

Neural population responses to taste and food in the parabrachial nucleus of the pons in the awake unrestrained rat using *in vivo* one-photon Ca^{2+} imaging

Flynn P. O'Connell¹, Jonathan D. Victor², Patricia M. Di Lorenzo¹

connell@Binghamton.edu; jdvicto@med.cornell.edu; diloren@Binghamton.edu

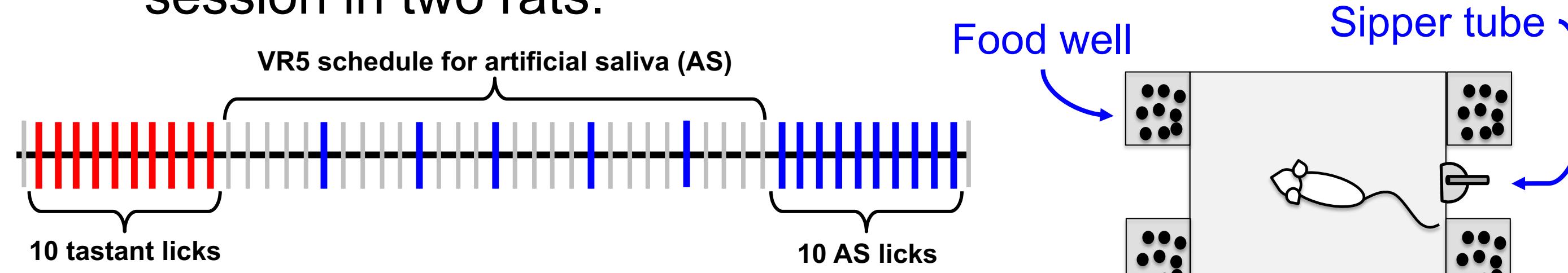
¹Dept. of Psychology, Binghamton University Binghamton, NY; ²Weill Cornell Medical College, New York, New York 10065.

I. INTRODUCTION

- ❖ Although brainstem responses to taste stimuli have been well-documented, the function of the taste system in natural feeding remains unexplored.
 - ❖ Electrophysiological data from the nucleus of the solitary tract (NTS) suggested that the taste system is strongly engaged while exploring food and sampling its taste and smell but becomes relatively disengaged when food is being consumed.
 - ❖ The **parabrachial nucleus of the pons (PbN)** receives direct input from the NTS and ultimately projects to the insular cortex through the ventromedial thalamus.
 - ❖ We used one-photon *in vivo* Ca^{2+} imaging to track the activity of a population of PbN cells as they licked tastants, smelled food and ate solid food.
 - ❖ We tested the **hypothesis** that PbN cells inherit key properties from NTS. In particular that:
 - ❖ Like NTS, PbN neurons would respond not only to the taste of food but also to its smell
 - ❖ Like NTS, PbN neurons would respond strongly while exploring and sampling, but relatively quiescent during eating and consumption

II. METHODS

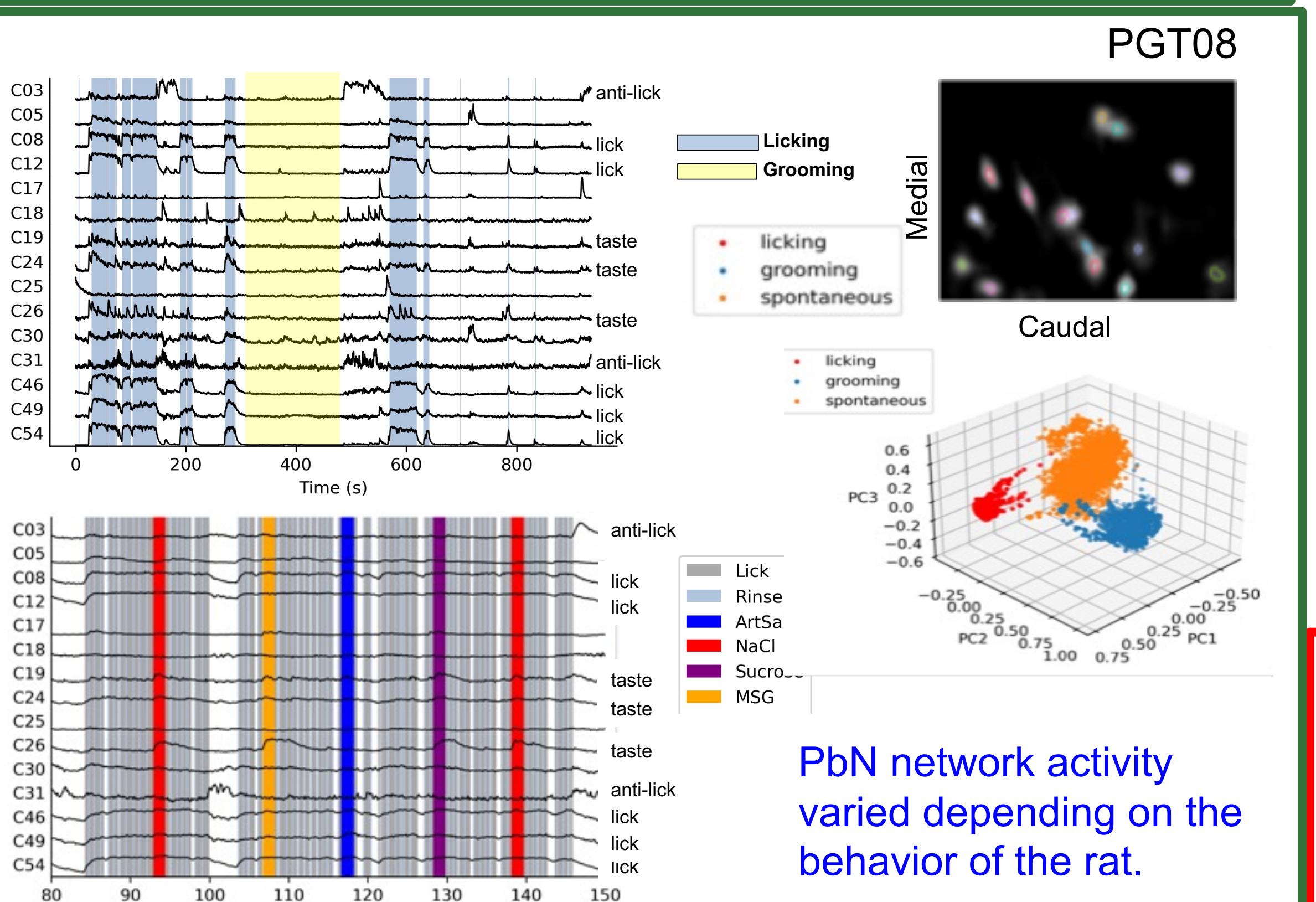
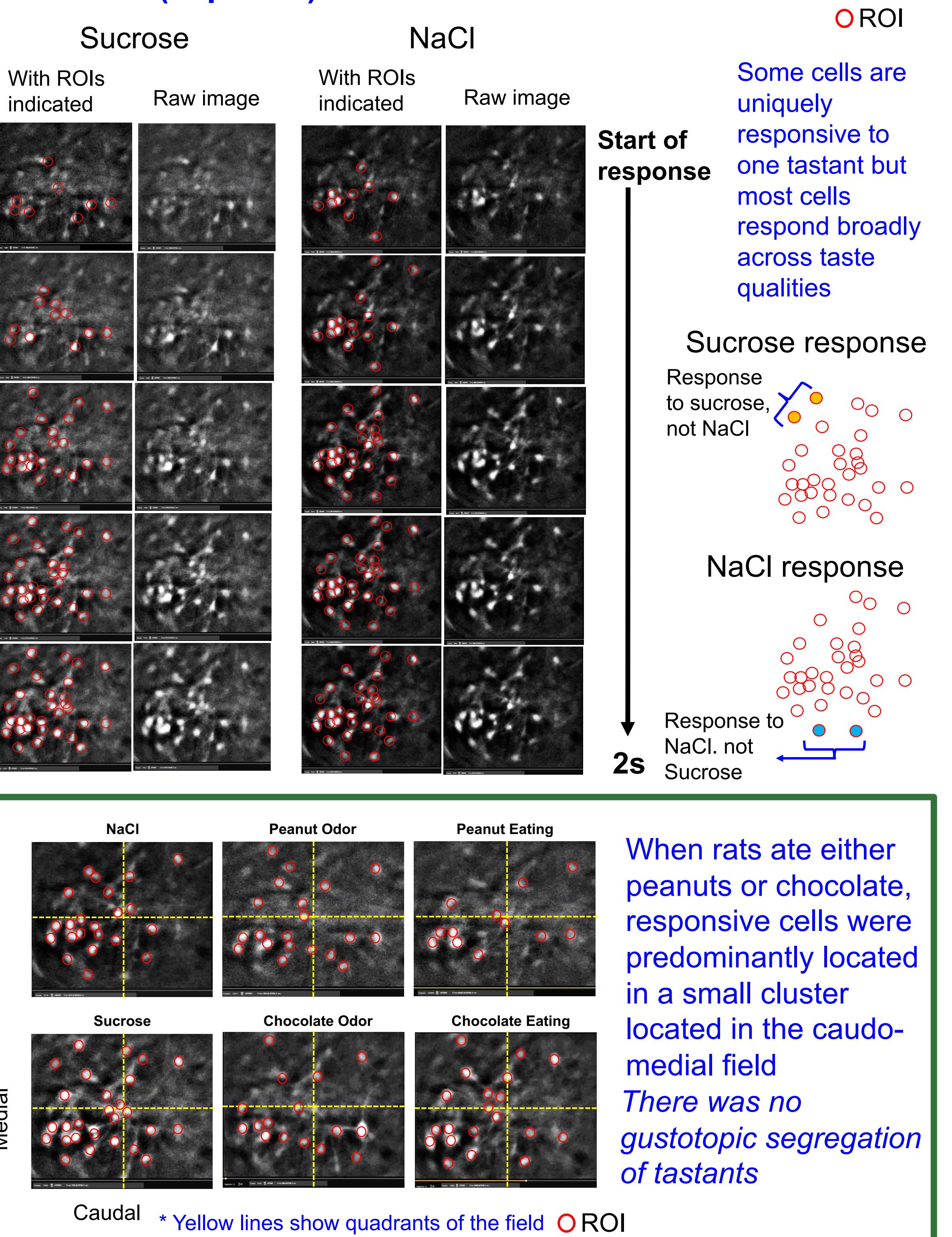
- ❖ Rats (n= 3) were prepared for Ca^{2+} imaging:
 - ❖ The PbN (11.3 A/P, 1.7 M/L, -5.6 D/V) was infused with AAV9 virus containing the GCaMP7s gene under the synapsin 1 promoter (Addgene #104487-AAV9; pGP-AAV-syn-jGCaMP7s-WPRE). Two weeks later, a GRIN lens was implanted with the tip positioned just above the PbN.
 - ❖ Following recovery, rats were water deprived and a miniscope (Inscopix, Inc.) was mounted to the surgical baseplate.
 - ❖ Rats were first placed in an operant chamber with free access to taste stimuli representing basic taste qualities. Two olfactory stimuli were also presented along with artificial saliva (AS): peanut or chocolate odor.
 - ❖ Tastants were dissolved in artificial saliva (AS) and included:
 - ❖ 0.1 M sucrose, 0.1 NaCl, 0.0167 M citric acid, 0.0001 M quinine) as well as AS presented alone or paired with either peanut or chocolate odor.
 - ❖ Solid foods (Granny Smith apple, salted peanuts or milk chocolate were available in food wells after the licking session in two rats.



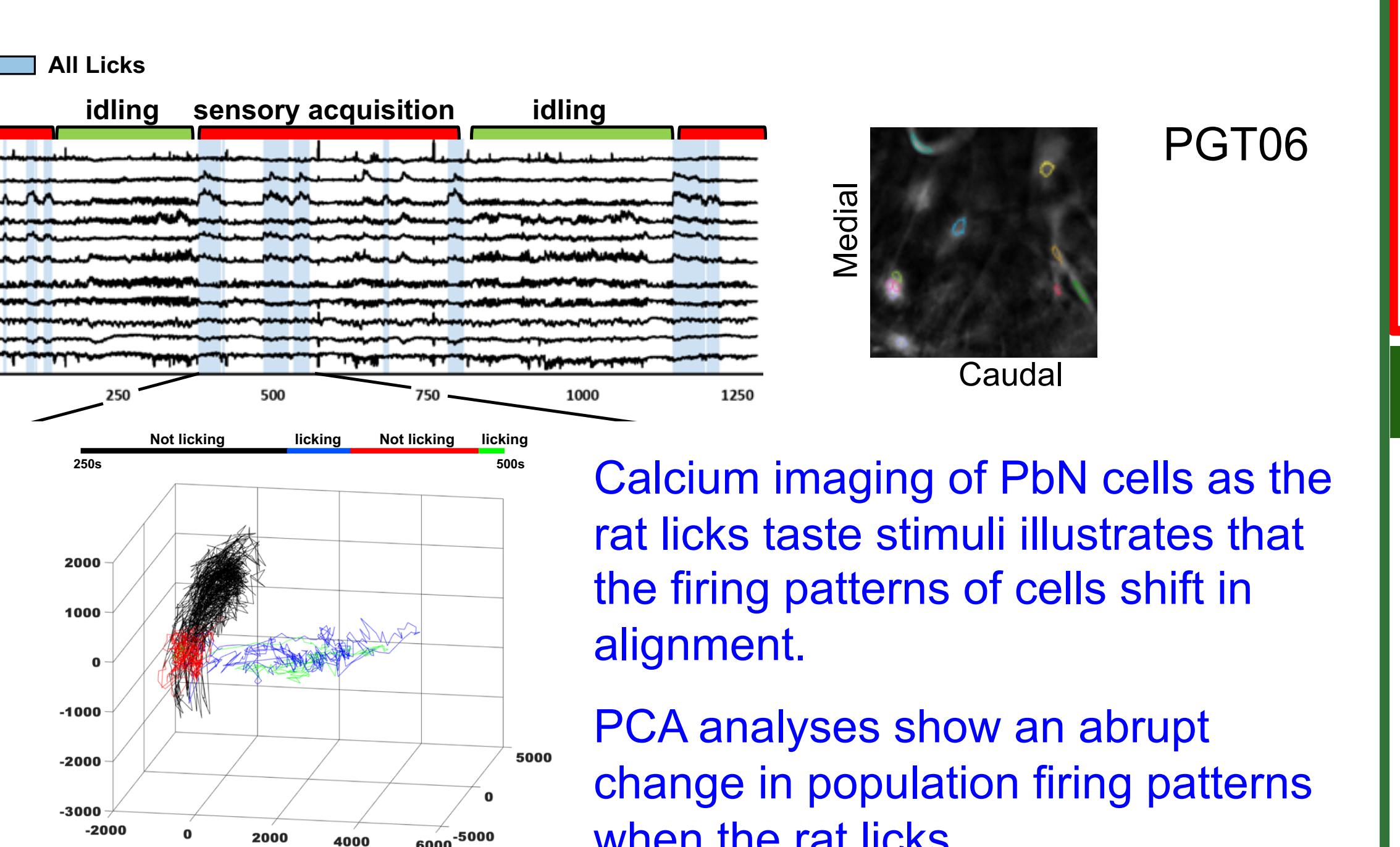
Data analysis:

- ❖ Video acquisition was done with nVista® software (Inscopix, Inc.) at a sample rate of 10 Hz
 - ❖ A single focal plane representing the sharpest image was selected.
 - ❖ Video processing with Inscopix Data Processing software (Inscopix DPS®, Inc.) in several steps:
 - ❖ Spatial downsampling (4x), bandpass filter, motion correction, manual ROI extraction, contour application
 - ❖ Final output was $\Delta F/F_0$ where F_0 where represents *mean* fluorescence across the

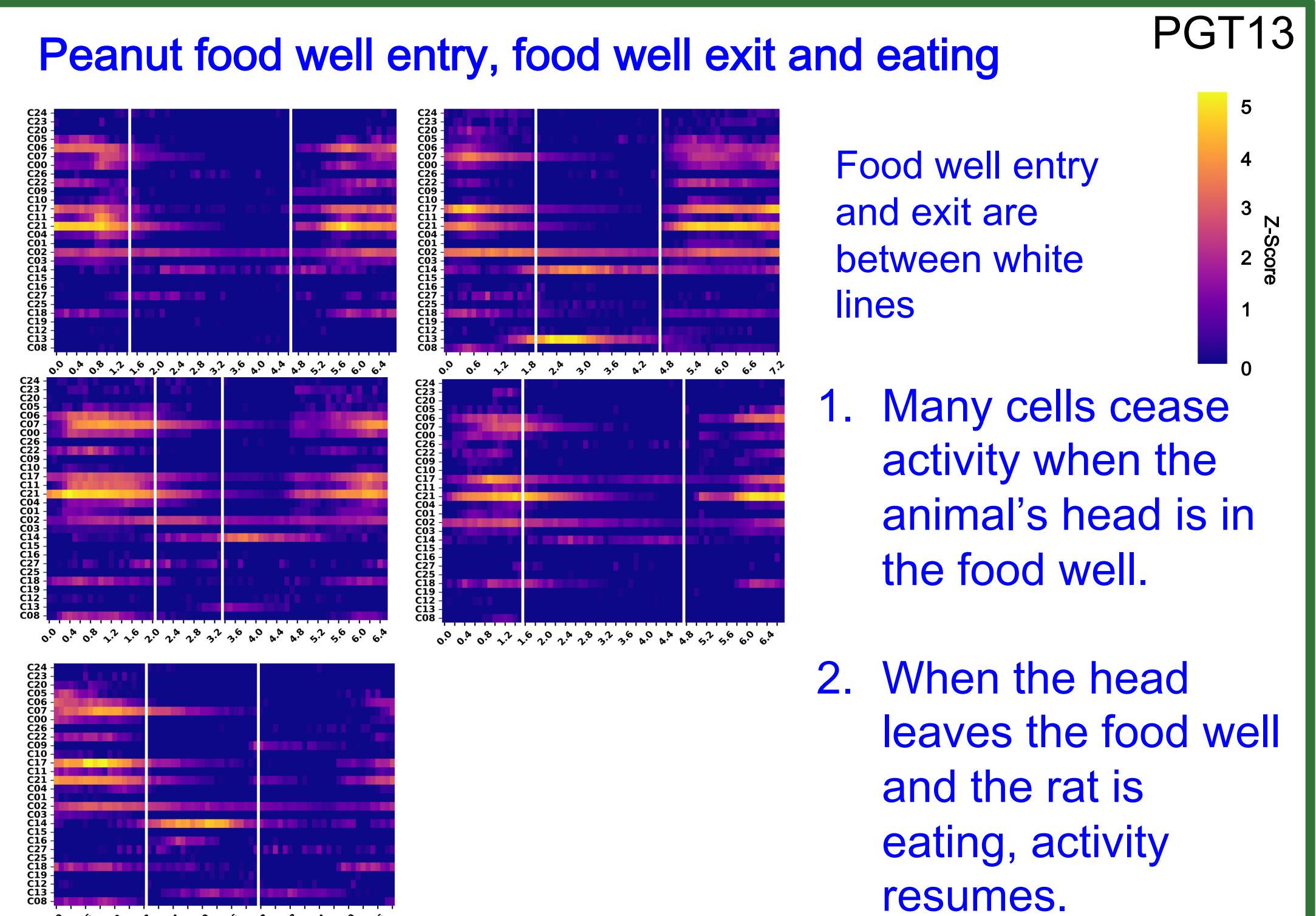
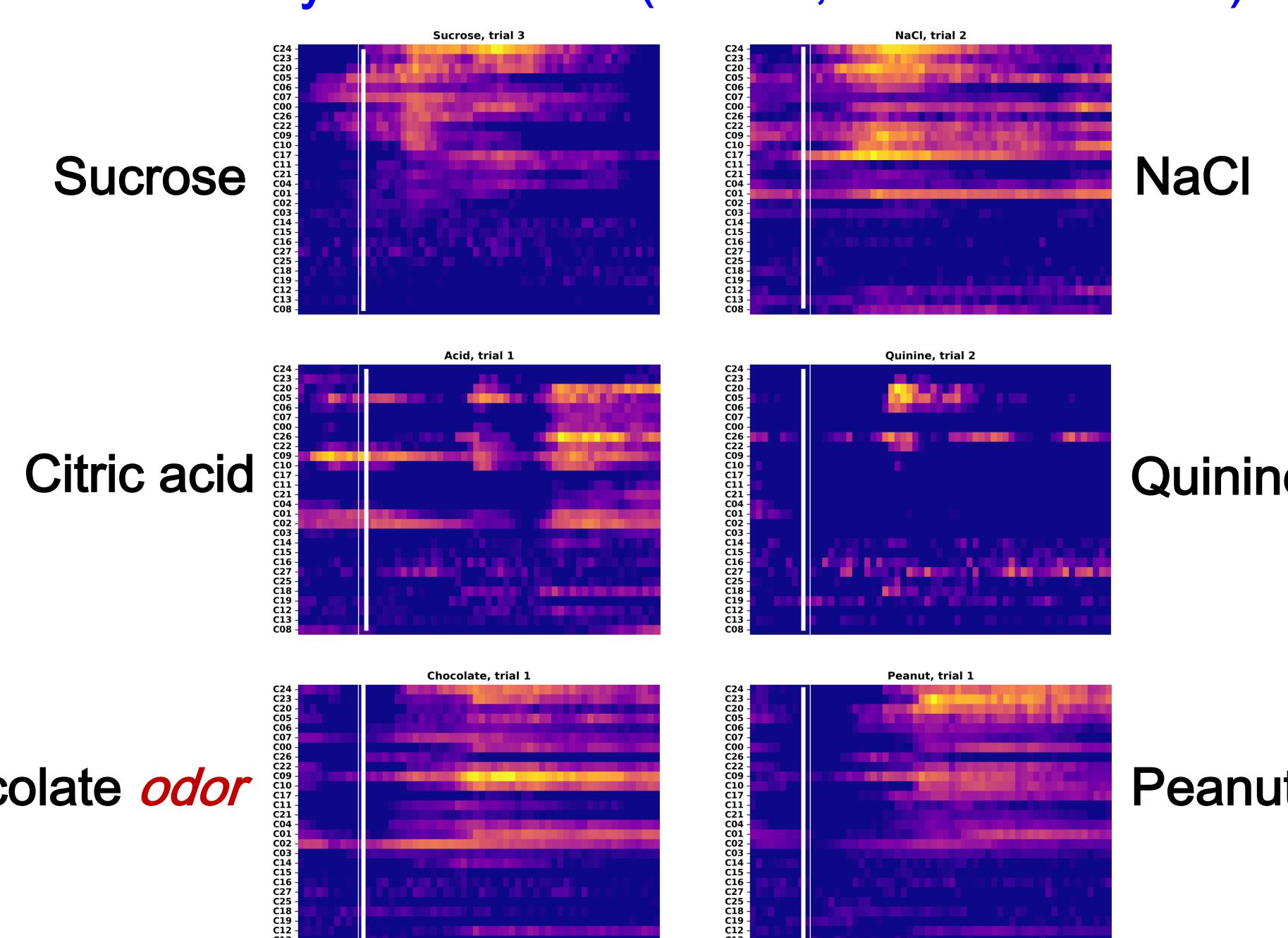
The spatial response to taste in the PbN evolves (expands) over time



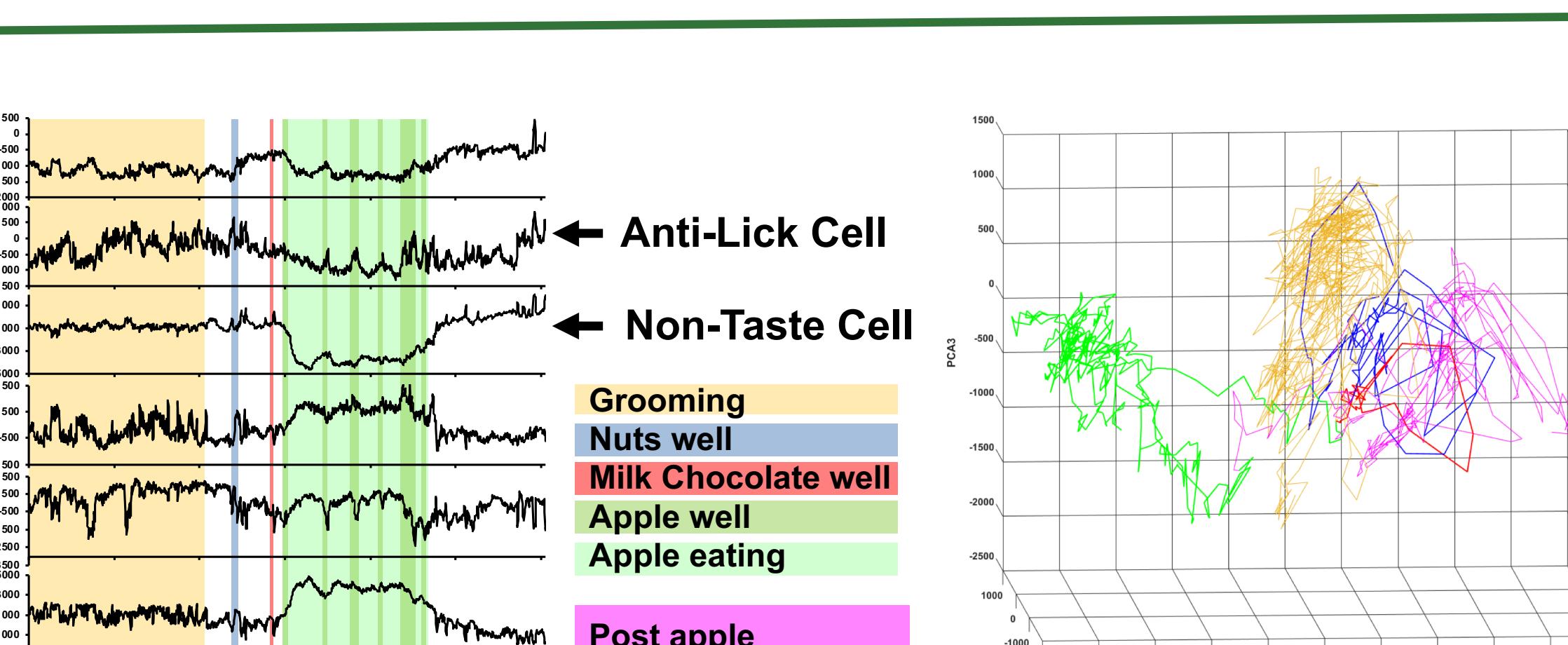
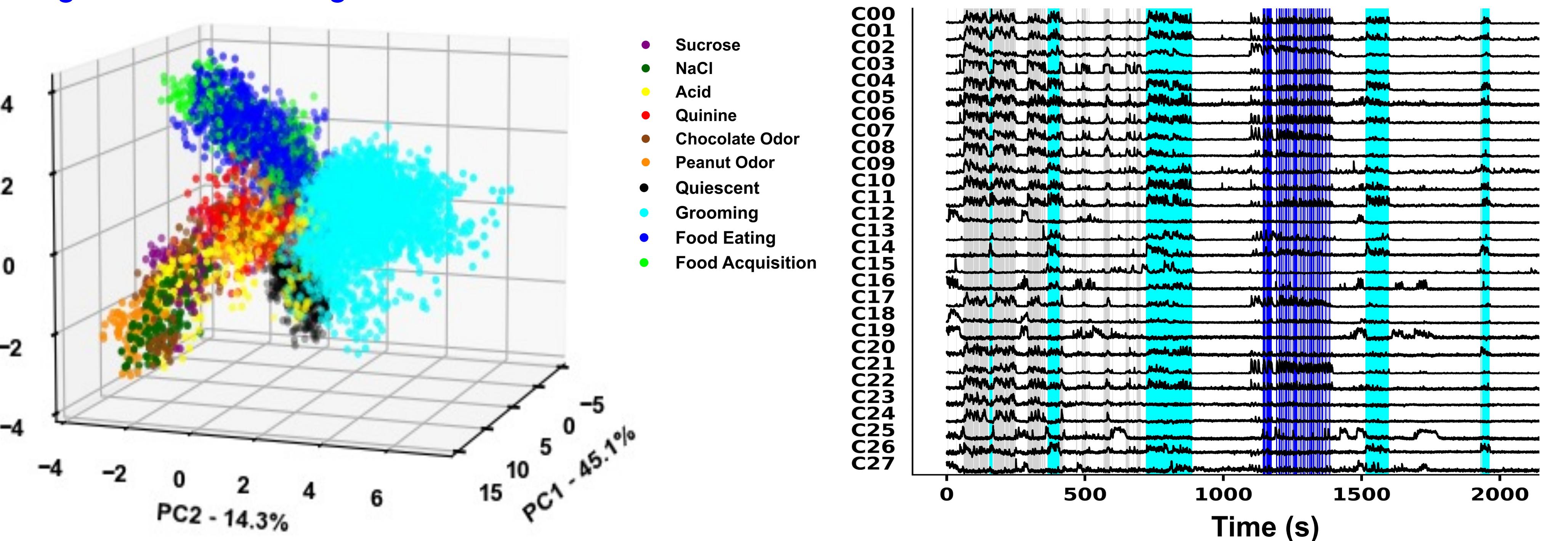
Taste-responsive cells were recorded alongside anti-lick and lick bout cells, as in electrophysiological experiments (see Roussin et al. 2012)



III. RESULTS



Network activity is different when the rat is licking tastants vs. eating.



IV. Conclusions

- ★ Taste responses in the PbN of awake, freely licking rats were recorded using one-photon Ca^{2+} imaging. Taste-responsive PbN cells were recorded from three rats. Records were obtained from two rats while they ate solid food.
 - ★ **Taste responses evolved over space and time without a clear gustotopic organization**
 - ★ In one rat (PGT13) records were obtained from 28 PbN cells as the rat licked tastants, smelled chocolate or peanut odor and ate chocolate and peanuts. **Results clearly indicate that PbN cells respond to odor as well as taste.**
 - ★ **PbN network activity changes abruptly when a rat licks or eats.** Activity during appetitive and consummatory behavior is likely *multimodal*, reflecting taste, olfactory and motor-related signals

V. Acknowledgements



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References