

# The ssSnake reference manual

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## 1 Introduction

ssNake (solid state NMR analysis and knowledge engine) is a versatile program for the processing and analysis of NMR data. It is aimed at solid state NMR. ssNake is written in the Python programming language, and is cross-platform and open-source (GPL3 licence). ssNake focusses at manual interactive processing of data (compared to other software which tries to automate as much as possible).

### 1.1 Documentation structure

Most of this documentation discusses the menu items as can be found in ssNake. Sometimes, extra Sections are inserted to explain underlying mechanisms of ssNake. For a general impression on how to work with ssNake, the tutorial that is bundled with ssNake is a better place to start. Detailed explanation of all the options can then be found in this Reference Manual. More advanced tutorials can be found online, on the dedicated github page: [https://github.com/smeerten/ssnake\\_tutorials](https://github.com/smeerten/ssnake_tutorials).

## 2 Running ssNake

### 2.1 Python and library versions

ssNake has been programmed to run on both the python 2.x and 3.x. ssNake should run on python versions starting from 2.7 and 3.4. For the library version, the following are needed:

- `numpy`  $\geq$  1.11.0
- `matplotlib`  $\geq$  1.5.0
- `scipy`  $\geq$  0.14.1

ssNake also needs the PyQt (version 4 or 5) and h5py libraries.

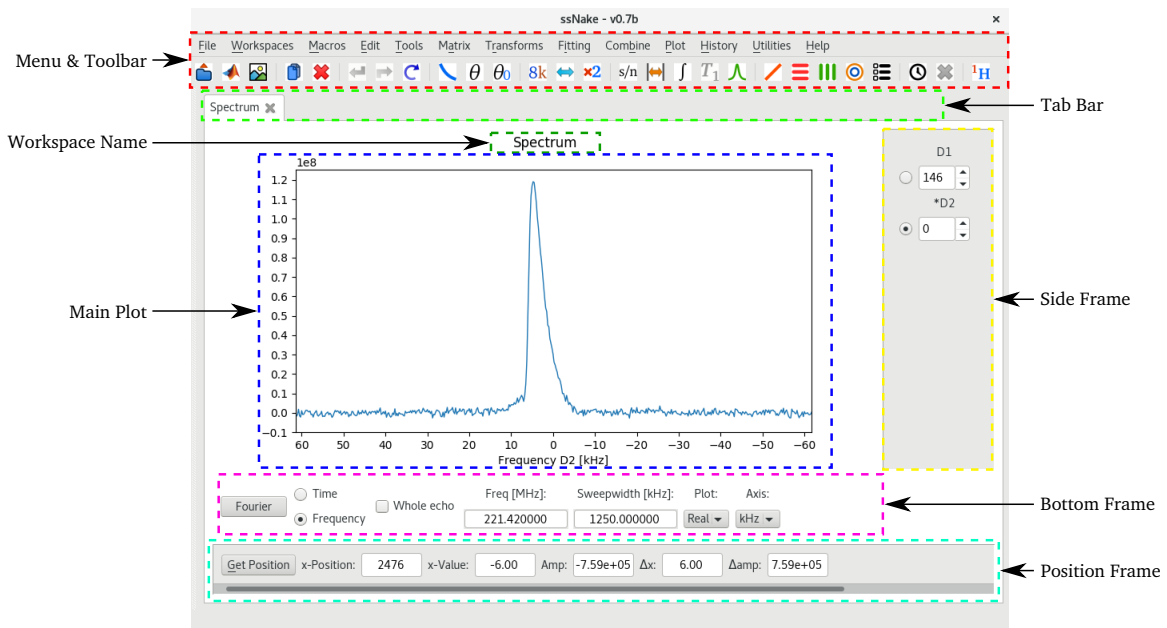
### 2.2 Installing

#### 2.2.1 Linux

On Linux, ssNake can be most efficiently run using the python libraries that are in the repositories. For Ubuntu, these can be installed by running:

```
sudo apt install python python-numpy python-matplotlib python-scipy  
python-pyqt5 python-h5py
```

Navigating to the ssNake/src directory in a terminal, ssNake can then be run by executing `python ssNake.py`.



**Figure 1:** Main interface of ssNake.

### 2.2.2 Windows

On Windows, the relevant python libraries can be installed by installing anaconda: <https://www.anaconda.com/download/>. If you do not have another python version installed already, make sure to add anaconda to the path during the install. In this case, ssNake can be run by executing the WindowsRun.bat file in the ssNake/src directory. Desktop and start menu shortcuts to this file can be created by executing the WindowsInstall.vbs file.

If you already have other version of python installed, adding anaconda to the path might create issues. Do not do this in this case. When not added to the path, the WindowsRun.bat should be edited in such a way that pythonw is replaced with the path to your pythonw.exe executable in the anaconda install directory. Note that, when updating ssNake, this file will be replaced.

### 2.2.3 OS X

On OS X, the relevant python libraries can be installed by installing anaconda: <https://www.anaconda.com/download/>. Navigating to the ssNake/src directory in a terminal, ssNake can then be run by `anacondapython ssNake.py`, with `anacondapython` replaced by the path to your anaconda python executable.

## 3 Main window

Figure 1 shows the main interface of ssNake. Below, each element will be described.

### 3.1 Menu & Toolbar

This area shows the menu and the toolbar. The toolbar holds a number of the actions that can be found in the Menu. The actions in the toolbar can be changed via the Preferences.

### 3.2 Tab Bar

Each data set that is opened in ssNake opens in its own tab (i.e. Workspace). Switching the active workspace can be done by clicking on the relevant tab, or by using keyboard shortcuts (found in the Workspaces menu). Closing workspaces can be done by clicking on the cross of the tab, or by clicking the middle mouse button when the mouse pointer is hovering over the tab name.

### 3.3 Main Plot

In the centre of the interface is the Main Plot. This shows the spectrum or FID of the current workspace (i.e. tab). The view of this plot can be changed by using the mouse controls described below. Some of the processing tools described later on rely on user input given by clicking in the Main Plot (i.e. selecting a region).

#### 3.3.1 Mouse controls

There are several ways to control the display of the current spectrum or FID in the main window using the mouse. Below, a list of these is given:

- Dragging a box while holding the left mouse button creates a zoombox, and will zoom to that region.
- Dragging while holding the right mouse button drags (i.e. pans) the spectrum. Doing this while holding the Control button pans only the x-axis, holding Shift pans only the y-axis.
- Double clicking with the right mouse button resets the view to fit the whole plot (both x and y).
- Scrolling with the mouse wheel zooms the y-axis. Doing this while also holding the right mouse button zooms the x-axis. By holding Control the zooming will use a larger step size.
- *Special*: Scrolling while holding shift changes the spacing between the different plots in the Array or Stack plot. In the Contour plot, the contour levels are zoomed.

Having an input window open (e.g. the phasing window, see below) does not prevent the use of these zoom controls. However, some tools require left clicking in the spectrum to select data points. In these cases, the zoom-box (dragging while holding the left mouse button) is disabled.



### 3.4 Bottom Frame

The bottom frame is the frame below the current data display. It consists of two parts: one with data parameters (and started by the 'Fourier' button), and the 'Get Position' frame. The features of these frames are explained below.

#### Data parameters

The data parameter frame features the following elements:

- The Fourier button: performs a forward or inverse fft depending on Time/Frequency toggle. If the current setting is Time, it does a forward transform (and fft shift), if it is Frequency it does an inverse transform.
- Time/Frequency toggle: displays whether the current data is an FID (Time) or Spectrum (Frequency). It is usually not necessary to manually change this, as this is done automatically.
- Whole echo: specifies if a whole echo transform should be done. This is set by using the swap echo tool from the Tools menu (see below). When enabled, several tools respond differently (e.g. Apodizing and Set Size).
- Freq [MHz]: The centre frequency of the current spectrum dimension in MHz. Note that this value can be different from the Reference frequency, which specifies the zero frequency of the spectrum.
- Sweepwidth [kHz]: The spectral width of the current spectrum dimension in kHz.
- Plot: A dropdown menu specifying how the data should be plot. It has four options: Real, Imag, Both and Abs. Real: plots only the real part of the complex data. Imag: plots only the imaginary part. Both: plots both the real and imaginary part of the data in different colours. Abs: plots the absolute value of the data. Note that this only toggles the view of the current data. Changing the data itself can be done via the Tools menu.
- Axis: Dropdown menu to change the units of the current x-axis. For a time axis, s, ms and  $\mu$  are supported. For the frequency axis: Hz, kHz, MHz and ppm.
- *Special*: Axis2: changes the unit of the y-axis in a 2D contour plot.

#### Position Frame

The Position frame is the frame in which the properties of a specific data point can be viewed. Pushing the 'Get Position' button and then left clicking on a specific point in the spectrum or FID prints this data. The 'Get Position frame' then displays the number of the data point you clicked on (note that in a spectrum the data goes from right to left!), the x and amplitude (y) value of

this data point and the difference with your last pick. In a Contour plot, the amplitude actually codes for the z value, and there now is a separate y value box (and corresponding y difference) as well as the index of the point along the y dimension. In most plot types, a dashed vertical line is shown at the position of the cursor to aid in the selection of a point. In the Contour plot, a horizontal line is also shown.

### 3.5 Side Frame

On the right side of the spectrum, there is an additional frame in some cases called the Side Frame. In this, special settings for the current plot type can be set, as well as the dimension along which the data should be plot. For a regular 1D spectrum of 1D data there are no additional options, and the Side Frame is not shown. The description of the elements that are shown in the Side Frame are explained in the section on the different plot types. However, the common feature for multidimensional data (i.e. the dimension selector) shall be explained here.

#### The dimension selector

For data that has more than 1 dimension, a dimension selector is shown in the Side Frame. For each dimension (labelled D1 to D $n$ , with  $n$  the number of dimensions) there is a radio button and an input box. The radio button indicates which dimension is plot in the main window. For 2D plots or arrayed plots, both the direct and the indirect dimension have to be selected. For all the dimensions of which the radio button is not selected, the input box is used to define which trace is plot. Take for example a series of ten 1D spectra with 1024 points per spectrum. The total data size is now 10x1024 (D1xD2). Note that the direct spectral dimension is always the last dimension. In a regular view of the first spectrum of this series, the radio button below D2 should be selected (as is the default setting). The number in the box next to D1 then determines which of the 10 spectra to plot. If this value is 0, the first spectrum is plot (note that ssNake uses the Python rules for array indexing, and that 0 is the first index). As we have ten spectra, 9 is the last. Similar to Python indexing, negative values can be used for reverse indexing (-1 is the last, -2 is the second to last, etc.). Using the up and down buttons next to the input box below D1, the spectra can be cycled. Also, scrolling with the scroll wheel when the mouse is on top of the box can be used.

For data with more dimensions, the idea is essentially the same. Note that ssNake always applies processing commands on the active (selected) dimension. If two dimensions need to be selected (as in an array or contour plot), the leftmost radio button is the selected dimension for operations.

### 3.6 Message bar

The bottommost part of the ssNake window is the message bar. Any error messages will be printed here in red. Warnings about wrong inputs will be displayed in black. For both type of

messages the same applies: they are shown for several seconds, and then disappear. If some tool is not responding, it is therefore wise to check the message bar for messages.

### 3.7 File Tree

On the left side of the Main Window, there is a file tree, which can be used to browse the computer, and open data. Data can be loaded by right clicking on a file or folder, and selecting 'Load Directory' or 'Load File'. Alternatively, clicking on the folder or file name using the middle mouse button opens it directly.

The File Tree can be closed by double clicking on the separator between the Tree and the Main Plot, or by clicking with the middle mouse button.

## 4 Pop-up windows

Many of the operations that can be performed in ssNake give a pop-up window in which the user can supply additional information. An example of this is the phasing window, in which the user can supply values for the first and second order phasing that should be applied to the NMR data. When such a pop-up window is active most of the Main Window is locked. When closing the pop-up window, the interface is enabled again. This makes sure that there is no interference between the different actions.

Most pop-up windows have an 'Ok' and a 'Cancel' button. Pushing 'Ok' applies the operation to the data set. Pushing 'Cancel' returns to the Main Window without any alteration of the data. In some cases, the effect of the current operation can be previewed in the plot. This allows the user to judge whether the operation (e.g. an amount of phasing) leads to the desired result. Choosing 'Cancel' returns to the original data. During the previewing, the mouse button zoom controls can still be used. However, in cases where the left mouse button is used to select a position in the plot, the 'zoom box' option is not available during the preview. Previewing is usually quite fast, as only the currently viewed part of the data is temporarily changed. On pushing 'Ok' the operation is applied to the full data set (if required).

### 4.1 Single slice

Some of the pop-up windows have a tickbox at the bottom that states 'Single slice'. When this is activated, the current operation only gets performed on the part of the data that is currently viewed. Phasing a single spectrum in a 2D data set can for example be performed in this way.

### 4.2 Keyboard modifiers

Some of the pop-up windows have left '<' and right '>' arrow buttons. These can be used to lower or raise the current value in an easy way. The step size of the buttons can be changed by holding specific keys when left clicking on the button: 'ctrl' gives x10 stepsize, 'shift' gives x100, and 'ctrl + shift' gives x1000.

## 5 File

Using the ssNake load tools is quite easy. Go to File → Open. Navigating and double clicking on the desired file then loads the data. Many formats supported by ssNake (like Bruker and Varian) have their data in a folder, in which several files with a fixed name are present. For these formats, loading any of the files in this directory is accepted (ssNake searches for the expected files in the selected directory). Loading large data files might take some time, depending on your computer hardware.

When loading the data, the user is prompted to give in a name by which the workspace is identified (see the Workspaces section for more on this). The default name is the name of the file (or folder for data in which the filename is fixed). If the name already exists as a workspace, the name spectrum $n$  is used (with  $n$  the lowest integer that is not in use).

### 5.1 Formats

ssNake is able to extract NMR data from a number of formats from several vendors. The following table summarizes the support.

Name	Specification
Varian/Agilent	VnmrJ 2 and newer. <code>fid</code> and <code>data</code> (spectrum)
Bruker	Topspin and XWinNMR ( <code>fid</code> & <code>ser,1r/i</code> & <code>2rr/ii</code> )
Bruker ParaVision	processed data (2dseq)
Bruker minispec	. <code>sig</code> file
JEOL Delta	1D and 2D only at the moment
Chemagnetics	–
Magritek	Both 1D and 2D data
SIMPSON	1D and 2D, -ascii and -binary
JCAMP	1D JCAMP-DX file
Siemens ima	single voxel only
JSON	ssNake JSON file (ascii)
Matlab	ssNake . <code>mat</code> file (binary)

Generally, ssNake loads the raw data and searches for some variables (e.g. spectral width and spectrometer frequency). The above data formats do not support data with more than two dimensions in a nice way. Getting the higher dimensions is therefore not straightforward. ssNake treats this data as 2D, after which you can split the data yourselves using Matrix → Split.

#### Varian/Agilent

Loading a Varian/Agilent `fid` file requires a *procpa*r file and a *fid* file. From the *procpa*r, the spectral frequency and the spectral width in one or two dimensions is extracted. The *fid* file is then checked for the version (VnmrJ 2 or 3) and the data is extracted from this file.

For processed data (*data*, in the *datdir* folder for example) it searches for a *procp* file in that directory or in the parent folder, from which additional parameters are extracted.

### **Bruker**

The Bruker load first checks for the *acqus* and *acqu2s* files and extracts the number of points in both dimensions, the spectral widths and the spectral frequency. Also, the bitorder is checked, which describes the way the data is saved in the *fid* or *ser* file. Using this, the data is extracted.

Bruker processed spectra can be loaded from *1r* and *1i* files for 1D data, *2rr* and *2ii* for 2D data, and *2rr* and *2ir* for hypercomplex data. It additionally takes parameters from *procs* or *proc2s* files in that directory. Moreover, the spectral frequency is extracted from the *acqus* and/or *acqu2s* files located *two directories up*.

### **Bruker ParaVision**

Loads processed data acquired via Bruker ParaVision. This feature is experimental at the moment. It need several files: *2dseq*, *d3proc* and *procs*.

### **Bruker minispec**

In some cases, the Bruker minispec also outputs a *.sig* data file. These can be loaded in *ssNake*, although this is experimental at the moment.

### **JEOL Delta**

JEOL Delta data is contained in a single *.jdf* file. Currently only 1D and 2D data is supported. This is because we have no access to a JEOL spectrometer for testing purposes. Please send us test files if you have any.

### **Chemagnetics**

Loading Chemagnetics data requires a *acq* (and *acq\_2* for 2D) and *data* file. From the former, the number of points in both dimensions is extracted, the spectral widths and the spectral frequency. All the data points are then extracted from the *data* file, and reshaped to a 2D array when necessary.

### **Magritek**

Magritek requires either two or three files: *acqu.par* for the spectral variables, and a file that ends with *.1d*. For 2D data, a file that ends in *.2d* is also needed. As with the other formats, the number of data points in the two dimensions is extracted from the parameter file (along with the spectral data) and the binary data in the *.1d* or *.2d* is unpacked.

## SIMPSON

SIMPSON data files are plain text files, *.fid* or *.spe* as file extension. From the file, the number of points in both dimensions is extracted, and the spectral widths. The spectral frequency is *not* included in the SIMPSON format, and is put at 0. SIMPSON files cannot contain a mixture of time and frequency data (both dimensions must be the same type). SIMPSON binary data is also supported, but not raw binary.

## JCAMP

JCAMP (or JCAMP-DX) is a general format for storing spectroscopic data. Although it can be applied to NMR data, it is inconvenient, especially for 2D data. ssNake therefore only supports 1D fid and spectrum data at the moment. As the JCAMP format support a wide variety of data formats, it depends on the specific file if it can be loaded into ssNake without errors. When encountering errors, do not hesitate to contact us.

## JSON

Loads a JSON file that is saved from ssNake. The file must have the *.json* or *.JSON* extension.

## Siemens ima

IMA files are dicom formatted files with additional entries from Siemens. Important: it requires pydicom > 1.0 python module to be installed. The problem in reading Siemens ima files is that there is limited documentation available on the relevant Siemens parts. In the current implementation it is assumed that all relevant spectral information is stored in the CSA Siemens header with dicom tag: '0029','1110' and that the binary signal data is stored in '7fe1','1010'. This might be different for other Siemens spectrometers currently tested. Furthermore, it is assumed that the data is in complex conjugate form; real minus imaginary. Acknowledgment is in place, this is a rewrite of the relevant part of the following source: VeSPA, versatile simulation pulses and analysis for magnetic resonance spectroscopy.

## Matlab

Loads a Matlab file that is saved from ssNake. The file must have the *.mat* or *.MAT* extension. Also supports *.mat* files that have been resaved to matlab v7.3 format.

## Unsupported file formats

Currently, we support all the data formats that we have access to. If you want your favourite data format to be support by ssNake, please send us a request along with some sample data (1D and 2D).

## 5.2 Open

When using ssNake's load function, ssNake analyses the selected file name and the names of other files in the directory to figure out the format you want to load. ssNake only needs to check for files that are actually needed for loading the data, so removing useless files from the format does not pose a problem. Below, the checks used by ssNake in this function can be found for each format.

Name	Folder contains files	File extension
Varian/Agilent fid	procpa, fid	
Varian/Agilent spectrum	data, procpa or ../procpa	
Bruker fid	acqu (and acqu2s for 2D), fid or ser	
Bruker spectrum	(1r/1i) or (2rr and 2ii/2ir) with procs/proc2s, ../../acqu, ../../acqu2s	
Bruker ParaVision	2dseq, procs and d3proc	
Bruker minispec		.sig
Chemagnetics	acq (and acq_2 for 2D) + data	
Magritek	acqu.par, *.1d or *.2d	
JEOL Delta		.jdf
NMRpipe		.fid and first byte is 0
SIMPSON		.fid or .spe
JCAMP		.jcamp or .dx or .jdx
JSON		.json or .JSON
Matlab		.mat or .MAT

## 5.3 Open & Combine

In case you want to load multiple data files and combine them into an  $n$ D array, Open & Combine can be used. It opens a window where a list of desired data can be made. This can be done by either navigating using the Browse button, or via drag-and-drop. Pressing OK loads and combines these data files, in a similar way as Workspaces → Combine does. In essence, Open & Combine does the same thing as loading all the data into separate workspaces, and then combining them. However, Open & Combine is more convenient, as it does not open separate workspaces for all the individual data files. Do note that only data files with the same shape can be combined (*i.e.* with the same number of data points). Also note that ssNake considers a 1D data file with  $n$  points to be different than a 2D data file with only one trace ( $n \times 1$  data points).

## 5.4 Save

Naturally, ssNake can also be used to save data. When saving data, the current data is saved, along with the spectral widths, frequencies and if the axis is in time or frequency units. Also, ppm references are saved, along with the processing history. Undo information is *not* saved.

### JSON

JSON (JavaScript Object Notation) is an ascii format (i.e. regular text) used to save data structures. Within ssNake this can be used to save the current data in a clear, human readable format. As JSON data is in ascii, it is not efficient in file size and in speed. If these are necessary then consider saving the data as a matlab binary.

### MATLAB

The MATLAB binary format is a open source format in which many different types of data can be saved. Also, it is the native format of MATLAB, which can be used for more special data manipulation, if necessary. As the format is binary, it is efficient in both speed and file size. The content of the file is the same as the JSON file.

## 5.5 Export

Using ssNake you can also export your data. Exporting does not save all information, so there will be loss of information when exporting and reloading the data. It is suggested you only use these formats for exchanging data to another program, but not for archiving.

### SIMPSON

SIMPSON is a general NMR simulation program with its own data format. The format only supports 1D and 2D data, and only if the data in both dimensions is of the same type (i.e. both spectrum, or both time). Apart from that, SIMPSON is an ascii format, and therefore has the same speed restrictions as the JSON format. The parameters that are saved within the SIMPSON data are restricted (no offset frequency), so when loading it back into ssNake some information is lost. We therefore advise to use the SIMPSON format only for transferring data to another program.

### ASCII (1D/2D)

This option saves the current data in a simple ascii format (text file). The first column is the axis in units of Hz or seconds. The next columns are alternating real and imaginary for all traces in the 2D data. For 1D data, only a single set of real and imaginary data is printed. The axis of dimension 1 of the 2D data is not saved.



## Figure

The Figure option can be used to export any Figure plot within ssNake. The title, axis labels and size of the Figure can be changed. Be aware that choosing a too small height can cause the x-axis label to disappear. After all the titles have been set, pushing ‘Save’ will save the Figure to a directory of your choice. The Figure can be saved in vector format (.eps, .pdf, .svg), in lossless pixel format (.png), and in lossy pixel format (.jpg). We advice to use vector formats to make high quality pictures.

The following settings can be changed:

- Title, x and y-axis labels
- Limits: the limits along the x and y-axis
- Dimensions: the size of the Figure in cm. Also the resolution (dots per inch, dpi) can be set.
- Font sizes: sizes of all the font (Main) or per item, when ‘Details’ is checked. This then allows the fonts sizes for the tile, x and y-labels, and x and y-ticks to be set individually.
- Legend: when ticked, shows the legend.
- Legend settings: allows the position of the legend to be set (allowed options: best, left, right, upper/lower left/right/center) as well as the order in the case of multiple spectra (multi plot). The order should be filled in as a list, with the first element being the topmost entry in the legend (an input of [1,0] will show element 1 on the top). Also the name can be set. The number input allows to switch between the different entries.

The legend can also be dragged around in the Figure to position it. Note that double-clicking on a plot line opens a window in which the line width and colour can be set. It also allows adding or changing plot markers.

## 5.6 Preferences

Opens a menu were some preferences can be set. Preferences are saved using the regular Qt settings files. Click on ‘Store’ to save the changes, and use ‘Reset’ (and ‘Store’) to reset to the default values. The ‘Plot’ and ‘Contour’ settings can also be changed for a specific workspace via the ‘Plot’ menu.

### 5.6.1 Window

- Default width and height of the ssNake window (in pixels). The window is never smaller than the ssNake logo shown on boot.
- Open maximized: make ssNake open with a maximized window

- Ask workspace name when loading: Whenever ssNake loads a file, it asks for a name (default is the file name, or spectrum0 etc.). When this box is unticked, files are loaded without asking for a name, but using the default name. Very useful when loading large sets of data.
- Show Shortcut Toolbar: Shows the toolbar below the menu.
- Edit Toolbar: Opens a window where, via drag-and-drop, items can be added or removed from the toolbar. Drag an item from left to right to add to the toolbar, and use the 'delete' button to remove items from the toolbar.

### 5.6.2 Plot

Can be used to set the linewidth (in pixels) and line colour. Also, x and y grids can be turned on. The default units can be set, and whether the spectrum should be in ppm by default. 'Scroll axis from zero' makes sure the  $y = 0$  position stays at the same spot on the screen when zooming y (default behaviour for most NMR software).

### 5.6.3 Contour

Change the default settings of a contour plot.

- Colourmap: defines the colourmap of the contours
- Constant Colours: makes all contour lines with the same sign the same colour (Colourmap is now ignored). Colours can be set with 'Positive colour' and 'Negative colour'.
- Ratio: sets the default ratio of the contour plot.

## 5.7 Quit

Closes the ssNake application. Quitting ssNake always asks for confirmation, to prevent data loss.

## 6 Workspaces

Within ssNake you can have multiple data files opened. Each loaded file gets its own environment in which you can process the data. These are called workspaces. All ssNake workspaces are independent, meaning that the data from workspace 1 cannot be changed while viewing another workspace. Going back to a workspace from another goes back to the same view as before: nothing is discarded when switching between workspaces. Also: every workspace has its own undo/redo information. Some tools can transfer data from other workspaces to the active workspace. You can find more about these tools later on, in their respective sections.

When loading data, ssNake prompts for a name. This will become the name of the workspace in which the data is loaded. The title of the graph will be set to this name, and the name will be

set in the list of opened workspaces in Workspaces → Active. You can also switch workspace using the tabs that are created for every workspace. Note that you cannot have multiple workspaces with the same name.

### 6.1 Duplicate

Makes a copy of the active workspace to a new workspace, of which the name is asked. All the data and the active view is copied to this new workspace. No undo and redo data is transferred. Also, the duplicated workspace will be active.

### 6.2 Slice to Workspace

Copy the current slice to a new workspace. The current slice is nothing more than the data that is currently shown on the screen. When viewing 2D data as a 1D plot, this tool copies only the shown 1D trace to a new workspace. This can be useful, for example, with T1 data, where you want the last spectrum as a separate workspace dataset (as it essentially is a good, quantitative spectrum). When viewing the data as a 2D plot (contour, array, etc), the new workspace is also 2D data. Note that when using it on an array plot with 'step = 2', half of the data is not copied to the new workspace. Only the data that is currently plotted is copied.

### 6.3 Delete

Deletes the active workspace and all references to it. The view is changed to the next workspace in the list. Be aware that there is no way to undo this!

### 6.4 Rename

Renames the active workspace. Note that two workspaces cannot have the same name.

### 6.5 Active

Shows a list of all workspaces. The active workspace can be changed by clicking on another workspace name.

### 6.6 Next

Makes the next workspace the active workspace.

### 6.7 Previous

Makes the previous workspace the active workspace.

## 6.8 Info

Shows information on the current workspace. This includes the number of dimension, sweep-widths, frequencies etc. When loading data, ssNake also saves some metadata in its data structure (number of scans, etc.). This can also be viewed in this window.

## 6.9 Keyboard shortcuts

ssNake features several keyboard shortcuts to work with the workspaces. Below is a list of all the combinations.

Combination	Effect
Ctrl + w	Close active workspace
Ctrl + n	Duplicate active workspace
F2	Rename active workspace
Ctrl + Page Up	Change active workspace to the one above in the list
Ctrl + Page Down	Change active workspace to the one below in the list

When the active workspace is the last in the list, and Ctrl + Page Down is pressed, the active workspace is set to the first in the list.

## 7 Macros

Macros can be used to save a particular series of action performed on the data, for later reuse. Say that you want to load several data files, and want to set the size of them to 4096 points and perform a Fourier transform. Doing this many times can be cumbersome. When you create a macro to do this, you only need to execute the macro for every data file. Thus saving time and making sure all are processed in an identical way. Also, macros are shared between workspaces: there is only one pool of macros. How to use macros is explained below.

### 7.1 Start recording

To create a macros, you must tell ssNake that you want to record a series of action, to be saved in the macro. Executing Macros → Start Recording prompts for a name of the future macro, and enables the recording. From now on, all the actions you perform on the data are recorded and saved in this macro.

### 7.2 Stop recording

After some actions have been performed (Fourier, zero fill, phasing...) the recording of the macro can be stopped using Macros → Stop Recording.

### 7.3 Run

Can be used to run the selected macro on the current data.

### 7.4 Delete

Delete a macro.

### 7.5 Save

Can be used to save a macro to the disk. This is a plain text file, in which you can see all the steps that are executed when running this macro.

### 7.6 Load

Loads a saved macro in ssNake. Execution of the macro can then be performed using Macros → Run.

### 7.7 Notes on the usage of macros

Note that ssNake only records the actions that are not undone. If you perform an action when recording is on, and you undo it while recording, the macro has no entries. However, performing two actions that cancel each other *are* recorded. For example, issuing Fourier twice has no net effect, but a macro of this includes both commands.

Note that it is possible to run a macro which has been recorded on a data set to be used on a data set with a different number of dimensions. For this, it is important to note that ssNake counts backwards to remember the dimension in which an action is performed. When there are 3 dimensions, ssNake classifies them as the second to last, first to last and last. A recorded macro can therefore be run at data that has all the dimensions in which some actions was performed, counting from the end. Only doing a Fourier transform in the last dimension is therefore possible on every data set, regardless of the fact that it might be D5 in some set, and D1 in another.

## 8 Edit

### 8.1 Undo

In ssNake, every action that causes a change to the data can be undone. When an action is performed, the undo list is increased with one entry, in which the information is stored as how to undo the performed action. Sometimes this involves the inverse operation (e.g. for fft/inverse fft), but for some actions the old data must be preserved and put back when an undo is requested. Duplicating data to another workspace does not copy the undo list. The undo/redo data can be cleared using Clear Undo/Redo List (see below).

Only changes to the data are recorded in the undo information. Changes to the view are not recorded and the undo can thus not be used to restore to a previous view of the data.

## 8.2 Redo

When Undo is used, the action that is undone is put in the redo list. Using Undo and then Redo therefore performs no net action. Every time a new action (i.e. something other than redo) is performed the redo list is cleared.

## 8.3 No Undo Mode

Activating this checkbox starts the No Undo Mode. This clears the current undo and redo list, and prevents any further actions from adding to the undo list. This way, no undo is possible for any further actions. Normally, this is undesirable, however, in some cases the undo information requires a large amount of computer memory, especially when processing large data sets. Having No Undo Mode activated makes it more convenient to process this data.

## 8.4 Clear Undo/Redo List

Clears the undo and redo lists of the current data. This clears the memory allocated to these lists, which might be used to reduce memory load when processing large data sets. Any operations performed after clearing the undo/redo lists will still be recorded.

## 8.5 Reload

This action reloads the data from the original source. All undo and redo information is retained, so you can undo back to before you reloaded the data.

## 8.6 Monitor

The monitor function can be used to automatically reload the data when the source file is changed (for example when measuring on the spectrometer). One or more macro's can be selected to be applied every time the data is reloaded. The Monitor action opens a window, in which the macro's can be selected (via dragging and dropping). Pushing 'Watch' then starts the monitoring, which can be stopped by returning to this window and pushing 'Unwatch'. Note that ssNake takes a cool-down period of half a second after the macro's have been executed, to avoid a lock-up of the program in the case of heavy macro's and fast changing data.

# 9 Tools

This section explains all the options located in the Tools menu.

## 9.1 Real

Real puts the imaginary values of all data points at zero. If the active dimension is hypercomplex the imaginary information corresponding to that dimension is removed. Any (hyper)complex data along other dimensions is kept.

### Background

In some NMR experiments, only the real value of the magnetization during a specific period is measured. In this case it is convenient to zero the imaginary part to force further processing to ignore this data.

## 9.2 Imag

Imag puts the real values of all data points at zero. If the active dimension is hypercomplex the real information corresponding to that dimension is removed. Any (hyper)complex data along other dimensions is kept.

## 9.3 Abs

Abs replaces all complex datapoints by the length of the vector they span in the 2D complex plane. That is:

$$F_{\text{new}} = \sqrt{\text{real}(F_{\text{old}})^2 + \text{imag}(F_{\text{old}})^2}$$

for all points in the complex data. These values are put as the real part. The imaginary part is zeroed. If the active dimension is hypercomplex the absolute values along that dimension are calculated. Any (hyper)complex data along other dimensions is kept.

## 9.4 Complex Conjugate

Multiplies the imaginary part of the data with  $-1$ . Performing this in the FID leads to a spectrum which is flipped along that dimension.

## 9.5 Apodize

Opens a tool that can be used to apply apodization (i.e. line broadening) to the current spectrum or FID. It supports Lorentzian, Gaussian,  $\cos^2$  and Hamming windows, with an optional shift or shifting (trace dependent shift).

### 9.5.1 Lorentzian

Lorentzian (i.e. exponential) apodization has the form:

$$f(t) = e^{-\pi \cdot \text{lor} \cdot |t|} \quad (1)$$

with  $lor$  the Lorentzian value in Hz, and  $t$  the time. In the frequency domain, this means that the spectrum is convoluted with the Fourier transform of this function:

$$F(\nu) = \frac{1}{\pi \cdot \gamma \cdot \left[1 + \frac{\nu^2}{\gamma^2}\right]} \quad (2)$$

with  $\gamma = 2 \cdot lor$  and  $\nu$  the frequency position. Lorentzian line shapes are characterized by a relative sharp peak, but with wide base.

### 9.5.2 Gaussian

Gaussian apodization has the form:

$$f(t) = e^{\frac{(-\pi \cdot g \cdot |t|)^2}{4 \log(2)}} \quad (3)$$

with  $g$  the Gaussian broadening in Hz. The Fourier transform of this is again a Gaussian function. Gaussian line shapes are broader (smoother) at the top when compared to Lorentzian shapes, but have more narrow base.

### 9.5.3 cos square

The  $\cos^2$  distribution is a squared cosine. The special feature of this distribution is that it involves no additional settings. It is a general  $\cos^2$  function. It is applied in such a way it goes smoothly to 0 at the end of the time signal. Therefore  $f(t) = \cos^2(\pi t / 2t_{\max})$  and the last point has a phase of  $90^\circ$  (or  $\pi/2$ ).

In ssNake, the number of periods (or half periods) of the  $\cos^2$  can be set (as the only input), with this value  $n$ , the equation becomes:

$$f(t) = \cos^2(n\pi t / 2t_{\max}) \quad (4)$$

This distribution is usually used for its easy use and certain decay to 0 (for  $n = 1$ ).

### 9.5.4 Hamming

The Hamming window is defined as:

$$f(t) = \alpha + (1 - \alpha) \cos(n\pi t / t_{\max}) \quad (5)$$

with  $\alpha = 0.53836$ , and  $n$  defined as with the  $\cos^2$  distribution (see above). It has a similar use as the  $\cos^2$ , but its Fourier transform has less intense ripples.

### 9.5.5 Shift

In some cases, the centre of the window function needs to be shifted from  $t = 0$  (in case of an echo experiment for example). This can be done using the Shift option (input in seconds). Note that all the window functions are symmetric around its centre.



### 9.5.6 Shifting

In some cases (i.e MQMAS experiments) the window function needs a trace-dependent shift. This usually means that, for 2D data, the  $t$ -shift of the window function along D2 depends on the position of that trace along D1. A 'Shifting' value of  $n$ , means that the shift for a trace is equal to  $n \cdot t_1$ , with  $t_1$  the time position of the specific trace along D1. For data with more than 2 dimensions, the shifting axis needs to be supplied. Shifting apodization can be best viewed from a Stack Plot view.

### 9.5.7 Whole echo

When the Whole Echo property is activated (see the section on the Bottom Frame), all the apodization windows are changed in such a way that they are symmetric around  $t = t_{\max}/2$ . This means that the window from  $t = t_{\max}/2$  to  $t = t_{\max}$  is equal to the time reverse of the window from  $t = 0$  to  $t = t_{\max}/2$ .

### 9.5.8 ssNake implementation

While using the Apodization tool, the effect can be previewed on the current plot. When used in the time domain, the windowing function is plot in green and the original data is shown in grey. When used from the frequency domain, it actually transforms the data to the time domain, applies the window, and transforms back to the frequency domain. Usually, this is fast and can be viewed live. However, especially for contour plots, the redrawing of the contour lines could take a long time. It is therefore advised to use this function from a 1D view (if previewing is needed).

The Lorentzian and Gaussian apodization have sliders and arrows. The step sizes of both are determined by the spectral width and number of points. Note that the coarseness of the steps when pushing on the arrow buttons can be altered by holding ctrl (x10), shift (x100), or ctrl + shift (x1000).

For most apodization types, negative line broadening parameters are allowed, but quickly lead to massive distortions.

## 9.6 Phasing

Opens a tool that can be use to change the phasing in the current spectrum or fid. Supports zero order and first order phasing. It also provides buttons for autophasing.

Selecting phasing in the tools menu opens the phasing window. In this windows, the zero and first order phasing can be set, as well as the pivot point (centre point) for the first order phasing. The values can be either filled in the boxes, changed by pushing the left and right buttons (click for  $\pm 1$ , ctrl+click for  $\pm 10$ , shift+click for  $\pm 100$ , and shift+ctrl+click for  $\pm 1000$ ), or set by dragging the slider to the left or the right. For zero order phase correction, the limits are -180 and 180 degrees. For first order correction, these are -540 and 540 degrees. Higher

values for the first order correction sometimes make sense, and can be filled in the box directly, or via clicking on the arrows. Note that the values for the zero order correction are circular, so that -180 and 180 degrees lead to the same effect.

While in this menu, the effect of a certain phasing can be immediately viewed in the ssNake window. When a good setting is reached, 'Ok' can be pushed to apply the phasing. Note that for multidimensional data, the preview only shows the current trace. Pushing 'Ok' the applies the same phasing correction on all the traces (might take some time for large data). If the phasing should only be applied on the current spectrum or fid, the box 'Single slice' can be ticked. Note that you still have to push 'Ok' to apply the phasing.

Apart from manual phasing, ssNake also supports two types of autophasing: zero order only, and zero and first order. When pushing these buttons, ssNake will search for the best phasing in the *current* trace, with only zero order phasing correction, or with both zero and first order. Do note that the setting that ssNake finds might not be the best: phasing an NMR spectrum is tricky and has no unique mathematical solution. However, for a fast analysis it can be useful.

Also, note that first order phase correction is always applied in the *frequency* domain. When first order phasing is applied when viewing the time domain signal, ssNake transforms to the spectral domain, does the phase correction, and transforms back. As Fourier transforms are quite fast, this provides no issues when viewing the effect of the phase correction.

### 9.7 Autophase 0

Performs a 0th order autophase on the active spectrum (and applies it to all traces). The only difference with the Autophase from the Phasing window is in the way macros handle the autophasing. The autophasing in the Phase windows is used to determine the phase which needs to be applied to obtain an in-phase spectrum. When a macro is being recorded it will only store the final phase values. This is unlike when the Autophase button is used from the Tools menu, which will store the autophasing operation to the macro. So when the macro is applied to another spectrum it will perform the autophasing again.

### 9.8 Autophase 0+1

Similar to Autophase 0, but this will also optimize the 1st order phase.

### 9.9 Swap echo

A tool to swap the time domain data symmetrically around a selected point. Echo swapping is done by left clicking on the echo maximum of the time signal. By default, the swapping is set at the centre of the time signal. After left clicking, a preview is shown. Pushing 'Apply' applies the transformation to all traces of this dimension. Also, the 'Whole echo' flag is set in the Bottom Frame. When this is on, Fourier transforming does not multiply the first point by 0.5 (as is needed for data that decays to zero), and line broadening is applied symmetrically.

### 9.10 Offset correction

Subtracts the average value of the selected data points from all data points. The Offset correction tool opens a new window. By default, the last 20% of the time data is selected as containing only noise (of which the average is the offset which should be subtracted). Left clicking two times in the graph sets the limits between which the average value is calculated. Pushing apply subtracts this value from all time data points. If the data has more dimensions than 1, the average value is subtracted from all data points. Take note that the average is only calculated for the current graph. To apply the offset correction only to the displayed data/traces you can use the 'Single slice' button.

### 9.11 Baseline correction

Fits a polynomial line of a given order through the current spectrum while ignoring the selected parts. This line is then subtracted from the spectrum. In ssNake, baseline correction is performed by fitting a  $n$ th degree polynomial through the spectrum, i.e.:

$$S_{\text{corr}} = \sum_{i=0}^n a_i x^i,$$

with  $a_i$  the amplitude of the  $i$ th degree polynomial. Fitting this to the spectrum is a mathematical operation with a single unique solution. Prior to doing this, specific parts of the spectrum can be selectively ignored in the fitting procedure. This can be done by left clicking in the spectrum at the two limits of the area that is to be ignored. The area is then marked with a red background. This way, multiple areas can be selected. Pressing 'Fit' then performs and shows the fit. Apply subtracts the line from the spectrum. Note that by default this single fit is subtracted from *all* the traces. This is useful if the background is identical in all of them. Alternatively, the box 'Single slice' can be ticked to only apply the correction on the displayed spectrum.

### 9.12 Subtract averages

Subtracts the average of a region from the respective data. The analysis is performed on every slices in the  $n$ D data. This effectively does the same as Offset Correction, but then for each trace independently. Opening this tool gives a window where the left and right limit of the selected data can be set. Alternatively, left clicking on the plot sets the start limit on the first click, and the end limit on the second click. Pushing apply performs the subtraction.

### 9.13 Reference deconvolution

Uses the FIDDLE algorithm to deconvolute a peak shape in the spectrum.

Using the 'Reference deconvolution' opens a window, in which the region of the reference signal has to be set (by left clicking, or by filling in the values). Additionally, the new linewidth of the peak has to be specified. Note that the algorithm cannot increase resolution, only improve

the lineshape. Using a linewidth that is too small will result in a distorted spectrum. Pushing apply performs the algorithm to all traces.

### 9.14 Correct digital filter

Corrects a fid to start at  $t = 0$  for data that has a digital filter delay. This is essentially the same as using a first order phase correction with the correct phase (i.e. number of points). For Bruker and JEOL data, the required phase shift is extracted from the parameters files. Failing to use this function for these data leads to a spectrum with a strong first order phasing error.

### 9.15 Scale SW

A function to scale the spectra width (i.e. the sweepwidth) of the current dimension. Essentially, this can also be done in the main windows, by filling in  $a * b$ , with  $a$  the current SW and  $b$  the multiplication factor. This tool is included for MQMAS data processing, where a scaling of SW of the indirect dimension can be used to get a true isotropic dimension (i.e. 1 ppm chemical shift corresponds to a 1 ppm peak shift). This scaling factor for this is  $Q$  and  $I$  dependent, and can be selected in this tool via an easy dropdown menu.

To automate this process, ssNake uses information from a text by W. M. Westler and F. Abildgaard. Every Bruker NMR spectrometer has its own characteristics, and therefore needs a different amount of first order phase correction. For older Bruker hardware, the amount of correction is found by extracting two parameters from the Bruker acquis file, and to use a lookup table supplied by Westler & Abildgaard. For newer hardware, this value can be read directly from the acquis file.

Using this tool opens a windows, in which you have to open (again) the current Bruker file. ssNake then finds the desired phase correction and applies it to all traces.

### 9.16 Reference

This is a collection of tools to set the current reference (0 frequency) of the current spectrum.

#### 9.16.1 Set reference

Opens a tool where the reference frequency can be set. It is easiest to click on the position in the spectrum which should be referenced, which fills in the frequency. In the lowest box ('Reference [ppm]') the desired ppm value of the clicked position can be filled in. Alternatively, a series of common solid state references are available via the drop-down menu called 'Secondary Reference'.

A name can be given to the reference if reuse is desired. The references saved this way are accessible via Tools → Reference → Apply.

### 9.16.2 Clear current reference

Removes the reference of the current spectrum. This means that 0 Hz (or ppm) is now at the centre of the spectrum.

### 9.16.3 Apply

Apply the selected saved reference.

### 9.16.4 Delete

Delete a saved reference.

### 9.16.5 Rename

Rename a saved reference.

### 9.16.6 Save

Save a saved reference to disk. The create file contains a single number: the zero frequency of the reference.

### 9.16.7 Load

Loads a reference from disk.

## 10 Matrix

This section describes the ssNake matrix manipulation tools. NMR data is nothing more than a data matrix with  $N$  dimensions. Using the matrix tools, you can change the size of a dimension, split the data into more dimension, integrate, multiply, mirror etc. In the following section, the different supported matrix manipulations are introduced.

### 10.1 Sizing

The sizing tool can be used to edit the number of points in the active dimension.

Selecting 'Matrix → Sizing' creates a sizing window. In here a single number has to be filled in the box titled 'Size'. Decimal values will be rounded to the nearest integer. Apart from numbers, the command 'k' (or 'K') is also supported, which stands for '1024'. When using this definition there has to be a number before the 'k'. A valid input would therefore be: '2k'. Which sets the number of points to  $2 \cdot 1024 = 2048$ . Pressing 'Apply' executes the sizing command on all traces in all dimensions. Also, the  $\pm 2^n$  buttons can be used to go to the nearest power of 2 (which makes subsequent Fourier transforms much faster).

If the supplied number is larger than the current size, zeroes are added at the right-hand side of the time signal. If the value is lower, points are removed from the right-hand side of the time signal. If the whole echo type is on, the zeroes are added in the centre of the time signal, or points are taken symmetrically from the centre, if the size is reduced. Alternatively, the position of the zero filling can be specified by left-clicking in the spectrum, or by manually filling in the desired offset value in the box 'Offset'.

ssNake always applies the sizing in the time domain. So when the size is changed within a spectrum, it is transformed to the time domain, the size is changed, and a forward Fourier transform gets the data back to the frequency domain.

## 10.2 Shift data

Shifts the data to the left or to the right, by removing data points on the left or on the right, and filling with zeroes on the opposite side. Negative values are shift left, positive values shift right.

Starting the shift data tool creates an input window. In this window, a integer number must be filled in that equals the number of data points of the desired *right* shift. Left shifts can be made by using negative values. Alternatively, the arrows on the right and left side of the input box can be used to shift the data in the direction of the arrow. The effect of the operation on the current 1D can be seen in the main window. When pushing 'Apply' the shift is performed on all traces of the nD data.

The shifting is always applied in the time domain. In the frequency domain Fourier transforms are used to transform to and from the time domain to apply the shifting.

## 10.3 Roll data

Rolls the data to the left or to the right, by removing data points from one side, and inserting them on the other side of the data. Negative values roll to the left, positive values to the right. Non-integer values can also be used. In this case, Fourier-based tricks are used to accomplish this.

Starting the roll data tool creates an input window. In this window, a number must be filled in that equals the number of data points of the desired *right* roll. Left rolls can be made by using negative values. Alternatively, the arrows on the right and left side of the input box can be used to roll the data in the direction of the arrow. The effect of the operation on the current 1D can be seen in the main window. When pushing 'Apply' the roll is performed on all traces of the nD data.

## 10.4 Integrate

Integrates a specific part of the active dimension on all traces.

When using the integration matrix tool, a window pops up, where you have to select the limits of the area you want to integrate in the active dimension. This can be done by typing in the numbers of the data points, or by left clicking in the spectrum. The first click sets the first limit, the second click the second limit. Clicking Apply performs the integration. If the input boxes are

left empty the entire length is integrated. If only a single set of limits is supplied the resulting data will have one less dimension than the original.

### **10.5 Sum**

Sums a specific part of the active dimension on all traces. This method is implemented in a way similar to integrating.

### **10.6 Max**

Takes the maximum value of the selected part in the active dimension on all traces. Implementation is similar to integration.

### **10.7 Min**

Takes the minimum value of the selected part in the active dimension on all traces. Implementation is similar to integration.

### **10.8 Max position**

Returns the x value of the maximum position (i.e. top of a peak) within the selected area for all traces.

### **10.9 Min position**

Returns the x value of the minimum position within the selected area for all traces. Works similar to Max position.

### **10.10 Average**

Returns the average of the selected region. Implementation is similar to integration.

### **10.11 Diff**

Differentiates the data in the active dimension (i.e. returns the difference between each data point and the next data point). This is equal to the slope between the points. Remember that NMR data in the frequency domain is always depicted from right to left, so the direction of differentiating is displayed 'in reverse'. The size of the data along that dimension will be reduced by one.

### 10.12 Cumsum

Performs a cumulative sum on the data in the active dimension. This means that each point is now equal to its old value, and those of all the previous points added. Remember (as with Diff) that the frequency domain is displayed in reverse.

### 10.13 Extract part

Deletes all parts of the active dimension that are outside the selected region.

This function can be used to delete parts from the active spectrum or fid. Left clicking on a prt sets the first limit on the first click, and the second limit on the second click. Apply deletes all parts outside this region on all traces. When doing this in the spectrum, the spectral width and zero point (reference frequency) are also changed to keep all peaks at the same frequency.

### 10.14 Flip L/R

Flips (i.e. mirrors) the left and right side of the active dimension. The axis is not flipped.

### 10.15 Delete

Deletes specific data points from the active dimension. The points can be given in array form. For example: [1,2,40] deletes data points 1, 2 and 40. Remember that ssNake uses the python way of array indexing: 0 is the first point. The above code therefore deletes the second, third and forty-first data point. Also note that the indexing goes from right to left in the frequency domain. Numpy commands can also be used to remove datapoints. For example, `arange(0,5000,2)` will remove every other datapoint for the first 5000 datapoints.

### 10.16 Split

Splits the active dimension in  $n$  parts with an equal number of data points. The newly created dimension is named D1, and all the other dimension numbers are increased by one (shifted one lower). This tool can only split the data in parts of equal size. It is therefore necessary to make sure that the size of the data in the active dimension is divisible by  $n$ .

### 10.17 Multiply

Multiplies the y-values of the data in the active dimension by a given number. If a single number is supplied, all the points are multiplied by this number. Alternatively, an array with the same length as the data can be given to specify a position dependent multiplication. For example, for a dimension with 3 data points: [1,3.34,2]. This multiplies the first data point by 1, second by 3.34, etc. Do not forget the about the mirroring in the frequency domain (see Diff)!



### 10.18 Normalize

A tool to scale a selected region to a particular maximum, minimum, or integral. This is essentially the same as the 'Multiply' tool, only ssNake calculates the multiplication factor for you.

### 10.19 Reorder

Reorders the data in the active dimension. Input is an array with the new, desired positions. For example, [0,2,1] keeps the position of the first element ('0'), but interchanges the second and the third. It is also possible to supply the input array by loading a file (as is used in Non-Uniform Sampling). If the new data length is longer than the old, the missing points are filled with zeros. Also, the length of the desired new dimension can be given. If the length of the final data is shorter than this value, some zeros are appended.

### 10.20 Regrid

Opens a window that can be used to regrid the spectrum. This involves linear interpolation if the new  $x$ -values fall between data points.

Using this tool opens a window in which the type of interpolation can be set (now only min/max). The min/max input then requires a new minimum and maximum to be set, and the number of points. Note that a power of 2 is advised for efficient Fourier transforms. The spectral width and centre frequency are adjusted to comply with the new axis (and the reference, when defined, stays the same).

### 10.21 Concatenate

Concatenates that data of dimension  $n$  after dimension  $n + 1$ . So if the original data is 10x5x3, and dimension 2 is selected, it becomes 10x15. This can be seen as the opposite of the 'Split' function. Note that after applying this function the numbers of the dimensions in the ssNake window changes accordingly.

### 10.22 Shearing

Shearing data is a function most commonly used for MQMAS data (in solid state NMR, that is). It is used to correct a frequency slope along a specific dimension. The amount of shearing necessary is defined by a shearing constant, which needs to be supplied by the user. For every trace in the current spectrum, the data is shifted (left or right, depending on the sign of the shearing constant) a specific amount. Any data point that goes outside the range of the data rolls to the other side (aliasing). The size of the shift is directly related to the  $x$ -value of that specific trace in the shearing direction.

Activating Shearing opens a menu, in which the shearing constant, direction and axis must be set.

- *Constant*: The amount of shearing necessary. The shift for each trace depends on this constant, and the spectral width in the shearing direction. For MQMAS data, the constant depends on the spin quantum number, and the quantum level that is used. A selection menu with these values can be found under Shearing Constant.
- *Direction*: The direction is the axis number, over which the amount of shearing differs. Usually, this is this indirect dimension (for 2D data this is D1).
- *Axis*: The axis over which the data must be rolled. In a 2D case, this is D2.

In ssNake the shearing is implemented in the time domain for the *Axis* dimension. For each trace in dimension *Direction* the time domain data is multiplied with a phase linear in *Axis*. This means that data can be sheared with non-integer values as well. If the *Axis* dimension is not in the time domain, an inverse and a forward Fourier transform is used.

## 11 Transforms

### 11.1 Fourier Transform

Performs a Fourier Transform (equal to the 'Fourier' button in the bottom frame). When the current data is in time units, it performs a regular Fourier transform, when in spectrum mode, it performs an inverse Fourier transform. The type (i.e. time or spectrum) of the data is modified accordingly.

For the Fourier transform, ssNake uses the Fast Fourier Transform (FFT) from the numpy library. It also executes the corresponding `fftshift` after the transformation. Note that FFT is the most efficient when the data size is a power of 2.

### 11.2 Real Fourier Transform

Performs a real Fourier transform. This is equivalent to first applying Tools → Real and then a complex Fourier transform. This makes the spectrum symmetric around the centre. In some Figures in literature, only the positive frequencies are shown for this reason. In ssNake the entire spectrum is kept.

### 11.3 Fftshift

Performs an `fftshift`. This interchanges the left and right part of the data. Normally, this function is not needed, as ssNake performs an `fftshift` after a Fourier transform.

### 11.4 Inv fftshift

Performs an inverse `fftshift`.

## 11.5 Hilbert Transform

Performs a Hilbert Transform along the active dimension. This removes the imaginary data, and creates new imaginary data from the real data via the Kramers–Kronig relations. The sign of the imaginary data is lost in this way. Most commonly, a Hilbert transform is used in algorithms that take some part of the real data, and the imaginary part needs to be reconstructed.

## 11.6 NUS

Non-uniform sampling (NUS) is a way to reduce measurement time of a 2D (or higher) experiment. It involves measuring only selected  $t_1$  delays, and using an algorithm to reconstruct the data. Reconstruction is necessary, as a regular Fourier transform can only deal with linearly sampled data. Most commonly, NUS is performed by choosing a regular delay time and number of points (i.e. a linear grid), and deleting selected values from this list. Processing NUS data is more involved than regularly measured data: the reconstruction process is not unique, and requires optimization.

Processing NUS data in ssNake requires the data to be reordered (using Matrix  $\rightarrow$  Reorder, and selecting the NUS list), in this way points that are not measured are taken as a 0. Now, phasing should be performed. The easiest way to do this is to do a regular Fourier transform. This will lead to ugly data, but is sufficient to perform some 0th order phasing. Do another Fourier transform to get back to the time domain data. After this, NUS processing can be performed using one of the following methods.

### 11.6.1 FFM

Fast Forward Maximum entropy (FFM, [1]) is a method to reconstruct the missing point in the FID by searching for the highest entropy. In our case, the entropy of the spectrum is defined as the sum of the absolute values of the complex data. FFM changes the values of the missing points in search for the highest entropy. This means that FFM reduces the noise in the spectrum (but adds uncertainty to the peak shapes and amplitudes) and favours narrow lines. Naturally, the points that *are* measured force the FFM analysis to create a spectrum that resembles the actual data.

For FFM, no additional parameters are needed, except for the NUS sampling scheme. Note that although the noise seems to be reduced by FFM, it is now equally contained in the peak shapes and amplitudes.

### 11.6.2 CLEAN

The CLEAN algorithm attempts to take single point in the spectrum, transfer part of it to a separate reconstructed spectrum, and subtract it from the main spectrum. The Fourier transform of the FID input mask (1's for measured points, 0's for skipped points) is used to not only subtract part of the spectrum maximum, but also the associated artifacts, that were introduced by the

NUS. After a maximum number of iterations (or if the residual spectrum maximum falls below a value) the loop is stopped, and the remainder is added to the reconstruction data. An inverse Fourier transform returns the reconstructed FID.

In short:

- Fourier transform to get the spectrum
- Select the maximum
- Multiply the maximum with the gamma factor and add it to the reconstructed spectrum (at that data point)
- Remove the  $\max * \gamma$  value (multiplied by the Fourier transform of the mask, and shifted for a peak at that position) from the spectrum

Steps 2–4 are repeated. Finally, the remained of the spectrum is added to the reconstruction.

The user needs to supply the sampling scheme, the gamma value (lower is better, but slower) and the threshold (stopping criterion in %, should be equal to the noise band for best results).

### 11.6.3 IST

Iterative Soft Thresholding (IST) is one of the methods to reconstruct the full FID from NUS sampled data. It involves the following steps:

- Fourier transform the FID
- Cut off the part of the spectrum that is above the threshold (in absolute mode)
- Add the cut off part to a separate array, and remove it from the spectrum
- Reconstruct the imaginary part of the spectrum via a Hilbert transform
- Inverse Fourier transform to get the FID
- Set the not measured parts back to zero
- (Repeat)

This is repeated for a specific maximum number of iterations, or until the residual spectrum is below a certain value. Then finally:

- The separate array is considered to be the reconstructed spectrum
- The remained of the spectrum is added to this (i.e. the noise)
- A Hilbert transform is used to reconstruct the imaginary data
- Inverse Fourier transform gives the reconstructed FID

The settings that need to be supplied by the user are:

- Positions of the spectra: list with the indices of the values that *have* been measured (the others will be zeroed every iteration).
- Threshold: at this relative intensity, the spectrum is cut (above goes to the separate array). Higher value (closer to 1) are more accurate, but more iterations are needed.
- Max. iterations: Maximum number of iterations, higher is better but slower.
- Stop when residual is below: stop when the spectrum has no points above value relative to the first spectrum (before any threshold steps). Value should be equal to the noise maximum. It is no problem if this value is too low (it might take longer). Having it too high leads to a bad reconstructions (if at 100%, no reconstruction is done at all!).

Reconstruction is done separately for every trace in the  $n$ D data. This also goes for the settings: the residual is compared to the first spectrum of *that* trace.

## 11.7 Hypercomplex

Hypercomplex data has a real and imaginary components along more than 1 dimension. For example, each datapoint from a 2D hypercomplex data set has four components: RR, RI, IR and II (R = Real and I = Imaginary). In NMR there are a number of ways to obtain the full hypercomplex data. This generally involves alternating the phase of some RF pulses. Depending on the way the data is recorded, a different procedure is required to construct the hypercomplex data. These procedures are supplied in the Hypercomplex menu.

When a specific dimension is hypercomplex, some methods will alter their behavior. For example, phasing along a hypercomplex dimension will rotate the real/imaginary values corresponding to that dimension. Also a Fourier transform will use the real/imaginary pairs of the dimension to be transformed (if available). The Side Frame shows whether a dimension is (hyper)complex by an asterix next to the dimension number. Using the Real, Imag or Abs routines will remove hypercomplex pairs from the active dimension. The last dimension is always complex and will not be removed, but instead the imaginary values will all be 0. In the case that the active dimension is not hypercomplex the complex pair of the last dimension is used for phasing, Fourier transforms, etc.

### 11.7.1 States

States conversion takes the alternating data set and makes new hypercomplex values. For 2D data this means that when performing states along D1, the two complex numbers are converted to a real part along that dimension:

$$RR_n + i \cdot RI_n = R_{2n} + i \cdot I_{2n} \quad .$$

$$IR_n + i \cdot II_n = R_{2n+1} + i \cdot I_{2n+1} \quad .$$

A new hypercomplex datapoint with index  $n$  is created from the two complex datapoints with indices  $2n$  and  $2n + 1$ .  $R$  is the real part and  $I$  the imaginary part.

### 11.7.2 States-TPPI

States-TPPI conversion takes the alternating data set and makes new complex values following the same procedure as States. After the procedure, the values at odd positions are multiplied with  $-1$  (and the even positions are retained).

### 11.7.3 Echo-antiecho

Echo-antiecho conversion takes the alternating data set and makes new complex values following:

$$RR_n + i \cdot RI_n = (R_{2n} + R_{2n+1}) + i \cdot (I_{2n} + I_{2n+1})$$

$$IR_n + i \cdot II_n = (I_{2n+1} - I_{2n}) + i \cdot (R_{2n} - R_{2n+1}) \quad .$$

## 12 Fitting

In NMR spectroscopy, the shape of the resonance lines can provide information on the strength of interactions that the nucleus experiences. ssNake can, as many other programs, fit some idealized lineshapes which are often encountered in real life spectra. Fitting in ssNake ranges from trivial ‘full width at half maximum’ (FWHM) fitting, to the complex quadrupolar Cziżek distribution. The following section will explain the way ssNake handles the different types of spectra it can fit, and which equations are used to simulate the different spectra.

### 12.1 Fitting basics

Roughly speaking, ssNake uses two types of fitting procedures: those with a mathematical solution, and those who have to be approximated iteratively. An example of the former type is the fitting of the width of a peak at half height (FWHM). This involves nothing more than finding the left and right side of the peak (at half height) and to calculate the width of this interval. This can be done in one easy mathematical procedure, and involves no iterative loop. The second type of fitting procedure is used for the powder lineshapes (e.g. quadrupolar and chemical shift lineshapes). With these problems, there is no direct mathematical solution. We are searching a parameter space for the solution that best fits the experimental spectrum. This is done by trying different settings, and qualifying how well these fit. Based on these attempts, a new setting is tested. This is continued until an optimum has been found. There are several ways to do such an iterative fitting: ssNake uses the Nelder-Mead algorithm [2] (also known as “Simplex”).

## Powder distributions and line shape fitting

ssNake supports fitting procedures for several idealized lineshapes commonly encountered in solid state NMR. In the following sections, the equations for these shapes will be given. In all cases, the fitted shape represents an ideal version assuming perfect excitation, and no mixed lineshape effects (e.g. quadrupolar and CSA at the same time). Note that the quality of the fit is determined by the digital resolution, which is the same as in the spectrum that is fitted. In many cases, convergence towards to optimum solution is difficult, as our fitting procedure is sensitive for local optima. It is therefore good practice to have a good initial guess. Use the ‘Sim’ button to simulate the lineshape, and set all variables to reasonable values.

## ZCW crystal files

Simulation of powder patterns requires averaging over the crystal angles. In practice, only a finite amount of angles (the polar angles  $\theta$  and  $\phi$ ) is selected and saved in the crystal file. There is no perfect way to do this, and therefore there exist multiple methods to do this. We use the ZCW method (named after Zaremba, Conroy, and Wolfsberg [3–5]), being both fast and accurate. The required input is a single number: the ‘Cheng’ number. This number specifies the amount of crystal orientations via the Fibonacci series, with Cheng(0) being ‘1’, Cheng(1) ‘2’ and subsequent values the sum of the two previous:

$$F_M = F_{M-1} + F_{M-2} \quad . \quad (6)$$

From this, a list  $js$  is made, going from 0 to 1 (not including) with the number of steps equal to the Fibonacci number. Phi is equal to:

$$\phi = \frac{2\pi}{c_2} \cdot \text{mod}(F_{M-1} \cdot js, 1) \quad (7)$$

with  $c_2$  a scaling factor (see below) and  $F_{M-1}$  the value of the Fibonacci series for Cheng – 1. In practice this means that (due to the ‘mod’), the list makes  $F_{M-1}$  passes through the region  $0:2\pi/c_2$ , with slightly different values for every pass. Theta is equal to:

$$\theta = \text{acos}(c_0(c_1 \cdot js - 1)) \quad . \quad (8)$$

In the above, the  $c$  parameters define the limits over which the angles are calculated. ssNake provides three options for this: a full sphere, a hemisphere (the upper half) and an octant (a quarter of the upper half). In the simulations that ssNake performs, the symmetry of the NMR interactions is high, and only the octant needs to be sampled. The  $c$  values are:

$$\mathbf{c} = \begin{cases} (1, 2, 1) & \text{full sphere} \\ (-1, 1, 1) & \text{hemisphere} \\ (-1, 1, 4) & \text{octant} \end{cases} \quad . \quad (9)$$

The source code of this can be found in `zcw.py`. For more information on powder averaging see the paper by Mattias Edén [6].

## Spherical tensors

The chemical shift anisotropy (CSA) and quadrupole simulation routines in ssNake make use of the spherical tensor description of these systems. Spherical tensors are a method of writing angular dependencies of spin interaction in such a fashion that rotations can be easily described. Rotations are necessary for simulations of powder patterns, where each possible orientation of the crystallites needs to be sampled (see the part on the ZCW scheme). This requires the NMR interaction equation to be rotated to the orientation of the specific crystallite.

If only this ‘crystal orientation’ rotation is considered, equations for CSA and the quadrupole interaction can be written down in a regular fashion (with sin and cosine terms), and are usually not too complicated. However, in cases where magic angle spinning is used, an extra rotation is required to account for the angle of the rotor, and the rotation of the rotor itself. The equations needed for this become complicated very quickly, and are rather inconvenient to handle.

For this purpose, it is more straightforward to work in the spherical tensor basis: a basis where rotations are easily defined. Writing a spin interaction (e.g. CSA) as a spherical tensor means we are decomposing the interaction into parts with different behaviour under rotation. For the chemical shift, this leads to a part that is constant under rotations (i.e. the isotropic part) with tensor rank 0, and a part that is a rank 2 tensor (which is influenced by rotations). Each of these parts has a spin term and a space term. Rotating the interaction rotates the space term of each rank.

In this way, every Hamiltonian can be described as a sum of inner products:

$$H = \sum_n T_n \bullet \Lambda_n \quad (10)$$

with  $T_n$  the spin part of rank  $n$ , and  $\Lambda_n$  the space part. For NMR interactions only ranks 0, 2 and 4 are relevant. Rotations are then easily described as a matrix multiplication:

$$\Lambda'_n = \Lambda_n \cdot D^n(\alpha, \beta, \gamma) \quad (11)$$

with  $D^n$  a Wigner rotation matrix, of which the elements are:

$$D^n_{m',m} = e^{-iam'} e^{-i\gamma m} d^n_{m',m}(\beta) \quad (12)$$

with  $d^n$  a Wigner small d-matrix. These terms can be found online. Note that rank 0 terms are not affected by rotations: they are scalars.

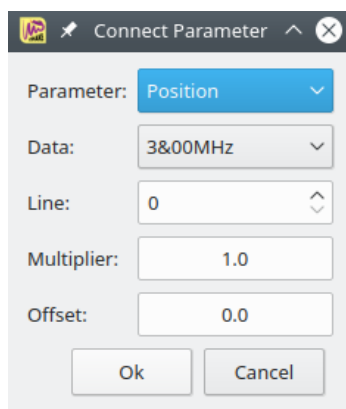
The Hamiltonian of a rotated interaction is therefore just calculated using the rotated spin part  $\Lambda'_n$  for each rank  $n$ , following [Equation 10](#).

## 12.2 Connecting parameters

In all iterative fitting routines supported by ssNake, it is possible to connect parameters, thereby using extra information that the user has about the system, and reducing the number of free parameters in the fit. This can for example be used to link integral values, which should be the same for multiple sites (i.e. in a solid material with a 1:1 ratio between different sites).



Linking parameters can be done by right-clicking on an input box in an iterative fitting routine, and selecting the option ‘Connect Parameter’. This gives the following pop-up window:



Here several values need to put in:

- Parameter: the name of the parameter that should be linked to
- Data: the name of the fitting data tab that should be linked to
- Line: the number of the row that should be linked to (first row is ‘0’)
- Multiplier: multiply the linked value by this number
- Offset: offset the linked value by this number

When doing this, the original parameter is replaced with the number ‘Multiplier ·  $x$  + Offset’ during the fit (with  $x$  the value of the parameter that is linked to).

When pushing ‘Ok’ in the pop-up window, the original value in the input box is replaced by a value like ‘(‘Integral’, 0, 1.0, 0.0, 0)’. The input represents: (‘Par name’, Line, Multiplier, Offset, Data index) The pop-up menu is essentially an easy tool to create this input.

Note that chain-linking parameters will lead to errors. That is, do not link par2 to par1, and par3 to par2. Always refer to the original parameter (link par2 to par1, and par3 to par1).

A tutorial on fitting using connected parameters can be found in the ‘MultiFitQuadrupole’ advanced tutorial.

### 12.3 Simultaneous fitting of multiple data sources

Apart from connecting parameters during a fit, as described in the previous section, ssNake also allows for the simultaneous fit of multiple data sources (i.e. spectra or time series). When fitting in such a case, ssNake tries to minimize the difference between all the experimental and simulated data. Fitting multiple data simultaneously only makes sense if there are connected parameters between the fitting parameters of the different data sets (otherwise the results are identical to the case where each data set is fit individually).

When in a fitting window, extra data can be added using the '+Add data+' tab that can be found in the vertical tabs on the left hand side of the window. Clicking this prompts a pop-up window, where the workspace name and the desired fitting routine can must be supplied. The new data tab will be added to the vertical tab bar.

The new data will have its own parameters. Note that in the 'extra' fitting tabs, several buttons are missing. These are universal buttons, which are only displayed in the first (i.e. master) fitting tab. Pushing 'Fit' fits all the data sets simultaneously.

Be very careful with the vertical scaling of the spectra. If spectrum1 has 10000x more intensity than spectrum2, spectrum2 will be almost ignored during the fit. To avoid this, scale the spectra to have the same integral (scaling can be done via Matrix → Normalize), or to have the same noise intensity. Tread carefully...

A tutorial on simultaneous fitting of multiple data sets can be found in the 'MultiFitQuadrupole' advanced tutorial.

## 12.4 S/N

S/N (or SNR) calculates the signal-to-noise ratio between a defined section of noise, and a peak. SNR is a useful way to describe the quality of a spectrum, with too low values being a sign for inaccurate data. In ssNake, opening the S/N tool creates a window, in which the limits of the section of noise has to be set, as well as the region with the peak of interest. Left clicking in the plot can set these values (noise limits first, then signal limits). Note that it is essential that the region that is specified as 'noise' really is noise. Any remaining signal will disrupt the SNR calculation. When the limits are set, ssNake directly calculates the SNR, using:

$$\text{SNR} = a_{\text{max}} / \text{std}(\text{NoiseRange}) \quad (13)$$

with  $a_{\text{max}}$  the maximum value within the range with the signal, and  $\text{std}(\text{NoiseRange})$  the standard deviation of the noise.

## 12.5 FWHM

FWHM, or Full Width at Half Maximum, is a tool to calculate the width of a peak. Selecting the tool in the menu creates a window, in which the boundaries of the peak of interest have to be set, for ssNake to search for the width of the peak. Make sure that there is only one peak in the selected window, otherwise the result might be inaccurate. The easiest way to select the regions is by left clicking in the spectrum. After valid input is given, ssNake directly calculates the FWHM in the units of the current axis. The order of input of the two limits is not important for the eventual calculation.

## 12.6 Centre of Mass

Calculates the centre of mass of a selected part of a spectrum or FID. For a symmetric peak, this is equal to the centre of the peak. However, for an asymmetric peak, identifying the centre is

not possible via visual methods and a centre of mass calculation is required. Opening the tool creates an input window, in which the limits have to be set (as with the FWHM tool). After these are set, the centre of mass is calculated in units of the current axis, using:

$$R = \frac{1}{M} \sum_i y_i \cdot x_i \quad , \quad (14)$$

with  $M = \sum y_i$  and  $y_i$  the y-value corresponding to x-value  $x_i$ .

### 12.7 Integrals

Using this tool, integral values of selected regions of the spectrum can be obtained. Click on the left and right side of a peak integrates this region, and displays the integral in the window, as well as drawing a curve in the plot. By overwriting one of the obtained integral values, all the results can be scaled: setting the first integral to 1 shows all the integrals relative to the first integral.

### 12.8 Relaxation Curve

This routine can be used to fit relaxation curves of time-domain data ( $T_1, T_2$ ). The equation used for this is:

$$y = \text{amp} \cdot (\text{const} + \text{coeff} \cdot \exp(-x/T)) \quad (15)$$

The standard values in ssNake are  $\text{const} = 1$  and  $\text{coeff} = -1$ , which are generally used in case of a saturation recovery  $T_1$  measurement. For inversion recovery,  $\text{const} = 1$  and  $\text{coeff} = -2$ . In case of a  $T_2$  experiment:  $\text{const} = 0$  and  $\text{coeff} = 1$ .

The axes of the plot can be set to a logarithmic scale. Negative number will not be shown in this view. When switching to the log axis, sometimes a recalculation of the curve is necessary to get a nice plot.

### 12.9 Diffusion Curve

This routine can be used to fit diffusion curves. The equation used for this is:

$$y = \text{amp} \cdot (\text{const} + \text{coeff} \cdot \exp(-(\gamma \delta x)^2 D (\Delta - \delta/3.0))) \quad (16)$$

Here,  $\gamma$  represent the  $\gamma$ -value of the isotope that is measured,  $\delta$  is the length of the gradient pulse,  $\Delta$  is the time between the gradient pulses (i.e. the time from the start of gradient pulse 1, to the start of gradient pulse 2).  $D$  is the diffusion constant, which is what we will fit. The 'const' and 'coeff' parameters have the same meaning as in the relaxation curve fit. Note that the x-axis should be in terms of the gradient strength (in Tesla). ssNake will still show 'seconds' as a unit though ( $1 \text{ s} = 1 \text{ T}$ ).

### 12.10 CSA

This fitting routine can be used to simulate lineshapes influence by chemical shift anisotropy (CSA), in both the static case, and under both infinite and finite MAS. An iterative fitting procedure is used for the fitting, so the initial guess should be somewhat close to the optimal solution. General values that need to be set are:

- The Cheng number (number of powder averages)
- The definition: the used chemical shift definition (see the part on on the CSA Utility for more information)
- The background (constant value added to all data points)
- The MAS method: static, finite or infinite
- The spinning speed in kHz (when MAS is on 'finite').
- The number of sidebands (when MAS is on 'finite'). This value must always be larger then the expected number of spinning sidebands. When too few sidebands are simulated, the sideband intensities will be off. Larger values take more time to simulate. For efficiency powers of two should be used.
- The rotor angle in radian (when MAS is not static). Default is the magic angle:  $\arctan(\sqrt{2})$

And for each site:

- The three chemical shift values (depend on the definition) in units of the current axis.
- Integral: total integral of the lineshape
- Lorentz: amount of Lorentzian linebroadening in Hz
- Gauss: amount of Gaussian linebroadening in Hz.

A maximum of 10 sites can be fitted simultaneously.

Peak picking can be used by left clicking on the spectrum. It takes three clicks (on different the discontinuities) to set the three chemical shift values. Note that this can only be used in the 11-22-33 or xx-yy-zz definition.

### Equations

The chemical shift interaction is described using the spherical tensor definitions discussed above. The equations are simplified by assuming that the magnetic field is along  $z$ , and by truncating the Hamiltonian to only parts that commute with the Zeeman interaction (i.e. commute with  $I_z$ ). (These are the regular approximations used in NMR)

The chemical shift has only a rank 0 and rank 2 tensor part. The terms are:

$$T_0 = \left( -\sqrt{\frac{1}{3}} I_z B_0 \right) \quad \Lambda_0 = \left( -\gamma \sqrt{\frac{1}{3}} (\sigma_{zz} + \sigma_{xx} + \sigma_{yy}) \right) \quad (17)$$

$$T_2 = \begin{pmatrix} 0 \\ 0 \\ \sqrt{\frac{1}{6}} 2I_z B_0 \\ 0 \\ 0 \end{pmatrix} \quad \Lambda_2 = \begin{pmatrix} \frac{1}{2} \gamma (\sigma_{xx} - \sigma_{yy}) \\ 0 \\ \gamma \sqrt{\frac{1}{6}} (2\sigma_{zz} - \sigma_{xx} - \sigma_{yy}) \\ 0 \\ \frac{1}{2} \gamma (\sigma_{xx} - \sigma_{yy}) \end{pmatrix} \quad (18)$$

Here, it is clear that the rank 0 term accounts for the isotropic chemical shift, while the rank 2 terms hold the anisotropic information. To calculate the actual NMR frequencies, the difference between the  $I_z$  value of two energy levels should be used. This is usually 1, as only single quantum transitions are observed.

### 12.11 Quadrupole

This fitting routine can be used to fit powder lineshapes of quadrupoles of any spin quantum number, either static, or under spinning (with finite or infinite spinning speed). Satellite transitions can be turned on or off. General values that need to be set are:

- The spin quantum number  $I$
- The MAS type: static, finite MAS or infinite MAS
- The satellites (on or off, note that for integer  $I$  values, they must be on)
- The Cheng number (number of powder averages)
- The rotor angle in radian (when MAS is not 'static'). Default is the magic angle:  $\arctan(\sqrt{2})$
- The number of sidebands (when MAS is on 'finite'). This value must always be larger than the expected number of spinning sidebands. When too few sidebands are simulated, the sideband intensities will be off. Larger values take more time to simulate. For efficiency powers of two should be used.
- The spinning speed in kHz (when MAS is on 'finite').
- The background (constant value added to all data points)

And for each site:

- Position: centre of the lineshape. This value is added to all  $x$ -values from the simulation. Scale is in Hz.
- $C_Q$ : quadrupolar coupling constant in MHz.

- $\eta$ : the asymmetry parameter
- Integral: total integral of the lineshape
- Lorentz: amount of Lorentzian linebroadening in Hz
- Gauss: amount of Gaussian linebroadening in Hz.

A maximum of 10 sites can be fitted simultaneously.

When simulating an infinite MAS spectrum, a more efficient routine can be used than for finite MAS. If spinning sidebands are not important in the calculation, it might be useful to switch to 'infinite' for speed considerations.

### Equations

Quadrupole systems are influenced by both the first and second order quadrupolar coupling. The first order quadrupolar coupling has only rank 2 terms, while the second order has rank 0, 2 and 4 terms.

*First order:*

$$T_2 = \begin{pmatrix} 0 \\ 0 \\ 3I_z^2 - I(I+1) \\ 0 \\ 0 \end{pmatrix} \quad \Lambda_2 = \frac{C_Q}{4I(2I-1)} \begin{pmatrix} \sqrt{\frac{1}{6}}\eta \\ 0 \\ 1 \\ 0 \\ \sqrt{\frac{1}{6}}\eta \end{pmatrix} \quad (19)$$

*Second order:*

$$T_0 = (I_z(I(I+1) - 3I_z^2)) \quad \Lambda_0 = B \left( -\frac{1}{5}(3 + \eta^2) \right) \quad (20)$$

$$T_2 = \begin{pmatrix} 0 \\ 0 \\ I_z(8I(I+1) - 12I_z^2 - 3) \\ 0 \\ 0 \end{pmatrix} \quad \Lambda_2 = B \begin{pmatrix} \frac{1}{7}\sqrt{\frac{3}{2}}\eta \\ 0 \\ \frac{1}{14}(\eta^2 - 3) \\ 0 \\ \frac{1}{7}\sqrt{\frac{3}{2}}\eta \end{pmatrix} \quad (21)$$

$$T_4 = \begin{pmatrix} 0 \\ 0 \\ 0 \\ 0 \\ I_z(18I(I+1) - 34I_z^2 - 5) \\ 0 \\ 0 \\ 0 \\ 0 \end{pmatrix} \quad \Lambda_4 = B \begin{pmatrix} \frac{1}{4\sqrt{70}}\eta^2 \\ 0 \\ \frac{3}{70}\sqrt{\frac{5}{2}}\eta \\ 0 \\ \frac{1}{140}(18 + \eta^2) \\ 0 \\ \frac{3}{70}\sqrt{\frac{5}{2}}\eta \\ 0 \\ \frac{1}{4\sqrt{70}}\eta^2 \end{pmatrix} \quad (22)$$

With  $B = \left( \frac{C_Q}{4I(2I-1)} \right)^2 \frac{2}{\nu_0}$ , where  $\nu_0$  is the Larmor frequency of the nucleus.

## 12.12 Czjzek

This fitting routine can be used to simulate the 1D spectrum of a quadrupolar nucleus that experiences a Czjzek distribution (normal or extended Czjzek). This fitting routine behaves differently than the ones described above, as it uses a library for simulating the spectra. Czjzek distributions are the sum of multiple quadrupolar spectra, with a different  $C_Q$  and  $\eta$  values, and scaled with the intensity that comes out of the distribution. To be able to simulate this in a reasonable time, it is convenient to precalculate all the individual spectra (for each  $C_Q$  and  $\eta$  pair). Fitting the spectrum leads to a new intensity distribution for each iteration, which, using the spectrum library, can easily be used to construct the Czjzek spectrum.

Using this fitting routine, first requires the generation of a library. Using the 'Library' button, a window is opened, in which some general quadrupolar settings have to be set (see the part about regular quadrupolar fitting). Apart from that, the  $C_Q$  and  $\eta$  grid have to be set: number of steps and minimum/maximum for each parameter. Note that the values of these parameters depend on the Czjzek distribution that is to be calculated (i.e. the  $\sigma$  value). When a distribution with a higher  $\sigma$  value is used, the largest  $C_Q$  value of the distribution should be higher, the properly sample the whole intensity distribution. To aid with this, the current distribution of each site can be viewed in this window (press 'Show' on the righthand side of the window to draw the contour plot). Press 'Generate' to generate the library, and then close the window.

The created library can now be used to simulate Czjzek spectra. For this, the following values need to be set:

- d: the order parameter (integer from 1 to 5)
- $\sigma$ : the width of the distribution in MHz

In case of an extended Czjzek (i.e. a distribution on top of a non-zero  $C_Q$  and  $\eta$  value):

- $C_Q0$ : the undistorted  $C_Q$ -value
- $\eta0$ : the undistorted  $\eta$ -value

Note that the extended Czjzek values will be activated when the ‘Type’ dropdown menu is changed to ‘Extended’. Extended Czjzek distributions take a long time to simulate.

Note that, if the parameters change much during a fit, the  $C_Q$  and  $\eta$  range of the Library might not be appropriate anymore. If this is the case, we advice to generate a new library with better settings, and then rerun the fit, to get the final convergence.

## Equations

The normal Czjzek distribution is defined as [7]:

$$A(C_Q, \eta) = \frac{C_Q^{d-1} \cdot \eta}{\sqrt{2\pi} \cdot \sigma^d} (1 - \eta^2/9) \cdot \exp \left[ \frac{-C_Q^2(1 + \eta^2/3)}{2\sigma^2} \right] \quad (23)$$

This equation gives the amplitude  $A$  for a specific  $C_Q$ ,  $\eta$ ,  $\sigma$  and  $d$ . ssNake scales the amplitude distribution to make the sum equal to 1.

For the extended Czjzek:

$$\begin{aligned} A(C_Q, \eta) = & \frac{C_Q^{d-1}}{\sigma^d} \eta \left( 1 - \frac{\eta^2}{9} \right) \int \int \int \sin(\beta) d\alpha d\beta d\gamma \\ & \exp \left\{ -\frac{1}{2\sigma^2} \left[ C_{Q,0}^2 \left( 1 + \frac{\eta_0^2}{3} \right) + C_Q^2 \left( 1 + \frac{\eta^2}{3} \right) \right. \right. \\ & \left. \left. - \frac{2}{\sqrt{3}} C_Q C_{Q,0} \left( \sqrt{3} a_{11} + \eta_0 a_{15} + \eta \left( a_{51} + \frac{\eta_0 a_{55}}{\sqrt{3}} \right) \right) \right] \right\} \end{aligned} \quad (24)$$

with

$$a_{11} = \frac{1}{2} (3 \cos^2 \beta - 1) \quad (25)$$

$$a_{15} = \frac{\sqrt{3}}{2} \sin^2 \beta \cos 2\alpha \quad (26)$$

$$a_{51} = \frac{\sqrt{3}}{2} \sin^2 \beta \cos 2\gamma \quad (27)$$

$$a_{55} = \frac{1}{2} \cos 2\alpha (1 + \cos^2 \beta) \cos 2\gamma - \sin 2\alpha \cos \beta \sin 2\gamma. \quad (28)$$

In this case, integration over three angles is required, which makes the calculation quite involved (and therefore slow). For the special case  $\eta_0 = 0$ , a faster equation can be used:

$$A(C_Q, \eta) = \frac{C_Q^{d-1}}{\sigma^d} \eta \left( 1 - \frac{\eta^2}{9} \right) \exp \left\{ -\frac{C_{Q,0}^2 + C_Q^2(1 + \frac{\eta^2}{3})}{2\sigma^2} \right\} \quad (29)$$

$$\cdot \int_0^1 I_0(z) \exp \left\{ \frac{C_Q C_{Q,0}}{2\sigma^2} (3t^2 - 1) \right\} dt \quad (30)$$



with  $I_0(z)$  a modified Bessel function, and

$$z = \eta \left| \frac{C_Q C_{Q,0}}{2\sigma^2} \right| (1 - t^2) \quad (31)$$

To speed-up the calculations of these distributions, ssNake can use the numba python library. With the extended Czjzek distribution, around a twofold speed increase can be gained in this way. ssNake tests for the numba package itself, so regular installation of this package should be sufficient.

### 12.13 External

Simulating NMR data for fitting can be straightforward, if idealized line shapes are required. These type of calculation are programmed into ssNake, and can be run using the different tools described above. However, if actual pulse sequences need to be calculated, the simulations become involved quite quickly. In order to be able to fit spectra which require a more involved calculation, the ‘External’ tool can be used to fit spectra using external programs.

Here we provide an example on how to use this tool together with the SIMPSON package [8–10]. For this, a SIMPSON input script is required which gives a single spectrum/FID as output. ssNake then loads this data, applies a Fourier transform (if necessary) and regrid the data to have the same  $x$ -axis as the experimental data.

As with the ‘Function Fit’ described below, the variables that we want to fit (or change) in the ssNake fit window must be defined as ‘@a@’ in the SIMPSON input file (where  $a$  can be any name). ssNake then recognizes the different variables defined in this way. When a simulation is run, ssNake replaces these variable names by the relevant number and performs a SIMPSON simulation using this input script. By default, ssNake also adds a scale factor, and Lorentzian and Gaussian apodization input boxes.

The window also features a dropdown box for the number of sites. When increasing this, new input boxes for all the relevant parameters appear. Note that ssNake is now running SIMPSON multiple times for each simulation: once for each site. Note that it is also possible to make SIMPSON run simulations with multiple sites in one go, but that this is really inefficient. Naturally, when spin-spin interactions are present (dipolar,  $J$ ) the relevant spins *should* be included in a single SIMPSON file.

If, for example, we want to fit the chemical shift anisotropy ( $\delta_{\text{aniso}}$ ) and asymmetry ( $\eta$ ), ( $\delta_{\text{aniso}}$ ) and asymmetry ( $\eta$ ), we could use the following SIMPSON spinsys:

```
spinsys {
  channels      1H
  nuclei        1H
  shift         1 100 @aniso@ @eta@ 0 0 0
}
```

Here, the isotropic shift ( $\delta_{\text{iso}}$ ) has been fixed at 100 Hz, while the anisotropy and eta can be changed from within ssNake. Note that the `@a@` variables can be used anywhere in the input file. Also, the used names can be anything (but be careful with special characters), but names like 'a' or 'agdv' might be difficult to recognise for the user in the ssNake window. A full input file could look like:

```
spinsys {
    channels      1H
    nuclei        1H
    shift         1 100 @aniso@ @eta@ 0 0 0
}

par {
    proton_frequency      600e6
    method                 direct
    sw                     1E6
    variable np           512
    crystal_file           zcw232
    gamma_angles           1
    start_operator         Inx
    detect_operator        Inp
    verbose                0000
    spin_rate              0
}

proc pulseq {} {
    global par
    delay [expr 1e6/$par(sw)]
    store 1
    reset
    acq $par(np) 1 0
}

proc main {} {
    global par
    set f [fsimpson]
    fzero fill $f 16384
    fft $f
    fsave $f $par(name).spe
}
```

Note that, when external files like user crystal files are used, the full path to this file must be included in the input file (ssNake copies the input file to a random directory, without any required files).

SIMPSON calculations can take a very long time. We therefore suggest to always optimize the input file as much as possible (minimum number of points and crystal orientations).

### 12.14 Function fit

While we try and include as many relevant fit function as possible, we cannot include everything in ssNake. In cases where the supplied functions are not useful, 'Function fit' can be used. Using this window, a function input by the user can be supplied. Pushing the 'Input Function' button opens a window where the required function can be supplied. The  $x$ -variable is the  $x$ -axis, and other variable can be supplied by the `@a@` notation (with  $a$  any variable name you like, with no special characters in the name). For example, let us say we want to fit a parabola with an offset. This would look like:

$$x ** 2 * @a@ + @b@ \quad (32)$$

with  $a$  and  $b$  the variable to be fit. Note the Python way of writing the square of  $x$  is `x ** 2`. Pushing Ok will load the function, and display  $a$  and  $b$  for the user to input (or fit). Note that the '@' signs are only used by ssNake to recognise which are variable in the equation.

## 13 Combine

The combine menu options allow the interaction between different workspaces like multiplication and addition. For some of these tools it is essential that the workspaces are the same size, or can be broadcasted. Broadcasting is described below.

### 13.1 Broadcasting

ssNake makes use of the general broadcasting rules of the numpy python package. Generally, data that has less dimensions than the target can be added (multiplied, etc) if the last dimensions agree on size. For example,  $3 \times 1024$  can be added to  $2 \times 3 \times 1024$  (the data is just stretched along a new third dimension). Alternatively, data that has a size 1 along a specific dimension can also be added:  $2 \times 1 \times 1024$  added to  $2 \times 10 \times 1024$  (again, the data is copied in the second dimension to reach length 10). Note that always the last dimensions are checked, so adding  $1024 \times 3$  to  $1024 \times 3 \times 10$  is not possible: checking is done from the right.

### 13.2 Combine Workspaces

Using Combine Workspaces opens a window that allows for the selection of multiple workspaces via dragging and dropping in the right-hand list. Only workspaces with the same size can be combined (and error will be given otherwise). The selected workspaces will be combined and

copied to a new workspaces, of which the name will be asked. Combining multiple sets of data this way requires the creation of a new dimension, which is always the new dimension D1. For example, combining two 1D data sets with 1024 points will lead to a combined data set of 2x1024 points (D1xD2). Combining two sets of 10x512 data points leads to a 3D set of 2x10x512.

### 13.3 Insert From Workspace

Insert From Workspace can be used to append data from another workspace to the current. For this, it is *not* necessary to have two Workspaces of exactly the same size. However, appending must be possible in some way. The way the data is appended depends on the active dimension. For example: the current data has size 2x1000 and D2 is viewed. Inserting a 1D data set of 500 points leads to a 2x1500 data set. Doing the same when viewing the D1 dimension gives an error. It would be possible to append a 1D set of 1000 points to this data from D1. This gives back a 3x1000 data set.

### 13.4 Add

Add the data of the selected workspace to the active workspace. The active dimension is not considered. Data of both workspaces must be the same size, or must be able to be broadcasted (see above).

### 13.5 Subtract

Same as add, but now subtracting.

### 13.6 Multiply

Same as add, but now multiplying the data sets. Note that ssNake by default has complex data. Using Multiply therefore does a complex multiplication. If this is not desirable, first use Tools → Real to get rid of the imaginary part of the data.

### 13.7 Divide

Same as add, but now dividing. Note that the data is complex, as described for Multiply.

## 14 Plot

The plot menu allows the user to change the view and nature of the current data. Several types, for both 1D and 2D views are supported. Apart from the view, the current x-axis can be changed, and a frequency reference can be set and saved.

### 14.1 1D Plot

Shows the default 1D plot with a solid line. The side window only shows the dimension selector, as there are no additional options.

### 14.2 Scatter Plot

This plot type is essentially the same as the 1D Plot. However, the data is now plot as points without a line. Scatter plots can be useful for viewing plots with only a couple of points.

### 14.3 Stack Plot

A stack plot is a way to view 2D data. Every trace is plot as a separate line, with a certain y-offset depending on its number. The y-axis in this view must be view with care, due to the offset given to each trace. In a stack plot, the dimension selector show two bullets: a left one for the main view, and the right one for the stack. For a T1 array, for example, having D2 as the first, and D1 as the right bullet shows a plot of the data along D2, with a separate spectrum (or FID) for every point in D1 (the arrayed recovery time in this case).

The side frame shows several options:

- From: index from where to start with the stack (zero is the start).
- To: index where to stop with the stack. Note that this should be 1 higher than the last point (it is to not including the shown value)
- Step: step size of the From to To array. 1 shows every spectrum, 2 only every second spectrum, etc.
- Spacing: the distance in units of y between each spectrum (or FID). This can also be scrolled using <shift – scroll>.

### 14.4 Array Plot

This plot is essentially the same as the Stack Plot, only now the spectra are plot next to each other. In this case, the labels of the x-axis are not shown anymore, as the do not make any sense. The label of the x-axis is changed to 'Frequency' or 'Time' to indicate a series of spectra or FID's. The x-limits of every subplot are taken from the view before changing to the Array Plot, and cannot be changed from the array plot.

The settings in the side frame are the same as with the Stack Plot.

### 14.5 Contour Plot

A contour plot shows the height profile of the data using constant height lines (like altitude lines on a map). This is the most common way to view true 2D data (data in which there are two NMR dimension, which are both Fourier transformed). Calculation of the contour lines takes some

time, especially for larger data sets. As with the Stack and Array plot, the dimension selector shows two bullets. Now they stand for the x-axis (left) and y-axis (right). The Get Position frame now shows some extra features, as each data point now has a x-value, y-value and amplitude (z-value). Note that you can change the colour scheme of the contour plot under Plot → Plot Settings.

The side frame has several options, which are described below.

### Contour type

- *Number*: the number of contour lines plot. Positive and negative lines are treated separately (so the real maximum is twice this number). Also, when the multiplier type is chosen, the total number of lines depends on whether or not the level maximum is exceeded.
- *Sign*: whether or not to plot only the positive contours (+ only), only the negative (− only) or both (Both).
- *Type*: Linear or multiplier. Linear has even spacing between the contour lines, and goes in ‘Number’ steps from the contour minimum to the maximum (for both the negative and positive contours). Multiplier takes the lowest contour and multiplies it with the Multiplier to get the 2nd contour. This is continues until the contour maximum, or the maximum number of lines is reached.
- *Multiplier (optional)*: Supplies the multiplier value as described above.

### Contour limits [%]

Sets the maximum and minimum contour. The maximum cannot be higher than 100%, and the minimum cannot be lower than 0%. Whenever the minimum is higher than the maximum, the meaning (but not the labels) is interchanged. The minimum contour level can be scrolled using the <shift – scroll> with the mouse wheel.

Two types are supported via the ‘Rel. to.’ dropdown menu: ‘Current 2D’ and ‘Full data’. ‘Current 2D’ sets the 100 % value of the contours equal to the maximum of the current shown 2D plot. ‘Full data’ displays it relative to the maximum of the entire dataset. This last option is particularly handy when view 3D data, were traces with no intensity compared to the data maximum are irrelevant, and should not show any contour levels.

### Projections

The projections are the regular 1D plots shown at the top and right of the contour plot. These can be of help in viewing the data. For both, several types are available:

- Sum: the sum of the data
- Max: the maximum of the data

- Min: the minimum of the data
- Off: no projection is displayed
- Trace: value of a particular trace

Sum, Max, and Min are applied along the axis opposite to the projection. So the top projection (which shows the x-axis) is summed (max/min) along the y-axis.

Alternatively, the Projection Ranges box can be toggled. This allows the user to specify between which data points the operation (sum/min/max) should be applied. This can be useful to get higher signal-to-noise level projections, by preventing contributions from noise-only areas.

The 'Trace' option sets the projection equal to a particular line in the 2D data. Using the 'Select Traces' button, a trace can be selected by clicking in the contour plot.

### Diagonal

Ticking this box draws a diagonal line across the 2D data at  $x = y$ . Note that the axis type is of importance: no scaling is performed. Therefore, if a data set with two identical axes is plot, only if the two units are the same, the diagonal is plot in a way that (probably) is the correct way. Alternatively, there is a box called 'Multiplier', in which a value can be put that multiplies the x-axis (only for the calculation of the diagonal).

## 14.6 2D Colour Plot

A 2D Colour plot shows the height profile of the data using a colour scheme. This plot type essentially show the same information as a Contour plot, but might be more applicable for certain types of data. It is used often for viewing NMR imaging (MRI) data.

As with the Contour plot, the 2D Colour plot shows projections, of which the settings can be changed in the side frame (see the Section on Contour Plot for more information).

## 14.7 Multi plot

A multi plot is an overlay of multiple workspaces. It shows the 1D Plot of the active workspace, and a button to add additional data (from other, or the same workspace). Things that can be changed are:

- Scale: linear multiplier of the signal (y value)
- Offset: gives an offset to the current data. The step size depends on the data, and is shown between brackets.
- Shift: horizontal shift in units of the current axis.
- Dimension selector: selects the dimension and trace to be shown (as with a regular 1D plot).

The added data can be removed by pushing the 'x' button.

## 14.8 User X-axis

This function opens a window where the current axis can be changed. This is commonly done in the time domain for relaxation data (as the axis is probably non-linear). The possible inputs are described below. Pushing OK applies the axis to the current data. To preview it, push enter while in one of the input boxes. The values are then printed in the table below.

Note that several actions on the data reset the  $x$ -axis (Fourier transform, for example). This is because these processes can only use the default axis. It is therefore wise to set the  $x$ -axis after all processing is done (i.e. before fitting, or printing a figure).

### 14.8.1 Expression

Allows input of a list of values via an input box. Allowed inputs are any Python commands that lead to a list of the correct length. Alternatively a space separated input can be given, e.g.: 0 0.1 0.2 0.4.

One extra command that we have included and regularly use is the 'euro' command. For relaxation measurements a list that follows the series of the euro coins is easy to remember and convenient. The series always start at 1, 2 or 5 (or 10 times lower/higher values). Having a euro series of length 5 starting at 0.1 therefore gives: 0.1, 0.2, 0.5, 1, 2. The euro series can be created with the command 'euro( $y$ )' with  $y$  the start value of the series.

### 14.8.2 Linear

A regular linear spacing. The start and end value needs to be supplied.

### 14.8.3 Logarithmic

A logarithmic spacing. The start and end value need to be supplied (and ssNake calculates the spacing). Note that negative values are not allowed.

## 14.9 Plot Settings

This opens a window where the plot settings of the active workspace can be changed. The input values are explained at File → Preferences.

# 15 History

## 15.1 History

Whenever ssNake performs an action on the data, a line is added to the history list to be able to show the processing history. In the History menu, these entries can be viewed. Saving the data (to a JSON or MATLAB file) also saves the history (but not the undo information). Note



that changing the view of the data is not an action, nor is fitting the data. Changing the spectral width and such is considered an action.

## 15.2 Error Messages

Whenever a wrong input is given to ssNake, a message is printed in the status bar (on the bottom of the main window). In Error Messages, these messages can also be found (printed in black), with a time stamp of their occurrence. Apart from these warning messages, there can also be more serious issues. These are python errors, and indicate that we did something wrong, or forgot to check if the input is correct. These messages are not displayed in the status bar, but only in the Error Messages window. They are printed in red, and when selected show some additional information in the bottom area of this window. Whenever a tool gives no, or an unpredictable result, consult this window for any errors. If such an error occurs you can email us with the specifics. For this, it is useful to be able to reproduce the error, otherwise it becomes difficult to find the cause.

## 16 Utilities

### 16.1 Chemical shift conversion tool

In NMR literature there are several definitions in use for describing the chemical shift tensor. This tool provides an easy way to inter convert them. The definitions are based on the very useful website of Klaus Eichele (<http://anorganik.uni-tuebingen.de/klaus/nmr/index.php?p=conventions/csa/csa>). In the most useful conventions, the three tensor values  $\delta_{xx}$ ,  $\delta_{yy}$  and  $\delta_{zz}$  are rewritten to an average value, a width and an asymmetry. Below follow the definitions of the different conventions.

The primary convention is called the ‘Standard Convention’. This convention just gives the three principal components of the tensor, without any restructuring, called  $\delta_{11}$ ,  $\delta_{22}$  and  $\delta_{33}$ . They are ordered in such a way that:  $\delta_{11} \geq \delta_{22} \geq \delta_{33}$ .

The second convention we call the ‘xyz Convention’. This is essentially the same as the Standard Convention, only the ordering is different. We now have  $\delta_{xx}$ ,  $\delta_{yy}$  and  $\delta_{zz}$ . With  $|\delta_{zz} - \delta_{iso}| \geq |\delta_{xx} - \delta_{iso}| \geq |\delta_{yy} - \delta_{iso}|$ . With  $\delta_{iso}$  the isotropic value, defined as the average of the three principal components. In this definition,  $\delta_{zz}$  is the value furthest away from the isotropic value,  $\delta_{yy}$  the closest, and  $\delta_{xx}$  the remaining value.

The third definition is the Haeberlen definition (or Haeberlen-Mehring-Spiess). Here, the principal components are ordered as in the xyz convention, only now they are redefined into a width, asymmetry and isotropic value. With  $\delta_{iso} = (\delta_{xx} + \delta_{yy} + \delta_{zz})/3$ ,  $\delta_{aniso} = \delta_{zz} - \delta_{iso}$  and  $\eta = (\delta_{yy} - \delta_{xx})/\delta_{aniso}$ . This way, the asymmetry parameter  $\eta$  always lies between 0 and 1. Not that due to the ordering of the principal components, the sign of the anisotropy (i.e. the width) is ill defined for  $\eta = 1$ .

The final definition is the Herzfeld-Berger definition. This convention uses the Standard Convention as a basis, and then defines the isotropic value as  $\delta_{\text{iso}} = (\delta_{11} + \delta_{22} + \delta_{33})/3$ , the span as  $\Omega = \delta_{11} - \delta_{33}$  and the skew as  $\kappa = 3(\delta_{22} - \delta_{\text{iso}})/\Omega$ . This way, the span is always positive, while the skew lies between -1 and 1.

### Use of the tool

Using the chemical shift conversion tool is quite easy. You can fill in all the necessary values for one convention, and push the ‘Go’ button next to it to convert it to all the other definitions. Note that ssNake also checks the definitions. So if you mess up the order of the Standard Convention, for example, it will be put in the correct order. The same is done for values that are outside the defined limits (e.g.  $\eta$  and  $\kappa$ ). Pushing ‘Reset’ will reset all the values to 0.

## 16.2 Quadrupole coupling conversion tool

Nuclei with a spin quantum number greater than 1/2 have an asymmetric charge distribution. This leads to a quadrupolar moment, and thus to a coupling between the nucleus and the electronic field gradient at the nuclear site. Just as for chemical shift, there are multiple convention in use to describe the resulting quadrupolar tensor. Apart from being a symmetric tensor, Laplace relation states that the sum of the field gradients in all direction should be zero (otherwise the nuclei would experience a net force). Due to this, there is no isotropic term, and the quadrupolar coupling can be described by two numbers: the strength and the asymmetry. Of these there are two conventions in use:  $C_Q$  and  $\omega_Q$ . When the field gradients in the three independent directions are  $V_{xx}$ ,  $V_{yy}$  and  $V_{zz}$  (with  $|V_{zz}| \geq |V_{yy}| \geq |V_{xx}|$ ),  $C_Q$  is defined as:

$$C_Q = \frac{eV_{zz}Q}{\hbar} = \frac{e^2qQ}{\hbar} \quad (33)$$

or the more commonly used  $C_Q/2\pi$  (which has Hz as unit):

$$C_Q/2\pi = \frac{eV_{zz}Q}{h} = \frac{e^2qQ}{h} \quad (34)$$

With  $e$  the elementary charge,  $Q$  the quadrupole moment (in  $\text{m}^2$ ),  $h$  the Planck constant (and  $\hbar$  the reduced Planck constant). Alternatively, a value that is scaled with the spin quantum number  $I$  is used:

$$\omega_Q/2\pi = \frac{3e^2qQ}{2I(2I+1)h} = \frac{3}{2I(2I+1)}C_Q/2\pi \quad (35)$$

For both definitions, the asymmetry is defined as:

$$\eta = \frac{V_{xx} - V_{yy}}{V_{zz}} \quad (36)$$

which, due to the ordering of the fields gradient components, is always between 1 and 0.

Generally, only the  $C_Q$  and  $\omega_Q$  definitions are used, and the field gradients themselves are not shown. However, if the quadrupolar coupling for a nucleus in a material is known, the field

gradients can be calculated by using the known quadrupolar moment of the nucleus. This way, the quadrupole coupling for another isotope at the same site in the material can be calculated by reversing the equation with a new moment and spin quantum number.

### Use of the tool

Using the quadrupole conversion tool is quite easy. You can fill in all the necessary values for one convention, and push the 'Go' button next to it to convert it to all the other definitions. Note that ssNake also checks the definitions. So if you have an asymmetry below 0, it will reorder the field gradients in such a way it lies between 1 and 0 again. To calculate the field gradients from the quadrupole coupling constant (or the other way around), the quadrupolar moment should be given (which starts at 'ND', being 'not defined'). When starting from the field gradients, ssNake checks if they sum to 0 (which they should). If not, the difference is subtracted from the three values, to make sure it is. Note that only the absolute quadrupolar coupling is found in NMR, so pay no heed to the absolute sign of the field gradients.

### 16.3 NMR table

This tool provides an interactive way to find and calculate NMR frequencies and other characteristic values. The main view is the periodic table, showing the NMR frequency in MHz at a specific magnetic field for the most common isotope of each element. Changing one of the values (and pressing enter) updates all other values. Most commonly, either the  $B_0$  field (in the top left corner) or the  $^1\text{H}$  frequency is supplied by the user. Each element also has a coloured box representing the spin quantum number (of the most common isotope). Apart from the  $B_0$  field, the top left corner also shows the electron frequency in GHz.

Additional details on specific element can be obtained by double clicking on the element title, or by clicking the 'Details' button. This creates a pop-up box showing these details. Navigating between the elements can be done from this box via the element number or abbreviation. Show values are:

- Mass
- Spin quantum number ( $I$ )
- Natural abundance and radio-active life-time (when applicable)
- Gyromagnetic ratio ( $\gamma$ ) in  $10^7 \text{ rad s}^{-1} \text{ T}^{-1}$
- Quadrupole moment ( $Q$ ) in  $\text{fm}^2$  (when applicable)
- NMR frequency at the given magnetic field in MHz (based on the frequency ratio  $\Xi$  with respect to  $^1\text{H}$ )
- Sensitivity relative to a specific nucleus. The sensitivity is taken to be proportional to  $\gamma^3 I(I + 1)$ , scaled with the natural abundance.

- Reference sample for ppm scale
- Reference sample condition
- Line-width factor for quadrupole nuclei ( $I > 1/2$ ). It is defined as  $\frac{Q^2(2I + 3)}{I^2(2I - 1)}$

The leftmost entry of the details window is a bullet, which can be used to set the preferred isotope of this element in the periodic table view.

The shown frequencies are those of 0 ppm for that nucleus, at the specified magnetic field. If an experimental frequency of TMS  $^1\text{H}$  is filled in the details box of  $^1\text{H}$ , the frequency of 0 ppm for all other nuclei can be read from their respective boxes (in the details, where enough digits are displayed).

## 17 Help

### 17.1 Update

This tool can be used to update ssNake via the github page. It show a dropdown menu with all the available versions. We recommend using that latest version (that has a version number). The 'develop' version is the version we are currently working on. This has the most recent features, but might be unstable. Note that updating via this tool does not always work (ssNake needs write permission on its own directory). If ssNake does not work after using the Update tool, you can download it from the github page: <https://github.com/smeerten/ssnake/>.

### 17.2 About

Opens the About box with information on the version, acknowledgements and license (GPL v3.0).

## 18 Contact

To contact the ssNake team write to `ssnake@science.ru.nl`.

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