



COMP90014

Algorithms for Bioinformatics

Week 2B - Sequence Alignment and Mapping II

Sequence Alignment and Mapping II



Local, Semi-Global alignment (previous lecture)

Scoring/Substitution Matrices

Gap penalties

Read mapping

Seed-extend (short reads)

Seed-chain-align (long reads)

BLAST

Academic Integrity Statement

DUE TOMORROW

Pairwise Alignment

Levenshtein distance

Forms the basis for the remainder of lecture

Global alignment

Local alignment Semi-global alignment

These add a few important variations:

Penalties rather than adding edits
Penalties are different for mismatch vs gap
The arrows are stored
(directions we took to calculate each cell)

SPLATTERING -> PATTERN

Global	SPLATTERING -P-ATTERN
Local	ATTER ATTER
Semi-global	PLATTERIN P-ATTER-N

Global Alignment

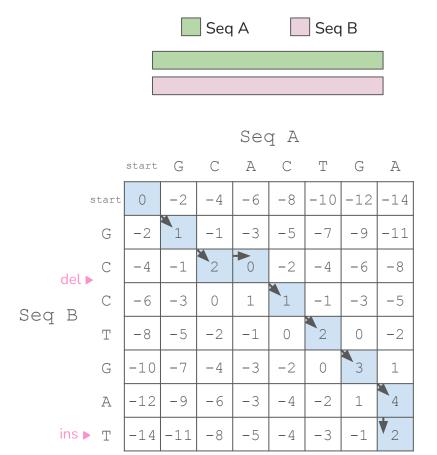
Algorithm: Needleman Wunsch

```
GCACTGAT
GC-CTGAT
```

Algorithm 2: Needleman-Wunsch(A, B)

return S(length(A), length(B))

```
\begin{split} g &\leftarrow GapPenalty; \\ \textbf{for } i &= 0 \textbf{ to } length(A) \textbf{ do} \\ &\mid S(i,0) \leftarrow g \times i; \\ \textbf{for } j &= 0 \textbf{ to } length(B) \textbf{ do} \\ &\mid S(0,j) \leftarrow g \times j; \\ \textbf{for } i &= 1 \textbf{ to } length(A) \textbf{ do} \\ &\mid Goldsymbol{o} for j &= 1 \textbf{ to } length(B) \textbf{ do} \\ &\mid Match \leftarrow S(i-1,j-1) + Scoring(A_i,B_j); \\ &\mid Insert \leftarrow S(i,j-1) + g; \\ &\mid Delete \leftarrow S(i-1,j) + g; \\ &\mid S(i,j) \leftarrow max(Match,Insert,Delete); \end{split}
```



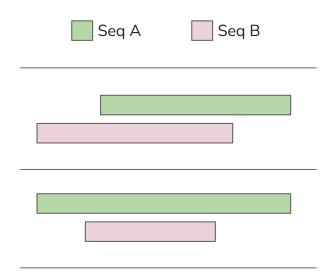
Alignment of complete sequences, where offset is not penalised

All of Seq A All of Seq B

Best when sequences are expected to have similar overlapping region

eg. Read overlaps in OLC assembly

Returns the full alignment, clipped by best offset



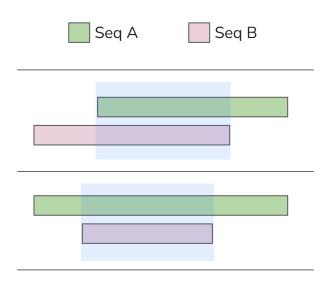
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All of Seq A All of Seq B

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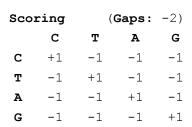
eg. Read overlaps in OLC assembly

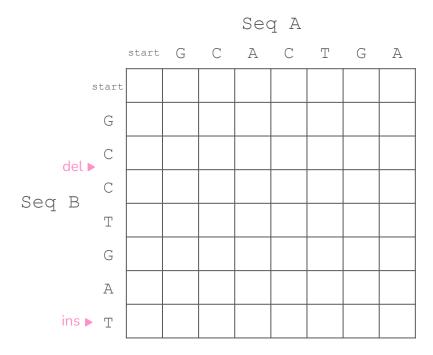
Returns the full alignment, clipped by best offset



Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:





Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

-> no offset penalties for top row, left column: Why?

 Scoring
 (Gaps: -2)

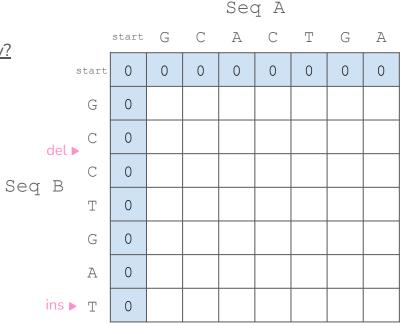
 C
 T
 A
 G

 C
 +1
 -1
 -1
 -1

 T
 -1
 +1
 -1
 -1

 A
 -1
 -1
 +1
 -1

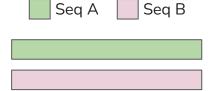
 G
 -1
 -1
 -1
 +1

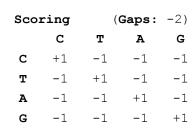


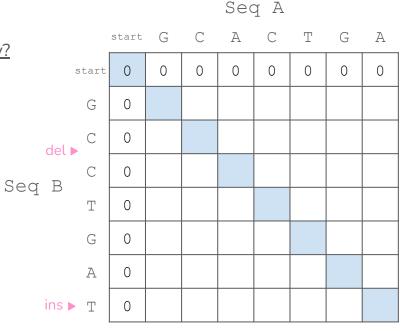
Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

-> no offset penalties for top row, left column: Why?





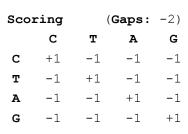


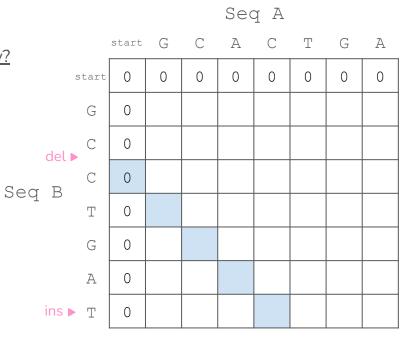
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Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

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Seq A Seq B

 Scoring
 (Gaps: -2)

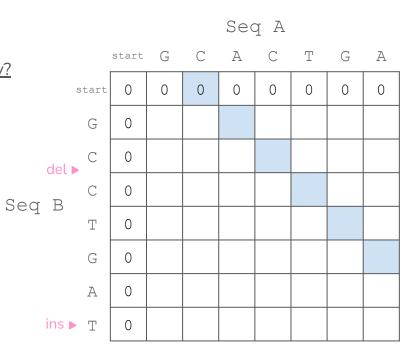
 C
 T
 A
 G

 C
 +1
 -1
 -1
 -1

 T
 -1
 +1
 -1
 -1

 A
 -1
 -1
 +1
 -1

 G
 -1
 -1
 -1
 +1



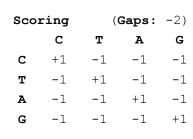
Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

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Seq A Seq B

Do not want to penalise beginning shifts



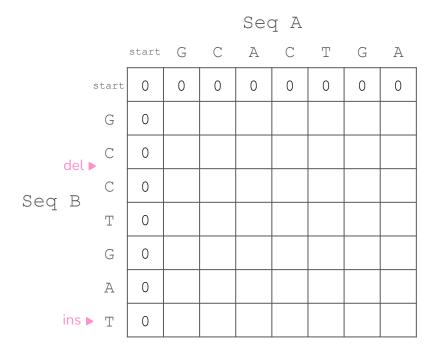
Seq A A start 0 G del ▶ Seq B ins ▶ T

Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

-> no offset penalties for top row, left column

Scoring		(Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1



Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

-> no offset penalties for top row, left column

Scoring		(1	Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1



Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

- -> no offset penalties for top row, left column
- -> the return score:

Max(bottom row), or Max(right column)

Scoring		(1	Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1

							_			
			start	G	С	A	С	T	G	A
		start	0	0	0	0	0	0	0	0
		G	0	1	-1	-1	-1	-1	1	-1
	del	С	0	-1	2	0	0	-1	-1	0
Seq I		С	0	-1	0	1	1	-1	-2	-2
beq	ם	Т	0	-1	-2	-1	0	2	0	-2
		G	0	1	-1	-3	-1	0	3	1
		A	0	-1	0	0	-2	-2	1	4
	ins	T	0	-1	-2	-1	-1	-1	-1	2

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Max(bottom row), or Max(right column)



Scoring		(Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1

							_			
			start	G	С	A	С	T	G	A
		start	0	0	0	0	0	0	0	0
		G	0	1	-1	-1	-1	-1	1	-1
	del	С	0	-1	2	0	0	-1	-1	0
		С	0	-1	0	1	1	-1	-2	-2
Seq	ם	T	0	-1	-2	-1	0	2	0	-2
		G	0	1	-1	-3	-1	0	3	1
		A	0	-1	0	0	-2	-2	1	4
	ins	T	0	-1	-2	-1	-1	-1	-1	2

Algorithm: Needleman Wunsch (variant)

Same as Needleman-Wunsch except:

- -> no offset penalties for top row, left column
- -> the return score:

Max(bottom row), or Max(right column)

-> the backtracking:

Starts at max cell

Ends when hit top row, or left column

GCACTGA

GC-CTGA

Scoring		((Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1

							_			
			start	G	С	A	С	T	G	A
		start	0	0	0	0	0	0	0	0
		G	0	1	-1	-1	-1	-1	1	-1
	del	С	0	-1	2	0	0	-1	-1	0
Seq		С	0	-1	0	1	1	-1	-2	-2
beq	ם	Т	0	-1	-2	-1	0	2	0	-2
		G	0	1	-1	-3	-1	0	3	1
		A	0	-1	0	0	-2	-2	1	4
	ins	T	0	-1	-2	-1	-1	-1	-1	2

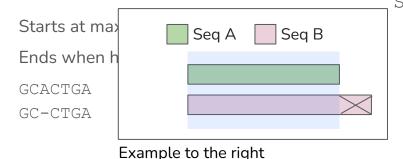
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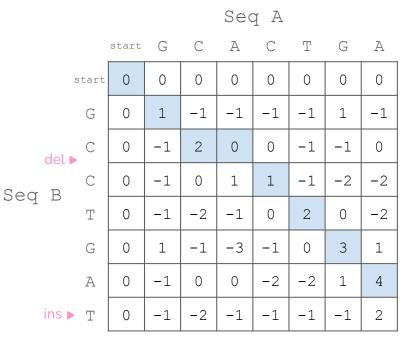
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С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1



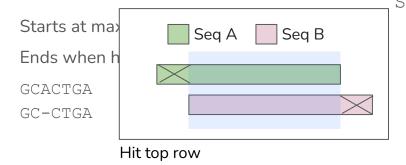
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С	+1	-1	-1	-1
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A	-1	-1	+1	-1
G	-1	-1	-1	+1



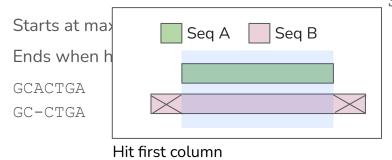
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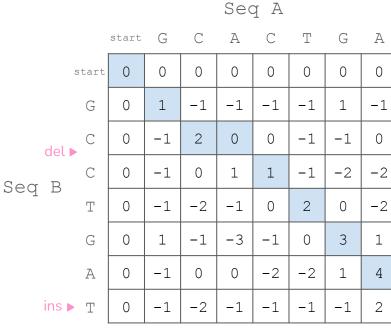
- -> no offset penalties for top row, left column
- -> the return score:

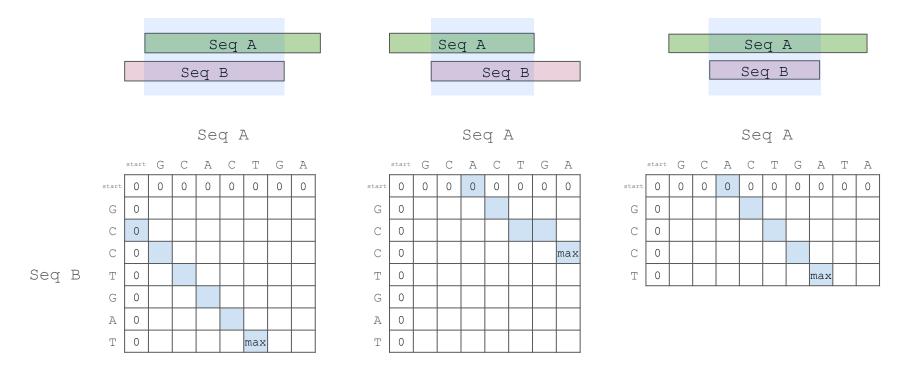
Max(bottom row), or Max(right column)

-> the backtracking:



Scoring		(Gaps:	-2)
	С	T	A	G
С	+1	-1	-1	-1
T	-1	+1	-1	-1
A	-1	-1	+1	-1
G	-1	-1	-1	+1





Region of best local similarity

Some of Seq A Some of Seq B

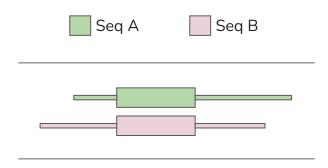
Best when sequences are dissimilar, but contain regions of similarity

eg. BLAST: gene homology

Best region dictated by penalty scores

Returns the alignment in highest scoring region

Algorithm: Smith Waterman



Gene homology between rat and human

Parts of the gene will be similar (due to evolutionary viability) (active sites, specific domains)

Parts of the gene will be dissimilar (those which are more permissive to mutation)

Region of best local similarity

Some of Seq A Some of Seq B

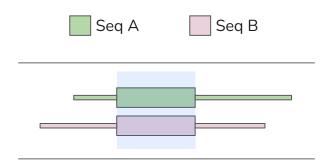
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Algorithm: Smith Waterman

Same as Needleman-Wunsch except:

- -> First row and first column set to 0
- -> Negative score set to 0
- -> the return score: max(S)
- -> the backtracking:

Starts at max(S)

Ends when hit score of zero

Sco	ring	(-2)	
	С	T	A	G
С	+2	-1	-1	-1
T	-1	+2	-1	-1
A	-1	-1	+2	-1
G	-1	-1	-1	+2

A 0 0
0
0
0
2
6
10
8

Algorithm: Smith Waterman

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Sco	ring	(-2)	
	С	T	A	G
С	+2	-1	-1	-1
T	-1	+2	-1	-1
A	-1	-1	+2	-1
G	-1	-1	-1	+2

						_			
		start	G	С	A	С	T	G	A
	start	0	0	0	0	0	0	0	0
	G	0	2	0	0	0	0	2	0
	del ▶ C	0	0	4	2	2	0	0	0
Seq	С	0	0	2	3	4	2	0	0
bed	T	0	0	0	1	2	6	4	2
	G	0	2	0	0	0	4	8	6
	А	0	0	1	0	0	2	6	10
	ins ▶ T	0	0	0	0	0	2	4	8

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Sco	ring	((Gaps:			
	С	T	A	G		
С	+2	-1	-1	-1		
T	-1	+2	-1	-1		
A	-1	-1	+2	-1		
G	-1	-1	-1	+2		

						_			
		start	G	С	A	С	Т	G	A
	start	0	0	0	0	0	0	0	0
	G	0	2	0	0	0	0	2	0
	del ▶ C	0	0	4	2	2	0	0	0
200	С	0	0	2	3	4	2	0	0
Seq	T	0	0	0	1	2	6	4	2
	G	0	2	0	0	0	4	8	6
	A	0	0	1	0	0	2	6	10
	ins ▶ T	0	0	0	0	0	2	4	8
DCq	T G A	0	2	0 1	0	0	4 2	8	-

Algorithm: Smith Waterman

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Sco	ring	((Gaps:			
	С	T	A	G		
С	+2	-1	-1	-1		
T	-1	+2	-1	-1		
A	-1	-1	+2	-1		
G	-1	-1	-1	+2		

							_			
			start	G	С	A	С	Т	G	A
		start	0	0	0	0	0	0	0	0
		G	0	2	0	0	0	0	2	0
	del	С	0	0	4	2	2	0	0	0
Seq		С	0	0	2	3	4	2	0	0
seq	ט	Т	0	0	0	1	2	6	4	2
		G	0	2	0	0	0	4	8	6
		А	0	0	1	0	0	2	6	10
	ins I	T	0	0	0	0	0	2	4	8

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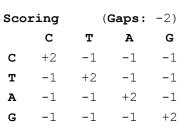
- -> First row and first column set to 0
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Ends when hit score of zero

GCACTGA

GC-CTGA



			-							
			start	G	С	A	С	T	G	A
		start	0	0	0	0	0	0	0	0
		G	0	2	0	0	0	0	2	0
	del ▶	С	0	0	4	2	2	0	0	0
Seq		С	0	0	2	3	4	2	0	0
seq	D	T	0	0	0	1	2	6	4	2
		G	0	2	0	0	0	4	8	6
		A	0	0	1	0	0	2	6	10
	ins >	Т	0	0	0	0	0	2	4	8

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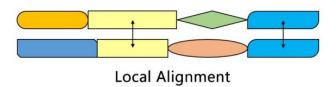
ACTGA

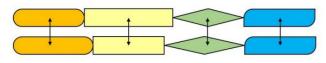
CCTGA

Sco	ring	(1	-2)	
	С	T	A	G
С	+2	-1	-1	-1
T	-1	+2	-1	-1
A	-1	-1	+2	-1
G	-1	-1	-1	+2

							_			
			start	G	С	A	С	T	G	A
		start	0	0	0	0	0	0	0	0
		G	0	2	0	0	0	0	2	0
	del	С	0	0	4	0	2	0	0	0
Seq		С	0	0	2	3	4	2	0	0
seq	ט	Т	0	0	0	1	2	6	4	2
		G	0	2	0	0	0	4	8	6
		А	0	0	1	0	0	2	6	10
	ins I	T	0	0	0	0	0	2	4	8

Local vs. Global

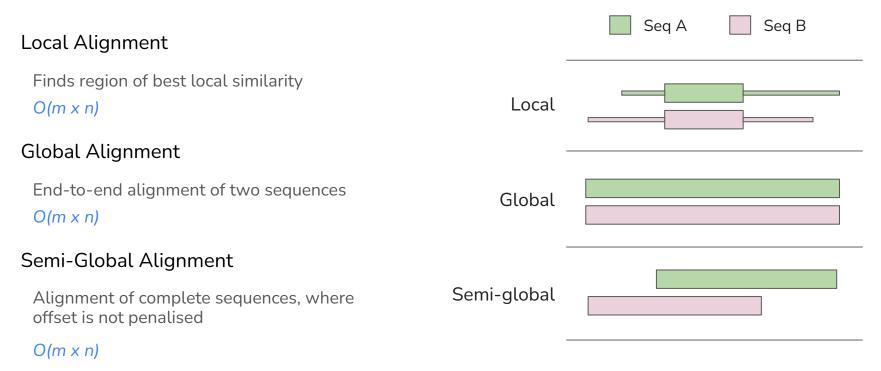




Global Alignment

	Smith-Waterman algorithm	Needleman-Wunsch algorithm
Initialization	First row and first column are set to 0	First row and first column are subject to gap penalty
Scoring	Negative score is set to 0	Score can be negative
Traceback	Begin with the highest score, end when 0 is encountered	Begin with the cell at the lower right of the matrix, end at top left cell
Complexity	$O(m \times n)$	$O(m \times n)$

Pairwise Alignment



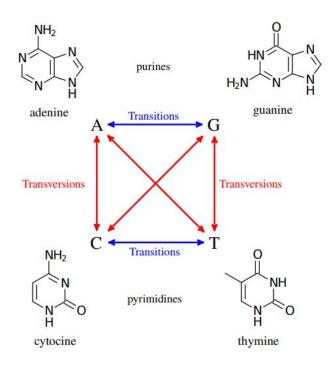
Does this scale to large datasets?

Alignment: Scoring / Substitution Matrices

Scoring Alignments

- The optimal alignment depends on our scoring system
 - Matches (reward)
 - Mismatches (penalty)
 - Starting/Extending Gaps (penalty)
- Simple match/mismatch score
 - Matches: +1
 - Mismatches/Gaps: -1
- Generalize to a substitution matrix
 - Assign a score to each pair of characters
 - N×N Symmetric matrix
 - N=4 Nucleic acids
 - N=20 Proteins
 - M(i,j) cost/reward to change from i to j

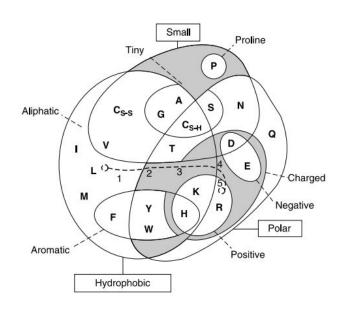
Why can't we just use a constant score for mismatches?



- two types of DNA substitution mutations:
 - transitions (bases with similar shape)
 - transversions (different number of rings)
- transition mutations occur more frequently
- transitions are less likely to result in amino acid substitutions (often synonyms)
- we can capture this in a substitution matrix

```
C T A G
C 2 1 -1 -1
T 1 2 -1 -1
A -1 -1 2 1
G -1 -1 1 2
```

Protein substitution matrix



- protein substitution matrices are more complex than DNA scoring matrices
- proteins are composed of 20 different amino acids
- varied physicochemical properties
 - compare a D to E with a D to W
- scoring matrices reflect:
 - chemical similarity
 - observed mutation frequencies
 - how protein sequences evolve

Protein substitution matrices

How often is one amino acid substituted for another in related proteins?

PAM

Point Accepted Mutation

BLOSUM

Blocks **Substitution Matrix**

PAM1 matrix

- Margaret Dayhoff in <u>Atlas of protein sequence and structure</u> (1978)
- the first widely-used amino acid substitution matrix



- derived from global alignments of closely related sequences
 - 71 groups of protein sequences
 - minimum 85% identity
 - functional proteins
 - 1572 amino-acid changes/mutations
- evolutionary model
 - assumes symmetry: $A \rightarrow B = B \rightarrow A$
 - assumes substitutions observed over short periods of time can be extrapolated to long periods of time (mathematically)

Extrapolating PAM matrices to longer distances

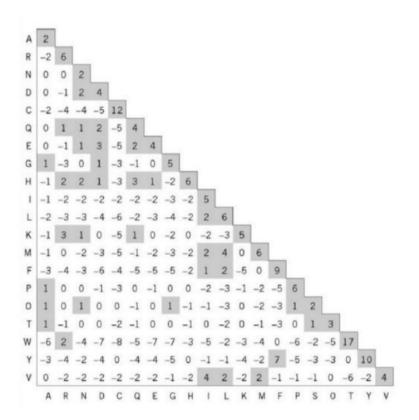
- Margaret Dayhoff in
 <u>Atlas of protein sequence and structure</u>
 (1978)
- Family of matrices: PAM1, PAM80, PAM120, PAM250
- evolutionary interval is the time taken for n mutations to occur per 100 amino acids
- the number represents the evolutionary distance between the sequences
- higher numbers denote greater distances

- PAM matrices for larger evolutionary distances are extrapolated from PAM1
- probabilities are calculated by matrix multiplication, e.g.

$$\begin{array}{ll} M_2 &= M_1 \times M_1 \\ M_{250} &= M_1^{250} \end{array}$$

 PAM250 is the substitution matrix calculated from M₂₅₀

Limitations of PAM matrices



- inferred from a small dataset with >85 % identity
- mainly small globular proteins
- doesn't account for different evolutionary rates between conserved and non-conserved regions

BLOSUM (BLOcks SUbstitution Matrix)

-	C	S	Т	Α	G	P	D	Е	Q	N	Н	R	K	M	I	L	V	W	Υ	F	
C	9																				C
S	-1	4																			S
Т	-1	1	5																		T
Α	0	1	0	4																	Α
G	- 3	0	-2	0	6																G
P	-3	-1	-1	- 1	-2	7															P
D	-3	0	-1	-2	-1	-1	6											$\overline{}$			STREET, SQUARE,
E	-4	0	-1	-1	-2	-1	2	5													DEO
0	- 3	0	-1	- 1	-2	-1	0	2	5												0
N	-3	1	0	-2	0	-2	1	0	0	6											N
Н	-3	-1	-2	-2	-2	-2	-1	0	0	1	8						_	\vdash			Ц
	170220		- 2	- 2	-2		- 33	0	0	1		-									H R
R	- 3	-	-1	- 1	- 5	-2	-2	0	1	0	0	5	-								K
K	- 3	0	-1	-1	-2	-1	-1	1	1	0	-1	2	5	-		_	_	-		_	K
M	-1	-1	-1	- 1	-3	-2	-3	- 2	0	- 2	-2	-1	-1	5	- 14						MI
I	-1	- 2	-1	-1	- 4	-3	- 3	- 3	- 3	-3	-3	- 3	- 3	1	4						I
L	-1	- 2	-1	-1	-4	-3	-4	- 3	- 2	-3	-3	- 2	- 2	2	2	4					L
٧	-1	- 2	0	0	- 3	- 2	- 3	- 2	- 2	- 3	-3	- 3	-2	1	3	1	4				٧
W	-2	- 3	- 2	- 3	- 2	-4	-4	- 3	-2	-4	-2	- 3	-3	-1	- 3	-2	-3	11			W
Y	- 2	- 2	-2	-2	-3	-3	- 3	-2	-1	-2	2	-2	-2	-1	- 1	-1	-1	2	7		Y
F	-2	- 2	-2	-2	-3	-4	- 3	-3	-3	-3	-1	-3	-3	0	0	0	-1	1	3	6	F
	C	S	Т	Α	G	P	D	E	Q	N	Н	R	K	M	I	L	V	W	Υ	F	

- Henikoff & Henikoff, Amino acid substitution matrices from protein blocks (1992). 10.1073/pnas.89.22.10915
- alignments of 500 distantly related protein families
- scores derived from frequencies of substitutions in blocks of ungapped local alignments
- BLOSUMx is based on sequences that share at least x% identity
 - e.g. BLOSUM62 was constructed from aligned sequences sharing no more than 62 % identity

Image: Paul Gardner via Wikimedia Commons

PAM vs. BLOSUM

BLOSUM80 PAM1

BLOSUM62 PAM120 BLOSUM45 PAM250

Less divergent



More divergent

- For closely related proteins: lower PAM matrices or higher BLOSUMs
- For distantly related proteins higher PAM matrices or lower BLOSUMs
- BLOSUM62 is commonly used for database searching (BLAST default)

- In general:
 - BLOSUMs perform well for local similarity searches
 - PAM matrices perform well for global alignments
- BLOSUMs are calculated from observed frequencies
- higher PAM matrices are extrapolated mathematically

Alignment: Gap Penalties

Why penalize gaps?

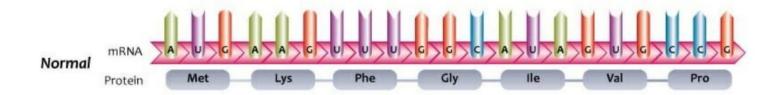
 allowing gaps with no cost results in misleading alignments

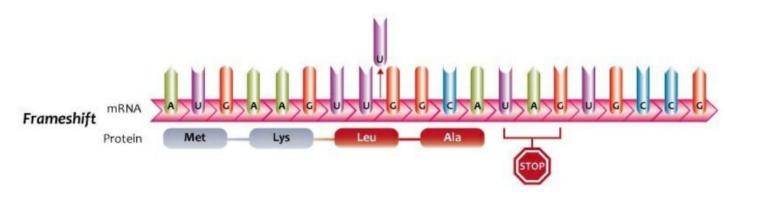
optimal alignment: **maximizes** the number of matches, and **minimizes** the number of gaps



- tradeoff: adding gaps reduces mismatches
- penalising gaps heavily forces alignments to have fewer gaps

Why penalize gaps?





Adapted from Campbell NA (ed). Biology, 2nd ed, 1990.

How do we penalise gaps?

Negative score:

- Gap penalty should be several times greater than the mismatch penalty
 - Proteins: an insertion/deletion could interrupt the entire polymer chain
 - DNA: shift the reading frame

A naïve approach:

- fixed penalty (σ) for every gap/indel
 - -σ for one indel
 - -2σ for two consecutive indels
 - -3σ for three consecutive indels
- what is the problem with that?

```
V D S - C Y
V E S L C Y
```

Fixed gap penalty

Alignment 1:

ATGTAGTGTATAGTACATGCA
ATGTAG-----TACATGCA

- an indel of length k is more likely to occur as a single event than as k events each of length 1
- i.e. alignment 1 is a better representation of homology

Alignment 2:

ATGTAGTGTATAGTACATGCA
ATGTA--G--TA---CATGCA

- a fixed gap penalty would give the same score for both
- treat gap initiation and gap extension differently

Affine gap penalties

affine: linear (in this context).

i.e. the penalty grows at the same rate as the length of the gap.

- large penalty for opening a gap
- much smaller penalty for gap extension
- \odot an indel of length k has a penalty W(k)
 - $W(k) = -(\rho + \sigma \times k)$
 - ρ: penalty to open a gap
 - σ: penalty to extend a gap
- e.g. used by BLAST

Recap

Kmers

Indexing

Subsequences of length K

Method to quickly identify matching regions of two sequences

Used extensively in bioinformatics

Key: Value store

Kmers: Occurrences

Alignment

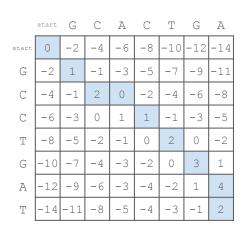
Finding optimal match between

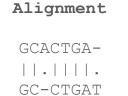
two sequences

Considers matches, mismatches,

gaps

donomo	-
donomo	
genome — — —	
kmer index hits	
Hash Table (index	()
CGT 3	
GCG 2	
GGG 8	
GGG 9	
G G G 6 10	
GTG 0	
GTG 4	
GTG 6	
kmers TGC 1	
TGG 7	
read TGT 5	





Mapping reads to human reference genome

Ideally, we could use semi-global alignment

Is this feasible for average dataset?

- -> each read is ~ 100 bp
- -> reference length ~ 3.2 billion bp
- -> number of reads ~ 300 million (30x coverage)

Semi-global alignment time and space complexity

- -> quadratic: $O(n \times m)$
- -> single read: 300 billion operations
- -> average dataset: 10²⁰

Not feasible!

We need a heuristic approach



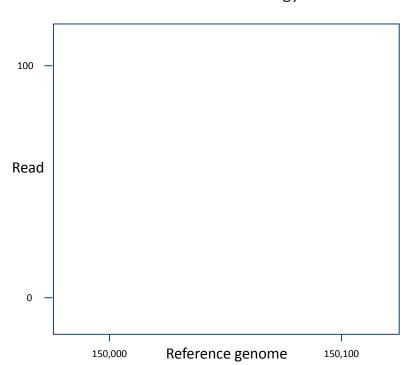
Needle in haystack via wikipedia commons (CC BY-SA 4.0)

Seed-Extend

Combines indexing and alignment

Used for aligning short, accurate sequences

Process



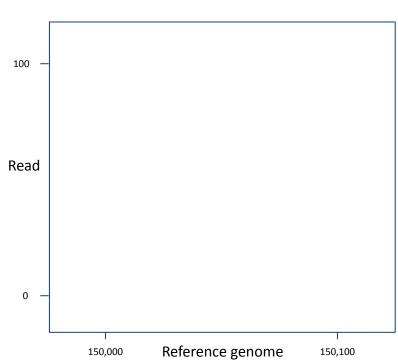
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Process

Index the reference genome using kmers



Seed-Extend

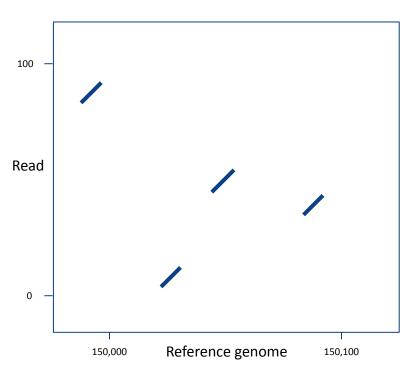
Combines indexing and alignment

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Process

Index the reference genome using kmers

Use the index to find "seed" matches for each read (kmer hits between read and index)



Seed-Extend

Combines indexing and alignment

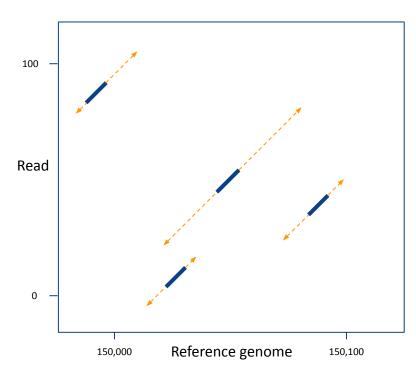
Used for aligning short, accurate sequences

Process

Index the reference genome using kmers

Use the index to find "seed" matches for each read (kmer hits between read and index)

Extend the match ends using alignment (Local alignment and / or gap-free alignment)



Seed-Extend

Combines indexing and alignment

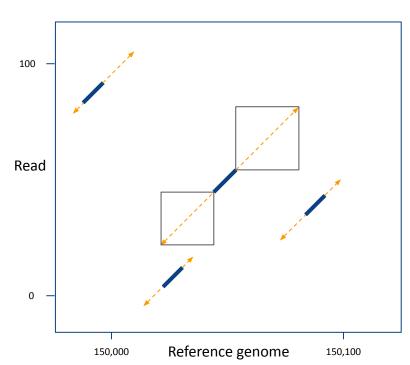
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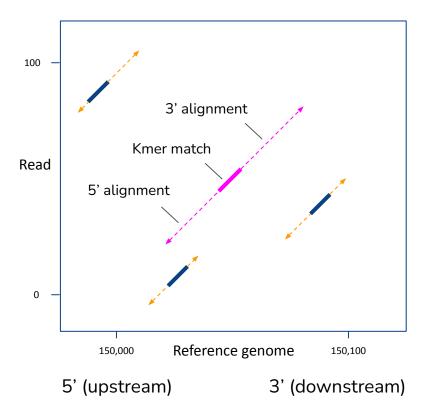
Process

Index the reference genome using kmers

Use the index to find "seed" matches for each read (kmer hits between read and index)

Extend the match ends using alignment (Local alignment and / or gap-free alignment)

Return the best location (5' alignment + kmer match + 3' alignment)

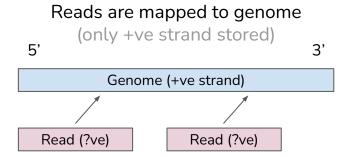


What about reverse strand?

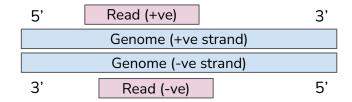
Genome: Double Stranded DNA (except some viruses)

DNA Seq. Reads: Can originate from +ve or -ve strand

Reference genome: Only the +ve strand



But reads originate from both strands of genome!



What about reverse strand?

Genome: Double Stranded DNA (except some viruses)

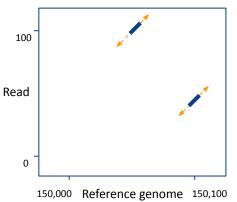
DNA Seq. Reads: Can originate from +ve or -ve strand

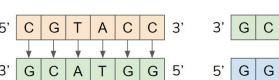
Reference genome: Only the +ve strand

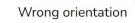
For each read (sequence to align)

- 1. Do Seed-Extend (original forward orientation).
- 2. Flip it (reverse complement).

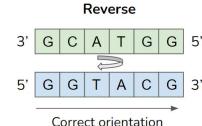
Read in Fwd (original) Orientation







Calculate Complement



What about reverse strand?

Genome: Double Stranded DNA (except some viruses)

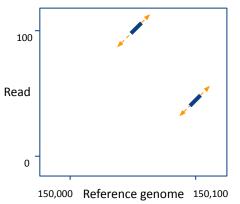
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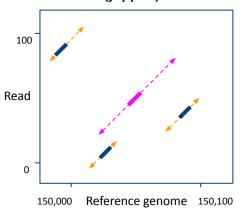
For each read (sequence to align)

- 1. Do Seed-Extend (original forward orientation).
- 2. Flip it (reverse complement).
- 3. Do Seed-Extend (flipped reverse orientation).
- 4. Take the best alignment from Fwd & Rev orientation.
- 5. If it's Rev, report seq as originating from the -ve strand.

Read in Fwd (original) Orientation



Read in Rev (flipped) Orientation



Aligning long, noisy sequences

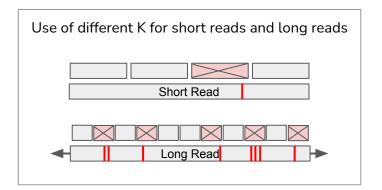
Long-read data has unique properties due to sequencing technology

Aligning long, noisy sequences

Long-read data has unique properties due to sequencing technology

Error rate

- -> Long-read data more noisy
- -> ~1 error every 10 bp (improved by 2023)
- -> Kmer size needs to be lower



Aligning long, noisy sequences

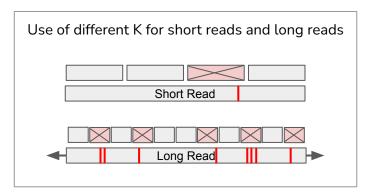
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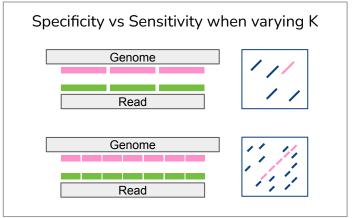
Error rate

- -> Long-read data more noisy
- -> ~1 error every 10 bp (improved by 2023)
- -> Kmer size needs to be lower

Read length

- -> Length ~10kb
- -> Many kmers -> seeds for a single read (thousands)
- -> Alignments for each seed: very expensive

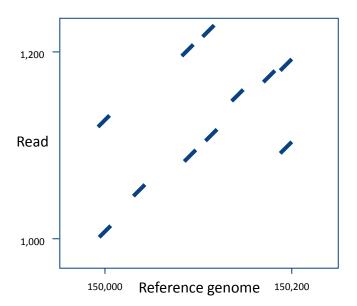




Can we use this to our advantage?

For a given read, many kmers will match the genome index

Expect the relative kmer positions to be similar in the read and genome

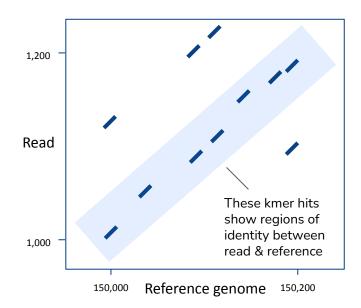


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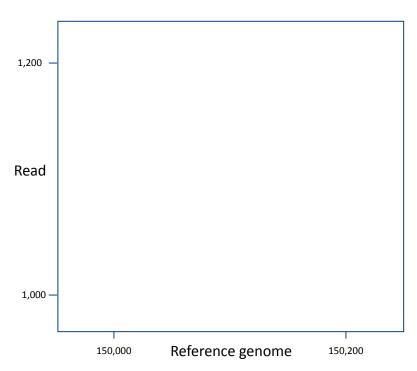
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Expect the relative kmer positions to be similar in the read and genome

Chaining

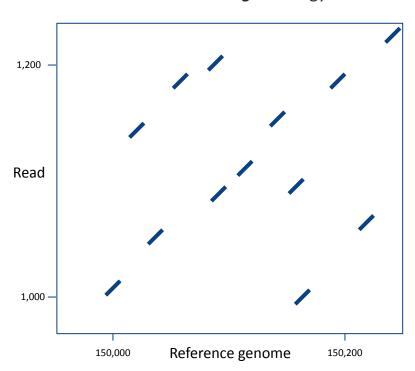


Seed-Chain-Align



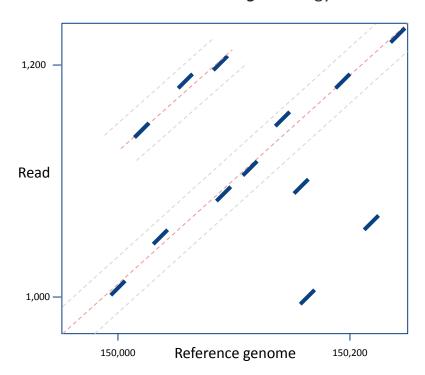
Seed-Chain-Align

1. Break the read into k-mers and look up their genomic positions in the index to find seeds Note: don't need to extract every possible kmer from the read – can jump a little in between. Common to extract a kmer every few – tens of base pairs.



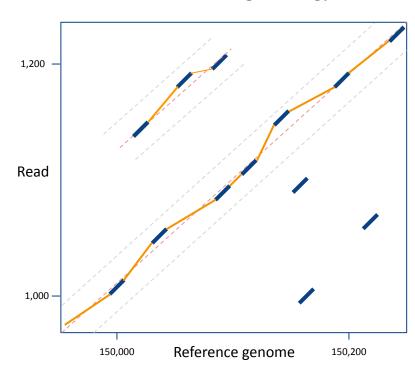
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- 2. Identify colinear chains



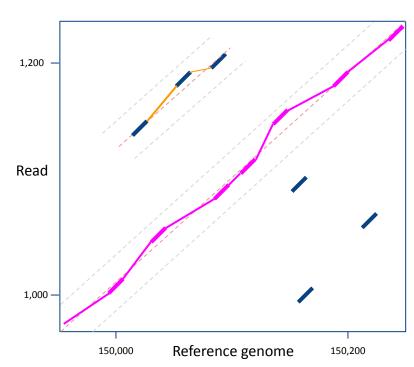
Seed-Chain-Align

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- 3. For each: base-level alignments to fill gaps



Seed-Chain-Align

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- 2. Identify colinear chains
- 3. For each: base-level alignments to fill gaps
- 4. Return the best location (gap-filled chain with highest score)

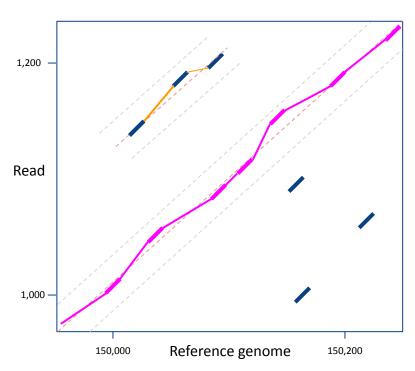


Seed-Chain-Align

- Break the read into k-mers and look up their genomic positions in the index to find seeds Note: don't need to extract every possible kmer from the read – can jump a little in between. Common to extract a kmer every few

Expanded upon in week 3 caugnments to fill gaps

arn the best location (gap-filled chain with highest score)

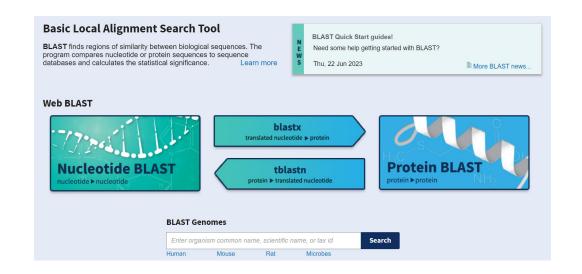


Basic Local Alignment Search Tool (BLAST) - Extreme efficiency heuristics!

Finding conserved sequences (eg. genes)

Query sequence -> Massive database

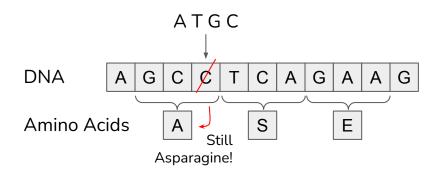
Somewhere between Seed-Extend and Seed-Chain-Align



Extreme efficiency heuristics

Extreme efficiency heuristics

- 1. DNA translated to protein seq for use
 - -> AA seq more conserved than DNA seq
 - -> Degeneracy / codon wobble
 - -> 3rd base in codon: multiple different nucleotides encode same AA
 - -> Eliminates meaningless DNA mismatches
 - -> More useful kmer hits (mismatches, but coding seq undisturbed)



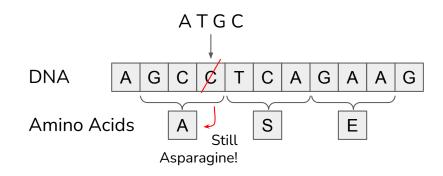
Extreme efficiency heuristics

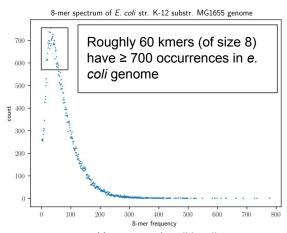
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2. Low-complexity regions removed

- -> Repetitive DNA
- -> Functional elements generally not repetitive
- -> Leads to uninformative kmer seeds (if retained)



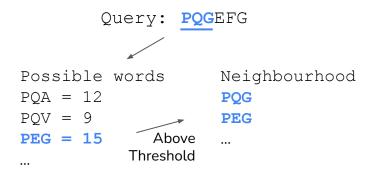


Ytngargar via wikipedia commons

Extreme efficiency heuristics

- 3. Kmers are fast, let's maximise their use
 - -> Generate kmer (word) 'neighbourhood' for query kmers
 - -> Referred to as
 - -> Allows mismatches in the seed step (prev. only exact kmer matches in this lecture)
 - -> Similar to short ungapped alignment
 - -> Kmer matches must be above threshold score (high scoring words)
 - -> Words in neighbourhood looked up in index

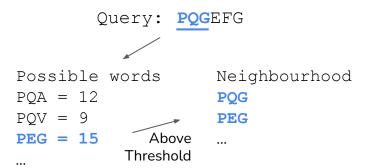
Generating Word Neighbourhood



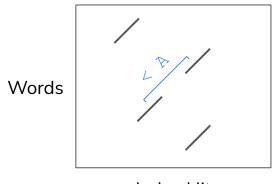
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 - -> Similar to short ungapped alignment
 - -> Kmer matches must be above threshold score (high scoring words)
 - -> Words in neighbourhood looked up in index
- 4. Identify nearby word hits on same diagonal
 - -> Similar to Chaining in Seed-Chain-Align
 - -> Word distance < A
 - -> Extends matching region to a: "High-scoring Segment Pair (HSP)"

Generating Word Neighbourhood



Matching words on diagonal



Index Hits

Extreme efficiency heuristics

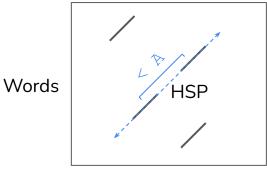
5. Ungapped extension

- -> Extend HSP on either end
- -> How far do we extend? Stop when score < threshold (efficiency)
- -> Rank HSPs by E-score & do cutoff

How likely this segment would appear in random sequence, same size of our database?

Probability that this segment appears by simple chance. Lower is better.

Ungapped extension



Index Hits

Extreme efficiency heuristics

5. Ungapped extension

- -> Extend HSP on either end
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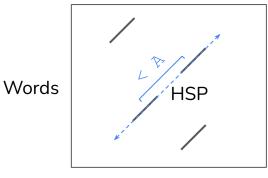
How likely this segment would appear in random sequence, same size of our database?

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6. Gapped alignment

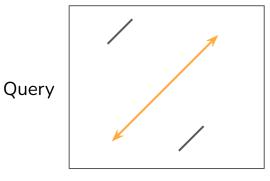
- -> Local-alignment for HSP + end extension (early termination)
- -> Recalculate E-score & report alignments greater than threshold

Ungapped extension



Index Hits

Final Alignment



Database

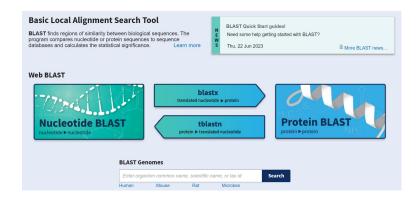
Summary

Iterative approach

Ruthlessly & Continuously reduces search space

Very fast!

Clever thought applied to properties of genomic data.



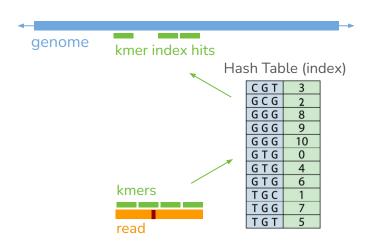
Sequence Alignment

Kmers

breaking sequence into smaller pieces

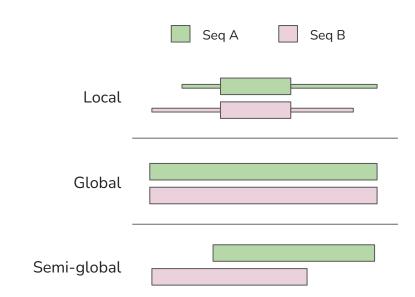
Indexing

Use of kmers + hash tables Fast lookup of subsequence matches



Alignment

Different variations on Levenshtein distance ...for different tasks







Thank you!

Don't forget your signed academic integrity statement **Due tomorrow!!**

Today: Sequence Alignment II

Next time: Comparing Sequences I