



KAGGLE MABe – SOCIAL ACTION RECOGNITION IN MICE PRESENTATION

<https://www.youtube.com/watch?v=tTyxZPQVIkM>

EN.605.742.81: Deep Neural Networks

Professor: Oleg Melnikov

Team Members: XiangFeng Liang, Lijian Gong, Swarup Sahu



JOHNS HOPKINS
WHITING SCHOOL
of ENGINEERING



Background

JHU-Team 1 Participation Overview

- Team members: Swarup Sahu, Xiang Feng Liang, Lijian Gong
- Competition started on September and concludes on December 15, 2025
- Prize pool totals \$50,000 with 1280 teams on leaderboard
- Consent granted to share presentations for educational use after December 2, 2025
- Project completed as part of Johns Hopkins University DNN 605.742 course



(AI generated image)

Key Acronyms in Our Project

CPU/GPU/TPU stand for central, graphical, and tensor processing units.

FP, FPIP, FPVP: course acronyms for to final, interim, and video project presentations.

RAM: short for random access memory in computing.

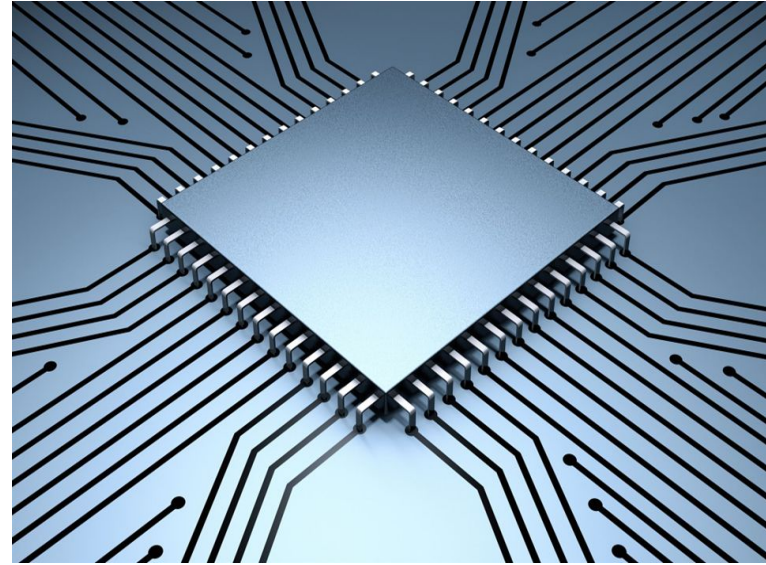
TF: Tensorflow, open-source machine learning library developed by Google.

URL: represents uniform resource locator, a web address.

MS PPT: Microsoft Powerpoint*

LB: leaderboard tracking within a competition's standings.

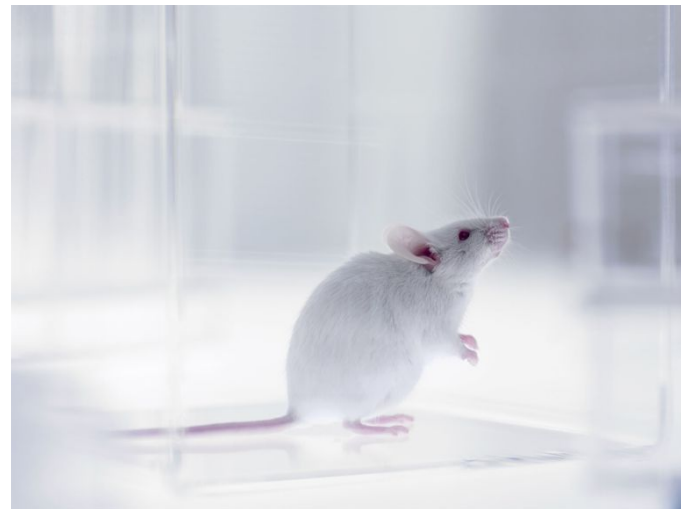
F1-Score: competition metric. Calculated by using recall & precision metrics



(AI-generated image)

Advancing Mouse Behavior Analysis with Machine Learning

- This competition is sponsored by Cornell University
- Develop a model to recognize over 30 mouse behaviors from video footage data
- Leverage a comprehensive dataset with 400+ hours of mice actions & tracked movements using 20+ different recording systems
- Overcome challenges of limited labeled data and rare mouse behaviors (ex: 'rest' or 'freeze')
- Aim for robust cross-lab generalization to ensure wide applicability
- Aim of competition is to automate large-scale behavior analysis to support neuroscience and ecology



(AI-generated image)

Training Infrastructure & Dataset Overview:

- Kaggle notebooks must complete runs within 9 hours on CPU or GPU
- Kaggle notebooks run in a fully offline environment — no internet access is allowed during execution.
- Typical RAM available: ~13 GB for CPU, ~16 GB for GPU notebooks
- Dataset includes 400+ hours of video from 20+ different recording methods
- Metadata stored as csv files & tracking & annotation data as .parquet files. The final submission data requires 7 fixed columns.
- Competition started in September. Key deadlines:
 - 12/7: Deadline to merge teams
 - 12/15: Final submission



(AI-generated image)

Team's Metric & Performance Overview

- Performance tracked across multiple modules from 5 to 14.
- Latest leaderboard scores range from 0.268 to 0.446.
- Team rank improved steadily, reaching 441 out of 1286 teams.
- Cumulative submissions increased from 5 to 59 over modules.

Module	9	10	11	12	13	14
LB score	0.281	0.362	0.370	0.268	0.378	0.446
rank/#teams	523/857	508/930	492/1005	474/123	456/1207	441/1286
Cumulative # of submissions	5	16	24	33	47	59

- Macro F scores are averaged across each lab & video
- Scores only specific behaviors and the mice that were annotated for a specific video
- Includes a weighting factor to balance recall / precision
- Macro-averaging across actions prevents long, frequent behaviors from overwhelming the metric (ex: 'sniff' vs 'rest')

$$F_{\beta}(a) = \frac{(1 + \beta^2) \cdot TP_a}{(1 + \beta^2) \cdot TP_a + \beta^2 \cdot FN_a + FP_a}$$

$$F_{\beta \text{ macro}} = \frac{1}{n} \sum_{a=1}^n F_{\beta}(a)$$

β = weighting factor
 a = single action (ex: sniff)

```
for action in distinct_actions:
    if tps[action] + fns[action] + fps[action] == 0:
        action_f1s.append(0)
    else:
        action_f1s.append(((1 + beta**2) * tps[action] / ((1
+ beta**2) * tps[action] + beta**2 * fns[action] + fps[action]))
return sum(action_f1s) / len(action_f1s)
```

Source: [MABe F Beta Notebook](#)

Data's Key Features

Data's Key Features:

- **Metadata**: provides general information about each video and the mice within it.
 - Teams received CSV metadata for all videos used for *training* | *test*. Categories of features are listed below:
 - **Identifiers** → the lab providing the data and the video id
 - **Video data** → frames per second, video duration, pixels per cm, etc.
 - **Arena** → the shape, type, dimensions, etc.
 - **Labels** → i.e tracked body parts (tail, ears, etc.), behaviors labeled (chase, attack, etc.), tracking method
- **Tracking**: "pose data" or where the mouse's body parts are within each frame.
 - All data for both *training* | *test* was in .parquet files (*more on this later*). Below are all of the features:
 - **video_frame** → Frame index
 - **mouse_id** → Mouse ID
 - **bodypart** → Body part tracked
 - **x, y** → X/Y pixel coordinates
- **Annotation**: behavior labels (i.e. what one mouse may be doing at a given time to itself or another mouse).
 - Same as the tracking data above, all annotation data was in .parquet format. Only training data was available:
 - **agent_id** → the mouse performing the behavior
 - **target_id** → the mouse receiving the behavior (or same mouse for self-directed actions like grooming)
 - **action** → the behavior type (e.g., grooming, chasing, sniffing)
 - **start_frame / stop_frame** → when the behavior occurs in the video

Data's Key Features:

Metadata Structure

video_id: 44566106
video_id: 44566106
frames_per_second: 30
video_duration_sec: 60
arena_shape: square

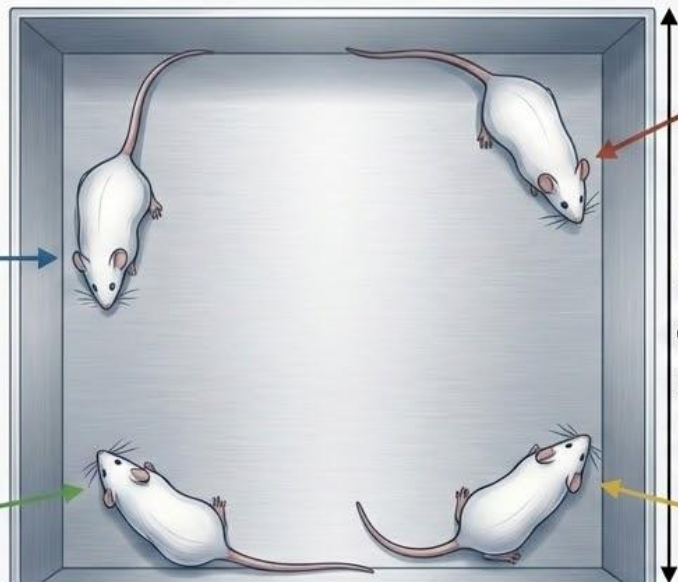
Mouse 1: Strain CD-1 (ICR)
Color: white
Sex: male
ID: 10.0, 8-12 weeks
Condition: wireless device

Mouse 3: Strain CD-1 (ICR)
Color: white
Sex: male
Age: 8-12 weeks
Condition: wireless device

Raw Data

video_id	lab_id	arena_type	mouse1_strain	mouse1_age	mouse2_strain	mouse2_age	mouse3_strain	mouse4_strain	mouse4_age
44566106	40602	open_field	CD-1 (ICR)	8-12 weeks	male male	8-12 weeks	CD-1 (ICR)	male male	8-12 weeks

Diagram and Raw Data



body_parts_tracked: nose, ears, body_center, tail, behaviors
_labeled: locomotion, pause, sniff_body, tracking_method

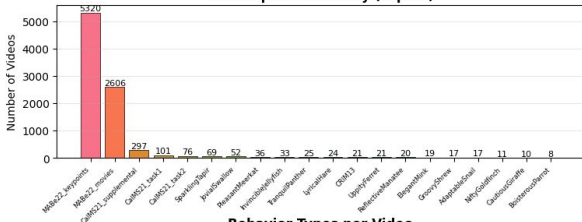
Raw Data

Mouse 2:
Strain: CD-1 (ICR)
Color: white
Sex: male
ID: 38.0
Age: 8-12 weeks
Condition: wireless device

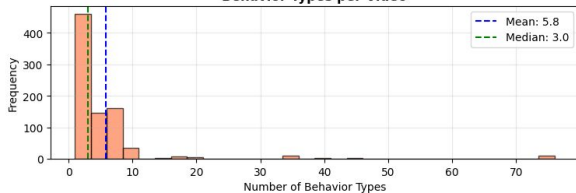
Mouse 4:
Strain: CD-1 (ICR)
Color: white
Sex: male
ID: 51.0
Age: 8-12 weeks
Condition: wireless device

MABE Dataset Comprehensive Overview

Videos per Laboratory (Top 20)



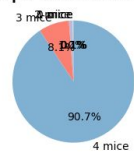
Behavior Types per Video



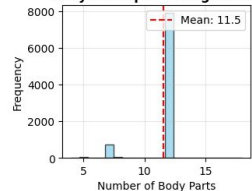
Training vs Test Dataset Comparison



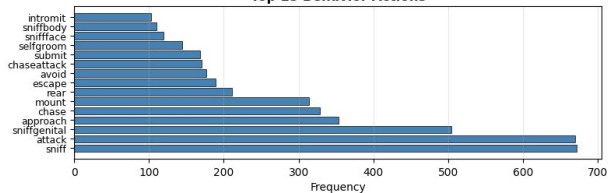
Mice per Video Distribution



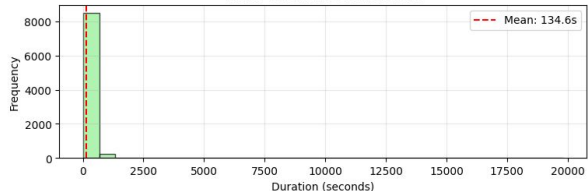
Body Parts per Configuration



Top 15 Behavior Actions



Video Duration Distribution



DATASET STATISTICS SUMMARY

TRAINING DATA:

- Total Videos: 8,789
- Videos with Missing Behaviors: 7941 (90.4%)
- Unique Laboratories: 21
- Average Mice per Video: 3.89
- Total Unique Body Part Configurations: 10

TEST DATA:

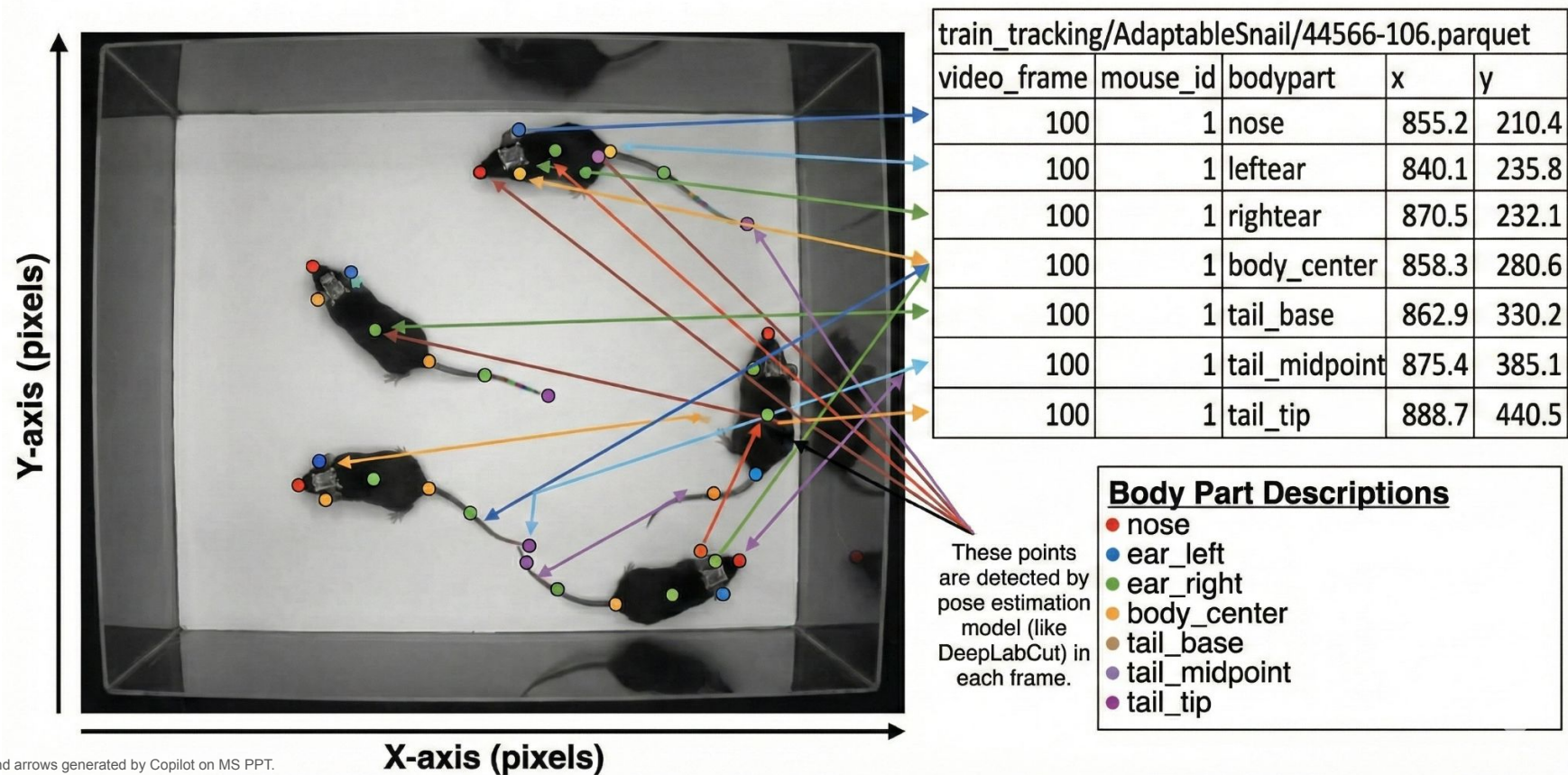
- Total Videos: 1
- Unique Laboratories: 1

BEHAVIOR ANALYSIS:

- Total Unique Behaviors: 37
- Most Common Behavior: sniff (672 occurrences)
- Average Behaviors per Video: 5.8
- Max Behaviors in Single Video: 76

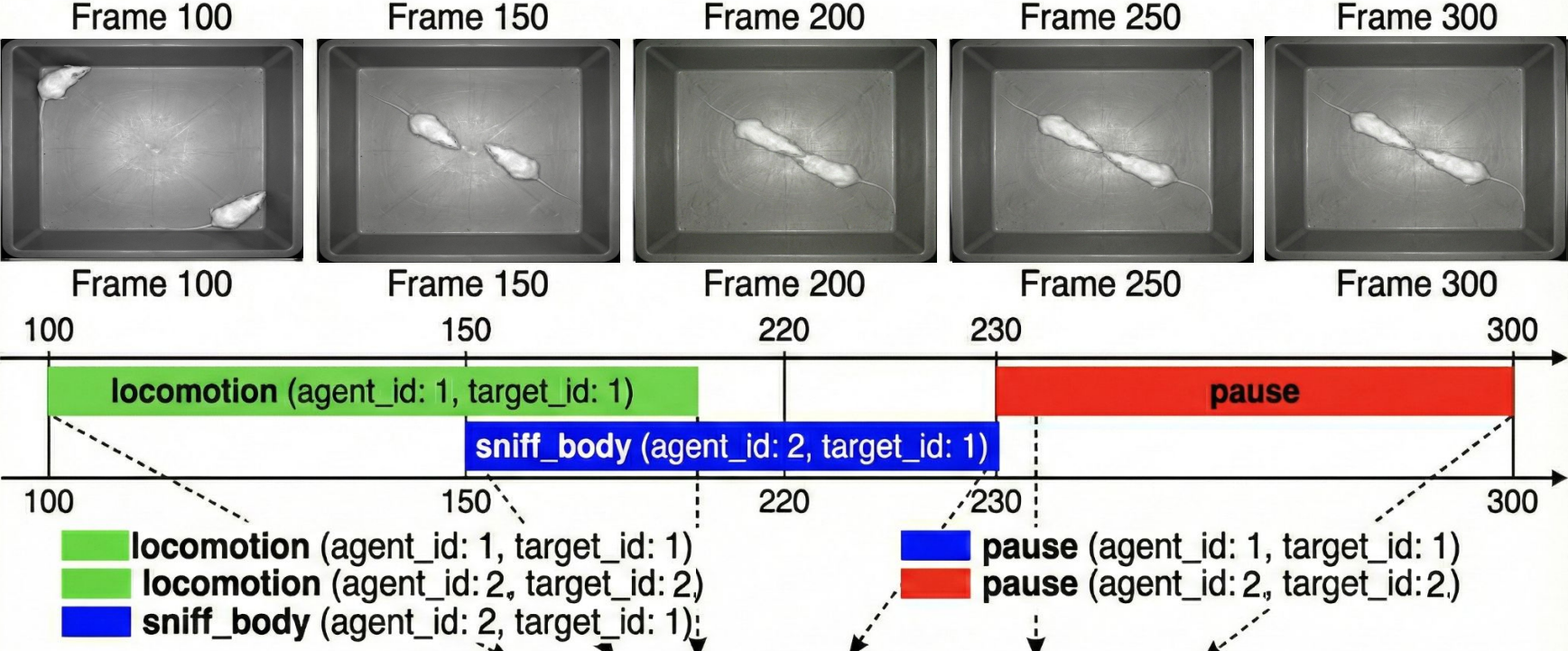
Data's Key Features:

Tracking Data and Body Part Calculation: Diagram and Raw Data ([train/test]_tracking/)



Data's Key Features—Target Variable

Behavioral Annotation Timeline: Diagram and Raw Data (train_annotation/)



video_id	agent_id	target_id	action	start_frame	stop_frame
44566106	1	1	locomotion	100	180
44566106	2	1	sniff_body	150	220

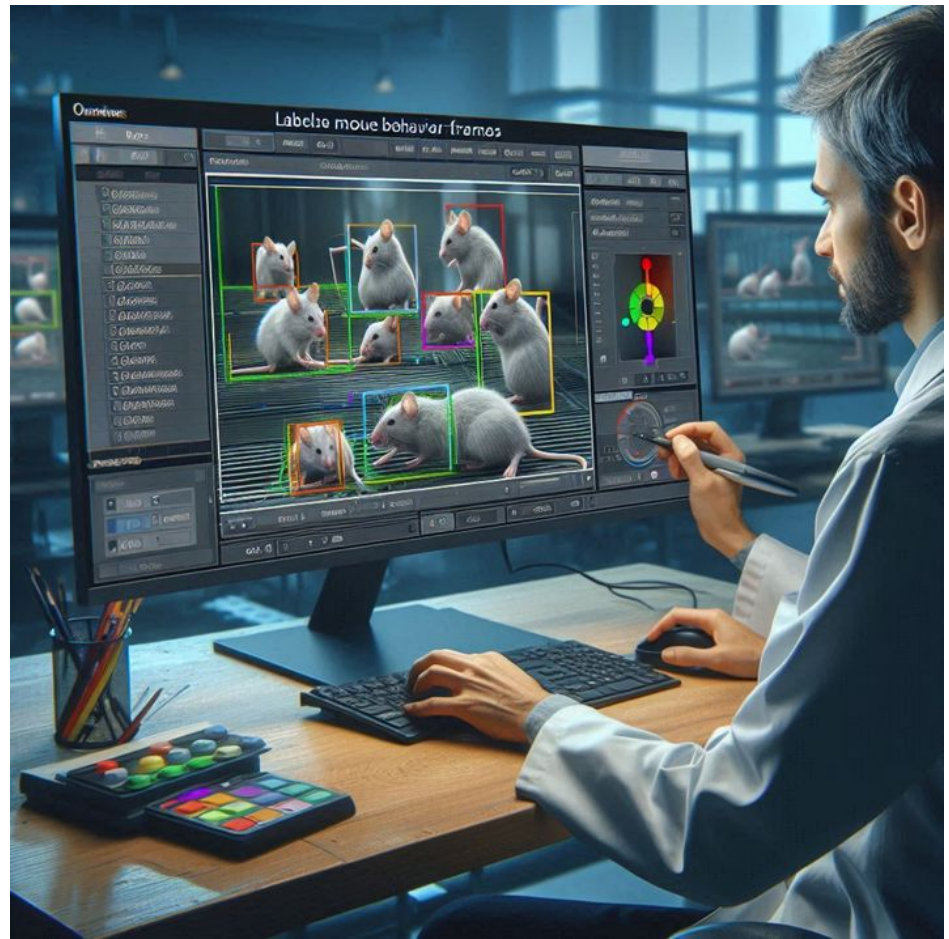
Submission Format: Raw Data (sample_submission.csv)

row_id	video_id	agent_id	target_id	➡action⬅	start_frame	stop_frame
1	44566106	1	1	locomotion	100	180
2	44566106	2	1	sniff_body	150	220
3	44566106	1	1	pause	230	300
4	44566106	3	3	grooming	350	400
5	44566106	4	3	chasing	420	480

Preprocessing Pipeline

Preprocessing and Organizing Mouse Behavior Data

- Organize tracking data capturing mouse positions per video frame
- Manage annotation data with detailed behavior labels for each frame
- Index data using video ID, agent and target mouse IDs, and frame number
- Track multiple body parts including ears, nose, neck, and tail bases
- Differentiate behaviors into self-actions and pairwise interactions
- Preprocess labels by extracting agents and behaviors from raw CSV files



(AI generated image)

Advanced Feature Engineering for Mouse Behavior Analysis

- Quantify self features such as distances, speeds, and body orientation.
- Analyze curvature, turning rates, and multiscale speed statistics.
- Capture movement states and transitions with smoothed position data.
- Extract pair features including distances and speeds between two mice.
- Aggregate interaction metrics like rolling distances for agent-target pairs.
- Implement a pipeline to extract and save features per video for training.



(AI generated image)

Preprocessing Code Snippets

```
def generate_speed_features(mouse_A, mouse_B, body_parts_tracked, interval=10):
    important_parts = ['tail_base', 'nose', 'ear_left', 'body_center']
    parts = list(set(important_parts) & set(body_parts_tracked))

    # Schema enforcement: fill missing parts with NaN
    for part in parts:
        if part not in mouse_A.columns:
            mouse_A[part] = np.nan
        if part not in mouse_B.columns:
            mouse_B[part] = np.nan

    features = {}

    # Precompute shifted positions
    shifted_A = {part: mouse_A[part].shift(interval) for part in parts}
    shifted_B = {part: mouse_B[part].shift(interval) for part in parts}

    # Precompute speeds
    speed_A = (part: np.square(mouse_A[part] - shifted_A[part]).sum(axis=1, skipna=False) for part in parts)
    speed_B = (part: np.square(mouse_B[part] - shifted_B[part]).sum(axis=1, skipna=False) for part in parts)

    for partA, partB in itertools.product(parts, repeat=2):
        # Speed features
        features[f'speed_{partA}'] = speed_A[partA]
        features[f'speed_{partB}'] = speed_B[partB]
        features[f'speed_{partA}[partB]'] = np.square(mouse_A[partA] - shifted_B[partB]).sum(axis=1, skipna=False)
        features[f'speed_{partB}[partA]'] = np.square(mouse_B[partB] - shifted_A[partA]).sum(axis=1, skipna=False)

    return pd.DataFrame(features)
```

Quantifies speed of both mice - this can help determine actions such as when mice chase or play with each other

```
# Process single mouse data
if len(single_mouse_list) > 0:
    single_mouse, single_mouse_label, single_mouse_meta = down_sampling(single_mouse_list)
    print(f"📄 Processing {len(single_mouse_list)} single mouse batches...")
    performance_metrics['single_mouse_batches'] += len(single_mouse_list)

    single_mouse = pd.concat(single_mouse_list)
    single_mouse_label = pd.concat(single_mouse_label_list)
    single_mouse_meta = pd.concat(single_mouse_meta_list)

    del single_mouse_list, single_mouse_label_list, single_mouse_meta_list

    X_tr = transform_single(single_mouse, body_parts_tracked)
    X_tr, single_mouse_label, single_mouse_meta = down_sampling(X_tr, single_mouse_label, single_mouse_meta)
    print(f"📄 Downsampling: {len(X_tr)} single mouse batches...")
    del single_mouse

    print(f"📄 Features shape: {X_tr.shape}")

    # Create feature analysis for first 2 configs
    if create_visualizations and section <= 2:
        # Create feature analysis plot(X_tr, 'single', section)
        if validate or submit == 'validate':
            thresholds = cross_validate_classifier_enhanced(
                binary_classifier, X_tr, single_mouse_label,
                single_mouse_meta, body_parts_tracked_str
            )
        else:
            submit(body_parts_tracked_str, 'single', binary_classifier, X_tr,
                    single_mouse_label, single_mouse_meta)

    del X_tr
    gc.collect()
```

```
# Process pair data
if len(mouse_pair_list) > 0:
    mouse_pair, mouse_pair_label, mouse_pair_meta = down_sampling(mouse_pair_list, mouse_pair_label_list, mouse_pair_meta_list)
    print(f"📄 Processing {len(mouse_pair_list)} mouse pair batches...")
    performance_metrics['pair_batches'] += len(mouse_pair_list)

    mouse_pair = pd.concat(mouse_pair_list)
    mouse_pair_label = pd.concat(mouse_pair_label_list)
    mouse_pair_meta = pd.concat(mouse_pair_meta_list)

    del mouse_pair_list, mouse_pair_label_list, mouse_pair_meta_list

    X_tr = transform_pair(mouse_pair, body_parts_tracked)
    X_tr, mouse_pair_label, mouse_pair_meta = down_sampling(X_tr, mouse_pair_label, mouse_pair_meta)
    print(f"📄 Downsampling: {len(X_tr)} mouse pair batches...")
    del mouse_pair

    print(f"📄 Features shape: {X_tr.shape}")

    # Create feature analysis for first 2 configs
    if create_visualizations and section <= 2:
        # Create feature analysis plot(X_tr, 'pair', section)
        if validate or submit == 'validate':
            thresholds = cross_validate_classifier_enhanced(
                binary_classifier, X_tr, mouse_pair_label,
                mouse_pair_meta, body_parts_tracked_str
            )
        else:
            submit(body_parts_tracked_str, 'pair', binary_classifier, X_tr,
                    mouse_pair_label, mouse_pair_meta)

    del X_tr
    gc.collect()
```

```
def estimate_ear_position(X, available_body_parts):
    center_parts = ['nose', 'head', 'neck', 'body_center', 'spine_1', 'spine_2', 'tail_base']
    ear_pairs = [('ear_left', 'ear_right'), ('ear_right', 'ear_left')]

    for ear_unknown, ear_known in ear_pairs:
        for head, tail in itertools.combinations(center_parts, 2):
            if head not in available_body_parts or tail not in available_body_parts:
                continue

            # Build mask for valid frames
            mask = (
                X[ear_unknown, 'x'].isna() | X[ear_unknown, 'y'].isna()
            ) & (
                X[tail, 'x'].notna() & X[tail, 'y'].notna()
            ) & (
                X[head, 'x'].notna() & X[head, 'y'].notna()
            )

            if not mask.any():
                return X

            # Midline vector
            dx = X.loc[mask, (head, 'x')] - X.loc[mask, (tail, 'x')]
            dy = X.loc[mask, (head, 'y')] - X.loc[mask, (tail, 'y')]
            norm = np.sqrt(dx**2 + dy**2) + 1e-6
            spine_dx = dx / norm
            spine_dy = dy / norm

            # Vector from tail to known ear
            ear_dx = X.loc[mask, (ear_known, 'x')] - X.loc[mask, (tail, 'x')]
            ear_dy = X.loc[mask, (ear_known, 'y')] - X.loc[mask, (tail, 'y')]

            # Project ear vector onto spine
            proj = ear_dx * spine_dx + ear_dy * spine_dy
            proj_x = proj * spine_dx
            proj_y = proj * spine_dy

            # Perpendicular component
            perp_x = ear_dx - proj_x
            perp_y = ear_dy - proj_y

            # Reflect across spine
            X.loc[mask, (ear_unknown, 'x')] = X.loc[mask, (tail, 'x')] + proj_x - perp_x
            X.loc[mask, (ear_unknown, 'y')] = X.loc[mask, (tail, 'y')] + proj_y - perp_y
```

```
def estimate_nose_position(mouse_df, scale=0.75):
    # Compute ear center
    ear_left_x = mouse_df[['ear_left', 'x']]
    ear_left_y = mouse_df[['ear_left', 'y']]
    ear_right_x = mouse_df[['ear_right', 'x']]
    ear_right_y = mouse_df[['ear_right', 'y']]

    center_x = (ear_left_x + ear_right_x) / 2
    center_y = (ear_left_y + ear_right_y) / 2

    # Ear-to-ear vector
    dx = -ear_right_x + ear_left_x
    dy = -ear_right_y + ear_left_y

    # Perpendicular direction (forward-facing)
    perp_x = -dy
    perp_y = dx
    norm = np.sqrt(perp_x**2 + perp_y**2) + 1e-6
    perp_x /= norm
    perp_y /= norm

    # Estimate nose position
    ear_dist = np.sqrt(dx**2 + dy**2)
    est_x = center_x + scale * ear_dist * perp_x
    est_y = center_y + scale * ear_dist * perp_y

    # Fill only missing values
    mask_x = mouse_df[['nose', 'x']].isna()
    mask_y = mouse_df[['nose', 'y']].isna()
    mouse_df.loc[mask_x, ('nose', 'x')] = est_x[mask_x]
    mouse_df.loc[mask_y, ('nose', 'y')] = est_y[mask_y]

    return mouse_df
```

Nose / Ear position can help determine mice's location within the arena as well as helping identifying "sniff"-related actions or whether they are turning towards or away from another mice.

We wrote custom code to track 1 mice (sometimes mice act on themselves) or pair of mice (ex: chase). Google Gemini was used to check our code & offer improvements.

Code such as generating nose positions and speed features are from the Kaggle community and have been cited under References.

LightGBM Model & Final Results

Our Most Invested Model

- Idea to use LightGBM based on upon a Kaggle community post with a 26% leaderboard score (Kongyanmiao)
- Implemented advanced feature engineering and preprocessing steps as discussed in previous sections
- Focused on hyperparameter tuning to enhance model F1-score and robustness
- Achieved an improved leaderboard score of of 38% (see below)

*Custom LightGBM implementation of code w/
hyperparameters*

```
# Create classifier
binary_classifier = make_pipeline(
    SimpleImputer(),
    TrainOnSubsetClassifier(
        lgb.LGBMClassifier(
            n_estimators=200,
            learning_rate=0.03,
            min_child_samples=40,
            num_leaves=31,
            max_depth=-1,
            subsample=0.8,
            colsample_bytree=0.8,
            verbose=-1,
            random_state=0
        )
    )
)
```



Competition Notebook

MABe Challenge - Social Action Recogni...

Public Score

0.376

Best Score

0.380 V31

Version 33 of 33

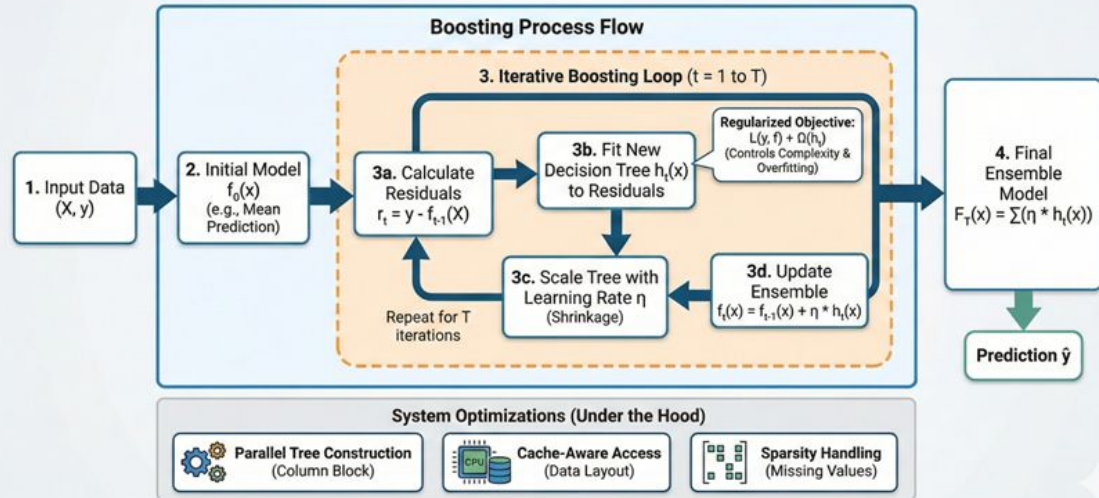


Runtime

▶ 55m 30s

Training Model Setup and Optimization

XGBoost Architecture (Extreme Gradient Boosting)



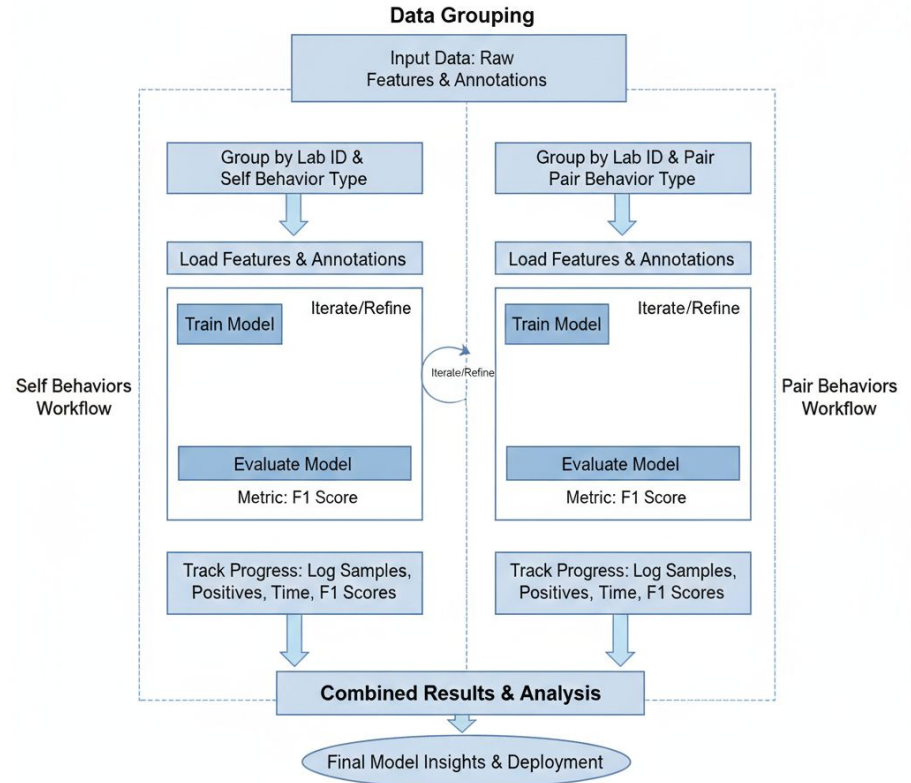
- Use XGBoost with binary logistic objective for classification
- Employ StratifiedGroupKFold cross-validation grouped by video ID.
- Address class imbalance using `scale_pos_weight` parameter.
- Hyperparameters:
 - Learning Rate = 0.05
 - Max depth = 6
 - Subsample | colsample = 0.8.
- Apply early stopping after 10 rounds without improvement to prevent overfitting.
- Generate outputs per fold including model files, thresholds, and performance plots.

(AI generated image using MS PPT Copilot)

Training Execution Workflow for Mouse Behaviors

- Group data by lab ID and behavior type for self and pair behaviors.
- Load respective features and annotations for each behavior group.
- Train and evaluate models per behavior with F1 scores logged.
- Track progress with detailed tables showing samples, positives, and elapsed time.
- Separate workflows for self behaviors and pairwise interactions ensure accuracy.

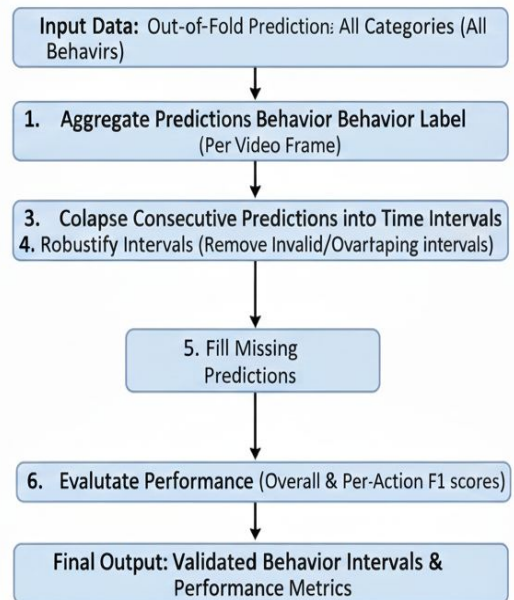
Behavior Analysis Modeling Workflow



(AI generated flow chart)

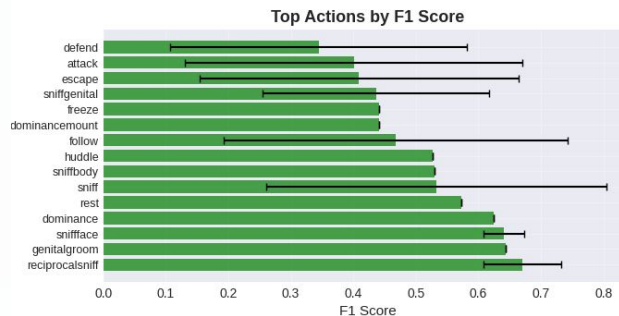
Validation & Post-Processing for Reliable Predictions

Behavior Analysis Post-Processing Pipeline



(AI generated image)

- Apply methods to remove invalid and overlapping intervals.
- Aggregate out-of-fold predictions across all behavior categories, and select the most confident behavior label for each video frame.
- Collapse consecutive identical predictions into defined intervals.
- Fill missing predictions for videos lacking output data.
- Evaluate performance using overall and per-action F1 scores.




Challenges in Mouse Behavior Analysis



(AI generated image)

- Inconsistent frame labeling by lab annotators and diverse mouse body part definitions complicate data normalization across labs
- System memory (even on Colab Pro) limits restrict features to under 300, demanding careful selection and fine tuning
- Missing key body part positions limits model choices to tree-based methods (excl. deep neural networks)
- Rare positive action cases (~10%) make differentiating behaviors very difficult
- Cross-validation results often do not correlate with leaderboard performance

#	Team	Members	Score	Entries	Last	Join
507	JHU-Team 1		0.446	65	2d	

References

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Appendix

TensorFlow Preprocessing Pipeline

- Major bottleneck - 8789 video tracking datasets available in .parquet format
- Max RAM available on Google Colab Pro is <13 GB
- Team's solution was to create a TF pipeline that loads directly from .parquet files. Main advantage:
 - 1) Compatibility with LightGBM package (our current model)
 - 2) Native-support with TF's deep learning models (next model for the team to try)
 - 3) Training data in batches in parallel that can be integrated with GPU | TPUs in Google Colab
-

Custom-built TF Flow for MABe Tracking Data

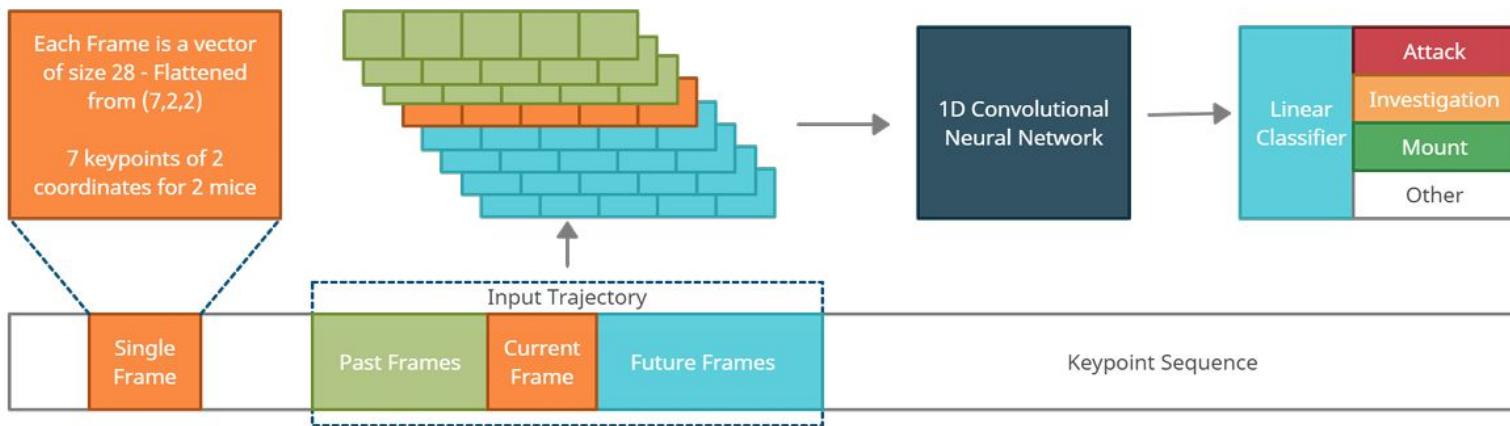
```
1 import pyarrow.parquet as pq
2 import tensorflow as tf
3
4 def parquet_generator(files, regex_start):
5     print(f"Concatenating {len(files)} files...")
6
7     for file in files:
8         print(f"Reading file: {file}")
9         parquet_file = pq.ParquetFile(file)
10
11         for batch in parquet_file.iter_batches(batch_size=1024, use_threads=True):
12             df = batch.to_pandas().assign(
13                 lab_id=re.search(regex_start + "/([^\s]+)/", file).group(1),
14                 video_id=re.search(r"(\d+)(?=\s\.parquet)", file).group(1))
15
16             for row in df.itertuples(index=False):
17                 yield dict(row._asdict())
18
19 dataset_tf = tf.data.Dataset.from_generator(
20     lambda: parquet_generator(files=train_tracking, regex_start="train_tracking"),
21     output_signature={
22         "video_frame": tf.TensorSpec(shape=(), dtype=tf.int32),
23         "mouse_id": tf.TensorSpec(shape=(), dtype=tf.int32),
24         "bodypart": tf.TensorSpec(shape=(), dtype=tf.string),
25         "x": tf.TensorSpec(shape=(), dtype=tf.float32),
26         "y": tf.TensorSpec(shape=(), dtype=tf.float32),
27         "lab_id": tf.TensorSpec(shape=(), dtype=tf.string),
28         "video_id": tf.TensorSpec(shape=(), dtype=tf.int32)
29     }).batch(128).prefetch(tf.data.AUTOTUNE)
```

DNN Architecture

- Using neural net models tend to outperform our LightGBM solution
- 1D CNN performs the best based on authors of the original study
- Features the authors considered: distances, velocities, and angles between coordinates relative to the agents' orientations.
- Hyperparameters considered includes number of input frames, frame skips, units per layer, and learning rate.

Method	Average F1	MAP
Fully Connected	.659 ± .005	.726 ± .004
LSTM	.675 ± .011	.712 ± .013
Self-Attention	.610 ± .028	.644 ± .018
1D Conv Net	.793 ± .011	.856 ± .010

Table 1: Class-averaged results on Task 1 (attack, investigation, mount) for different baseline model architectures. The value is shown of the mean and standard deviation over 5 runs.



All images are from Sun et. al from their paper: MABe22: A multi-species multi-task benchmark for learned representations of behavior.

Note: MAP - mean average precisions results - averages the number of