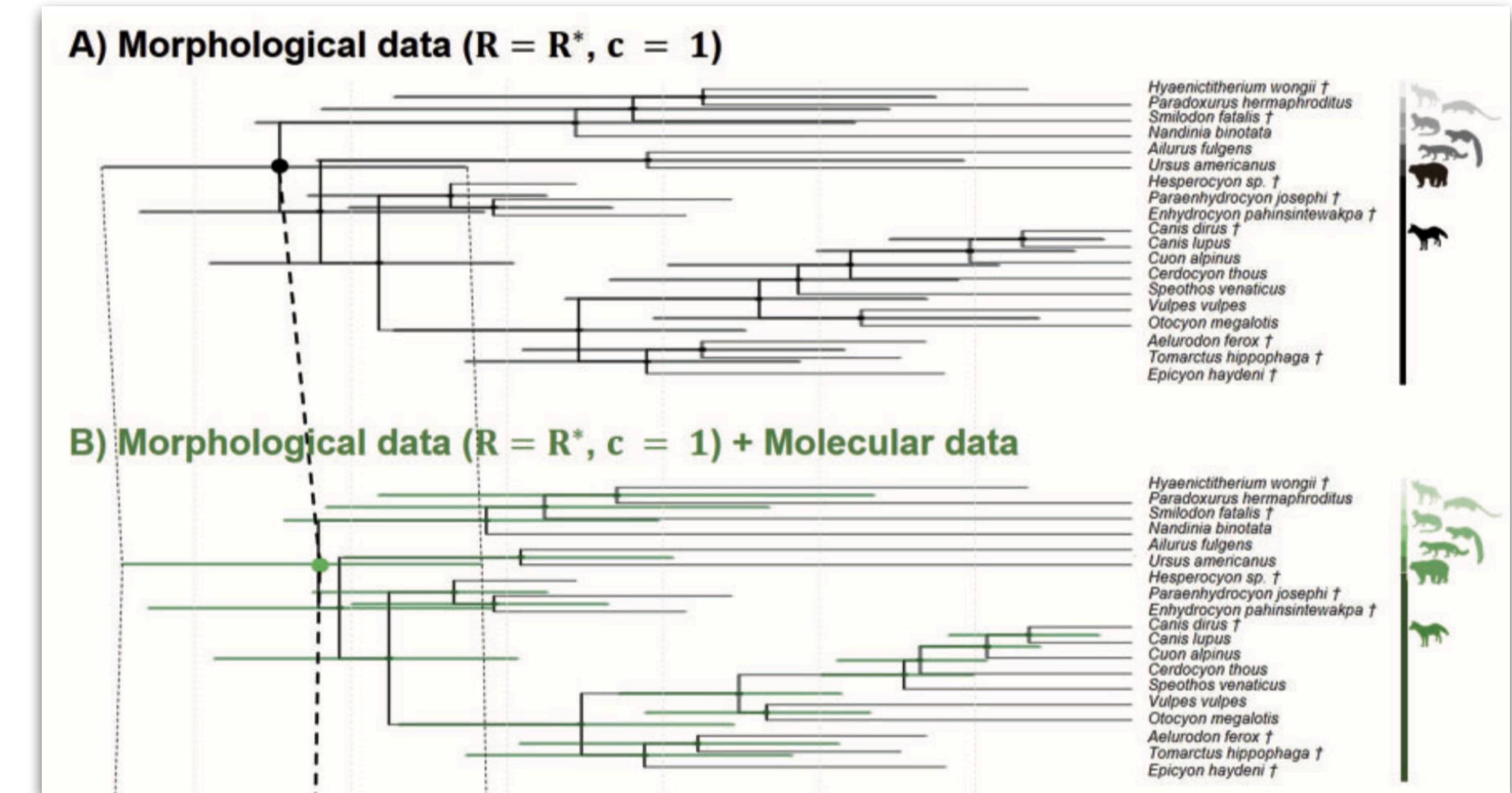
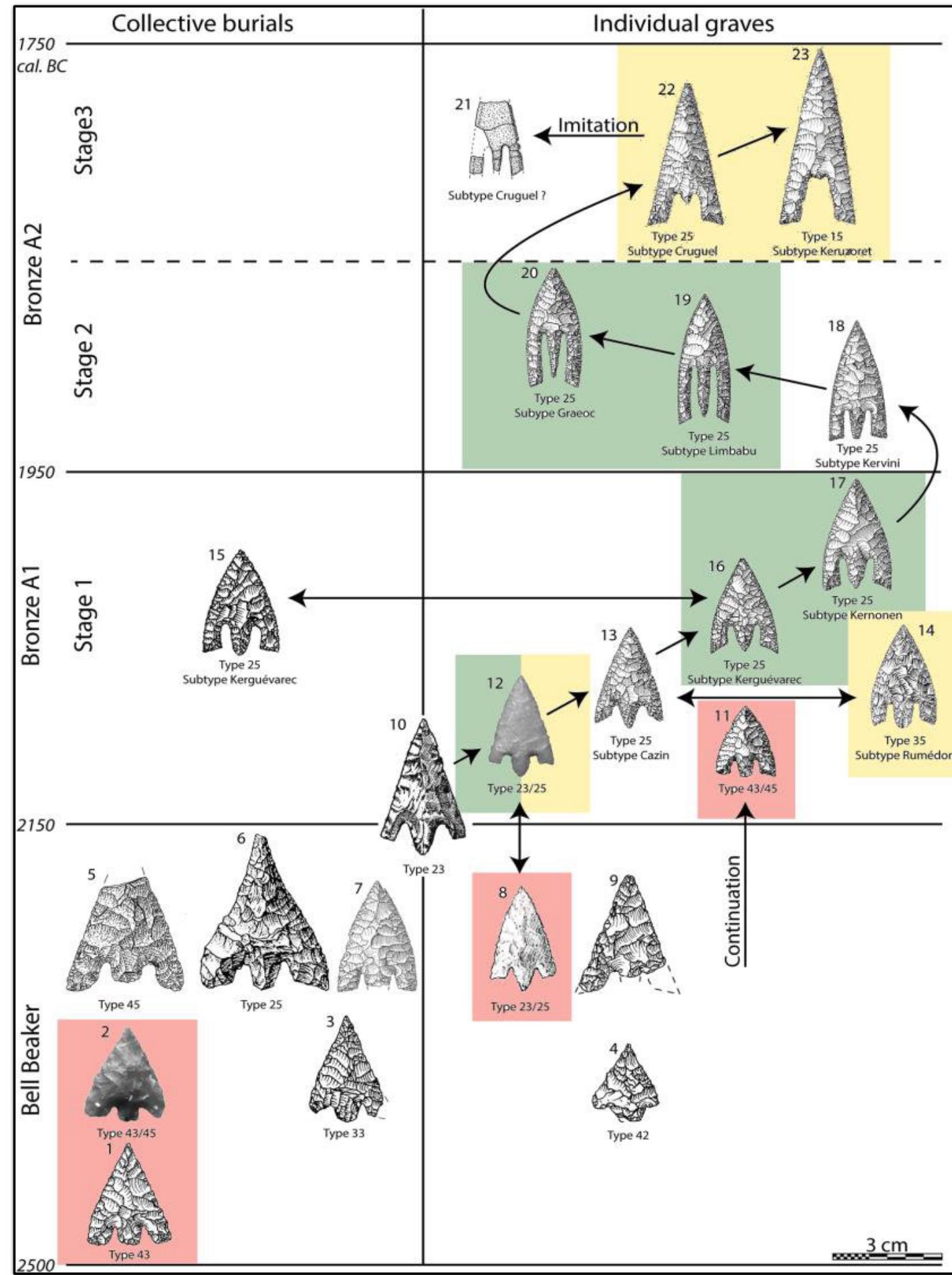


Image source Adams & Collyer ([2019](#))



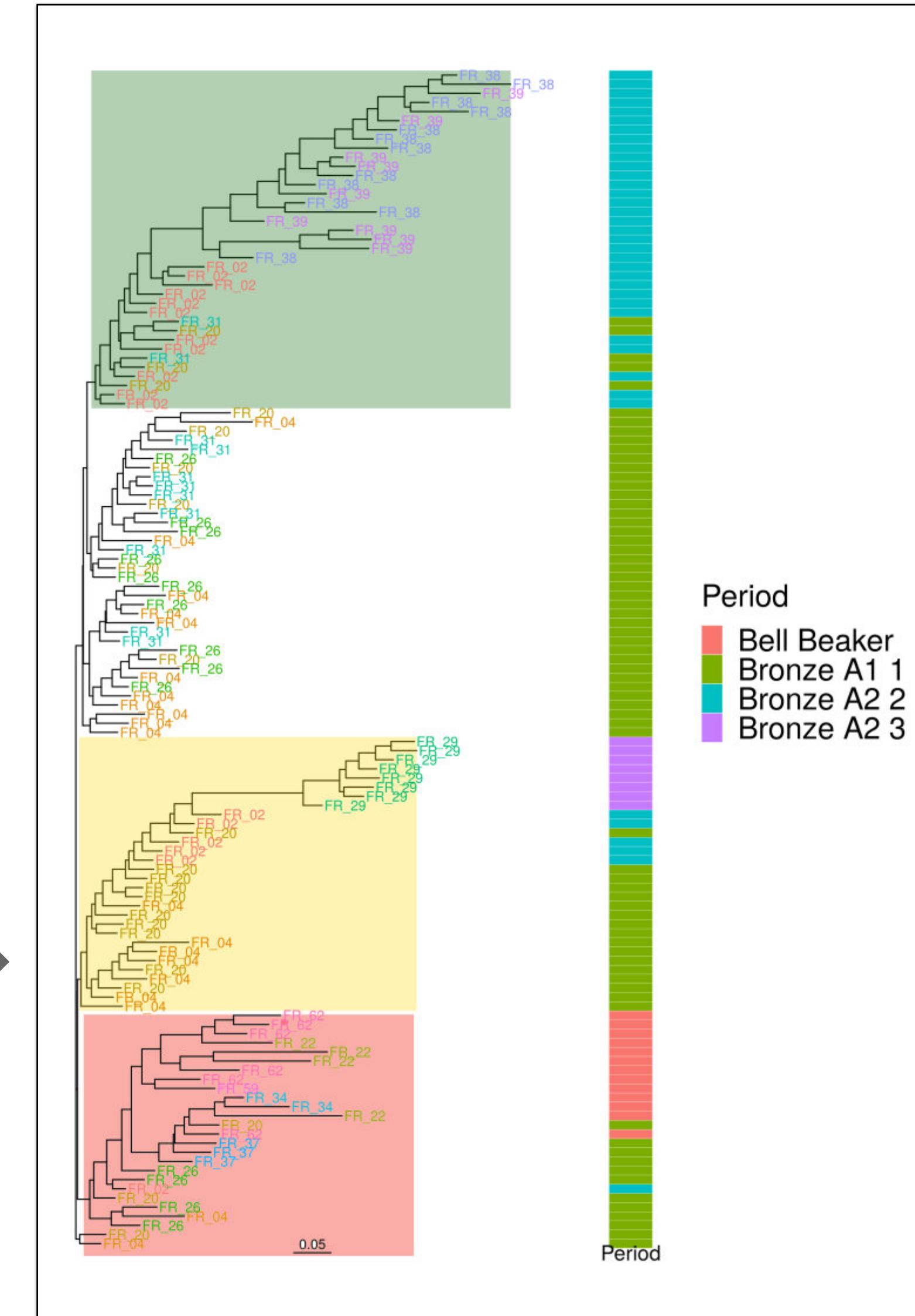
Álvarez-Carretero et al. ([2019](#)) Bayesian Estimation of Species Divergence Times Using Correlated Quantitative Characters

Cultural evolution



← Typo-Chronology of
Palaeolithic stone tools

Outline based NJ tree →

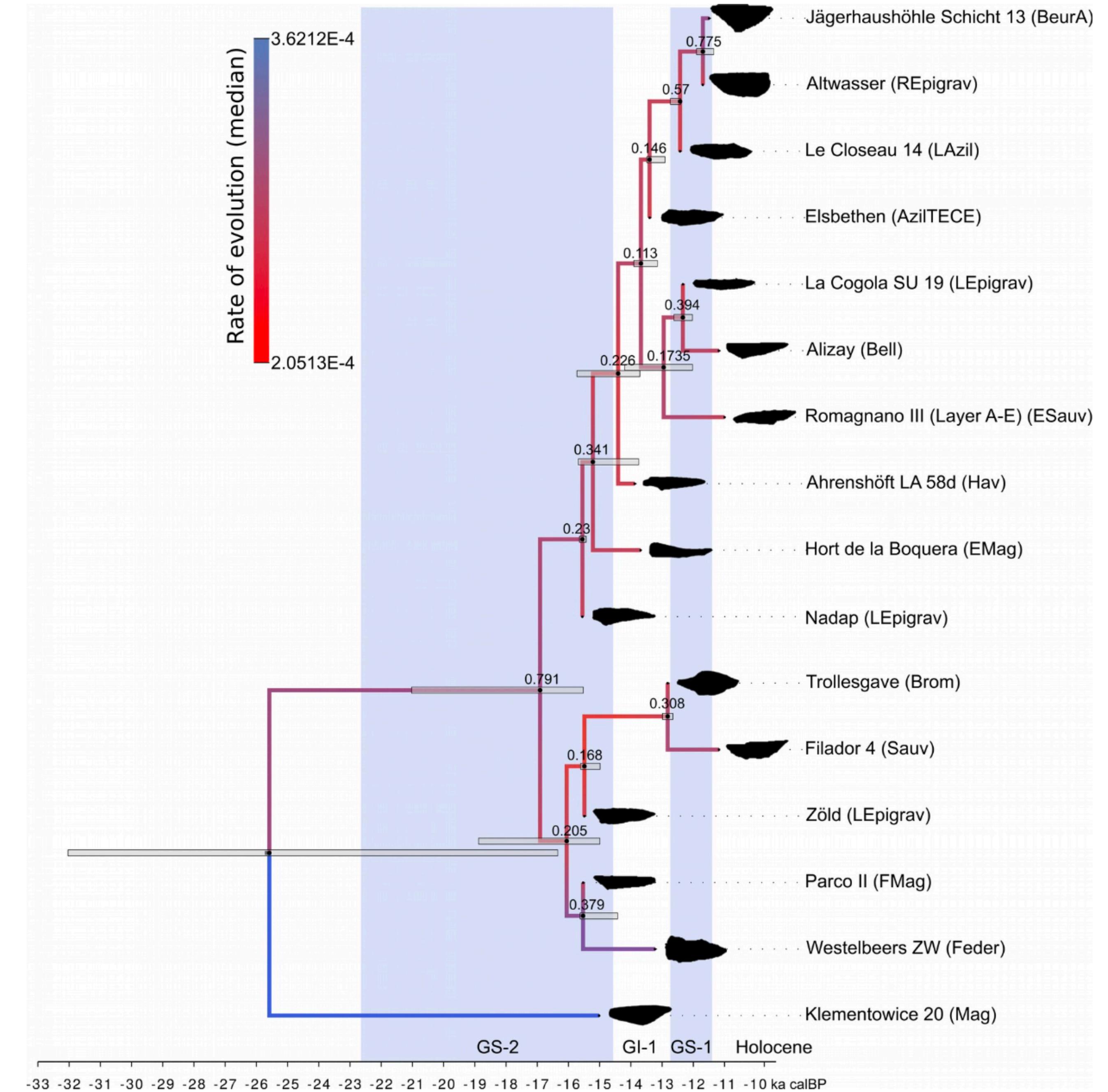


Matzig et al. 2021.

After Nicolas (2017)

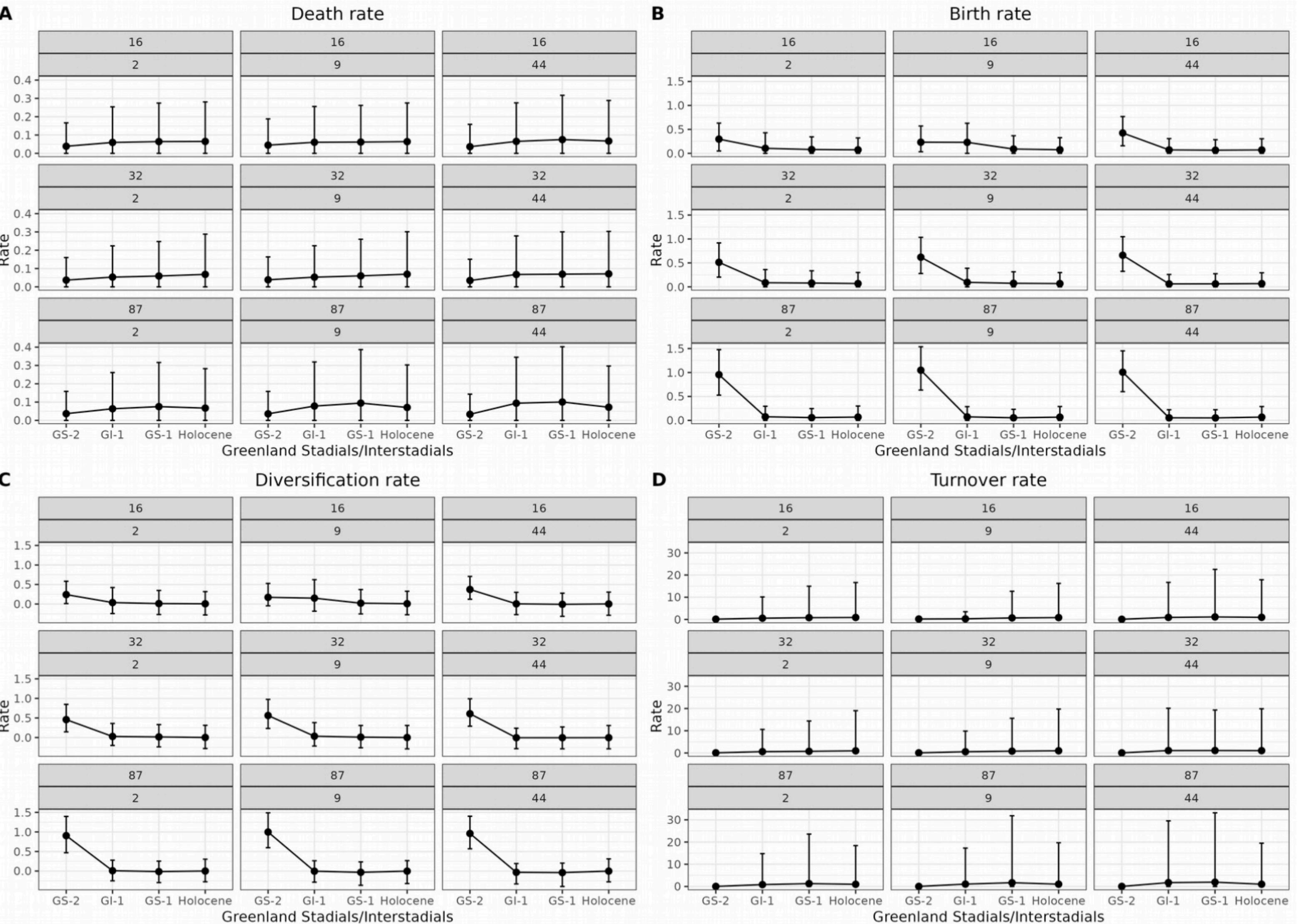
The tree topology of stone tools exhibits a lot of uncertainty

Matzig et al. (in review) A macroevolutionary analysis of European Late Upper Palaeolithic stone tool shape using a Bayesian phylodynamic framework (preprint available)



Sensitivity analyses

Birth, death, and sampling rates are impacted by trait and taxon sampling



Joint phylogenetic estimation of geographic movements and biome shifts



Global diversity of *Viburnum*

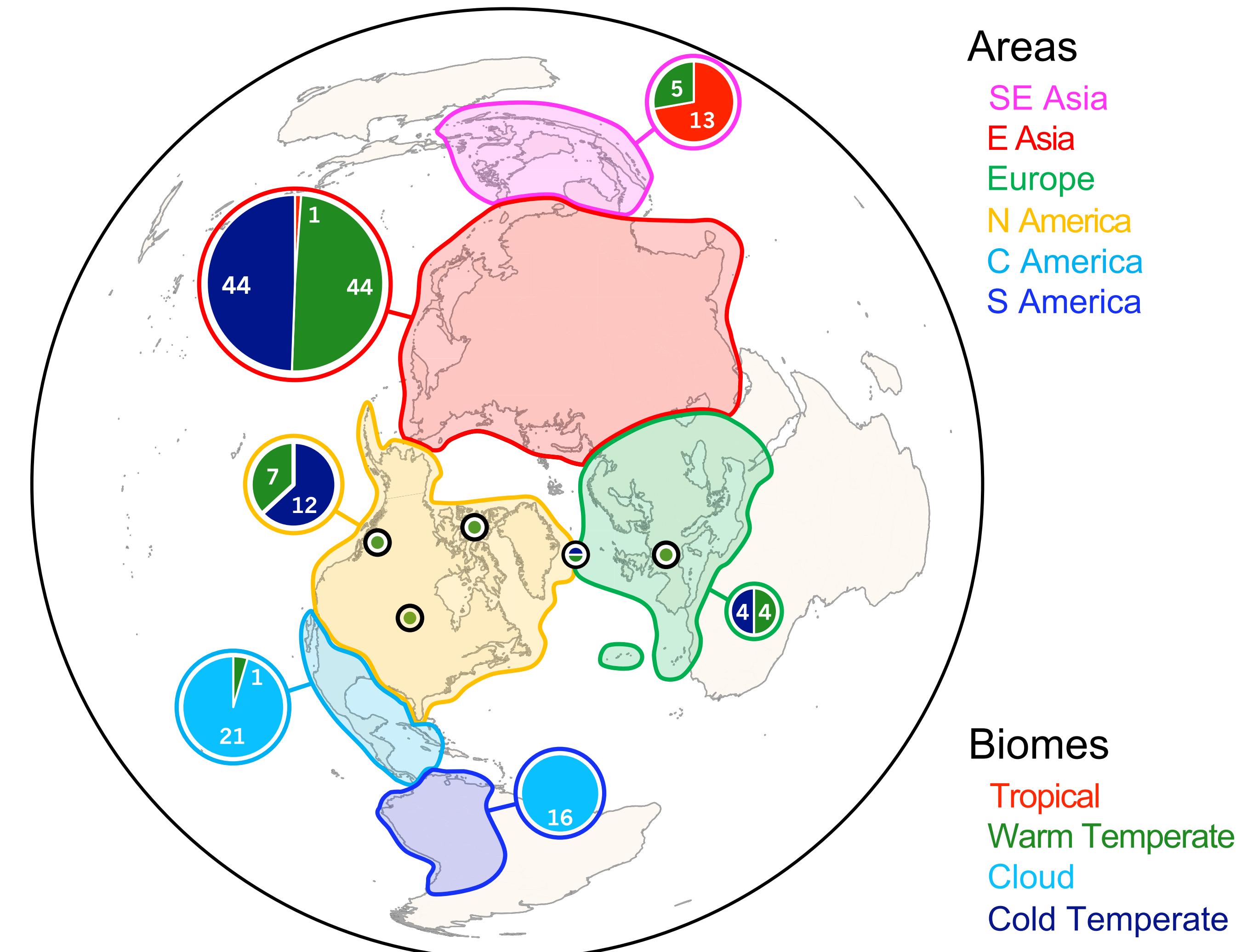
Data

163 extant species (127 with DNA)

5 fossils (with taxonomic constraints)

6 geographic areas

4 biomes



Areas

- SE Asia
- E Asia
- Europe
- N America
- C America
- S America

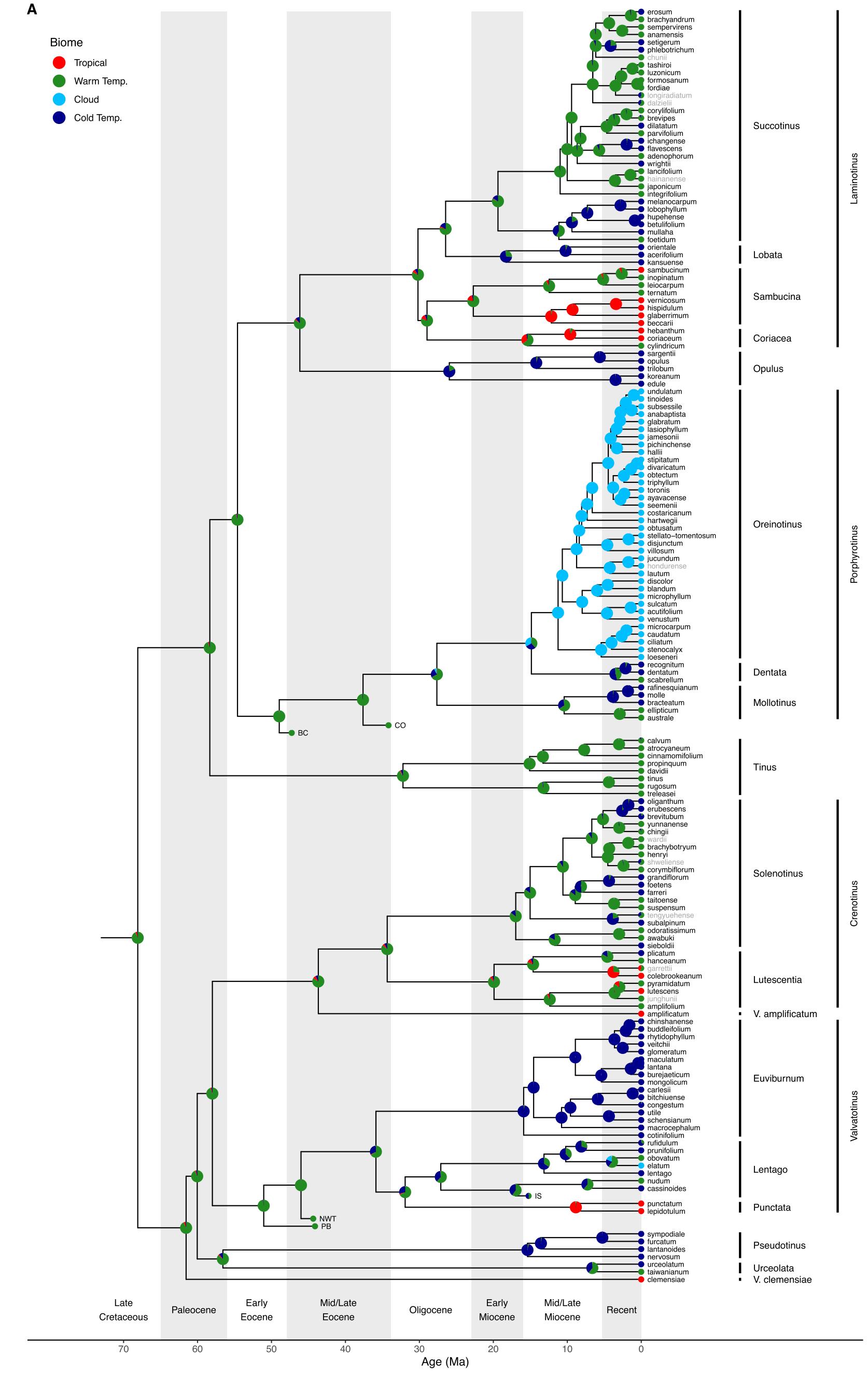
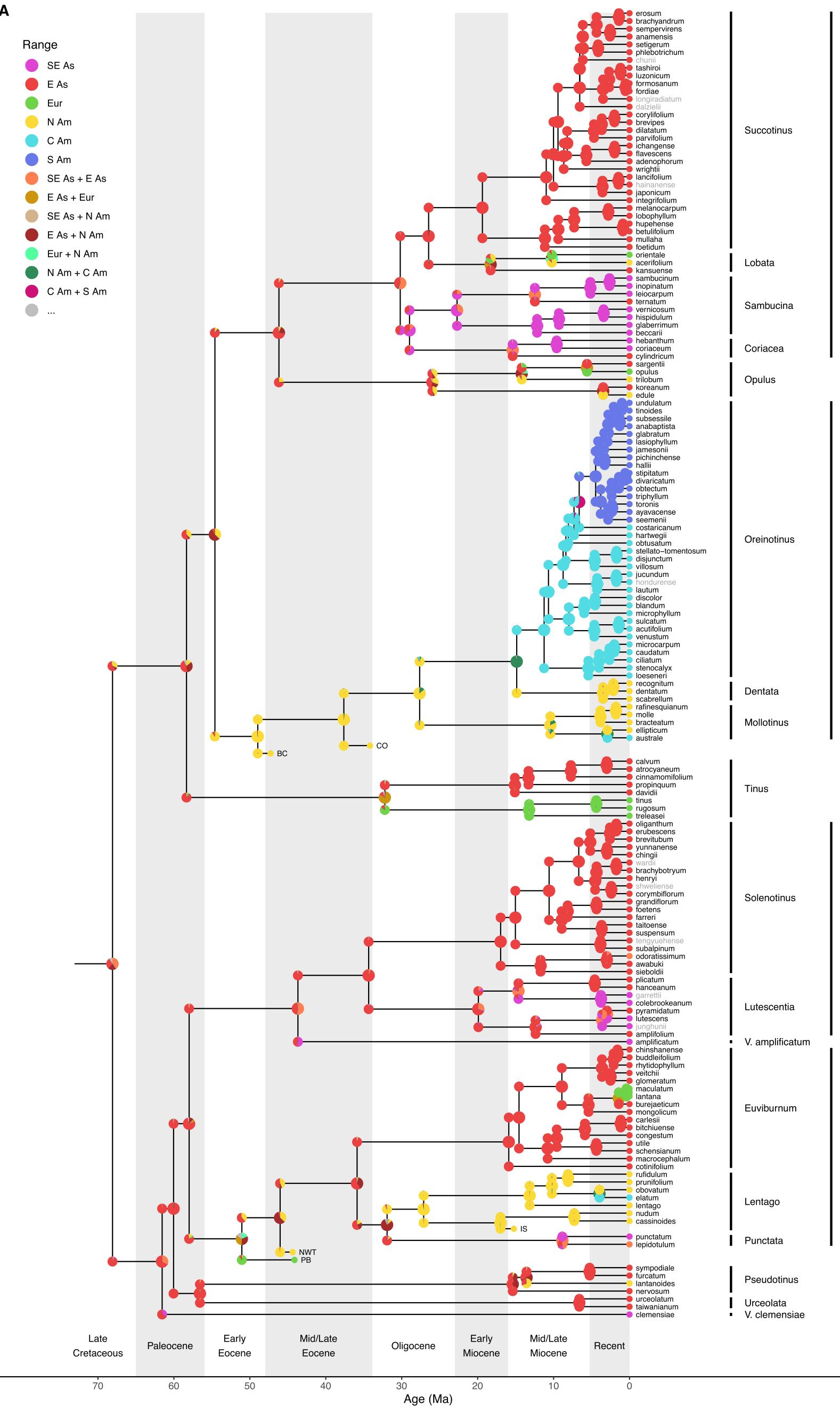
Biomes

- Tropical
- Warm Temperate
- Cloud
- Cold Temperate

- Can we integrate **geographic range** and **biome data** into analysis using the FBD model?
- What can we learn about the **diversification history** of the *Viburnum*?

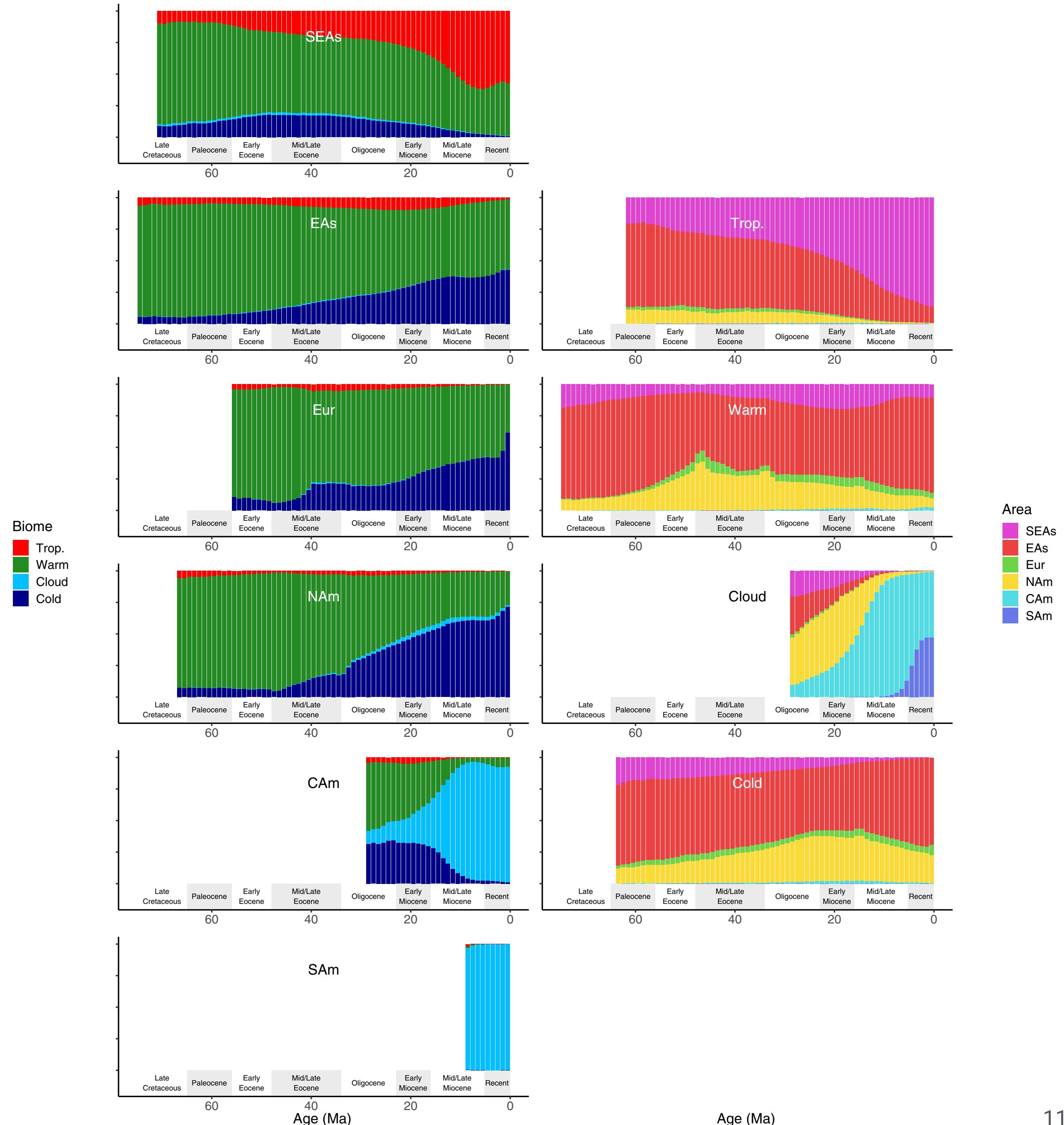
Analysis

1. Estimate the extant topology using maximum likelihood
2. Joint inference divergence times, biogeographic and biome history
(normally we *first* infer a dated tree, and *then* separately infer biogeographic history)
3. Ancestral state reconstruction
4. + various sensitivity analyses



Results summary

- Joint inference using the FBD and biogeographic models allows us to estimate a rich diversification history
- Major lineages of *Viburnum* likely originated in warm / temperate regions and later adapted to the cold
- Fossils can change the results



*“It is, it must be admitted, a **humbling** task to infer ancient events, and the results in many cases are tenuous at best. Given the obvious limitations of working with extant species and few, if any, fossils, **it is necessary to integrate all of the available sources of evidence** if we hope to produce assuring answers.”*

Landis et al. (2023) *Systematic Biology*
Joint phylogenetic estimation of geographic movements and biome shifts

Decoding Genomes: From Sequences to Phyldynamics

*Tanja Stadler, Carsten Magnus,
Timothy Vaughan, Joëlle Barido-Sottani,
Veronika Bošková, Jana S. Huisman,
Jūlija Pečerska*

Illustrated by Cecilia Valenzuela Agúí

Edited by Jūlija Pečerska

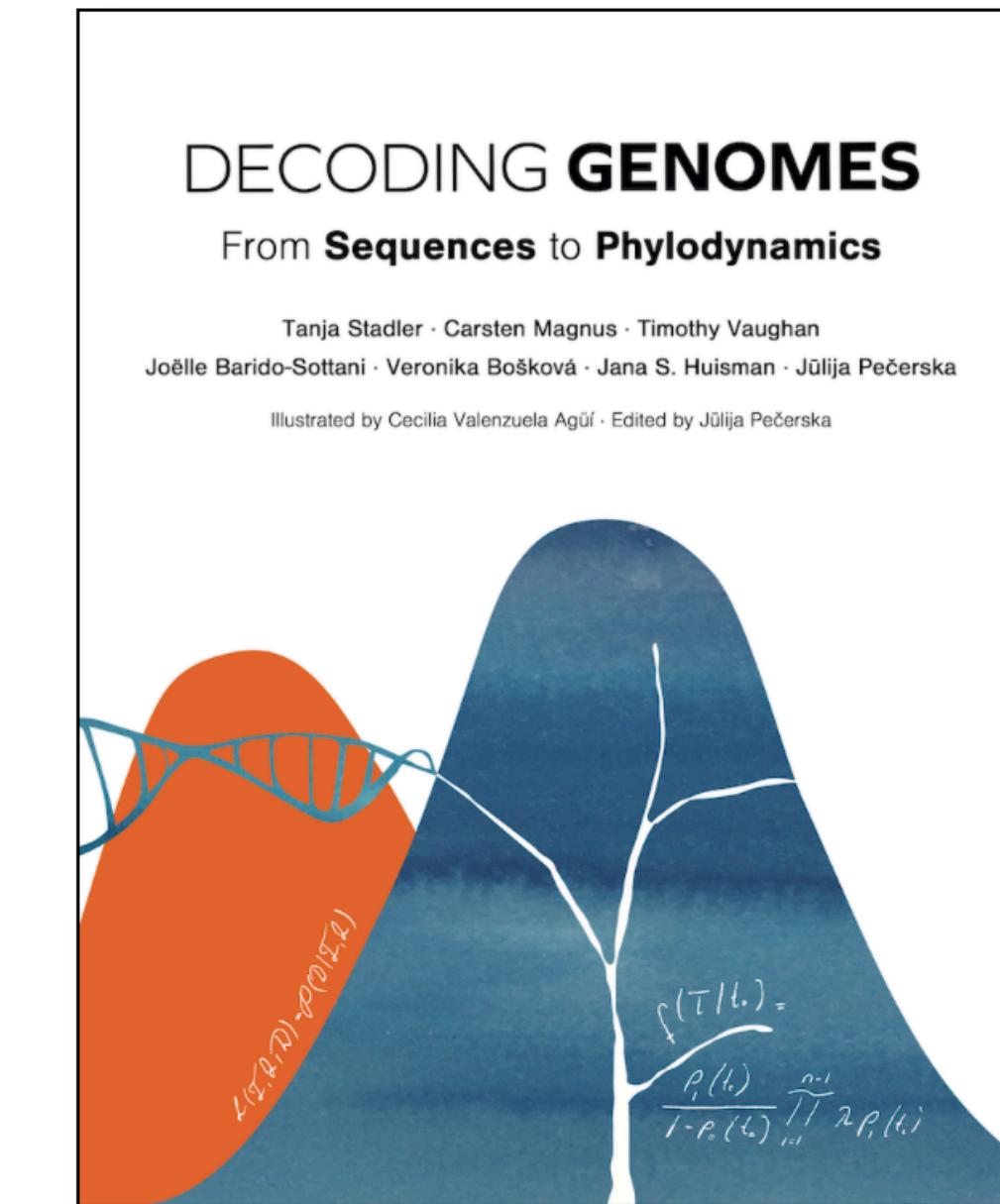
Obtaining the book

You will shortly be able to purchase a hard copy from Amazon. (Quality testing currently in progress.)

Alternatively, you can [Download](#) the complete PDF of the book free of charge. (See below for license information.)

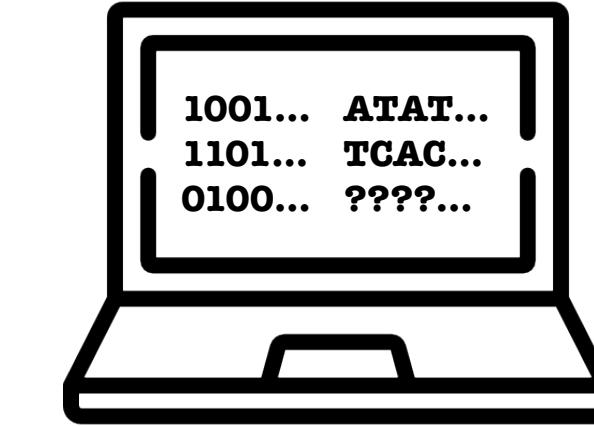
About the book

Decoding Genomes demonstrates how to uncover information about past evolutionary and population dynamic processes based on genomic samples. The last decades have seen considerable theoretical and methodological advances in this area. These enable the assessment of critical scientific questions such as the impact of environmental changes on biodiversity and the evolution of pathogens during recent epidemics. The book gives the reader a detailed understanding of the whole process: from genome sampling to obtaining biological insights by applying sophisticated statistical and computational analyses. In particular, sequencing of genomic samples, the alignment of sequences, molecular evolution models, phylogenetics, and phyldynamics are core topics. Statistical and computational approaches discussed include dynamic programming, maximum likelihood, Bayesian statistics, and model selection, to name a few. The concepts introduced and applied throughout the book enable readers to answer



A few notes on software

for Bayesian time tree estimation. All open source.



- MCMCTree – BDSS process, continuous trait models. Best option for large sequence alignments and trees. Requires a fixed tree. Language: C
- PhyloBayes – for extant time tree inference. Good for amino acid data. C++
- MrBayes – FBD model, some unique clock models. Easy to use. C++

For increased modularity & flexibility:



- BEAST2 – FBD model, lots of flexible tree and character evolution models. More widely used in epidemiology. Java. (Sister software BEAST 1.8)
- RevBayes – FBD model, lots of flexible tree and character evolution models. C++. Uses graphical models. Developed by folk closer to macroevolution



When do use different software?

Scenario	Software
Large datasets of extant taxa and node calibrations	MCMCTree
If want (or have to) fix the tree topology	MCMCTree
If fossil sampling is sparse or complex	MCMCTree
If you have abundant fossil data, or are interested in the topological position of fossils	BEAST2, RevBayes
If you're interested in the phylodynamic parameters	BEAST2, RevBayes
If you want to use a specific model	BEAST2, RevBayes, MCMCTree