For all migration experiments dendritic cells were used at **11 days** after differentiation. Oil droplets are made from soybean oil in an aqueous continuous phase containing 15 % w/w of Poloxamer 188 block polymer surfactant and **2 % w/w** sodium alginate. The rough emulsion was sheared in a Couette cell apparatus at a controlled shear rate of **250 rpm**. With this condition diameter of droplets is of 4-5 micrometer (eventually, we also have 5-6 micrometer drops, made with 200 rpm).  For live imaging studies droplets were first **“blocked and stained”** with Nile Red in cell growth complete media containing 10µM of Nile Red for 30 minutes.

PDMS chips chosen are composed of parallels channels of **5x5 micrometers** fresh glowed on Fluorodish F35. Immediately after mounting chips were let stay **30 min under vacuum.** This latest step can be done in parallelof the drop “block and stain” step.After such pre-incubation drops were charged at a 1:100 dilution into **dry** chip **(immediately after vacuum**).  Note that no fibronectin was used.

Cells were loaded after drops, at a concentration of **105 cells per well of 2,5 mm** diameter and let recover in incubator for **3h** before imaging.

The event we wish to monitor, the first contact between the cell and the drop it is **quite rapid** (5-7 minutes). Thus for movies, **a 30’’ time frame** is the minimum.