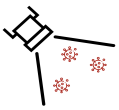
Certification Run Assessment v1.1

FloodLAMP.bio



SOP

- ☐ Only approved personnel can perform this assessment
- ☐ Watch video and check items
- ☐ Note timestamp of any deviations
- ☐ Determine PASS or FAIL
- ☐ Log assessment with Form

	Certification Run Assessment Form Link	version 1.1
	Video Link:	
Assessor Name:		
Date and Time:		
PASS or FAIL		

Safety

- ☐ Proper PPE (Gloves, Lab Coat, N-95 Mask, and Goggles / Face Shield)

Contamination

- ☐ Pen and sharpie labeled with green tape
- ☐ Separation of Post-Amp area

Contamination Infractions

- ☐ PPE lapse/removal
- ☐ Use green labeled pen/marker with bare hands and not putting it to be cleaned afterwards
- ☐ Use unmarked pen/marker with clean gloves (without cleaning gloves after or changing gloves)
- ☐ Bare hands on anything improper, except in desk area
- ☐ Reuse pipette tip improperly
- ☐ Touch pipette tip to anything and then not discard it
- ☐ Touch anything dirty then go back to work (face, mask, uncleaned phone, uncleaned laptop)

Prep

- ☐ Turn on heaters, heat indicating light
- ☐ Alcohol wipe the collection tubes

Documentation

- ☐ Read each item then check it off after completing it
- ☐ Writing legible

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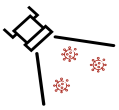

Inactivation

- ☐ Write down on log sheet the amounts of saline and 100X prior to pipetting
- ☐ Label 1XISS and logging the preparation correctly
- ☐ Volumes correct for Saline and 100X IS
- ☐ Mixing correct for 1XISS
- ☐ Volume correct for 1XISS added to samples (1mL)
- ☐ Sample tubes opened and closed properly
- ☐ Vortex of samples correct
- ☐ Timing correct for inactivation heating and cooling

Reaction Mix Prep (strip8 tubes)

- ☐ Write down entry on log sheet prior to starting prep
- ☐ Calc volumes correctly for PGS and CLAMP based on number of reactions
- ☐ CLAMP and PGS thawed properly, so not to get too warm
- ☐ Spin down reagents and reaction mix
- ☐ Correct volumes for CLAMP and PGS
- ☐ Get PGS and CLAMP source tubes back in freezer quickly
- ☐ Vortex and spin down reaction mix tube
- ☐ Marking the top tube in strip with 1,2,3, etc
- ☐ Correct volume of reaction mix in each pcr tube (23uL)
- ☐ Pipetting at the bottom of tubes
- ☐ No blowout while dispensing reaction mix
- ☐ Volume check strip tubes
- ☐ Keep reactions covered strip tubes with rack lid to prevent dust

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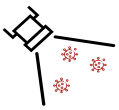


Amp Run (strip8 tubes)

- ☐ Get Cold PCR Block out ~10min before adding tubes
- ☐ Reaction mix tubes ready (thawed but not too warm (<10 min)
- ☐ Strip 8 tubes labeled on top tube of each strip
- ☐ Tips setup properly to align with strips8 tubes to be used
- ☐ Add negative control 1st
- ☐ Using a lookup rack to keep continuity of sample order
- ☐ Visually checking the 2uL in the tip from each inactivated sample
- ☐ Amp pipetting technique correct - pipet up and down 5 times, blowout in liquid, tip touch
- ☐ Well position correct and correspondence between tip, strip tube# and, where sample tube is put in lookup rack
- ☐ Transfer strip to regular PCR block (so they aren't cold going to Amp)
- ☐ Only touching positive control (TPC) with one hand
- ☐ Glove change after putting the positive control (TPC) back in freezer and before caps
- ☐ Not touching the top of strip8 tubes or inside of a strip8 cap
- ☐ Completely sealed strip tubes - check at eye level
- ☐ Change gloves again after touching loading tubes/plate onto amp heater
- ☐ Set main timer for 25min
- ☐ Set phone timer for 24min
- ☐ Put sample tubes in fridge during amp heating
- ☐ Wait at least 1 minute after pulling off the amp heater
- ☐ Take photo in lightbox
- ☐ For photo apply crop and vivid filter
- ☐ Logging amp run sheet properly with google form entry

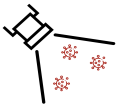
App

- ☐ Intake tubes correctly
- ☐ Process and Result tubes correctly

ONLY COMPLETE THIS LAST SECTION IF TRAINING FOR PLATE SCALE TESTING

Dispenser Demonstration

- ☐ Unpack an electric dispenser
- ☐ Properly prime the dispenser
- ☐ Dispense 1mL into tube without touching the tip



Amp Run (Abbreviated Plate)

- [ ] Prepare reaction Mix following all previous guidelines
- [ ] Pipette Reaction Mix into a reservoir
- [ ] Use Multichannel Pipette to fill Column 1
- [ ] Fully seal plate with foil seal
- [ ] Pierce B1 to add negative control
- [ ] Pierce C1 - H1 to add samples
- [ ] Pierce A1 to add TPC, following all TPC guidelines
- [ ] Glove change
- [ ] Use a second foil seal on top of the first
- [ ] Tight seal, no delamination
- [ ] Put on Amp heater
- [ ] Glove change