



# **Single cell RNA sequencing data analysis**

## **Practical exercises**

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## Practical on zoom

- Work in groups of 5, the idea is that you can help each other out and discuss results.
- One zoom breakout room per group, you should have already been assigned to a group in slack. Please stay online in the breakout room for your group.
- Come to main room for question or tell us on slack and we can come to your breakout room.
- We have some extra rooms where we can sit with a student if there are specific questions.

# <https://nbisweden.github.io/workshop-scRNAseq/exercises>

## Tutorial



Quality Control

[Seurat\\_qc \(.Rmd\)](#)

[Scater\\_qc \(.Rmd\)](#)

[ScanPY\\_qc \(.ipynb\)](#)



Dimensionality reduction

[Seurat\\_dr \(.Rmd\)](#)

[Scater\\_dr \(.Rmd\)](#)

[Scanpy\\_dr \(.ipynb\)](#)



Data integration

[Seurat\\_integr \(.Rmd\)](#)

[Scater\\_integr \(.Rmd\)](#)

[Scanpy\\_integr \(.ipynb\)](#)



Clustering

[Seurat\\_clust \(.Rmd\)](#)

[Scater\\_clust \(.Rmd\)](#)

[Scanpy\\_clust \(.ipynb\)](#)



Differential expression

[Seurat\\_dge \(.Rmd\)](#)

[Scater\\_dge \(.Rmd\)](#)

[Scanpy\\_dge \(.ipynb\)](#)



Celltype prediction

[Seurat\\_ct \(.Rmd\)](#)

[Scater\\_ct \(.Rmd\)](#)

[Scanpy\\_ct \(.ipynb\)](#)



Spatial transcriptomics

[Seurat\\_ST \(.Rmd\)](#)

[Scater\\_ST \(.Rmd\)](#)

[Scanpy\\_ST \(.ipynb\)](#)



Trajectory inference

[Slingshot\\_ti \(.Rmd\)](#)

[Slingshot\\_ti](#)

[PAGA\\_ti](#)

# Three main pipelines for analysing single cell data:

- Seurat:
  - R based, centered around Seurat objects.
  - Mainly developed for droplet based data
  - Easy to use, recommended for R beginners
  - Cons: uses a LOT of memory
- Scrان:
  - R based, centered around SingleCellExperiment objects
  - Has more different statistical methods
  - Can handle spike-ins
  - Cons: More complicated than Seurat to run.
- Scanpy:
  - Python based
  - Handles large datasets better.
  - Cons: Requires quite some python knowledge. Does not yet have all the functionality of the R based tools

# Seurat object

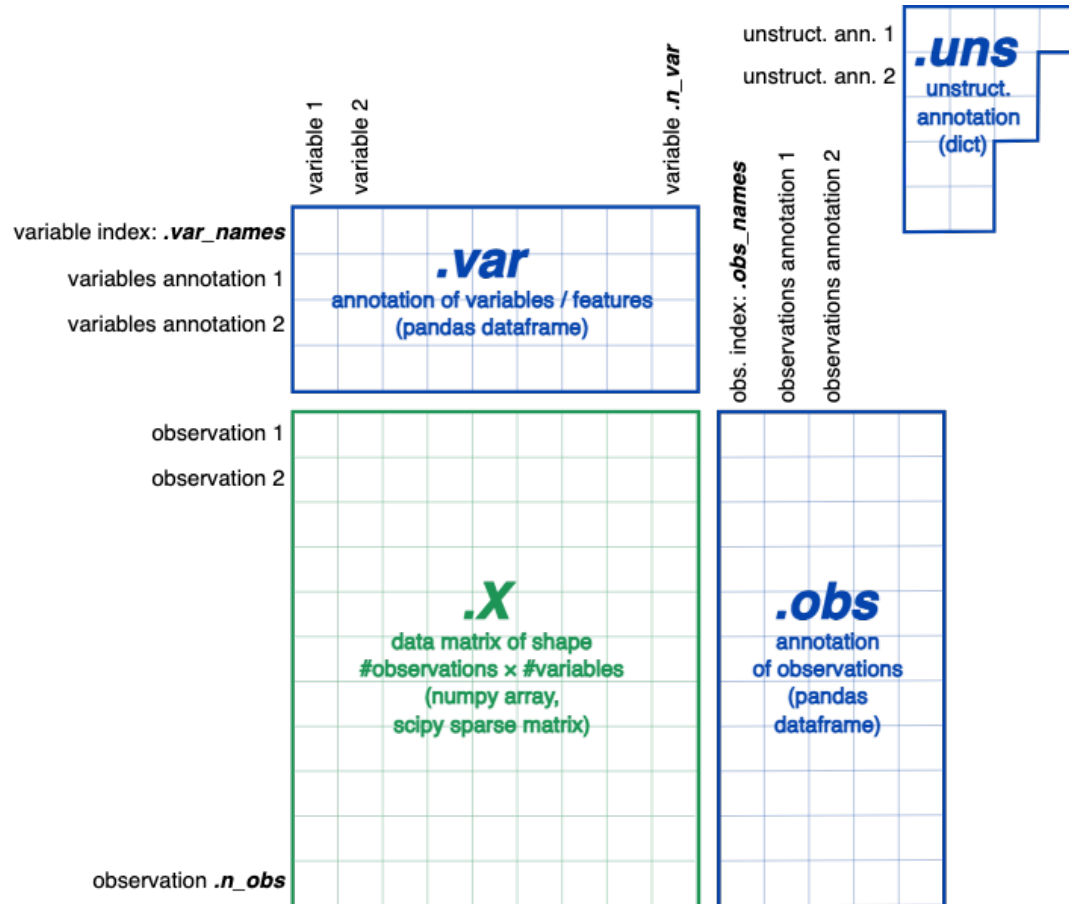
| Slot                      | Function  |
|---------------------------|---|
| <code>assays</code>       | A list of assays within this object   |
| <code>meta.data</code>    | Cell-level meta data  |
| <code>active.assay</code> | Name of active, or default, assay   |
| <code>active.ident</code> | Identity classes for the current object   |
| <code>graphs</code>       | A list of nearest neighbor graphs   |
| <code>reductions</code>   | A list of DimReduc objects  |
| <code>project.name</code> | User-defined project name (optional)  |
| <code>tools</code>        | Empty list. Tool developers can store any internal data from their methods here |
| <code>misc</code>         | Empty slot. User can store additional information here                          |
| <code>version</code>      | Seurat version used when creating the object                                    |

# SingleCellExperiment (SCE) objects

```
## class: SingleCellExperiment
## dim: 611 379
## metadata(2): SuppInfo which_qc
## assays(3): tophat_counts logcounts counts
## rownames(611): 0610007P14Rik 0610009B22Rik ... 9930111J21Rik1
##    9930111J21Rik2
## rowData names(0):
## colnames(379): SRR2140028 SRR2140022 ... SRR2139341 SRR2139336
## colData names(22): NREADS NALIGNED ... Animal.ID passes_qc_checks_s
## reducedDimNames(2): PCA TSNE
## altExpNames(3): ERCC RIKEN original
```

<https://bioconductor.org/packages/release/bioc/vignettes/SingleCellExperiment/inst/doc/intro.html>

# AnnData (Scanpy) objets



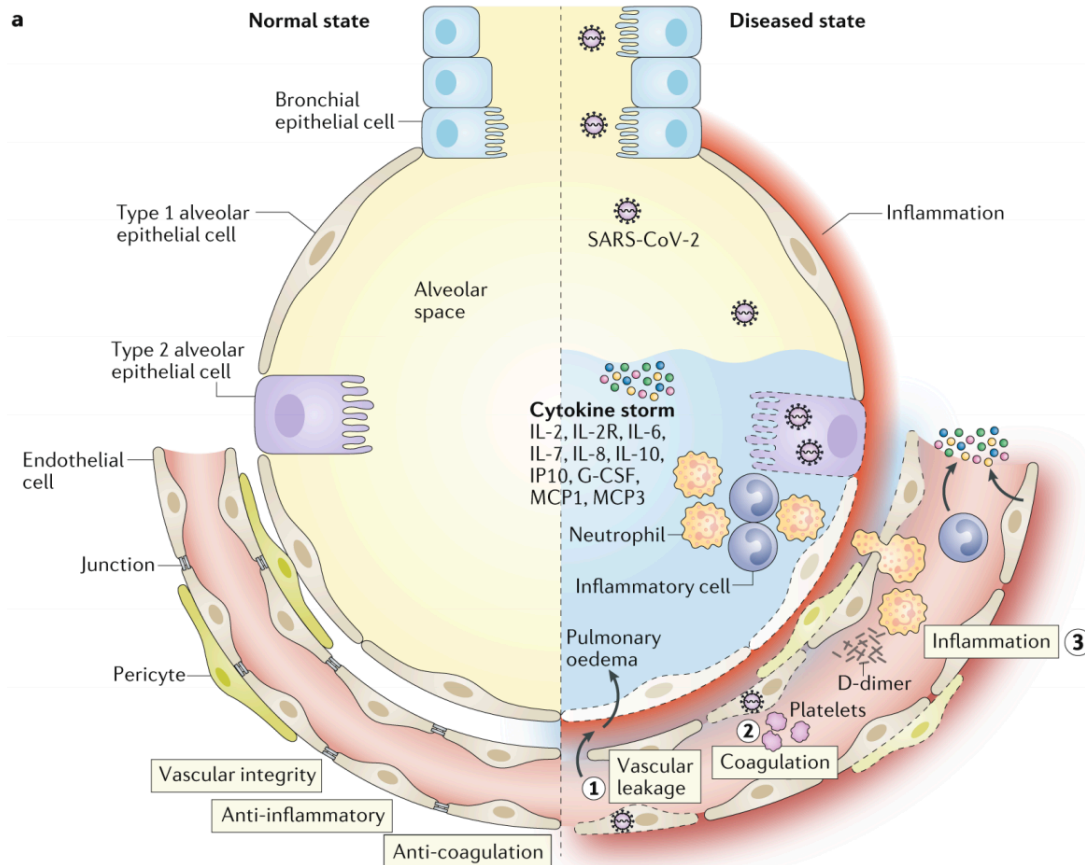
[https://anndata.readthedocs.io/en/latest/ann\\_data.AnnData.html](https://anndata.readthedocs.io/en/latest/ann_data.AnnData.html)

# What to chose?

- It is recommended that you go through all the steps with one pipeline as each exercise depends on saved objects from the previous step.
- Everyone works in very different pace. Focus on one of the pipelines first. If you have time left over, you can also try out the other ones.



# The datasets – Covid-19 PBMCs



Teuwen et al (2020) *Nat reviews Immunology*

Elderly patients usually develop severe lung inflammation and lung disfunction.

Many cell types orchestrate the immune response to the virus.

Their relative contribution at the single-cell resolution is still unclear

# The datasets – Covid-19 PBMCs

- Data from paper: "Immunophenotyping of COVID-19 and influenza highlights the role of type I interferons in development of severe COVID-19" Lee et al. Sci Immuno
- We have selected 3 controls and 3 severe covid samples and subsampled to 1500 cells per subject for computational speed/memory.
- ST and trajectory lab will be with other datasets.

# Installation of all packages

- We have created a conda environment for the course that should contain all packages you need for the exercises
- However, for slingshot trajectory inference lab, there is an additional conda environment that needs to be installed.
- If you chose to instead work with standard R installations, you can use the list of required packages in the environment file and install them on your own.

# Why conda?

- Often easier installations compared to traditional R installation for packages with C-compilation etc.
- Good way to manage different versions of packages in different projects.
- There are other ways of managing packages. E.g packrat for R, pyenv for python etc.

## The code:

- All code for the exercises is available as R-markdown documents, or jupyter notebooks, in the folder:  
**workshop-scRNAseq/labs/compiled/**
- Please report to us if you find any errors in the code!
  - Slack channel **#exercises**
  - An Issue on the github page.
- We may find bugs and update the code – in that case, update your git repo with command `"git pull"`

# Reproducible coding

- You should always be able to find and recreate the results.
  - Scripts should be able to run from input files to create the output.
  - Never work with saved R sessions!
- Name your scripts with relevant names so you can find them 2 years later 😊
- Always backup code – good idea to use github that also gives you version control.

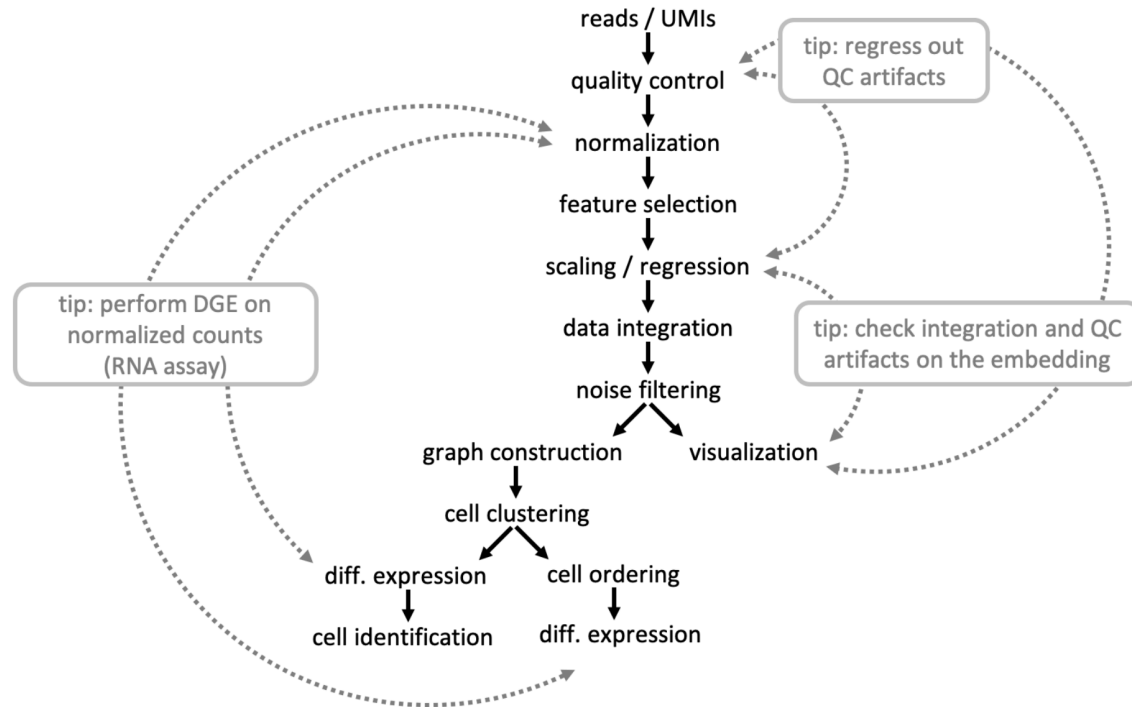
# Memory issues in R

- scRNAseq datasets are often large, think about how you code. Avoid duplicating objects!
- Remove unused matrices and clear memory with `gc()`.
- Most packages store scRNAseq as sparse matrices, will require package Matrix or similar for many standard matrix operations.
- If you still have issues with memory in R, test setting e.g. `R_MAX_VSIZE=70Gb` in the `.Renv` file. Default is 16Gb.

# Troubleshooting

- Slack channel - **#exercises**
- It is important that you learn how to troubleshoot yourselves.
  - Look at your error messages, perhaps the answer is there?
  - If not – Google is your best friend! Forums like Seqanswers, Stackexchange, Bioconductor support forum, specific forums (or github issues) for each package may have the answer.
- TAs are there to answer any questions and give suggestions, but we may not always have the answer.





## Downloading data

► Running bash code in RStudio

## Seurat Objects

# Rmarkdown demonstration