Phylogenetics in R

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Introduction

This is an R Markdown document that will guide you through some basic R code for phylogenetic analysis.

Getting set up

```
setwd("~/Github/CCAP_course/main/examlpes")

library(seqinr) #to read in fasta files
library(msa) #for sequence alignments
library(phangorn) #to build phylogenies
library(ips)
```

Alignment

```
#fasta <- readDNAStringSet("~/Github/CCAP_course/main/examlpes/arb-silva
#conversion between formats
#fasta <- as.DNAbin(fasta)
#trimEnds(fasta)

# Import data [fasta.dna=https://pastebin.com/8Mt3QUTV]
#fasta <- read.dna("~/Github/CCAP_course/main/examlpes/arb-silva.de_2021

fasta <- as.DNAbin(read.alignment("~/Github/CCAP_course/main/examlpes/arfasta <- trimEnds(fasta)
fasta_phyDat <- phyDat(fasta, type = "DNA")</pre>
```

```
## Warning in phyDat.DNA(data, return.index = return.index, ...): Found
## characters. Deleted sites with with unknown states.
```

```
# Subset (first ten)
fasta10 <- subset(fasta_phyDat, 1:10)
fasta10_phyDat <- phyDat(fasta10, type = "DNA", levels = NULL)</pre>
```

model testing

One option is to convert the alignment into a pairwise distances, which will speed up the inference of a tree. We'll need to decide on the model of nucleotide evolution that best fits the data, performing a likelihood ratio test as implemented in modelTest.

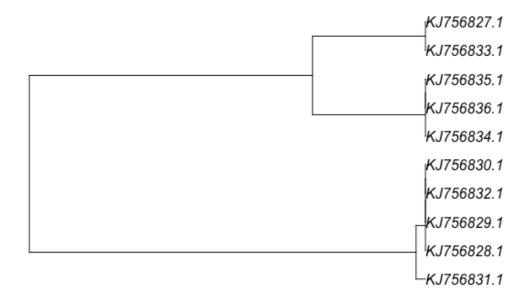
```
mt <- modelTest(fasta10)</pre>
## negative edges length changed to 0!
## [1] "JC+I"
## [1] "JC+G"
## [1] "JC+G+I"
## [1] "F81+I"
## [1] "F81+G"
## [1] "F81+G+I"
## [1] "K80+I"
## [1] "K80+G"
## [1] "K80+G+I"
## [1] "HKY+I"
## [1] "HKY+G"
## [1] "HKY+G+I"
## [1] "SYM+I"
## [1] "SYM+G"
## [1] "SYM+G+I"
## [1] "GTR+I"
## [1] "GTR+G"
## [1] "GTR+G+I"
print(mt)
```

```
AIC
                                                      AICc
##
        Model df
                    logLik
                                             AICw
                                                                    AICcw
           JC 17 -1849.785 3733.571 6.526902e-166 3733.586 6.552528e-166
## 1
## 2
         JC+I 17 -1849.785 3733.571 6.526903e-166 3733.586 6.552528e-166
## 3
         JC+G 18 -1849.785 3735.571 2.401010e-166 3735.587 2.408351e-166
       JC+G+I 18 -1849.785 3735.571 2.401010e-166 3735.587 2.408351e-166
## 4
## 5
          F81 20 -1468.953 2977.906 8.038422e-02 2977.927
                                                            8.047890e-02
        F81+I 20 -1468.953 2977.906 8.038423e-02 2977.927
                                                            8.047891e-02
## 6
        F81+G 21 -1468.953 2979.906 2.957079e-02 2979.929
## 7
                                                            2.957573e-02
## 8
     F81+G+I 21 -1468.953 2979.906 2.957079e-02 2979.929
                                                            2.957574e-02
## 9
          K80 18 -1846.292 3728.583 7.902693e-165 3728.600 7.926855e-165
        K80+I 18 -1846.292 3728.583 7.902694e-165 3728.600 7.926856e-165
## 10
        K80+G 19 -1846.292 3730.583 2.907157e-165 3730.602 2.913382e-165
## 11
## 12 K80+G+I 19 -1846.292 3730.583 2.907157e-165 3730.602 2.913382e-165
         HKY 21 -1466.718 2975.436 2.765137e-01 2975.458
                                                           2.765599e-01
## 13
## 14
        HKY+I 21 -1466.718 2975.436 2.765138e-01 2975.458
                                                            2.765600e-01
## 15
        HKY+G 22 -1466.718 2977.436 1.017215e-01 2977.460
                                                            1.016309e-01
## 16 HKY+G+I 22 -1466.718 2977.436 1.017215e-01 2977.460
                                                           1.016309e-01
          SYM 22 -1841.096 3726.191 2.613486e-164 3726.215 2.611158e-164
## 17
        SYM+I 22 -1841.096 3726.191 2.613486e-164 3726.215 2.611159e-164
## 18
        SYM+G 23 -1841.096 3728.191 9.614253e-165 3728.218 9.595070e-165
## 19
## 20 SYM+G+I 23 -1841.096 3728.191 9.614254e-165 3728.218 9.595071e-165
          GTR 25 -1466.184 2982.369 8.633666e-03 2982.400
                                                           8.596155e-03
## 21
## 22
        GTR+I 25 -1466.184 2982.369 8.633670e-03 2982.400
                                                            8.596159e-03
## 23
        GTR+G 26 -1466.184 2984.369 3.176088e-03 2984.402
                                                            3.158336e-03
## 24 GTR+G+I 26 -1466.184 2984.369 3.176089e-03 2984.402
                                                           3.158337e-03
dna_dist <- dist.ml(fasta10, model="JC")</pre>
```

```
Neighbor Joining and UPGMA We can estimate trees from distance matrices using neighbor-joining and UPGMA algorithms
```

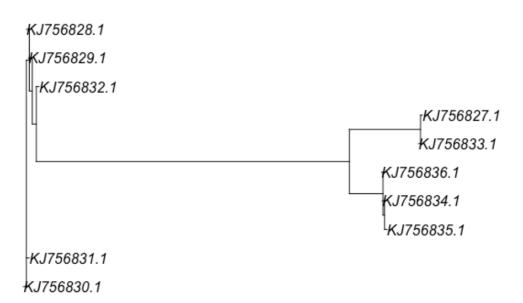
```
fasta_UPGMA <- upgma(dna_dist)
fasta_NJ <- NJ(dna_dist)
plot(fasta_UPGMA, main="UPGMA")</pre>
```

UPGMA



```
plot(fasta_NJ, main="NJ")
```

NJ



Which tree firts our data best? we can use parsimony() function to compare their respective parsimony scores. The function like optim.parsimony() and prachet() takes this a little further, and are worth exploring

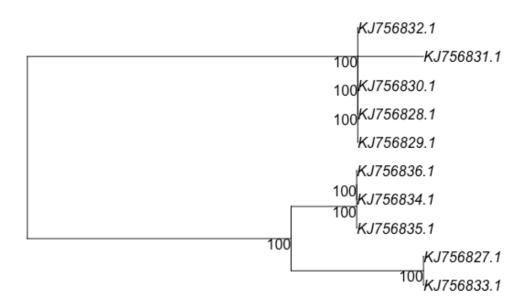
Maximum likelihood and bootstrapping

calculate the likelihood of a given tree using pml(), and use the function optim.pml() to optimize tree tolopogies and branch lengths for your selected model of nucleotide evolution

```
fit <- pml(fasta_NJ, fasta10)</pre>
## negative edges length changed to 0!
print(fit)
##
## loglikelihood: -1850.391
##
## unconstrained loglikelihood: -7555.037
##
## Rate matrix:
## acgt
## a 0 1 1 1
## c 1 0 1 1
## g 1 1 0 1
## t 1 1 1 0
##
## Base frequencies:
## 0.25 0.25 0.25 0.25
fitJC <- optim.pml(fit, model = "JC", rearrangement = "stochastic")</pre>
```

```
## optimize edge weights: -1850.391 --> -1849.785
## optimize edge weights: -1849.785 --> -1849.785
## optimize topology: -1849.785 --> -1849.785
## optimize topology: -1849.785 --> -1849.785
## optimize topology: -1849.785 --> -1849.785
## 2
## optimize edge weights: -1849.785 --> -1849.785
## optimize topology: -1849.785 --> -1849.785
## 0
## [1] "Ratchet iteration 1 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 2 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 3 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 4 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 5 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 6 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 7 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 8 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 9 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 10 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 11 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 12 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 13 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 14 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 15 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 16, best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 17, best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 18 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 19 , best pscore so far: -1849.78533514409"
## [1] "Ratchet iteration 20 , best pscore so far: -1849.78533514409"
## optimize edge weights: -1849.785 --> -1849.785
## optimize topology: -1849.785 --> -1849.785
## 0
## optimize edge weights: -1849.785 --> -1849.785
```

```
bs <- bootstrap.pml(fitJC, bs=100, control = pml.control(trace=0))
plotBS(midpoint(fitJC$tree), bs, p = 50, type="p")</pre>
```



Exporting trees

```
write.tree(bs, file="bootstrap_example.tre")
```