

MotilA – A Python pipeline for the analysis of microglial fine process motility in 3D time-lapse multiphoton microscopy data

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Summary

MotilA is a Python-based image analysis pipeline for quantifying fine process motility of microglia from 3D time-lapse two-channel fluorescence microscopy data. Developed for high-resolution multiphoton *in vivo* imaging datasets, *MotilA* enables both single-file and batch processing across multiple experimental conditions. It performs image preprocessing, segmentation, and motility quantification over time, using a pixel-based change detection strategy that yields biologically interpretable metrics such as the turnover rate (TOR) of microglial fine processes. While originally designed for microglial imaging, the pipeline can be extended to other cell types and imaging applications that require analysis of dynamic morphological changes. *MotilA* is openly available, platform-independent, and includes extensive documentation, tutorials, and example data to facilitate adoption by the broader scientific community. It is released under the GPL-3.0 open-source license.

Statement of need

Microglia are innate immune cells of the central nervous system and exhibit highly dynamic, motile processes that continuously scan their environment (M. Fuhrmann et al., 2010; Nebeling et al., 2023; Nimmerjahn et al., 2005; Prinz et al., 2019; Tremblay et al., 2010). Quantifying microglial motility at the level of fine processes is crucial for studying their function in health and disease, including neurodegeneration, inflammation, and synaptic remodeling. However, despite the biological importance of this analysis, there is currently no dedicated open-source tool tailored for this task.

To date, researchers typically quantify microglial motility manually (see., e.g., Nebeling et al. (2023)) using general-purpose image processing software such as Fiji/ImageJ (Schindelin et al., 2012) or ZEISS ZEN (Carl Zeiss Microscopy GmbH, Accessed 2025). While these approaches are well established in the field, they are time-consuming, lack reproducibility, and are not well suited for batch processing or cohort-level comparisons. They often focus on individual microglia, whereas *MotilA* enables analysis of the full field of view, allowing for more comprehensive and scalable quantification. Manual workflows are also more susceptible to human bias, limiting their scalability and objectivity (Brown, 2017; Lee et al., 2024; Misra et al., 2015; Wall et al., 2018).

MotilA addresses these limitations by providing an end-to-end, user-friendly, and batch-capable pipeline specifically designed for 3D time-lapse two-channel microscopy data. It supports standardized workflows for single- and multi-channel datasets, integrates essential preprocessing steps (registration, spectral unmixing, histogram normalization), and derives biologically meaningful motility metrics from binarized pixel dynamics. The method builds on

41 strategies used in prior studies but automates the workflow in a reproducible, scalable, and
42 open-source manner. *MotilA* thus fills a critical gap in neuroimaging analysis pipelines and is
43 particularly valuable for labs working with multiphoton *in vivo* imaging.

44 What does *MotilA* do?

45 *MotilA* is a modular and customizable image analysis pipeline written in Python that quantifies
46 microglial fine process motility from time-lapse fluorescence microscopy data, typically acquired
47 with two-photon (Denk et al., 1990; Helmchen & Denk, 2005) or three-photon *in vivo* imaging
48 (F. Fuhrmann et al., 2024; Horton et al., 2013). Although it was originally developed for
49 microglial analysis, the pipeline is adaptable to other cell types and imaging contexts involving
50 dynamic morphological changes over time.

51 At its core, *MotilA* extracts sub-volumes from 3D time-stacks, performs 2D maximum intensity
52 projections, and segments the resulting images to classify pixel-wise changes in microglial
53 morphology. These changes are quantified frame-by-frame and used to calculate biologically
54 interpretable metrics, including the turnover rate (TOR). The design is tailored to biological
55 imaging data, with particular attention to typical issues such as z-axis projection loss, channel
56 bleed-through, motion artifacts, photobleaching, and signal heterogeneity.

57 The pipeline supports both single-file processing and large-scale batch analysis. Parameters are
58 highly customizable either programmatically or via metadata files, and all results are automati-
59 cally logged, saved, and summarized for downstream statistical analysis. The outputs include
60 segmented image series, intermediate diagnostics (e.g. histograms, projections, brightness
61 traces), and Excel spreadsheets with motility metrics.

62 To accommodate large-scale, high-resolution imaging data, *MotilA* supports memory-efficient
63 file handling via the Zarr format (Miles et al., 2025), enabling processing of large TIFF files
64 using memory mapping to avoid RAM overload.

65 *MotilA* can be run via Python scripts or Jupyter notebooks, and it includes extensive docu-
66 mentation, examples, and a tutorial dataset to make onboarding straightforward.

67 We welcome community contributions and issue reports via the GitHub repository: <https://github.com/FabrizioMusacchio/motila>.
68

69 How is “motility” determined?

70 *MotilA* quantifies motility by analyzing pixel-wise changes in microglial fine processes’ morphol-
71 ogy over time. The pipeline first extracts a sub-volume around a user-defined z-axis center from
72 each 3D image stack and applies a 2D maximum intensity projection to reduce dimensionality.
73 Although this sacrifices some z-axis information, it enables efficient segmentation and pixel-level
74 tracking while maintaining biological interpretability.

75 At each time point t_i , the projected and binarized image $B(t_i)$ is compared to the next time
76 point $B(t_{i+1})$. A temporal variation map $\Delta B(t_i)$ is computed as:

$$\Delta B(t_i) = 2 \times B(t_i) - B(t_{i+1})$$

77 Based on this difference image, each pixel is classified as:

- 78 ■ **Stable (S)** if $\Delta B = 1$
- 79 ■ **Gained (G)** if $\Delta B = -1$
- 80 ■ **Lost (L)** if $\Delta B = 2$

81 From these categories, *MotilA* calculates the microglial **fine process turnover rate (TOR)**, a
82 central metric representing the fraction of pixels that changed:

$$TOR = \frac{G + L}{S + G + L}$$

This approach allows for a quantitative assessment of microglial process dynamics at each time point and across the full recording session. The same principle can be extended to other motile cell types or dynamic cellular structures where morphological changes manifest as gain or loss of segmented pixels over time.

The implementation is based on analytical strategies described in previous studies such as M. Fuhrmann et al. (2010) and Nebeling et al. (2023), with added flexibility for batch processing, filtering, and parameter tuning.

Key features

MotilA offers a combination of modularity, reproducibility, and scalability specifically tailored to motility analysis in multiphoton *in vivo* imaging. Its key features include:

- **Automated preprocessing pipeline**
Includes optional steps for image registration (2D and 3D), spectral unmixing, histogram equalization for contrast enhancement within time points, and histogram matching for brightness normalization across time points (e.g. to correct for photobleaching), as well as noise reduction via median and Gaussian filtering.
- **Flexible segmentation and thresholding**
Supports multiple adaptive thresholding methods (e.g. Otsu, Li, Triangle) and customizable blob detection settings to isolate fine microglial processes or similar structures.
- **Pixel-based motility quantification**
Tracks pixel-wise changes between time points to classify stable, gained, and lost pixels, allowing biologically interpretable metrics like the turnover rate (TOR).
- **Batch processing capabilities**
Enables large-scale processing of multiple datasets with a standardized folder structure and parameter metadata sheets, suitable for cohort-level studies.
- **User-defined projection settings**
Allows flexible extraction of sub-volumes and z-projection around multiple centers to avoid overlapping cells and vascular artifacts.
- **Memory-efficient file handling**
Supports memory mapping of large TIFF files via the Zarr format, enabling efficient processing of high-resolution time-lapse datasets without exhausting system RAM.
- **Metadata integration and parameter logging**
Automatically reads per-dataset settings from metadata files (e.g. Excel sheets), and stores processing parameters and outputs in structured result folders.
- **Cross-platform compatibility**
Runs on Windows, macOS, and Linux, tested with Python 3.9 and compatible with common scientific computing environments via Conda.
- **Tutorials and example data included**
Comes with Jupyter notebooks, example datasets, and clear documentation to help new users get started quickly.

Pipeline steps

The *MotilA* pipeline follows a modular sequence of image processing and analysis steps designed for robust and reproducible quantification of motility from multi-dimensional imaging data. It supports both single-file and batch workflows and includes options for fine-grained customization at each step.

127 Core pipeline steps

128 For single datasets, *MotilA* executes the following sequence (Figure 1a)):

- 129 1. **Load image data**
130 Supports TIFF files (Gohlke, 2025) in TZCYX (multi-channel) or TZYX (single-channel)
131 format, following ImageJ/Fiji conventions (T: time, Z: z-axis, C: channel, Y: height, X:
132 width).
- 133 2. **Extract sub-volumes**
134 Selects a z-stack around a projection center for each time point, allowing focused analysis
135 and optional multiple projections per stack.
- 136 3. **(Optional) Register sub-volumes**
137 Applies 3D motion correction (Anuta, 1970; Guizar-Sicairos et al., 2008; Kuglin &
138 Hines, 1975) to each time series stack using user-defined template strategies (e.g. mean,
139 median).
- 140 4. **(Optional) Perform spectral unmixing**
141 Removes signal bleed-through between channels, especially relevant for two-channel
142 imaging setups.
- 143 5. **Z-projection**
144 Projects each 3D sub-volume into a 2D image via maximum intensity projection to
145 simplify segmentation and speed up processing.
- 146 6. **(Optional) Register projections**
147 Aligns the 2D projections across time to correct for lateral motion artifacts.
- 148 7. **(Optional) Apply histogram equalization**
149 Enhances local contrast within each projection using contrast-limited adaptive histogram
150 equalization (CLAHE) (Pizer et al., 1987; Walt et al., 2014).
- 151 8. **(Optional) Apply histogram matching**
152 Normalizes brightness across time points to mitigate bleaching effects or intensity drift
153 (Walt et al., 2014).
- 154 9. **(Optional) Apply filtering**
155 Reduces noise with optional median filtering (square or circular kernel) and/or Gaussian
156 smoothing (Harris et al., 2020; Virtanen et al., 2020).
- 157 10. **Segment microglial processes**
158 Applies adaptive thresholding (Glasbey, 1993; Li & Tam, 1998; Otsu, 1979; Prewitt &
159 Mendelsohn, 1966; Ridler et al., 1978; Yen et al., 1995; Zack et al., 1977) and blob
160 filtering (Fiorio & Gustedt, 1996; Walt et al., 2014; Wu et al., 2005) to identify and
161 isolate morphologically relevant structures.
- 162 11. **Analyze motility**
163 Quantifies pixel-level changes over time to classify stable, gained, and lost regions, from
164 which motility metrics are derived.

165 All intermediate outputs and metrics are saved for validation and further analysis.

166 Batch processing steps

167 *MotilA* supports fully automated batch processing using a standardized folder structure and
168 Excel-based metadata configuration. This enables reproducible cohort-level analysis across
169 many animals or experimental conditions.

- 170 1. **Define a project folder**
171 Each dataset is placed in an ID-specific subdirectory, containing imaging files, metadata,
172 and optional result directories.
- 173 2. **Run the batch process**
174 The core pipeline is executed for each dataset using shared or per-dataset parameters
175 defined in metadata.xls.
- 176 3. **Save results**
177 Segmentation outputs, projections, and motility metrics are stored in structured result

178 folders for each dataset.

179 4. Batch-collect metrics

180 Aggregates metrics across datasets into cohort-level Excel files for downstream statistical
181 analysis.

182 This design enables large-scale, reproducible quantification of microglial motility with minimal
183 manual intervention.

184 Assessing results and analyzing outputs

185 *MotilA* provides rich output in the form of diagnostic plots, intermediate image files, and
186 structured Excel tables to support both per-dataset assessment and cohort-level statistical
187 analysis.

188 Per-dataset assessment

189 For each processed image stack, *MotilA* generates:

- 190 ■ **Segmented images and overlays** showing gained, lost, and stable regions across time
191 points.
- 192 ■ **Histogram plots** for brightness, pixel area, and thresholding diagnostics.
- 193 ■ **Motility metrics table (*motility.xlsx*)** containing:
194 – Gained pixels (G)
195 – Lost pixels (L)
196 – Stable pixels (S)
197 – turnover rate (TOR) per time point
- 198 ■ **Brightness metrics (*brightness.xlsx*)** tracking average pixel intensity over time.
- 199 ■ **Cell area metrics (*cell_pixel_area.xlsx*)** reporting the segmented microglial pixel area
200 per time point.

201 These outputs help assess segmentation quality, evaluate photobleaching or signal loss, and
202 refine preprocessing parameters as needed.

203 Cohort-level batch analysis

204 During batch processing, *MotilA* can aggregate key metrics from all datasets into shared
205 summary files, including:

- 206 ■ *all_motility.xlsx* — All G/L/S/TOR metrics across datasets.
- 207 ■ *all_brightness.xlsx* — Mean brightness per dataset and time point.
- 208 ■ *all_cell_pixel_area.xlsx* — Segmented area per dataset and time point.
- 209 ■ *average_motility.xlsx* — Dataset-wise average motility metrics across the full record-
210 ing.

211 These results allow for statistical comparison of motility dynamics across experimental conditions
212 and facilitate downstream visualization and modeling in tools like Python, R, or Excel.

213 This multi-level output strategy ensures both technical validation and biological insight, making
214 *MotilA* suitable for both exploratory and hypothesis-driven studies.

215 Main functions

216 The three main entry points for the pipeline are:

- 217 ■ *process_stack* — Processes a single image stack, performing the full pipeline.
- 218 ■ *batch_process_stacks* — Executes the pipeline across multiple datasets in a project
219 folder.
- 220 ■ *batch_collect* — Gathers motility metrics from all datasets for cohort-level analysis.

221 Each function supports extensive parameterization via arguments or metadata files.

222 A complete overview of configurable parameters for single-file processing, batch workflows, and
223 image enhancement is provided in the [MotiLA README](#).

224 Useful helper functions

225 Several additional functions assist with data preparation and quality control, including:

- 226 ▪ `tiff_axes_check_and_correct` — Automatically adjusts TIFF axis order to TZCYX/
227 TZYX if needed.
- 228 ▪ `hello_world` — Verifies a successful import of the *MotiLA* module.
- 229 ▪ `logger_object` — Initializes logging for the current analysis session.

230 Applications and limitations

231 *MotiLA* was designed with a primary focus on the analysis of microglial fine process motility
232 *in vivo*, using high-resolution 3D time-lapse two-channel fluorescence microscopy data. Its
233 modular design and general image processing framework, however, make it applicable to a
234 broader range of dynamic imaging contexts.

235 Applications

- 236 ▪ **Microglial dynamics**
237 Quantification of process turnover during surveillance, neuroinflammation, or disease
238 models such as neurodegeneration and injury.
- 239 ▪ **Neuronal structural plasticity**
240 While *MotiLA* is optimized for microglial processes, its pixel-based change detection
241 framework can in principle be adapted to analyze dynamic changes in dendrites or axons
242 — such as growth, retraction, or remodeling — provided the structures can be reliably
243 segmented across time.
- 244 ▪ **Two-channel *in vivo* imaging**
245 Effective for experiments involving simultaneous imaging of microglia and neurons (e.g.,
246 Cx3Cr1-GFP with Thy1-YFP), with spectral unmixing to reduce bleed-through from
247 overlapping channels or fluorophores.
- 248 ▪ **Cohort-level studies**
249 Designed to analyze and compare motility metrics across large experimental groups,
250 enabling high-throughput, statistically robust results.
- 251 ▪ **Teaching and prototyping**
252 The example datasets and tutorials make *MotiLA* a useful tool for training purposes or
253 prototyping new analysis approaches.

254 Limitations

- 255 ▪ **Loss of z-resolution**
256 The use of 2D maximum intensity projections simplifies processing but sacrifices z-axis
257 information. This may lead to overlapping structures and limits spatial specificity.
- 258 ▪ **Segmentation-dependent**
259 Accuracy depends on appropriate thresholding and image quality. Overlapping processes,
260 blood vessels, or low signal-to-noise ratios can reduce segmentation performance.
- 261 ▪ **Limited spectral unmixing**
262 The current unmixing approach is a simple channel subtraction. More advanced unmixing
263 strategies may be required for some experimental setups.
- 264 ▪ **Not a general-purpose tracking tool**
265 *MotiLA* is optimized for pixel-level process motility, not for full cell tracking or object-based
266 morphological quantification over time.

267 ▪ **Assumes TIFF input with standardized axis order**
268 Input images must conform to TZCYX or TZYX structure; other formats require
269 conversion.

270 Despite these limitations, *MotilA* provides a powerful, reproducible framework for analyzing
271 microglial motility and similar biological processes, especially in experimental setups where
272 manual analysis would be impractical.

273 Real-world example

274 To demonstrate its practical utility, *MotilA* includes a fully compatible example dataset of *in*
275 *vivo* two-photon time-lapse imaging stacks from the mouse frontal cortex (Gockel et al., 2025).
276 These data were acquired to assess microglial fine process motility under control conditions
277 and during complement C4 overexpression, a genetic risk factor for schizophrenia.

278 The dataset contains two 5D TIFF stacks with the following structure:

- 279 ▪ **T**: 8 time points (5-minute intervals over 35 minutes)
- 280 ▪ **C**: 2 imaging channels (Cx3cr1-GFP for microglia, tdTomato for neurons)
- 281 ▪ **Z**: ~60 optical sections (1 μm step size)
- 282 ▪ **Y, X**: $\sim 1200 \times 1200$ px ($\sim 125 \times 125 \mu\text{m}^2$ field of view)

283 The files are formatted for direct use with *MotilA*, requiring no manual reorganization or
284 preprocessing.

285 **Figure 1** summarizes the full *MotilA* workflow as applied to the example dataset. For visu-
286 alization purposes, the original full-field dataset was cropped around a single microglial cell
287 to reduce background clutter and allow detailed inspection of each processing step. **Panel a)**
288 outlines the core and optional steps in the processing pipeline. **Panel b)** shows z-projections of
289 the microglial cell at time points t_0 and t_1 , including raw data, contrast enhancement, and
290 filtering prior to segmentation. **Panel c)** displays the delta image used for motility quantification
291 and the corresponding pixel-wise classification into stable (S, blue), gained (G, green), and lost
292 (L, red) pixels. **Panel d)** tracks the average brightness of the segmented cell relative to the
293 first time point, which helps assess signal stability and potential bleaching. **Panel e)** presents
294 the turnover rate (TOR) across all time points, capturing the dynamics of microglial process
295 remodeling.



Figure 1: Step-by-step illustration of the *MotiLA* pipeline using the included test dataset. **a)** Overview of the image processing pipeline, showing core and optional steps. **b)** Example projections of a cropped microglial cell at time points t_0 and t_1 , including raw, histogram-equalized, filtered (median and Gaussian), and binarized versions. **c)** Binarized pixel-wise comparison between t_0 and t_1 , with classification into stable (S, blue), gained (G, green), lost (L, red), and background (BG, white) pixels, along with the corresponding pixel statistics. **d)** Normalized cell brightness over time, relative to t_0 , used to assess bleaching and signal stability. **e)** Turnover rate (TOR) plotted across all time points for the same cell, representing process-level motility dynamics. All microglial image panels in **b** and **c** are shown at the same scale. Scale bar in the top-left image of panel **b** represents 10 μm .

Past and ongoing projects

MotilA has already been successfully applied in multiple neuroscience studies involving *in vivo* imaging of microglia and neurons in the mouse brain.

The following published and preprint works used *MotilA* to analyze fine process motility in physiological and pathological contexts:

- **Crux et al. (2024)**
Investigated the role of actin depolymerizing factors ADF/Cofilin1 in microglial motility and memory formation. *MotilA* was used to quantify reduced motility in knockout mice.
→ <https://doi.org/10.1101/2024.09.27.615114>
- **F. Fuhrmann et al. (2024)**
Employed deep three-photon imaging of microglia in the medial prefrontal cortex to measure sub-cellular process dynamics in awake mice. *MotilA* was used to quantify microglial turnover at depths beyond 1 mm.
→ <https://doi.org/10.1101/2024.08.28.610026>
- **Gockel et al. (2025)**
Generated and published the example dataset accompanying this pipeline, which was used to demonstrate microglial motility changes in response to complement C4 overexpression.
→ <https://doi.org/10.5281/zenodo.15061566>

These studies showcase the pipeline's suitability for both targeted microglial investigations and large-scale, high-resolution imaging projects. Ongoing work continues to extend *MotilA*'s application to additional brain regions, genetic perturbations, and imaging modalities, including multi-channel and high-speed two-photon datasets.

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All animal procedures related to the example dataset were conducted in compliance with institutional, national, and international regulations. Experiments were approved by the relevant animal care and use committees at DZNE (Germany), following guidelines equivalent to the ARRIVE 2.0 framework. All efforts were made to reduce the number of animals used and to refine experimental conditions in accordance with the 3Rs (Replacement, Reduction, and Refinement) principles.

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References

Anuta, P. E. (1970). Spatial registration of multispectral and multitemporal digital imagery using fast fourier transform techniques. *IEEE Transactions on Geoscience Electronics*, 8(4),

- 353–368. <https://doi.org/10.1109/TGE.1970.271435>
- 342 Brown, D. L. (2017). Bias in image analysis and its solution: Unbiased stereology. *Journal of*
343 *Toxicologic Pathology*, 30(3), 183–191. <https://doi.org/10.1293/tox.2017-0013>
- 344 Carl Zeiss Microscopy GmbH. (Accessed 2025). *ZEISS ZEN Microscopy Software*. <https://www.zeiss.com/metrology/en/software/zeiss-zen-core.html>.
345
- 346 Crux, S., Roggan, M. D., Poll, S., Nebeling, F. C., Schiweck, J., Mittag, M., Musacchio, F.,
347 Steffen, J., Wolff, K. M., Baral, A., Witke, W., Gurniak, C., Bradke, F., & Fuhrmann,
348 M. (2024). Deficiency of actin depolymerizing factors ADF/Cfl1 in microglia decreases
349 motility and impairs memory. *bioRxiv*. <https://doi.org/10.1101/2024.09.27.615114>
- 350 Denk, W., Strickler, J. H., & Webb, W. W. (1990). Two-photon laser scanning fluorescence
351 microscopy. *Science*, 248(4951), 73–76. <https://doi.org/10.1126/science.2321027>
- 352 Fiorio, C., & Gustedt, J. (1996). Two linear time union-find strategies for image processing.
353 *Theoretical Computer Science*, 154, 165–181. [https://doi.org/10.1016/0304-3975\(94\)](https://doi.org/10.1016/0304-3975(94)00262-2)
354 [00262-2](https://doi.org/10.1016/0304-3975(94)00262-2)
- 355 Fuhrmann, F., Nebeling, F. C., Musacchio, F., Mittag, M., Poll, S., Müller, M., Giovannetti, E.
356 A., Maibach, M., Schaffran, B., Burnside, E., Chan, I. C. W., Lagurin, A. S., Reichenbach,
357 N., Kaushalya, S., Fried, H., Linden, S., Petzold, G. C., Tivosanis, G., Bradke, F., &
358 Fuhrmann, M. (2024). Three-photon in vivo imaging of neurons and glia in the medial
359 prefrontal cortex with sub-cellular resolution. *bioRxiv*. [https://doi.org/10.1101/2024.08.](https://doi.org/10.1101/2024.08.28.610026)
360 [28.610026](https://doi.org/10.1101/2024.08.28.610026)
- 361 Fuhrmann, M., Bittner, T., Jung, C. K. E., Burgold, S., Page, R. M., Mitteregger, G., Haass,
362 C., LaFerla, F. M., Kretschmar, H., & Herms, J. (2010). Microglial Cx3cr1 knockout
363 prevents neuron loss in a mouse model of alzheimer's disease. *Nature Neuroscience*, 13(4),
364 411–413. <https://doi.org/10.1038/nn.2511>
- 365 Glasbey, C. A. (1993). An analysis of histogram-based thresholding algorithms. *CVGIP:*
366 *Graphical Models and Image Processing*, 55(6), 532–537. [https://doi.org/10.1006/cgip.](https://doi.org/10.1006/cgip.1993.1040)
367 [1993.1040](https://doi.org/10.1006/cgip.1993.1040)
- 368 Gockel, N., Nieves-Rivera, N., Druart, M., Jaako, K., Fuhrmann, F., Rožkalne, R., Musacchio,
369 F., Poll, S., Jansone, B., Fuhrmann, M., & Magueresse, C. L. (2025). *Example datasets for*
370 *microglial motility analysis using the MotiLA pipeline*. Zenodo. [https://doi.org/10.5281/](https://doi.org/10.5281/zenodo.15061566)
371 [zenodo.15061566](https://doi.org/10.5281/zenodo.15061566)
- 372 Gohlke, C. (2025). *TiffFile: Read and write image data from and to TIFF files* (Version
373 2025.3.13). Zenodo. <https://doi.org/10.5281/zenodo.6795860>
- 374 Guizar-Sicairos, M., Thurman, S. T., & Fienup, J. R. (2008). Efficient subpixel image
375 registration algorithms. *Opt. Lett.*, 33(2), 156–158. [https://doi.org/10.1364/OL.33.](https://doi.org/10.1364/OL.33.000156)
376 [000156](https://doi.org/10.1364/OL.33.000156)
- 377 Harris, C. R., Millman, K. J., Walt, S. J. van der, Gommers, R., Virtanen, P., Cournapeau, D.,
378 Wieser, E., Taylor, J., Berg, S., Smith, N. J., Kern, R., Picus, M., Hoyer, S., Kerkwijk,
379 M. H. van, Brett, M., Haldane, A., Río, J. F. del, Wiebe, M., Peterson, P., ... Oliphant,
380 T. E. (2020). Array programming with NumPy. *Nature*, 585(7825), 357–362. <https://doi.org/10.1038/s41586-020-2649-2>
381
- 382 Helmchen, F., & Denk, W. (2005). Deep tissue two-photon microscopy. *Nature Methods*,
383 2(12), 932–940. <https://doi.org/10.1038/nmeth818>
- 384 Horton, N. G., Wang, K., Kobat, D., Clark, C. G., Wise, F. W., Schaffer, C. B., & Xu, C.
385 (2013). In vivo three-photon microscopy of subcortical structures within an intact mouse
386 brain. *Nature Photonics*, 7(3), 205–209. <https://doi.org/10.1038/nphoton.2012.336>
- 387 Kuglin, C. D., & Hines, D. C. (1975). The phase correlation image alignment method.

- 388 *Proceedings of the IEEE International Conference on Cybernetics and Society*, 163–165.
- 389 Lee, R. M., Eisenman, L. R., Khuon, S., Aaron, J. S., & Chew, T.-L. (2024). Believing is
390 seeing – the deceptive influence of bias in quantitative microscopy. *Journal of Cell Science*,
391 137(1), jcs261567. <https://doi.org/10.1242/jcs.261567>
- 392 Li, C. H., & Tam, P. K. S. (1998). An iterative algorithm for minimum cross entropy
393 thresholding. *Pattern Recognition Letters*, 18(8), 771–776. [https://doi.org/10.1016/](https://doi.org/10.1016/S0167-8655(98)00057-9)
394 [S0167-8655\(98\)00057-9](https://doi.org/10.1016/S0167-8655(98)00057-9)
- 395 Miles, A., jakirkham, Hamman, J., Orfanos, D. P., Stansby, D., Bussonnier, M., Moore,
396 J., Bennett, D., Augspurger, T., Rzepka, N., Cherian, D., Verma, S., Bourbeau, J.,
397 Fulton, A., Abernathey, R., Lee, G., Spitz, H., Kristensen, M. R. B., Jones, M., &
398 Schut, V. (2025). *Zarr-developers/zarr-python: v3.0.6* (Version v3.0.6). Zenodo. <https://doi.org/10.5281/zenodo.3773449>
- 400 Misra, I., Zitnick, C. L., Mitchell, M., & Girshick, R. B. (2015). Seeing through the human
401 reporting bias: Visual classifiers from noisy human-centric labels. *2016 IEEE Conference*
402 *on Computer Vision and Pattern Recognition (CVPR)*, 2930–2939. [https://doi.org/10.](https://doi.org/10.1109/cvpr.2016.320)
403 [1109/cvpr.2016.320](https://doi.org/10.1109/cvpr.2016.320)
- 404 Nebeling, F. C., Poll, S., Justus, L. C., Steffen, J., Keppler, K., Mittag, M., & Fuhrmann,
405 M. (2023). Microglial motility is modulated by neuronal activity and correlates with
406 dendritic spine plasticity in the hippocampus of awake mice. *eLife*, 12, e83176. <https://doi.org/10.7554/eLife.83176>
- 407
- 408 Nimmerjahn, A., Kirchhoff, F., & Helmchen, F. (2005). Resting microglial cells are highly
409 dynamic surveillants of brain parenchyma in vivo. *Science*, 308(5726), 1314–1318. <https://doi.org/10.1126/science.1110647>
- 410
- 411 Otsu, N. (1979). A threshold selection method from gray-level histograms. *IEEE Transactions*
412 *on Systems, Man, and Cybernetics*, 9(1), 62–66. [https://doi.org/10.1109/TSMC.1979.](https://doi.org/10.1109/TSMC.1979.4310076)
413 [4310076](https://doi.org/10.1109/TSMC.1979.4310076)
- 414 Pizer, S. M., Amburn, E. P., Austin, J. D., Cromartie, R., Geselowitz, A., Greer, T., Haar
415 Romeny, B. ter, Zimmerman, J. B., & Zuiderveld, K. (1987). Adaptive histogram equaliza-
416 tion and its variations. *Computer Vision, Graphics, and Image Processing*, 39(3), 355–368.
417 [https://doi.org/10.1016/S0734-189X\(87\)80186-X](https://doi.org/10.1016/S0734-189X(87)80186-X)
- 418 Prewitt, J. M. S., & Mendelsohn, M. L. (1966). The analysis of cell images. *Annals of the*
419 *New York Academy of Sciences*, 128(3), 1035–1053. [https://doi.org/10.1111/j.1749-6632.](https://doi.org/10.1111/j.1749-6632.1965.tb11715.x)
420 [1965.tb11715.x](https://doi.org/10.1111/j.1749-6632.1965.tb11715.x)
- 421 Prinz, M., Jung, S., & Priller, J. (2019). Microglia biology: One century of evolving concepts.
422 *Cell*, 179(2), 292–311. <https://doi.org/10.1016/j.cell.2019.08.053>
- 423 Ridler, T. W., Calvard, S., & name, F. (1978). Picture thresholding using an iterative
424 selection method. *IEEE Transactions on Systems, Man, and Cybernetics*, 8(8), 630–632.
425 <https://doi.org/10.1109/TSMC.1978.4310039>
- 426 Schindelin, J., Arganda-Carreras, I., Frise, E., Kaynig, V., Longair, M., Pietzsch, T., Preibisch,
427 S., Rueden, C., Saalfeld, S., Schmid, B., & others. (2012). Fiji: An open-source platform
428 for biological-image analysis. *Nature Methods*, 9(7), 676–682. [https://doi.org/10.1038/](https://doi.org/10.1038/nmeth.2019)
429 [nmeth.2019](https://doi.org/10.1038/nmeth.2019)
- 430 Tremblay, M.-È., Lowery, R. L., & Majewska, A. K. (2010). Microglial interactions with
431 synapses are modulated by visual experience. *PLOS Biology*, 8(11), 1–16. <https://doi.org/10.1371/journal.pbio.1000527>
- 432
- 433 Virtanen, P., Gommers, R., Oliphant, T. E., Haberland, M., Reddy, T., Cournapeau, D.,
434 Burovski, E., Peterson, P., Weckesser, W., Bright, J., Walt, S. J. van der, Brett, M.,
435 Wilson, J., Millman, K. J., Mayorov, N., Nelson, A. R. J., Jones, E., Kern, R., Larson, E., ...

- Contributors, S. 1.0. (2020). SciPy 1.0: Fundamental algorithms for scientific computing in python. *Nature Methods*, 17, 261–272. <https://doi.org/10.1038/s41592-019-0686-2>
- Wall, E., Blaha, L. M., Paul, C. L., Cook, K., & Endert, A. (2018). Four perspectives on human bias in visual analytics. In G. Ellis (Ed.), *Cognitive biases in visualizations* (pp. 29–42). Springer International Publishing. https://doi.org/10.1007/978-3-319-95831-6_3
- Walt, S. van der, Schönberger, J. L., Nunez-Iglesias, J., Boulogne, F., Warner, J. D., Yager, N., Gouillart, E., Yu, T., & contributors, the scikit-image. (2014). Scikit-image: Image processing in python. *PeerJ*, 2, e453. <https://doi.org/10.7717/peerj.453>
- Wu, K., Otoo, E., & Shoshani, A. (2005). *Optimizing connected component labeling algorithms* (Technical Report LBNL-56864). Lawrence Berkeley National Laboratory, University of California. <https://doi.org/10.1117/12.596105>
- Yen, J.-C., Chang, F.-J., & Chang, S.-J. (1995). A new criterion for automatic multilevel thresholding. *IEEE Transactions on Image Processing*, 4(3), 370–378. <https://doi.org/10.1109/83.366472>
- Zack, G. W., Rogers, W. E., & Latt, S. A. (1977). Automatic measurement of sister chromatid exchange frequency. *Journal of Histochemistry & Cytochemistry*, 25(7), 741–753. <https://doi.org/10.1177/25.7.70454>

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