

# Week 3: Case Studies, *R* package JM and Extensions of the Bivariate Joint Model

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[Previous lecture](#)

- Bivariate Joint Model for a Longitudinal Outcome and a Terminal Time-to-Event Outcome (*R* package **JM**)
- Extension of the Bivariate Joint Model for a Longitudinal Outcome and a Terminal Time-to-Event Outcome (*R* package **JM**)

### Outline of this lecture

- Case Study = AIDS data
- Case Study = PBC data
- Extensions of the Bivariate Joint Models
- Presentation of the *R* package JM

## References

- **Joint Models for Longitudinal and Time-to-Event Data With Applications in *R*** from Dimitris Rizopoulos. Chapman & Hall/CRC biostatistics series. CRC Press, Taylor & Francis Group. 2012.
  - Chapter 4 includes extensive comparisons between JM and the extended Cox model.
  - Chapter 5 presents the different parametrizations of the biomarker, interaction effects, stratified models and competing risks model.
  - Chapter 6 includes the JM related diagnostic plots.

# 1 Case Studies: AIDS data

## 1.1 Data description

AIDS: 467 HIV infected patients who had failed or were intolerant to zidovudine therapy (AZT) (Abrams et al., NEJM, 1994)

The aim of this study was to compare the efficacy and safety of two alternative antiretroviral drugs, didanosine (ddI) and zalcitabine (ddC)

### Outcomes of interest

- time to death
- randomized treatment: 230 patients ddI and 237 ddC
- CD4 cell count measurements at baseline, 2, 6, 12 and 18 months
- prevOI: previous opportunistic infections

```
head(aids[c("patient", "start", "stop", "event", "CD4", "obstime", "drug")], 7)
```

	patient	start	stop	event	CD4	obstime	drug
1	1	0	6.00	0	10.677078	0	ddC
2	1	6	12.00	0	8.426150	6	ddC
3	1	12	16.97	0	9.433981	12	ddC
4	2	0	6.00	0	6.324555	0	ddI
5	2	6	12.00	0	8.124038	6	ddI
6	2	12	18.00	0	4.582576	12	ddI
7	2	18	19.00	0	5.000000	18	ddI

## 1.2 Comparison extended Cox model vs. JM

- Extended Cox model

$$h_i(t) = h_0(t)R_i(t) \exp\{\gamma ddI_i + \alpha y_i(t)\},$$

```
data('aids')
td.Cox<-coxph(Surv(start, stop, event)~drug+CD4, data=aids)
summary(td.Cox)

Call:
coxph(formula = Surv(start, stop, event) ~ drug + CD4, data = aids)

      n= 1405, number of events= 188

              coef exp(coef) se(coef)      z Pr(>|z|)
drugddI  0.30948   1.36271  0.14653   2.112  0.0347 *
CD4      -0.19343   0.82412  0.02437  -7.937 2.08e-15 ***
---
Signif. codes:  0 *** 0.001 ** 0.01 * 0.05 . 0.1 1

              exp(coef) exp(-coef) lower .95 upper .95
drugddI    1.3627    0.7338    1.0225    1.8161
CD4         0.8241    1.2134    0.7857    0.8644

Concordance= 0.696 (se = 0.018 )
Likelihood ratio test= 94.62 on 2 df,  p=<2e-16
Wald test               = 65.7 on 2 df,  p=5e-15
Score (logrank) test = 73.34 on 2 df,  p=<2e-16

>
```

⇒ the **CD4 cell count** has a strong association with the risk for death, where a unit **decrease** in CD4 cell count corresponds to a  $\exp(-(-0.19)) = 1.20$  fold increase in the risk for death (95% CI: 1.16-1.27).

⇒ the **drug ddI** has also a significant association with the risk of death, where having the drug ddI vs. ddC corresponds to a  $\exp(0.31) = 1.36$  fold increase in the risk for death (95% CI: 1.02-1.82).

- JM to account for the endogeneity of CD4 cell count

```
aids.id<-aids[!duplicated(aids$patient),]
```

```
lmeFit <- lme(CD4 ~ obstime + obstime:drug,
random = ~ obstime | patient, data = aids)
coxFit <- coxph(Surv(Time, death) ~ drug, data = aids.id, x = TRUE)
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
method = "piecewise-PH-aGH")
summary(jointFit)
```

- The argument *timeVar* of *jointModel()* is used to specify the name of the time variable in the LMM, which is required in the internal computation of  $m_i(t)$ .
- The *method* argument specifies the type of baseline risk function.

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
method = "piecewise-PH-aGH")
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with piecewise-constant baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-4328.261	8688.523	8754.864

Variance Components:

	StdDev	Corr
(Intercept)	4.5839	(Intr)
obstime	0.1822	-0.0468
Residual	1.7377	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	7.2203	0.2218	32.5537	<0.0001
obstime	-0.1917	0.0217	-8.8374	<0.0001

```
obstime:drugddI 0.0116 0.0302 0.3834 0.7014
```

```
Event Process
```

```

      Value Std.Err  z-value p-value
drugddI  0.3348  0.1565   2.1397  0.0324
Assoct   -0.2875  0.0359  -8.0141 <0.0001
log(xi.1) -2.5438  0.1913 -13.2953
log(xi.2) -2.2722  0.1784 -12.7328
log(xi.3) -1.9554  0.2403  -8.1357
log(xi.4) -2.5011  0.3412  -7.3297
log(xi.5) -2.4152  0.3156  -7.6531
log(xi.6) -2.4018  0.4007  -5.9941
log(xi.7) -2.4239  0.5301  -4.5725
```

- Comparison of results: JM vs. Cox model

Table 1: Parameter estimates for [time to death](#)

	JM			Cox PH		
	logHR	SE	p-value	logHR	SE	p-value
Treat	0.33	0.16	0.032	0.31	0.15	0.035
CD4	-0.29	0.04	< 0.0001	-0.19	0.02	< 0.0001

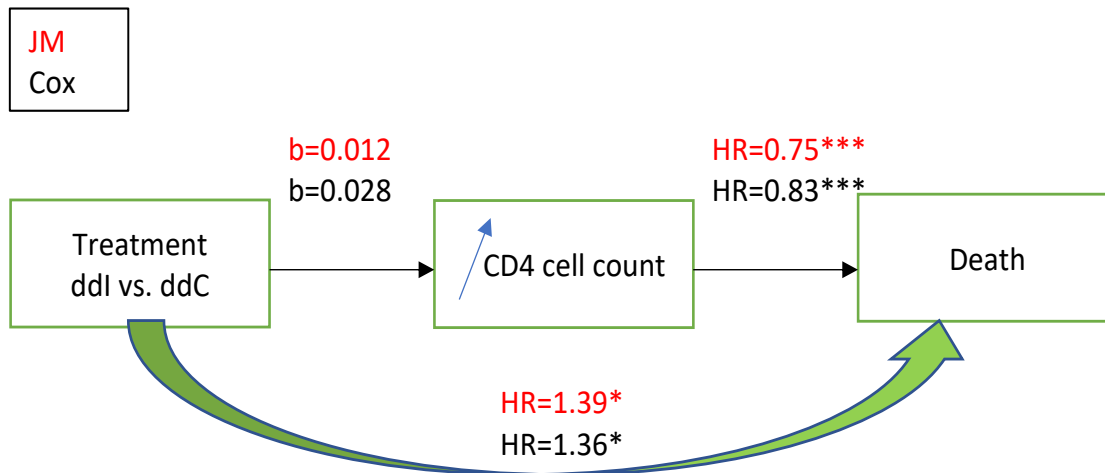
- Induced causal relationships

Remember:

Table 2: Parameter estimates for [CD4 cell count](#)

	JM			LMM <a href="#">linear mixed model</a>		
	$\beta$	SE	p-value	$\beta$	SE	p-value
Time	-0.19	0.02	< 0.0001	-0.16	0.02	< 0.0001
Time $\times$ ddI	0.012	0.03	0.70	0.028	0.03	0.34

Figure 1: Causal relationships in AIDS data

Causal relationships

direct effect: residual direct effect

indirect effect?

- Conclusion: The magnitude of CD4 cell count effect on the overall survival is reduced in the Cox model compared to the JM.

can't do causal inference: conditional on random effects

relationship/associations are time-dependent

**1.3 Effect of the baseline hazard function**

Available options are:

- 'piecewise-PH-GH': PH model with piecewise-constant baseline hazard

5 knots: 7 parameters

$$h_0(t) = \sum_{i=1}^{n_{int}} I_{\{t \in (t_{i-1}, t_i)\}} c_i$$

The interval  $[0, \tau]$  (with the last observed time among  $N$  individuals) is divided into  $n_{int}$  subintervals (6 by default). To change the default number, the control argument *lng.in.kn* can be used, whereas the argument *knots* control the position of the knots.



- 'spline-PH-GH': PH model with B-spline-approximated log baseline hazard

$$h_0(t) = \sum_{i=1}^m \zeta_i B_i(.),$$

The default number of knots is 5 with equally spaced percentiles of the observed event times. The control argument *lng.in.kn* can be used to finer control.

- 'weibull-PH-GH': PH model with Weibull baseline hazard  
parametric

$$h_0(t) = \sigma t^{\sigma-1}.$$

- 'weibull-AFT-GH': AFT model with Weibull baseline hazard

$$\log T_i^* = \gamma^T w_i + \epsilon_i$$

- 'Cox-PH-GH': PH model with unspecified baseline hazard

$$h_0(t) = \begin{cases} \xi_q, & t = T_q^* \\ 0, & t \neq T_q^*, \end{cases}$$

where  $T_q^*$  denotes the  $q$ th unique true event time. It is equivalent to assuming  $h_0(.)$  is discrete with point masses at the unique event times, i.e.,

## different baseline hazards

```
## Baseline = piecewise
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH")
summary(jointFit)

## Baseline = spline
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "spline-PH-GH")
summary(jointFit)

## Baseline = Unspecified
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "Cox-PH-GH")
summary(jointFit)

## Baseline = Weibull-PH-GH
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "weibull-PH-GH")
summary(jointFit)

## Baseline = Weibull-AFT-GH
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "weibull-AFT-GH")
summary(jointFit)
```

Table 3: Parameter estimates for [time to death](#)

	Treatment	Assoc	
	Estimate (SE)	Estimate (SE)	AIC
Piecewise hazard	0.33 (0.16)	-0.29 (0.04)	8688
Spline	0.35 (0.16)	-0.29 (0.04)	8706
Unspecified	0.32 (0.13)	-0.27 (0.03)	9549
Weibull	0.36 (0.16)	-0.29 (0.04)	8701
Weibull AFT	0.28 (0.12)	-0.23 (0.04)	8702

[not proportional hazard - AFT is a bit different](#)  
[may use AIC to select which one is the best](#)

```
## Baseline = piecewise 5 knots
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH")
summary(jointFit)

## Baseline = piecewise 10 knots
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH", lng.in.kn=10)
summary(jointFit)
```

Table 4: Parameter estimates for [time to death](#)

	Treatment	Assoc	
	Estimate (SE)	Estimate (SE)	AIC
Piecewise hazard (5 knots)	0.33 (0.16)	-0.29 (0.04)	8688
Piecewise hazard (10 knots)	0.34 (0.16)	-0.28 (0.04)	8691

## 1.4 Control of numerical integration [for the survival functions and random effects](#)

- The function `jointModel()` provides 3 control arguments that allow for a fine control of the numerical integration algorithms.
- The argument `GKk` controls the number of Gauss-Kronrod quadrature points of the survival function.
- The argument `GHk` controls the number of quadrature points of the Gauss-Hermite algorithm.
- The first 2 parts of the character string supplied in the `method` argument specify the baseline risk function and the type of survival model, and the last part the type of numerical integration. `GH` stands for the Gauss-Hermite rule and `aGH` for pseudo-adaptive rule.
- Examples:

if you have convergence, try these.

If not, stay with default

```
jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH", GHk=15)

jointFit <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH", GHk=21, GKk=15)
```

## 1.5 Sensitivity of inferences for the longitudinal process to the choice of the parameterization for the AIDS data

- We use the same [mixed model](#) as before, i.e.,

$$\begin{aligned} y_i(t) &= m_i(t) + \epsilon_i(t) \\ &= \beta_0 + \beta_1 t + \beta_2 \{t \times ddI_i\} + b_{i0} + b_{i1} t + \epsilon_i(t) \end{aligned}$$

```
aids.id<-aids[!duplicated(aids$patient),]
lmeFit <- lme(sqrt(CD4) ~ obstime + obstime:drug,
             random = ~ obstime | patient, data = aids)
```

and the following five survival submodels

- [Model I \(current value - default\)](#)

$$h_i(t) = h_0(t) \exp\{\gamma ddI_i + \alpha_1 m_i(t)\}$$

```
coxFit <- coxph(Surv(Time, death) ~ drug, data = aids.id, x = TRUE)
jointFit1 <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                      method = "piecewise-PH-aGH")
summary(jointFit1)

Call:
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
           method = "piecewise-PH-aGH")
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3\%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with piecewise-constant  
baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-2101.983	4235.965	4302.307

Variance Components:

	StdDev	Corr
(Intercept)	0.8699	(Intr)
obstime	0.0371	0.0834
Residual	0.3673	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5145	0.0425	59.1329	<0.0001
obstime	-0.0423	0.0045	-9.3082	<0.0001
obstime:drugddI	0.0052	0.0064	0.8092	0.4184

Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3484	0.1533	2.2723	0.0231
Assoct	-1.0906	0.1163	-9.3736	<0.0001
log(xi.1)	-1.6711	0.2477	-6.7461	
log(xi.2)	-1.3556	0.2381	-5.6942	
log(xi.3)	-1.0401	0.2848	-3.6513	
log(xi.4)	-1.5962	0.3727	-4.2829	
log(xi.5)	-1.4869	0.3490	-4.2597	
log(xi.6)	-1.4506	0.4274	-3.3937	
log(xi.7)	-1.4847	0.5436	-2.7311	

Integration:

method: (pseudo) adaptive Gauss-Hermite

quadrature points: 5

Optimization:

Convergence: 0

## of the biomarkers

- Model II (current value + current slope)  
slope of the biomarkers - speed of progression of CD4 cell count

$$h_i(t) = h_0(t) \exp\{\gamma ddI_i + \alpha_1 m_i(t) + \alpha_2 m'_i(t)\}$$

where  $m'_i(t) = \beta_1 + \beta_2 ddI_i + b_{i1}$

To fit this model we need to specify the 'derivForm' argument, which is a list with first component the derivative of the fixed-effects formula of 'lmeFit' with respect to 'obstime', second component the indicator of which fixed-effects coefficients correspond to the previous defined formula, third component the derivative of the random-effects formula of 'lmeFit' with respect to 'obstime', and fourth component the indicator of which random-effects correspond to the previous defined formula.

```
dForm <- list(fixed = ~ 1 + drug, indFixed = c(2, 3), random = ~ 1, indRandom = 2)
jointFit2<-jointModel(lmeFit, coxFit, timeVar = "obstime", method = "spline-PH-aGH",
  parameterization = "both", derivForm = dForm)
```

```
summary(jointFit2)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
  parameterization = "both", method = "spline-PH-aGH", derivForm = dForm)
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with spline-approximated  
baseline risk function

Parameterization: Time-dependent + time-dependent slope

log.Lik	AIC	BIC
-2095.27	4228.539	4307.32

Variance Components:

	StdDev	Corr
(Intercept)	0.8704	(Intr)
obstime	0.0369	0.0968

Residual      0.3678

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5147	0.0426	59.0379	<0.0001
obstime	-0.0429	0.0049	-8.6995	<0.0001
obstime:drugddI	0.0048	0.0064	0.7548	0.4504

Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3687	0.1591	2.3171	0.0205
Assoct	-1.0732	0.1210	-8.8722	<0.0001
Assoct.s	-2.8447	5.7456	-0.4951	0.6205
bs1	-3.1669	0.6800	-4.6569	<0.0001
bs2	-0.4143	0.6925	-0.5983	0.5497
bs3	-3.2779	0.7468	-4.3890	<0.0001
bs4	-0.5019	0.5378	-0.9333	0.3507
bs5	-2.1079	0.5808	-3.6292	0.0003
bs6	-1.2158	0.8094	-1.5020	0.1331
bs7	-2.1258	1.4947	-1.4222	0.1550
bs8	0.0168	2.9599	0.0057	0.9955
bs9	-8.0163	9.0693	-0.8839	0.3768

association of current value of biomarkers:  
increase in CD4 cell count decrease the chance  
of dying.

association of current slope: not significant

Integration:

method: (pseudo) adaptive Gauss-Hermite

quadrature points: 5

Optimization:

Convergence: 0 model is convergence

if it's 1, it means you have a convergence problem.

- Model III (random slope)

$$h_i(t) = h_0(t) \exp\{\gamma ddI_i + \alpha_3 b_{i1}\}$$

```
dForm <- list(fixed = ~ 1, indFixed = c(2), random = ~ 1, indRandom = 2)
jointFit3<-jointModel(lmeFit, coxFit, timeVar = "obstime", method = "spline-PH-aGH",
                      parameterization = "slope", derivForm = dForm)
```

```
summary(jointFit3)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
           parameterization = "slope", method = "spline-PH-aGH", derivForm = dForm)
```

## Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

## Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with spline-approximated  
baseline risk function

Parameterization: Time-dependent slope

log.Lik	AIC	BIC
-2121.845	4279.689	4354.323

## Variance Components:

	StdDev	Corr
(Intercept)	0.8255	(Intr)
obstime	0.0381	0.5317
Residual	0.3837	

random effects will have a normal dist with mean = 0,  
standard deviation of 0.04

## Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5227	0.0407	61.9196	<0.0001
obstime	-0.0484	0.0049	-9.8429	<0.0001
obstime:drugddI	0.0023	0.0065	0.3590	0.7196

## Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.2701	0.1755	1.5383	0.1240
Assoct.s	-27.4063	4.5162	-6.0685	<0.0001
bs1	-7.1770	0.7231	-9.9248	<0.0001
bs2	-4.2147	0.6864	-6.1399	<0.0001
bs3	-6.8020	0.7007	-9.7074	<0.0001
bs4	-3.7536	0.4652	-8.0685	<0.0001
bs5	-5.2156	0.5042	-10.3439	<0.0001
bs6	-4.3429	0.7565	-5.7404	<0.0001
bs7	-4.7486	1.4110	-3.3654	0.0008
bs8	-3.3367	2.6684	-1.2504	0.2111
bs9	-7.5859	6.4539	-1.1754	0.2398

## Integration:

method: (pseudo) adaptive Gauss-Hermite

can switch to the normal Gauss-Hermite



```
quadrature points: 5
```

```
Optimization:
Convergence: 1
```

**Note:** The previous model did not converge

⇒ Convergence: 1

We refit the model with more iterations `iter.qN=500` and using **Gauss-Hermite** integration algorithm

```
jointFit3<-jointModel(lmeFit, coxFit, timeVar = "obstime", method = "spline-PH-GH",
                      parameterization = "slope", derivForm = dForm, iter.qN=500)
```

```
summary(jointFit3)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
           parameterization = "slope", method = "spline-PH-GH", derivForm = dForm,
           iter.qN = 500)
```

Data Descriptives:

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Number of Groups: 467

Joint Model Summary:

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Event Process: Relative risk model with spline-approximated  
baseline risk function

Parameterization: Time-dependent slope

log.Lik	AIC	BIC
-2128.688	4293.375	4368.009

Variance Components:

	StdDev	Corr
(Intercept)	0.8215	(Intr)
obstime	0.0417	0.4883
Residual	0.3906	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5535	0.0321	79.5239	<0.0001
obstime	-0.0498	0.0051	-9.8409	<0.0001
obstime:drugddI	0.0039	0.0067	0.5898	0.5554

Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.2888	0.1819	1.5873	0.1124
Assoct.s	-26.9728	4.6533	-5.7965	<0.0001
bs1	-7.2946	0.7668	-9.5129	<0.0001
bs2	-4.3234	0.7157	-6.0406	<0.0001
bs3	-6.9239	0.7252	-9.5476	<0.0001
bs4	-3.8530	0.4838	-7.9645	<0.0001
bs5	-5.2167	0.5132	-10.1645	<0.0001
bs6	-4.3433	0.7540	-5.7602	<0.0001
bs7	-4.7304	1.3909	-3.4009	0.0007
bs8	-3.3588	2.6362	-1.2741	0.2026
bs9	-7.7152	6.3591	-1.2133	0.2250

Integration:

method: Gauss-Hermite

quadrature points: 15

Optimization:

Convergence: 0 everything is fine

- Model IV (area)

$$h_i(t) = h_0(t) \exp\{\gamma ddI_i + \alpha_4 \int_0^t m_i(s) ds\}$$

where  $\int_0^t m_i(s) ds = \beta_0 t + \frac{\beta_1}{2} t^2 + \frac{\beta_2}{2} \{t^2 \times ddI_i\} + b_{i0} t + \frac{b_{i1}}{2} t^2$

```
cForm <- list(fixed = ~ -1 + obstime + I(obstime^2/2) + I((obstime^2 * (drug=="ddI"))/2),
              indFixed = 1:3, random = ~-1 + obstime + I(obstime^2/2), indRandom = 1:2)
jointFit4<-jointModel(lmeFit, coxFit, timeVar = "obstime", method = "spline-PH-GH",
                     parameterization = "slope", derivForm = cForm)
```

```
summary(jointFit4)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
  parameterization = "slope", method = "spline-PH-GH", derivForm = cForm)
```

Data Descriptives:

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Joint Model Summary:

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Event Process: Relative risk model with spline-approximated  
baseline risk function

Parameterization: Time-dependent slope

log.Lik	AIC	BIC
-2116.463	4268.927	4343.561

Variance Components:

	StdDev	Corr
(Intercept)	0.8686	(Intr)
obstime	0.0383	0.0023
Residual	0.3752	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5587	0.0381	67.2408	<0.0001
obstime	-0.0394	0.0046	-8.6512	<0.0001
obstime:drugddI	0.0066	0.0065	1.0111	0.3120

Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3416	0.1502	2.2741	0.0230
Assoct.s	-0.1018	0.0132	-7.7221	<0.0001
bs1	-5.2835	0.5549	-9.5215	<0.0001
bs2	-1.9026	0.5706	-3.3344	0.0009
bs3	-3.6702	0.6372	-5.7595	<0.0001
bs4	-0.0388	0.4479	-0.0867	0.9309
bs5	-0.9667	0.5358	-1.8042	0.0712
bs6	-0.1028	0.8081	-0.1272	0.8988
bs7	0.4689	1.4811	0.3166	0.7516
bs8	0.6588	3.0512	0.2159	0.8291
bs9	-4.4165	9.5502	-0.4625	0.6438

association of biomarkers and time to death  
(cumulative associated effects)

biomarkers are decreasing the chance of dying

Integration:  
 method: Gauss-Hermite  
 quadrature points: 15

Optimization:  
 Convergence: 0

- Model V: Weighted Cumulative Effects (convolution):

$$h_i(t) = h_0(t) \exp\{\gamma ddI_i + \alpha_5 \int_0^t w(t-s)m_i(s)ds\}$$

We will use the normal function:  $w(x) = \frac{1}{\sqrt{2\pi}} \exp(-x^2/2)$ .

In order to construct the weighted cumulative effect, we are required to evaluate integrals of the form:

$$\int_0^t w(t-s)s^j ds = \frac{1}{\sqrt{2\pi}} \int_0^t s^j \exp\{-(t-s)^2/2\} ds,$$

for  $j = 0, 1$  because remember:

$$m_i(t) = \beta_0 + \beta_1 t + \beta_2 \{t \times ddI_i\} + b_{i0} + b_{i1} t.$$

These integrals do not have a closed-form so we need to use the *integrate()* function in *R*. We define the function  $g(.)$  that calculates the integrals numerically.

putting more weight at the end of the study

```
g<-function(u, pow=0){
  f<-function(t)
    integrate(function(s) s^pow*dnorm(t-s),0,t)$value
  sapply(u,f)
}
```

remove the default of intercept

```
cFormW <- list(fixed = ~ -1 + I(g(obstime)) + I(g(obstime,1))
               + I(g(obstime,1)*(drug=="ddI")), indFixed = 1:3,
               random = ~-1 + I(g(obstime)) + I(g(obstime,1)), indRandom = 1:2)
jointFit5<-jointModel(lmeFit, coxFit, timeVar = "obstime", method = "spline-PH-GH",
                      parameterization = "slope", derivForm = cFormW)
```

```
summary(jointFit5)      weighted cumulative effect
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
           parameterization = "slope", method = "spline-PH-GH", derivForm = cFormW)
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with spline-approximated  
baseline risk function

Parameterization: Time-dependent slope

log.Lik	AIC	BIC
-2102.956	4241.913	4316.547

Variance Components:

	StdDev	Corr
(Intercept)	0.8683	(Intr)
obstime	0.0387	0.0494
Residual	0.3750	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5554	0.0374	68.3079	<0.0001
obstime	-0.0414	0.0046	-9.0433	<0.0001
obstime:drugddI	0.0054	0.0065	0.8344	0.4040

Event Process

	Value	Std.Err	z-value	p-value	
drugddI	0.3512	0.1528	2.2983	0.0215	
Assoct.s	-2.1819	0.2384	-9.1510	<0.0001	for the weighted cumulative effect
bs1	-4.5668	0.5111	-8.9350	<0.0001	one unit increase in CD4 will decrease the risk of
bs2	0.5956	0.6131	0.9714	0.3313	dying
bs3	-3.5503	0.6318	-5.6192	<0.0001	
bs4	-0.1037	0.4147	-0.2501	0.8025	
bs5	-1.9903	0.4762	-4.1798	<0.0001	
bs6	-0.9439	0.7402	-1.2753	0.2022	
bs7	-1.9685	1.4528	-1.3550	0.1754	
bs8	0.0886	2.8690	0.0309	0.9754	
bs9	-7.1028	8.3292	-0.8528	0.3938	

Integration:

method: Gauss-Hermite

quadrature points: 15

Optimization:

Convergence: 0

- Effects of parametrization on LMM and event time components

	Model I Current value	Model II Current value + Current slope	Model III Random slope	Model IV Cumulative	Model V Weighted Cum.
AIC	4236.0	4228.5	4293.4	4268.9	4241.9
<b>LMM component</b>					
Time	-0.04 (0.004) $p < 0.0001$	-0.04 (0.005) $p < 0.0001$	-0.05 (0.005) $p < 0.0001$	-0.04 (0.005) $p < 0.0001$	-0.04 (0.005) $p < 0.0001$
Time $\times$ drug	0.005 (0.006) $p = 0.42$	0.005 (0.006) $p = 0.45$	0.005 (0.004) $p = 0.55$	0.007 (0.007) $p = 0.31$	0.005 (0.006) $p = 0.40$
<b>Event time component</b>					
Drug	0.35 (0.15) $p = 0.023$	0.37 (0.16) $p = 0.020$	0.29 (0.18) $p = 0.11$	0.34 (0.15) $p = 0.023$	0.35 (0.15) $p = 0.021$
Assoc.	-1.09 (0.11) $p < 0.0001$	-1.07 (0.12) $p < 0.0001$	- -	- -	- -
Assoc. (slope)	- -	-2.85 (5.75) $p = 0.62$	-27.0 (4.65) $p < 0.0001$	-0.10 (0.01) $p < 0.0001$	-2.18 (0.24) $p < 0.0001$

Which model is the best?

According to AIC, Model II is best but the current slope parameter is not significant in this model. What's wrong?

## 1.6 Diagnostic plots

- To assess different formulations of the JM, residual plots can be used.
- For JMs, they are available for both the longitudinal and time-to-event components.
- The default `plot()` function provides four types of plots:
  - Residuals vs. fitted values from LMM
  - QQ plots of residuals from LMM
  - Marginal survival function
  - Marginal cumulative hazard function

- Example in Figure 2:

```
pdf("Model1_plot.pdf")
par(mfrow=c(2,2))
plot(jointFit1)
dev.off()
```

- For the [LMM component](#), the standardized marginal residuals vs. marginal fitted values  $X_i\hat{\beta}$  can be obtained with the commands

```
resMargY.aids<-residuals(jointFit1, process="Longitudinal", type="Marginal")
fitMargY.aids<-fitted(jointFit1, process="Longitudinal", type="Marginal")

plotResid<-function(x,y, col.loess="black",...){
  plot(x,y,...)
  lines(lowess(x,y), col=col.loess, lwd=2)
  abline(h=0, lty=3, col="grey", lwd=)
}

plotResid(fitMargY.aids, resMargY.aids, xlab="Fitted values", ylab="Marginal residuals")
```



Figure 2: Diagnostic plots for Model I: Current value of the biomarker

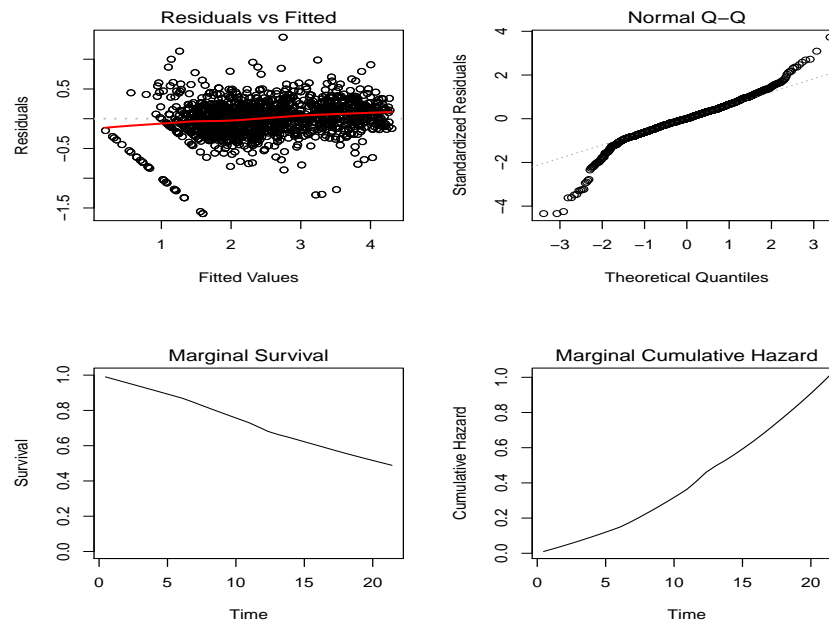
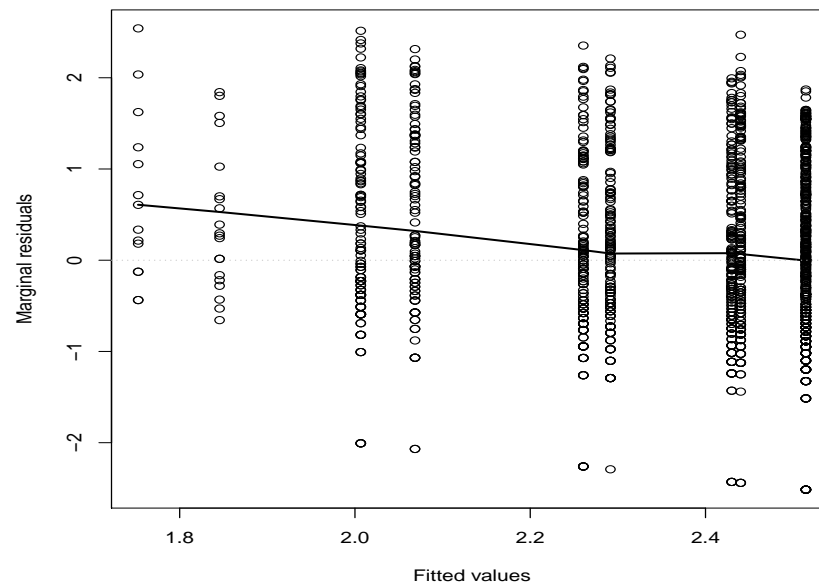


Figure 3: Diagnostic plots for Model I: Fitted values vs. marginal residuals



- For the [survival component](#), the martingale residuals can be used. They are defined as:

$$r_i^{tm}(t) = N_i(t) - \int_0^t R_i(s) h_i(s | \hat{\mathcal{M}}_i(s); \hat{\theta}) ds$$

- They can be viewed as the difference between the observed number of events for  $i$ th subject by time  $t$  and the expected number by the same time based on the fitted model.
- They can help identify individuals who are not well fitted by the model and evaluate appropriate functional forms for the covariates
- Another type of residuals for survival models is the Cox-Snell residuals. They are calculated as the value of the estimated cumulative risk function evaluated at the observed event time  $T_i$

difference here: hazard depends on the biomarkers

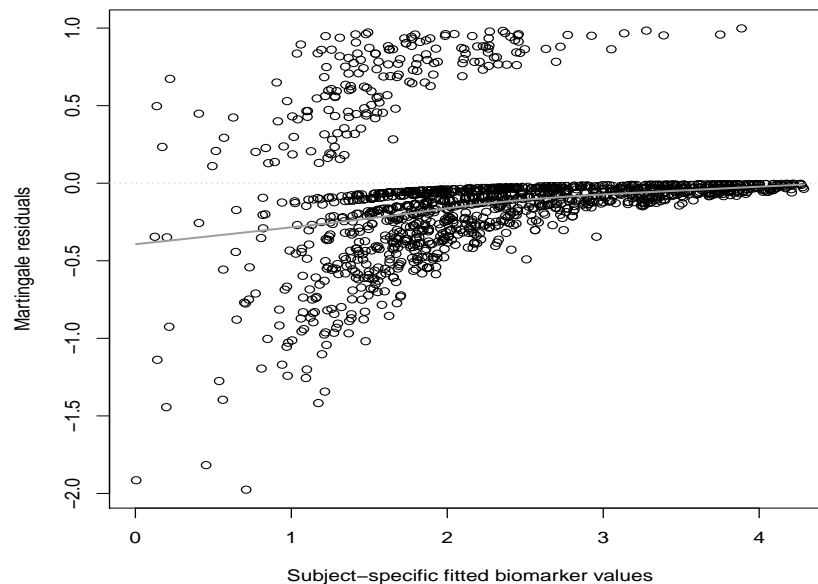
$$\begin{aligned} r_i^{tcs} &= \int_0^{T_i} h_i(s | \hat{\mathcal{M}}_i(s); \hat{\theta}) ds \\ &= \int_0^{T_i} \hat{h}_0(s) \exp\{\hat{\gamma}^T w_i + \hat{\alpha} \hat{m}_i(s)\} ds. \end{aligned}$$

- For a model that is well fitted, the Cox-Snell residuals are unit exponentially distributed.
- Example:

```
martRes <-residuals(jointFit1, process="Event")
mi.t <-fitted(jointFit1, process="Longitudinal", type="EventTime")
plotResid(mi.t, martRes, col.loess="grey62", ylab="Martingale residuals",
xlab="Subject-specific fitted biomarker values")
```

- The option `type="EventTime"` indicates to calculate the fitted values of the biomarker at all time points where the event time is observed.

Figure 4: Diagnostic plots for Model I: Martingale residuals



- We can also check the martingale residuals by conditioning on baseline covariates, e.g. treatment group.

```
library(lattice)
xyplot(martRes ~ mi.t | drug, data=aids, type=c("p","smooth"), col="black",
       lwd=3, ylab="Martingale residuals", xlab="Subject-specific fitted biomarker values")
```

- We can then assess the overall fit of the model by using the Cox-Snell residuals. To calculate the survival function of the Cox-Snell residuals, we use the function `survfit()` from the package `survival`.

```
resCST<-residuals(jointFit1, process="Event", type="CoxSnell")
sfit<-survfit(Surv(resCST, death)~1, data=aids.id)
plot(sfit, mark.time=F, conf.int=T, xlab="Cox-Snell residuals",
     ylab="Survival probability", main="Survival fct. of Cox-Snell residuals")
curve(exp(-x), from=0, to=max(aids.id$Time), add=T, col="grey62", lwd=2)
```

Figure 5: Diagnostic plots for Model I: Martingale residuals by treatment

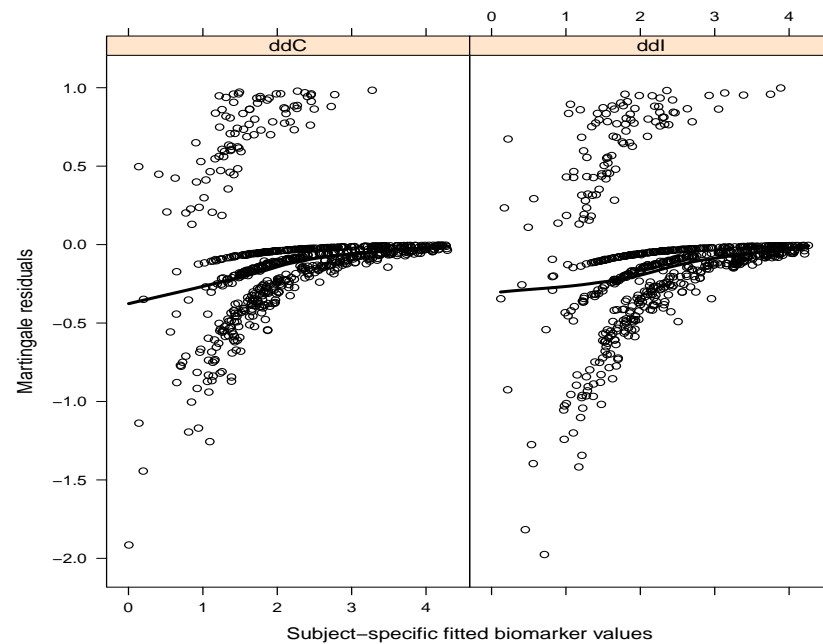
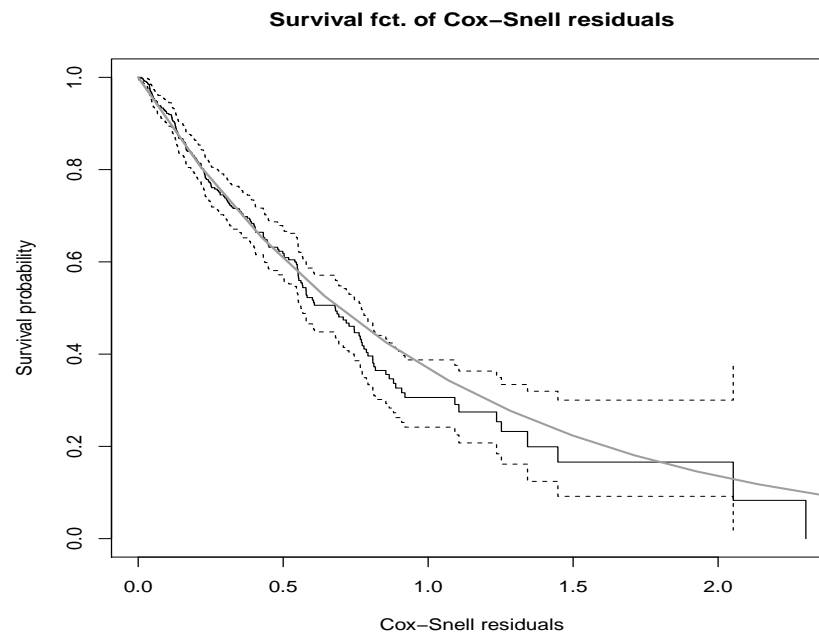


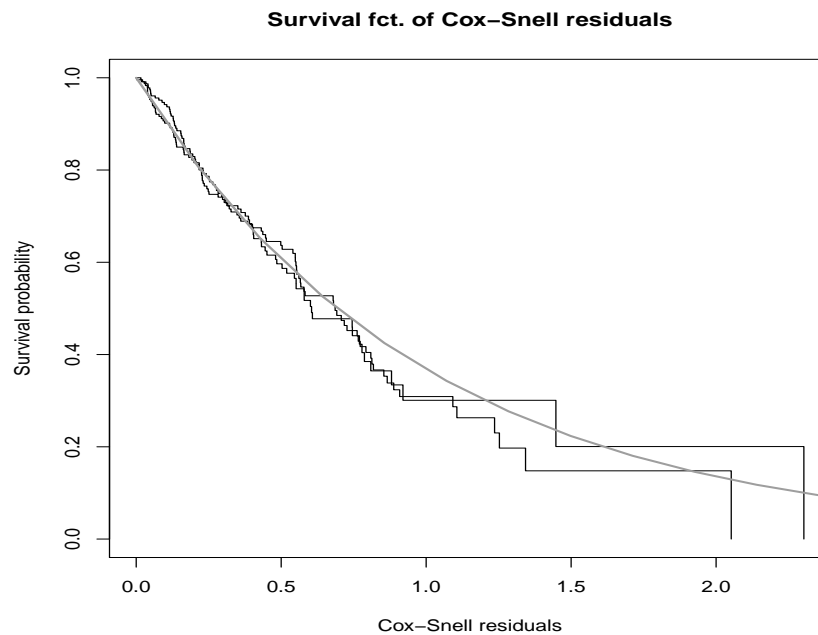
Figure 6: Diagnostic plots for Model I: Cox-Snell residuals



- We can also stratify the residuals by treatment group

```
sfit.drug<-survfit(Surv(resCST, death)~drug, data=aids.id)
pdf("Cox_snell_residuals_drug.pdf")
plot(sfit.drug, mark.time=F, conf.int=F, xlab="Cox-Snell residuals",
      ylab="Survival probability", main="Survival fct. of Cox-Snell residuals")
curve(exp(-x), from=0, to=max(aids.id$Time), add=T, col="grey62", lwd=2)
dev.off()
```

Figure 7: Diagnostic plots for Model I: Cox-Snell residuals by treatment effect



## 1.7 Effect of lag time

The hazard for an event at  $t$  is associated with the level of the marker at a previous time point:

$$h_i(t|\mathcal{M}_i(t)) = h_0(t) \exp\{\gamma ddI_i + \alpha m_i(t_+^c)\}$$

where

$$t_+^c = \max(t - c, 0)$$

```
## Standard model
jointFit.nolag <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                             method = "piecewise-PH-aGH")

## Lag =1 for the time-dependent biomarker ##
jointFit.lag1 <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                             method = "piecewise-PH-aGH", lag=1)

## Lag =2 for the time-dependent biomarker ##
jointFit.lag2 <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                             method = "piecewise-PH-aGH", lag=2)

## Lag =3 for the time-dependent biomarker ##
jointFit.lag3 <- jointModel(lmeFit, coxFit, timeVar = "obstime",
                             method = "piecewise-PH-aGH", lag=3)

## anova comparisons
anova(jointFit.nolag, jointFit.lag1, test=F)
anova(jointFit.nolag, jointFit.lag2, test=F)
anova(jointFit.nolag, jointFit.lag3, test=F)

> anova(jointFit.nolag, jointFit.lag1, test=F) best model is nolag model

      AIC      BIC log.Lik df
jointFit.nolag 4235.97 4302.31 -2101.98
jointFit.lag1  4236.78 4303.12 -2102.39  0

> anova(jointFit.nolag, jointFit.lag2, test=F) best model is nolag model

      AIC      BIC log.Lik df
jointFit.nolag 4235.97 4302.31 -2101.98
jointFit.lag2  4237.82 4304.16 -2102.91  0
```



```
interFact = list(value=~ prevOI, data=aids.id))
summary(jointFit.interac)
```

I want these variables to be included

```
anova(jointFit.nolag, jointFit.interac)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "obstime",
  method = "piecewise-PH-aGH", interFact = list(value = ~prevOI,
    data = aids.id))
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with piecewise-constant baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-2093.584	4223.167	4297.801

Variance Components:

	StdDev	Corr
(Intercept)	0.8697	(Intr)
obstime	0.0368	0.0791
Residual	0.3673	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5142	0.0425	59.1383	<0.0001
obstime	-0.0423	0.0046	-9.2892	<0.0001
obstime:drugddI	0.0062	0.0064	0.9811	0.3265

Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3369	0.1535	2.1948	0.0282
prevOIAIDS	1.6663	0.5832	2.8570	0.0043
Assoct	-0.6330	0.2173	-2.9135	0.0036
Assoct:prevOIAIDS	-0.4579	0.2623	-1.7458	0.0808
log(xi.1)	-3.2313	0.5529	-5.8446	
log(xi.2)	-2.8734	0.5444	-5.2784	
log(xi.3)	-2.5280	0.5638	-4.4837	

increase in CD4 decrease chance of dying



```
log(xi.4)      -3.0637  0.6114 -5.0109
log(xi.5)      -2.9294  0.5928 -4.9416
log(xi.6)      -2.8598  0.6373 -4.4876
log(xi.7)      -2.8376  0.7131 -3.9791
```

Integration:

method: (pseudo) adaptive Gauss-Hermite

quadrature points: 5

Optimization:

Convergence: 0

```
> anova(jointFit.nolag, jointFit.interac) in favor of interaction model
```

	AIC	BIC	log.Lik	LRT	df	p.value
jointFit.nolag	4235.97	4302.31	-2101.98			
jointFit.interac	4223.17	4297.80	-2093.58	16.8	2	2e-04

## 1.9 Stratified JM survival model is stratified

### baseline hazard function - particular strata

- In many applications, it is not realistic to assume that the samples come from a homogeneous population.
- For instance, in multicentre clinical trials, the different centres are expected to have different baseline survival functions.
- A standard extension of survival models specifies multiple strata, where each stratum has its own baseline hazard function but common values for the regression coefficients  $\gamma$  and  $\alpha$ .
- The hazard function is then expressed as

$$h_{ik}(t) = h_{0k}(t) \exp\{\gamma^T w_i + \alpha^T m_i(t)\},$$

where  $h_{0k}(t)$  denotes the baseline hazard function for stratum  $k$ .

- Joint models with stratification are available within the [JM](#) package under the B-spline approximated baseline risk function.
- In the following, we use the variable 'gender' as a possible stratification risk factor in the aids data.

```
### Model with stratification #####
lmeFit <- lme(sqrt(CD4) ~ obstime + obstime:drug, random = ~ obstime | patient,
data = aids)
coxFit.strata <- coxph(Surv(Time, death) ~ drug+strata(gender), data = aids.id, x = TRUE)

jointFit.strata <- jointModel(lmeFit, coxFit.strata, timeVar = "obstime",
                             method = "spline-PH-aGH")
```

```
summary(jointFit.strata)
```

```
anova(jointFit.nolag, jointFit.strata, test=F)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit.strata, timeVar = "obstime",
           method = "spline-PH-aGH")
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Stratified relative risk model with spline-approximated baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-2090.868	4235.735	4347.686

Variance Components:

	StdDev	Corr
(Intercept)	0.8707	(Intr)
obstime	0.0370	0.0734
Residual	0.3671	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5138	0.0426	59.0668	<0.0001
obstime	-0.0418	0.0045	-9.2102	<0.0001
obstime:drugddI	0.0053	0.0064	0.8303	0.4063

## Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3541	0.1536	2.3052	0.0212
Assoct	-1.0669	0.1159	-9.2012	<0.0001
bs1(female)	-1.7118	1.0718	-1.5971	0.1102
bs2(female)	0.9373	1.4934	0.6276	0.5303
bs3(female)	-4.4104	2.2629	-1.9490	0.0513
bs4(female)	-0.9001	1.3728	-0.6556	0.5121
bs5(female)	-1.1110	1.4456	-0.7685	0.4422
bs6(female)	-2.1772	2.8416	-0.7662	0.4436
bs7(female)	0.0058	5.2523	0.0011	0.9991
bs8(female)	0.4899	11.8006	0.0415	0.9669
bs9(female)	-1.6987	71.6775	-0.0237	0.9811
bs1(male)	-3.4587	0.7010	-4.9338	<0.0001
bs2(male)	-0.3442	0.6587	-0.5226	0.6013
bs3(male)	-3.0389	0.6775	-4.4852	<0.0001
bs4(male)	-0.3319	0.4184	-0.7933	0.4276
bs5(male)	-2.0195	0.4915	-4.1087	<0.0001
bs6(male)	-1.0795	0.7646	-1.4118	0.1580
bs7(male)	-1.9343	1.5088	-1.2820	0.1998
bs8(male)	-0.7939	2.9840	-0.2661	0.7902
bs9(male)	-5.7604	7.7282	-0.7454	0.4560

spline is the baseline hazard function

## Integration:

method: (pseudo) adaptive Gauss-Hermite  
 quadrature points: 5

## Optimization:

Convergence: 0

> anova(jointFit.nolag, jointFit.strata, test=F) not a big improvement when use stratification

	AIC	BIC	log.Lik	df
jointFit.nolag	4235.97	4302.31	-2101.98	
jointFit.strata	4235.74	4347.69	-2090.87	11

- The function `wald.strata()` can be used to compare the stratified to un-stratified versions of the JM.

`wald.strata(jointFit.strata)`

```
> wald.strata(jointFit.strata)
```

Wald Test for Stratification Factors

```
X^2 = 11.3151, df = 9, p-value = 0.2547 p value is not significant - not keep stratification
alternative hypothesis: spline coefficients for the baseline risk
function are not equal among strata
```

- We conclude that the stratification criterion is not significant.
- We can also extend the previous model by allowing the covariates to be specific to the strata. This leads to the following hazard model

$$h_{ik}(t) = h_{0k}(t) \exp\{\gamma_k^T w_{ik} + \alpha_k^T m_{ik}(t)\},$$

strata is the gender

```
coxFit.strata.inter <- coxph(Surv(Time, death) ~ drug*gender + strata(gender),
                             data = aids.id, x = TRUE)
jointFit.strata.inter <- update(jointFit.strata, survObject=coxFit.strata.inter,
                               interFact = list(value=~ gender, data=aids.id))
summary(jointFit.strata.inter)

anova(jointFit.strata, jointFit.strata.inter)

Call:
jointModel(lmeObject = lmeFit, survObject = coxFit.strata.inter,
           timeVar = "obstime", method = "spline-PH-aGH", interFact = list(value = ~gender,
                                     data = aids.id))
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1405 Number of Events: 188 (40.3%)

Number of Groups: 467

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Stratified relative risk model with spline-approximated baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-2090.447	4238.894	4359.138

## Variance Components:

	StdDev	Corr
(Intercept)	0.8709	(Intr)
obstime	0.0369	0.0742
Residual	0.3673	

## Coefficients:

## Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	2.5137	0.0426	59.0506	<0.0001
obstime	-0.0419	0.0045	-9.2267	<0.0001
obstime:drugddI	0.0053	0.0064	0.8250	0.4094

## Event Process

	Value	Std.Err	z-value	p-value
drugddI	0.3979	0.4798	0.8293	0.4069
drugddI:gendermale	-0.0414	0.5056	-0.0818	0.9348
Assoct	-0.8716	0.2884	-3.0227	0.0025
Assoct:gendermale	-0.2272	0.3143	-0.7230	0.4697
bs1(female)	-2.0735	1.2060	-1.7193	0.0856
bs2(female)	0.6174	1.5776	0.3914	0.6955
bs3(female)	-4.8466	2.3420	-2.0694	0.0385
bs4(female)	-1.2068	1.4643	-0.8242	0.4098
bs5(female)	-1.5900	1.5618	-1.0180	0.3087
bs6(female)	-2.1748	2.7649	-0.7866	0.4315
bs7(female)	-0.8457	5.2583	-0.1608	0.8722
bs8(female)	0.9138	11.6729	0.0783	0.9376
bs9(female)	-1.7227	69.3872	-0.0248	0.9802
bs1(male)	-3.4048	0.7060	-4.8230	<0.0001
bs2(male)	-0.2832	0.6637	-0.4267	0.6696
bs3(male)	-2.9914	0.6822	-4.3848	<0.0001
bs4(male)	-0.2740	0.4253	-0.6442	0.5195
bs5(male)	-1.9806	0.4982	-3.9757	0.0001
bs6(male)	-0.9827	0.7752	-1.2678	0.2049
bs7(male)	-2.0415	1.5601	-1.3086	0.1907
bs8(male)	-0.2472	3.1940	-0.0774	0.9383
bs9(male)	-7.5766	9.6129	-0.7882	0.4306

treatment effect:  
baseline is female

## Integration:

method: (pseudo) adaptive Gauss-Hermite

quadrature points: 5

Optimization:

Convergence: 0

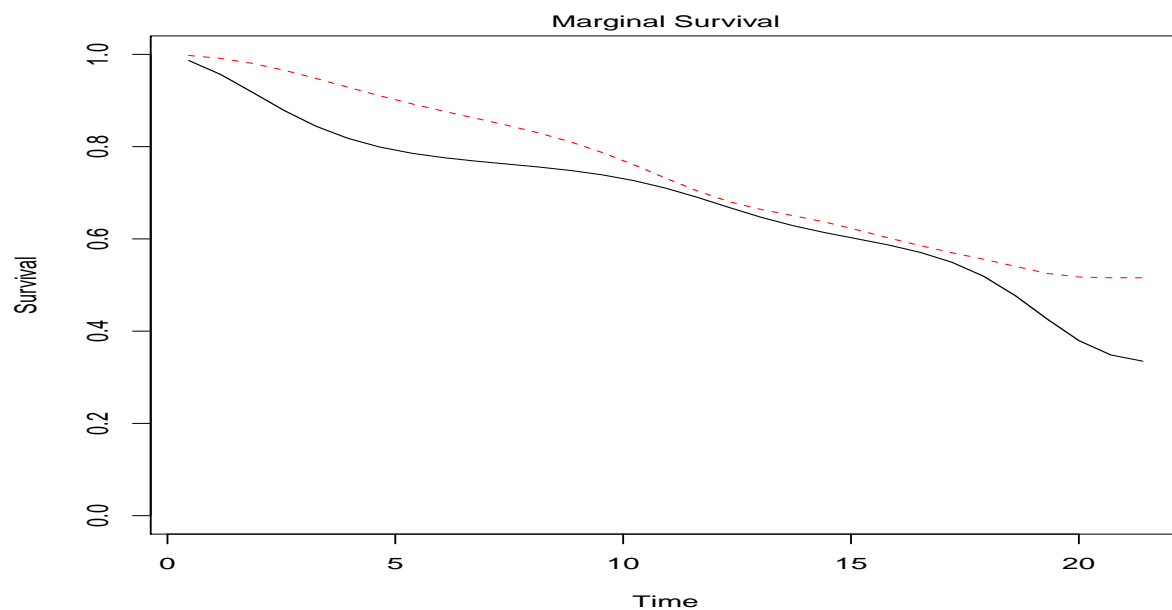
```
> anova(jointFit.strata, jointFit.strata.inter)
```

	AIC	BIC	log.Lik	LRT	df	p.value
jointFit.strata	4235.74	4347.69	-2090.87			
jointFit.strata.inter	4238.89	4359.14	-2090.45	0.84	2	0.6566

AIC is increasing for new model and p value testing the difference between these models is not significant

- Plot of survival function

Figure 8: Marginal survival functions (dashed line is for males and solid line for females)



marginal survival for these two groups are relatively similar

indication of the stratification does not give you a big difference in marginal survival

```
pdf("Survival_plot")
plot(jointFit.strata.inter, which=3)
dev.off()
```

## 2 Case study: Primary Biliary Cirrhosis

### 2.1 Data description

- **PBC: Primary Biliary Cirrhosis**

- A chronic, fatal but rare liver disease
- Characterized by inflammatory destruction of the small bile ducts within the liver

- **Data collected** by Mayo Clinic from 1974 to 1984 (Murtaugh et al., Hepatology, 1994)

- **Outcomes of interest**

- Time to death and/or time to liver transplantation
- Randomized treatment: 158 patients received D-penicillamine and 154 placebo
- Longitudinal serum bilirubin levels

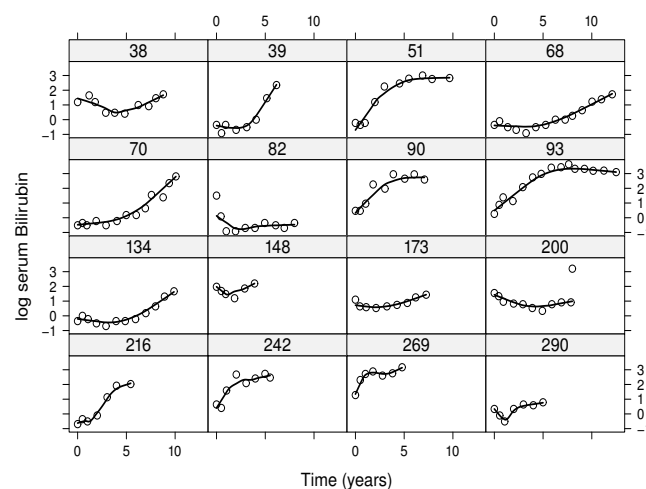


Figure 9: Longitudinal profiles of serum bilirubin levels

```
data("pbc2.id")
head(pbc2.id,5)
```

	id	years	status	drug	age	sex	year	ascites	hepatomegaly	spiders
1	1	1.095170	dead	D-penicil	58.76684	female	0	Yes	Yes	Yes
2	2	14.152338	alive	D-penicil	56.44782	female	0	No	Yes	Yes
3	3	2.770781	dead	D-penicil	70.07447	male	0	No	No	No
4	4	5.270507	dead	D-penicil	54.74209	female	0	No	Yes	Yes
5	5	4.120578	transplanted	placebo	38.10645	female	0	No	Yes	Yes

	edema	serBilir	serChol	albumin	alkaline	SGOT	platelets	prothrombin	histologic	status2
1	edema despite diuretics	14.5	261	2.60	1718	138.0	190	12.2	4	1
2	No edema	1.1	302	4.14	7395	113.5	221	10.6	3	0
3	edema no diuretics	1.4	176	3.48	516	96.1	151	12.0	4	1
4	edema no diuretics	1.8	244	2.54	6122	60.6	183	10.3	4	1
5	No edema	3.4	279	3.53	671	113.2	136	10.9	3	0

## 2.2 Comparison JM vs. Cox model

- Cox model

```
coxFit <- coxph(Surv(years, status2) ~ log(serBilir) + drug + sex + age,
               data = pbc2.id)
summary(coxFit)
```

Call:

```
coxph(formula = Surv(years, status2) ~ log(serBilir) + drug +
      sex + age, data = pbc2.id)
```

n= 312, number of events= 140

	coef	exp(coef)	se(coef)	z	Pr(> z )
log(serBilir)	1.085856	2.961975	0.092587	11.728	< 2e-16 ***
drugD-penicil	-0.127653	0.880159	0.176473	-0.723	0.469
sexfemale	-0.004180	0.995829	0.233030	-0.018	0.986
age	0.046376	1.047468	0.008222	5.640	1.7e-08 ***

---  
Signif. codes: 0 \*\*\* 0.001 \*\* 0.01 \* 0.05 . 0.1 1

	exp(coef)	exp(-coef)	lower .95	upper .95
log(serBilir)	2.9620	0.3376	2.4704	3.551
drugD-penicil	0.8802	1.1362	0.6228	1.244
sexfemale	0.9958	1.0042	0.6307	1.572
age	1.0475	0.9547	1.0307	1.064

Concordance= 0.811 (se = 0.018 )

Likelihood ratio test= 167.8 on 4 df, p=<2e-16

Wald test = 174.7 on 4 df, p=<2e-16

Score (logrank) test = 199.9 on 4 df, p=<2e-16



- JM

```
lmeFit <- lme(log(serBilir) ~ year+drug+year:drug, random = ~ year | id, data=pb2)
coxFit <- coxph(Surv(years, status2) ~ drug + sex + age, data = pb2.id, x = TRUE)
jointFit <- jointModel(lmeFit, coxFit, timeVar = "year", method = "piecewise-PH-aGH")
summary(jointFit)
```

Call:

```
jointModel(lmeObject = lmeFit, survObject = coxFit, timeVar = "year",
  method = "piecewise-PH-aGH")
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1945 Number of Events: 140 (44.9%)

Number of Groups: 312

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Relative risk model with piecewise-constant  
baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-1888.845	3815.689	3886.806

Variance Components:

	StdDev	Corr
(Intercept)	0.9986	(Intr)
year	0.1797	0.4313
Residual	0.3473	

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	0.5610	0.0826	6.7921	<0.0001
year	0.1864	0.0188	9.9328	<0.0001
drugD-penicil	-0.1353	0.1162	-1.1642	0.2443
year:drugD-penicil	-0.0033	0.0255	-0.1295	0.8970

Event Process

	Value	Std.Err	z-value	p-value
drugD-penicil	-0.0179	0.1856	-0.0966	0.9230
sexfemale	0.0992	0.2485	0.3994	0.6896
age	0.0627	0.0092	6.7812	<0.0001

```

Assoct      1.3438  0.1014  13.2542 <0.0001
log(xi.1)   -7.9743  0.7049 -11.3133
log(xi.2)   -7.7223  0.6963 -11.0898
log(xi.3)   -7.9591  0.7102 -11.2067
log(xi.4)   -7.9096  0.7325 -10.7981
log(xi.5)   -7.4819  0.6998 -10.6908
log(xi.6)   -7.1774  0.7231  -9.9254
log(xi.7)   -7.7526  0.7811  -9.9253

```

Integration:

method: (pseudo) adaptive Gauss-Hermite  
 quadrature points: 5

Optimization:

Convergence: 0      **clinical trial may not be randomized  
 treatment effect at baseline**

- LMM component

Table 5: Parameter estimates for **log bilirubin**

	JM			LMM		
	$\beta$	SE	$p$ -value	$\beta$	SE	$p$ -value
Drug	0.18	0.02	< 0.0001	0.19	0.02	< 0.0001
Years	-0.13	0.12	0.25	-0.14	0.12	0.24
Years $\times$ drug	-0.004	0.03	0.86	-0.003	0.03	0.90

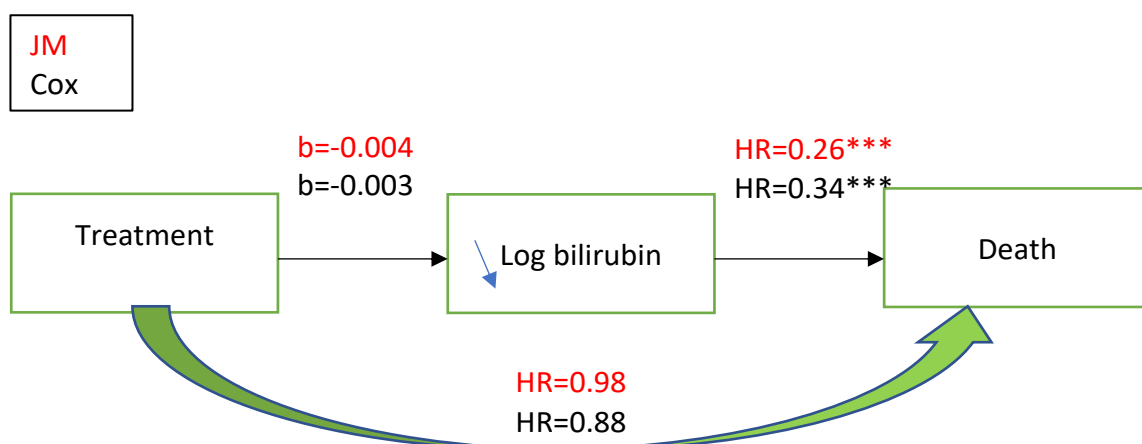
- Survival component

Table 6: Parameter estimates for time to death

	JM			Cox PH		
	logHR	SE	p-value	logHR	SE	p-value
Drug	-0.018	0.19	0.92	-0.13	0.18	0.47
Sex	0.099	0.25	0.69	-0.004	0.23	0.99
Age	0.063	0.009	< 0.0001	0.046	0.008	< 0.0001
log(bilirubin)	1.34	0.10	< 0.0001	1.09	0.09	< 0.0001

- Induced causal relationships

Figure 10: Causal relationships in PBC data

Causal relationships

decreasing log of bilirubin, decrease risk of dying (HR is lower than 1 = 0.26) from the disease, so improving the overall survival - direct effect is very significant  
hazard is close to 1 and it's not significant  
indirect and treatments and death. just a direction of indirect effect here.

## 2.3 Competing risks model

- In longitudinal studies, we often observed more than one time-to-event outcomes.
- Competing risks concern the situation where more than one cause of failure is possible (Putter, Fiocco, and Geskus, 2007).
- A classical example relates to several causes of death (e.g. from cancer) where the occurrence of any cause of death prevents the event of interest from occurring.
- Treating the events of the competing causes as censored observations will lead to biased estimates of the survival function for the event of interest when we are in the presence of competing risks (Putter et al., 2007).
- In the PBC data, several outcomes can "compete" to be the first event observed, e.g. time to death or time to transplantation. It could be interesting to distinguish these 2 events and estimate specific effects of baseline covariates for each of them.
- If these covariates are endogenous time-dependent covariates, the JM approach should be used.
- Notations:
  - We assume  $K$  different causes of failure
  - We let  $T_{i1}^*, \dots, T_{iK}^*$  be the true failure times
  - The observed event time is  $T_i = \min(T_{i1}^*, \dots, T_{iK}^*, C_i)$
  - The event indicator is  $\delta_i \in \{0, 1, \dots, K\}$ , where 0 corresponds to censoring and  $1, \dots, K$  to the competing events.

- For each of the  $K$  causes of failure, the hazard function is

$$h_{ik}(t) = h_{0k}(t) \exp\{\gamma_k^T w_i + \alpha_k m_i(t)\}.$$

- It includes baseline covariates and current values of the biomarker that affect the **cause-specific hazard** function.
- The contribution of the event time to the likelihood, conditional on the biomarker, can be written as

probability of having one event of interest given you survival all possible events before time  $t$  if someone has no event of interest, he only has the survival component

$$p(T_i, \delta_i | b_i; \theta_t, \beta) = \prod_{k=1}^K \left[ h_{0k}(T_i) \exp\{\gamma_k^T w_i + \alpha_k m_i(T_i)\} \right]^{I(\delta_i=k)} \\ \times \exp \left( - \sum_{k=1}^K \int_0^{T_i} h_{0k}(s) \exp\{\gamma_k^T w_i + \alpha_k m_i(s)\} ds \right).$$

two events of interest - so need to create the data framework

- For the data format, each subject has  $K$  rows, one for each possible cause of failure. The function `crLong()` can be used to create the 'competing risks long format' data set.
- Usual data format (for survival analysis)

```
head(pbc2.id[c("id", "years", "status")], 5)
```

```
> head(pbc2.id[c("id", "years", "status")], 5)
```

	id	years	status
1	1	1.095170	dead
2	2	14.152338	alive
3	3	2.770781	dead
4	4	5.270507	dead
5	5	4.120578	transplanted

an event for both - transplantation status and death status

I need to create a long format dataset - use crLONG - two rows for one individual - one for death status and one for transplantations status

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- New format

```
pbcr2.idCR<-crLong(pbc2.id, statusVar="status", censLevel="alive", nameStrata="CR")
head(pbc2.idCR[c("id","years","status","CR","status2")],10)
```

```
> head(pbc2.idCR[c("id","years","status","CR","status2")],10)
```

	id	years	status	CR	status2
1	1	1.095170	dead	dead	1
1.1	1	1.095170	dead	transplanted	0
2	2	14.152338	alive	dead	0
2.1	2	14.152338	alive	transplanted	0
3	3	2.770781	dead	dead	1
3.1	3	2.770781	dead	transplanted	0
4	4	5.270507	dead	dead	1
4.1	4	5.270507	dead	transplanted	0
5	5	4.120578	transplanted	dead	0
5.1	5	4.120578	transplanted	transplanted	1

- Note that each patient has 2 rows (i.e. 2 causes of failure, death or transplantation) but the time variable 'years' is similar.
- The variable 'CR' denotes the cause for the specific line of the dataset.
- LMM for the PBC data. We assume now a more complicated model

polynomial function with quadratic terms

$$y_i(t) = \beta_0 + \beta_1 \text{drug} + \beta_2 t + \beta_3 t^2 + \beta_4 \{\text{drug} \times t\} + \beta_5 \{\text{drug} \times t^2\} + b_{i0} + b_{i1} t + b_{i2} t^2 + \epsilon_i(t).$$

```
lmeFit.pbc <- lme(log(scrBilir) ~ drug*(year + I(year^2)),
                 random = ~ year + I(year^2) | id, data=pbc2)
summary(lmeFit.pbc)
```

```
> summary(lmeFit.pbc)
Linear mixed-effects model fit by REML
Data: pbc2
      AIC      BIC    logLik
2929.358 3001.767 -1451.679
```

Random effects:

```
Formula: ~year + I(year^2) | id
Structure: General positive-definite, Log-Cholesky parametrization
          StdDev      Corr
(Intercept) 0.99941237 (Intr) year
year         0.30424697  0.171
I(year^2)    0.02525548  0.011 -0.882
Residual     0.30486888
```

Fixed effects: log(serBilir) ~ drug \* (year + I(year^2))

	Value	Std.Error	DF	t-value	p-value
(Intercept)	0.5855771	0.08268683	1629	7.081867	0.0000
drugD-penicil	-0.1435083	0.11623757	310	-1.234612	0.2179
year	0.1624491	0.03112533	1629	5.219194	0.0000
I(year^2)	0.0015047	0.00314793	1629	0.477995	0.6327
drugD-penicil:year	0.0072035	0.04387470	1629	0.164182	0.8696
drugD-penicil:I(year^2)	-0.0023505	0.00448354	1629	-0.524245	0.6002

Correlation:

	(Intr) drgD-p year	I(y^2) drgD-
drugD-penicil	-0.711	
year	0.047 -0.034	
I(year^2)	0.088 -0.062 -0.842	
drugD-penicil:year	-0.033 0.050 -0.709 0.597	
drugD-penicil:I(year^2)	-0.061 0.085 0.591 -0.702 -0.842	

Standardized Within-Group Residuals:

Min	Q1	Med	Q3	Max
-4.847446422	-0.453308510	0.000722912	0.446415136	4.557128170

Number of Observations: 1945

Number of Groups: 312

- For the cause-specific hazard models, we assume

$$h_{i1}(t) = h_{01}(t) \exp\{\gamma_{11}\text{drug}_i + \gamma_{12}\text{age}_i + \alpha_1 m_i(t)\},$$

baseline hazard function to the first event

$$h_{i2}(t) = h_{02}(t) \exp\{(\gamma_{11} + \gamma_{21})\text{drug}_i + (\gamma_{12} + \gamma_{22})\text{age}_i + (\alpha_1 + \alpha_2)m_i(t)\},$$

baseline hazard function and parameters for the first event

biomarkers will be specific to the second event

where event 1 denotes the risk of transplantation and event 2 the risk of death.

all the variables will be dependent on the event indicator

hazard baseline function will also dependent on the event indicator

- Model fitting is obtained by

```
coxFit.pbc <- coxph(Surv(years, status2)~(drug + age) * CR + strata(CR),
                    data=pb2.idCR, x=T)
```

at the end, we are getting two baseline hazard functions and two survival functions

main effect variables in the model - that's not estimated

```
coxph(formula = Surv(years, status2) ~ (drug + age) * CR + strata(CR),
      data = pb2.idCR, x = T)
```

```
n= 624, number of events= 169
```

	coef	exp(coef)	se(coef)	z	Pr(> z )
drugD-penicil	-0.23680	0.78915	0.37723	-0.628	0.530
age	-0.09649	0.90802	0.02265	-4.259	2.05e-05 ***
CRdead	NA	NA	0.00000	NA	NA
drugD-penicil:CRdead	0.07473	1.07759	0.41480	0.180	0.857
age:CRdead	0.14221	1.15281	0.02419	5.878	4.15e-09 ***

---  
Signif. codes: 0 '\*\*\*' 0.001 '\*\*' 0.01 '\*' 0.05 '.' 0.1 ' ' 1

	exp(coef)	exp(-coef)	lower .95	upper .95
drugD-penicil	0.7891	1.2672	0.3768	1.6530
age	0.9080	1.1013	0.8686	0.9492
CRdead	NA	NA	NA	NA
drugD-penicil:CRdead	1.0776	0.9280	0.4779	2.4296
age:CRdead	1.1528	0.8674	1.0994	1.2088

```
Concordance= 0.646 (se = 0.021 )
```

```
Likelihood ratio test= 51.47 on 4 df, p=2e-10
```

```
Wald test = 47.58 on 4 df, p=1e-09
```

```
Score (logrank) test = 49.58 on 4 df, p=4e-10
```

- For fitting the JM, only the option 'spline-PH-aGH' or 'spline-PH-GH' are available.

indicate that we have compete risk

```
jointFit.pbc <- jointModel(lmeFit.pbc, coxFit.pbc, timeVar="year",
                          method="spline-PH-aGH", CompRisk=T,
                          interFact=list(value=~CR, data=pb2.idCR))
```



```
summary(jointFit.pbc)
```

Call:

```
jointModel(lmeObject = lmeFit.pbc, survObject = coxFit.pbc, timeVar = "year",
  method = "spline-PH-aGH", interFact = list(value = ~CR, data = pbc2.idCR),
  CompRisk = T)
```

Data Descriptives:

Longitudinal Process Event Process

Number of Observations: 1945 Number of Events: 169 (54.2%)

Number of Groups: 312

Joint Model Summary:

Longitudinal Process: Linear mixed-effects model

Event Process: Competing risks relative risk model with spline-approximated baseline risk function

Parameterization: Time-dependent

log.Lik	AIC	BIC
-1896.393	3866.785	4005.276

treatment for the first event - which is transplantation

Variance Components:

	StdDev	Corr
(Intercept)	0.9966	(Intr)
year	0.3090	0.1981
I(year^2)	0.0251	0.0233
Residual	0.3027	-0.8590

Coefficients:

Longitudinal Process

	Value	Std.Err	z-value	p-value
(Intercept)	0.5841	0.0753	7.7540	<0.0001
drugD-penicil	-0.1448	0.1069	-1.3545	0.1756
year	0.1650	0.0313	5.2646	<0.0001
I(year^2)	0.0031	0.0033	0.9383	0.3481
drugD-penicil:year	0.0064	0.0430	0.1494	0.8813
drugD-penicil:I(year^2)	-0.0025	0.0043	-0.5736	0.5662

Event Process

	Value	Std.Err	z-value	p-value
transplantation				
drugD-penicil	-0.2986	0.3844	-0.7766	0.4374
age transplantation	-0.0855	0.0245	-3.4952	0.0005
drugD-penicil:CRdead	0.2721	0.4213	0.6461	0.5182
age:CRdead	0.1516	0.0259	5.8603	<0.0001
Assoct	1.0189	0.1917	5.3146	<0.0001
Assoct:CRdead	0.4505	0.2182	2.0647	0.0389

people have a unit increase in bilirubin have

i

bs1(transplanted)	-4.8115	4.2422	-1.1342	0.2567
bs2(transplanted)	-3.9341	2.4143	-1.6295	0.1032
bs3(transplanted)	0.0880	1.7452	0.0504	0.9598
bs4(transplanted)	-2.3802	1.4399	-1.6530	0.0983
bs5(transplanted)	0.0968	1.4255	0.0679	0.9458
bs6(transplanted)	-2.2655	1.8350	-1.2346	0.2170
bs7(transplanted)	-4.2260	5.4507	-0.7753	0.4381
bs8(transplanted)	-4.2056	10.5547	-0.3985	0.6903
bs9(transplanted)	-3.7498	13.5878	-0.2760	0.7826
bs1(dead)	-8.0430	0.7625	-10.5483	<0.0001
bs2(dead)	-8.7315	0.8341	-10.4687	<0.0001
bs3(dead)	-7.9882	0.8535	-9.3599	<0.0001
bs4(dead)	-8.3107	0.7464	-11.1341	<0.0001
bs5(dead)	-8.2169	0.7947	-10.3393	<0.0001
bs6(dead)	-8.1018	0.7966	-10.1710	<0.0001
bs7(dead)	-6.2104	1.2790	-4.8557	<0.0001
bs8(dead)	-10.0470	2.1937	-4.5800	<0.0001
bs9(dead)	-6.9176	2.3545	-2.9380	0.0033

Integration:

method: (pseudo) adaptive Gauss-Hermite

quadrature points: 5

Optimization:

Convergence: 0

- The `interFact` option is used to include in the survival model the interaction between the biomarker  $m_i(t)$  and the variable 'CR'.
- The results indicate that a one unit increase of the current value of the log serum bilirubin is associated with a  $\exp(1.02) \approx 2.8$  fold increase (95% CI: 1.9-4.1) in a patient's risk of transplantation and a  $\exp(1.02+0.45) \approx 4.3$  fold increase (95% CI: 1.9-9.9) in the patient's risk of death.
- Other parametrizations for the effect of the biomarker can be introduced as well, e.g. effect of current value and current slope.