

# Genome-wide assessment of differential translations with ribosome profiling data – the xtail package

Zhengtao Xiao<sup>1–3</sup>, Qin Zou<sup>1,3</sup>, Yu Liu<sup>1–3</sup>, and Xuerui Yang<sup>1–3</sup>

<sup>1</sup>MOE Key Laboratory of Bioinformatics,

<sup>2</sup>Tsinghua-Peking Joint Center for Life Sciences,

<sup>3</sup>School of Life Sciences, Tsinghua University, Beijing 100084, China.

May 4, 2016

## 1 Introduction

---

This package, Xtail, is for identification of genes undergoing differential translation across two conditions with ribosome profiling data. Xtail is based on a simple assumption that if a gene is subjected to translational dysregulation under certain experimental or physiological condition, the change of its RPF abundance should be discoordinated with that of mRNA expression. Specifically, Xtail consists of three major steps: (1) modeling of ribosome profiling data using negative binomial distribution (NB), (2) establishment of probability distributions for fold changes of mRNA or RPF (or RPF-to mRNA ratios), and (3) evaluation of statistical significance and magnitude of differential translations. The differential translation of each gene is evaluated by two pipelines: in the first one, Xtail calculated the posterior probabilities for a range of mRNA or RPF fold changes, and eventually established their probability distributions. These two distributions, represented as probability vectors, were then used to establish a joint probability distribution matrix, from which a new probability distribution were generated for differential translation. The P-values, point estimates and credible intervals of differential translations were then calculated based on these results. In the other parallel pipeline, Xtail established probability distributions for RPF-to-mRNA ratios in two conditions and derived another distribution for differential translation. The more conserved set of results from these two parallel pipelines was used as the final result. With this strategy, Xtail performs quantification of differential translation for each gene, i.e., the extent to which a gene's translational rate is not coordinated with the change of the mRNA expression.

By default, Xtail adapts the strategy of DESeq2 [1] to normalize read counts of mRNA and RPF in all samples, and fits NB distributions with dispersions  $\alpha$  and  $\mu$ .

This guide provides step-by-step instructions on how to load data, how to execute the package and how to interpret output.

## 2 Data Preparation

The Xtail package uses read counts of RPF and mRNA, in the form of rectangular table of values. The rows and columns of the table represent the genes and samples, respectively. Each cell in the  $g$ -th row and the  $i$ -th columns is the count number of reads mapped to gene  $g$  in sample  $i$ .

Xtail takes in raw read counts of RPF and mRNA, and performs median-of-ratios normalization by default. This normalization method is also recommend by Reddy R. [2]. Alternatively, users can provide normalized read counts and skip the built-in normalization in Xtail.

In this vignette, we select a published ribosome profiling dataset from human prostate cancer cell PC3 after mTOR signaling inhibition with PP242 [3]. This dataset consists of mRNA and RPF data for 11391 genes in two replicates from each of the two conditions("treatment" vs. "control").

## 3 An Example

Here we run Xtail with the ribosome profiling data described above. First we load the library and data.

```
library(xtail)
data(xtaildata)
```

Next we can view the first five lines of the mRNA (mrna) and RPF (rpf) elements of xtaildata.

```
mrna <- xtaildata$mrna
rpf <- xtaildata$rpf
head(mrna,5)

##               control1 control2 treat1 treat2
## ENSG000000000003      825      955      866      1039
## ENSG000000000419     1054      967      992      888
## ENSG000000000457       71       75      139       95
## ENSG000000000460      191      162      199      201
## ENSG000000000971       81        2       88       11

head(rpf,5)

##               control1 control2 treat1 treat2
## ENSG000000000003      143      302      197      195
## ENSG000000000419      234      481      383      306
## ENSG000000000457       12       17       17       15
## ENSG000000000460       45       88       63       37
## ENSG000000000971       31        7       36        2
```

We assign condition labels to the columns of the mRNA and RPF data.

```
condition <- c("control","control","treat","treat")
```

Next, we run the main function, xtail(). By default, the second condition (here is "treat") would be compared against the first condition (here is "control"). Those genes with the minimum average expression of mRNA counts and RPF counts among all samples larger than 1 are used (can be changed by setting minMeanCount). All the available CPU cores are used for running program. The argument "bins" is the number of bins used for calculating the probability densities of log2FC and log2R. This paramater will determine accuracy of the final pvalue. Here, in order to keep the run-time of this vignette short, we will set bins to "1000". Detailed description of the arguments of the xtail function can be found by typing ?xtail or help(xtail) at the R prompt.

```
test.results <- xtail(mrna,rpf,condition,bins=1000)

## Calculating the library size factors
## 1. Estimate the log2 fold change in mrna
## 2. Estimate the log2 fold change in rpf
## 3. Estimate the difference between two log2 fold changes
## 4. Estimate the log2 ratio in first condition
```

```
## 5. Estimate the log2 ratio in second condition
## 6. Estimate the difference between two log2 ratios
## Number of times the log2FC and log2R used in determining the final p-value
## log2FC: 2199
## log2R: 9192
```

Now we can examine the first five lines of the results produced by the 'xtail' run.

```
head(test.results,5)

##           log2FC_TE_v1 pvalue_v1 log2FC_TE_v2 pvalue_v2 pvalue_final pvalue.adjust
## ENSG000000000003      0.05867991 0.8162414      0.0623457 0.8079550      0.8162414      0.9957191
## ENSG000000000419      0.35599462 0.1536697      0.3592684 0.1360275      0.1536697      0.9957191
## ENSG000000000457     -0.29931892 0.5865027     -0.2897093 0.6409663      0.6409663      0.9957191
## ENSG000000000460     -0.31079662 0.4203029     -0.3109160 0.4249511      0.4249511      0.9957191
## ENSG000000000971     -0.62232332 0.7105031     -0.6457720 0.7443768      0.7443768      0.9957191
##           log2FC_TE_final
## ENSG000000000003      0.05867991
## ENSG000000000419      0.35599462
## ENSG000000000457     -0.28970931
## ENSG000000000460     -0.31091597
## ENSG000000000971     -0.64577202
```

The results of first pipeline are named with suffix "\_v1", which are generated by comparing mRNA and RPF log2 fold changes: The element log2FC\_TE\_v1 represents the log2 fold change of TE; OVL\_v1 is overlap coefficient, which quantify the statistical confidence of difference between two distributions, here is difference of log2 fold change of mRNA and RPF. The pvalue\_v1 represent statistical significance. The second pipeline are named with suffix "\_v2", which are derived by comparing log2 ratios between two conditions: log2FC\_TE\_v2, OVL\_v2, and pvalue\_v2 are log2 ratio of TE, overlap coefficient, and pvalues. Finally, the more conserved results (with larger-Pvalue) was select as the final assessment of differential translation, which are named with suffix "\_final". The pvalue.adjust is the estimated false discovery rate corresponding to the pvalue\_final. The CI is the credible interval of log2FC\_TE\_final (95% by default), and the level of confidence can be changed by setting ci in xtail function.

Finally, the plain-text file of the results can be exported using the functions `write.csv` or `write.table`.

```
write.table(test.results, "test_results.txt", quote=F, sep="\t")
```

## Session Info

```
sessionInfo()

## R version 3.2.4 (2016-03-10)
## Platform: x86_64-apple-darwin13.4.0 (64-bit)
## Running under: OS X 10.11.4 (El Capitan)
##
## locale:
## [1] C
##
## attached base packages:
## [1] parallel stats4      stats      graphics  grDevices  utils      datasets  methods
## [9] base
##
## other attached packages:
## [1] xtail_1.1.4           DESeq2_1.10.1          RcppArmadillo_0.6.100.0.0
## [4] Rcpp_0.12.1           SummarizedExperiment_1.0.0 Biobase_2.30.0
## [7] GenomicRanges_1.22.0  GenomeInfoDb_1.6.0      IRanges_2.4.1
## [10] S4Vectors_0.8.0       BiocGenerics_0.16.0     knitr_1.11
##
## loaded via a namespace (and not attached):
```

```
## [1] RColorBrewer_1.1-2    formatR_1.2.1      futile.logger_1.4.1  highr_0.5.1
## [5] plyr_1.8.3           XVector_0.10.0     futile.options_1.0.0  tools_3.2.4
## [9] zlibbioc_1.16.0      rpart_4.1-10       digest_0.6.8         RSQLite_1.0.0
## [13] annotate_1.48.0       evaluate_0.8        gtable_0.1.2         lattice_0.20-33
## [17] DBI_0.3.1            proto_0.3-10       gridExtra_2.0.0      genefilter_1.52.0
## [21] cluster_2.0.3        stringr_1.0.0      locfit_1.5-9.1       nnet_7.3-12
## [25] grid_3.2.4           AnnotationDbi_1.32.0 XML_3.98-1.3         survival_2.38-3
## [29] BiocParallel_1.4.0   foreign_0.8-66     latticeExtra_0.6-26  Formula_1.2-1
## [33] geneplotter_1.48.0   ggplot2_1.0.1      reshape2_1.4.1       lambda.r_1.1.7
## [37] magrittr_1.5         Hmisc_3.17-0       scales_0.3.0         splines_3.2.4
## [41] MASS_7.3-45          xtable_1.7-4       BiocStyle_1.8.0      colorspace_1.2-6
## [45] stringi_1.0-1        acepack_1.3-3.3    munsell_0.4.2
```

## References

---

- [1] Love MI, Huber W, Anders S: *Moderated Estimation of Fold Change and Dispersion for RNA-Seq Data with DESeq2*. Genome Biology 2014, 15:550. A Comparison of Methods: Normalizing High-Throughput RNA Sequencing Data.
- [2] Reddy R: *A Comparison of Methods: Normalizing High-Throughput RNA Sequencing Data*. Cold Spring Harbor Labs Journals. bioRxiv 2015:1-9.
- [3] Hsieh AC, Liu Y, Edlind MP, et al.: *The translational landscape of mTOR signaling steers cancer initiation and metastasis*. Nature 2012, 485:55-61.