JDRF Sensitivity / Benchmark

Dror Berel 2019-03-29

Benchmarking on pre-processing parameters:

- 1. Univariate cuttof at each assay (top_n)
- 2. clustering minimum correlation (h)

A. Setup

1. Load raw data sets and scale

```
data_task # mlr's task format. currently non-functional, so require sep
## Supervised task: mlr.Data
## Type: regr
## Target: cpep_model_decayrate
## Observations: 31
## Features:
##
                   factors
                               ordered functionals
      numerics
         76746
##
## Missings: TRUE
## Has weights: FALSE
## Has blocking: FALSE
## Has coordinates: FALSE
# pre-processing: Scaling
task_j<-data_task %>>% cpoScale()
```

2. mlr's learner setup

```
lrn.glmnet.1.orig<-makeLearner(cl= "regr.cvglmnet", par.vals = list(alpha=1, s='lambda.min') ) # s will</pre>
lrn_PreProcess_glmnet<-Fun_lrn_univ_Clusters_All_makePrep_MaG(lrn.glmnet.1.orig,</pre>
                                                                train_F = F_PreProc_3_UnivClust_Train_M
                                                                Predict_F = F_PreProc13_BOTH_Predict_MaG,
                                                                param.Univ.filt.top.n.features = NA,
                                                                param.UnivClustRankTopN
                                                                param.cluster_method_KH
                                                                                                = NA
                                                                param.corrplot.n.clusters.k
                                                                                                = NA
                                                                param.corrplot.n.clusters.h
                                                                                                = NA
                                                                parame.gene.or.module
                                                                param.LASSO.n.features.arbitrary=NA)
Assay.Analyte.sep<-'.ZZZ.'
is.numeric(param.impute.knn.k<-20)</pre>
```

[1] TRUE

```
param.assay.type.vec<-c('Short', 'Long', 'Short', 'rep('Long', 11))
lrn_PreProcess_glmnet$par.vals[['param.assay.type.vec']]<-param.assay.type.vec
lrn_PreProcess_glmnet$next.learner$properties %<>% c(., 'missings') # ok to add only because
lrn_PreProcess_glmnet

## Learner regr.cvglmnet.preproc from package glmnet

## Type: regr

## Name: ; Short name:

## Class: PreprocWrapper

## Properties: numerics,factors,missings,weights

## Predict-Type: response

## Hyperparameters: alpha=1,s=lambda.min
```

3. Benchmarking / sensitivity parameter setup

```
## 'gold standard'
param.corrplot.n.clusters.h<-0.3
param.Univ.filt.top.n.features<-30</pre>
```

3.1 pre-processing filtering h, top_n {fixed s=cv.glmnet 'lambda.min'}

```
param.LASSO.n.features.arbitrary<-6
## Primary(Univ + cluster_rank=1)
## Secondary(univariate only, without clustering).
h_{seq}<-c(seq.int(0, 20, by=1)/20)[-c(17:21)]; top_h_sqe<-seq.int(10, 60, by = 5)
# h_seq<-c(0.1, 0.2, 0.25, 0.3, 0.35, 0.4, 0.5, 0.75); top_h_sqe<-c(5, 10, 20, 30, 40, 50)
# cs \leftarrow seq(0,0.75, 0.05); Ns \leftarrow seq(5, 60, 5)
# 1-h_seq
bmr_tib<-expand.grid(</pre>
 hclust_cutree_h = h_seq,
               = top_h_sqe) %>% as_tibble
 Univ_top_n
# bmr_tib %<>% add_row(hclust_cutree_h = 0.3, Univ_top_n = 30, .before = 1)
## 0. setup
bmr tib %<>%
 mutate(args_vec_i = map2(Univ_top_n, hclust_cutree_h, ~list(.x, 1, 'method.h', 0, .y, 'gene', param.')
 mutate(lrd_ID_i = paste0('lrn_', 1:n())) %>%
 mutate(lrn_i = map2(args_vec_i, lrd_ID_i, ~Func_update_args_univ_clusters(lrn = lrn_PreProcess_glmnet
```

B. Run:

B.1: naive, manual loop (via appply-like / map), broken to pre-processed/baked -> s.control (cv.glmnet min) -> ML

```
# bmr_tib_naive<-bmr_tib %>% mutate(whole_run = lrn_i %>% imap(~whole_run_function(task = task_j, lrn =
# save(bmr_tib_naive, file = 'data/bmr_tib_naive.rdata')
# load(file = 'Z:/R_rhino/JDRFCAV/data/bmr_tib_naive.rdata') #
bmr_tib_naive
## # A tibble: 176 x 6
    hclust_cutree_h Univ_top_n args_vec_i lrd_ID_i lrn_i
##
                                                         whole run
             <dbl>
##
                      <dbl> <list>
                                    <chr>
                                            t>
                                                         t>
                                            <S3: PreprocWr~ <list [7~
## 1
              0
                        10 <list [7] > lrn_1
                        10 <list [7]> lrn_2
## 2
             0.05
                                           <S3: PreprocWr~ <list [7~
## 3
             0.1
                        10 <list [7]> lrn_3
                                           <S3: PreprocWr~ <list [7~
                                           <S3: PreprocWr~ <list [7~
## 4
             0.15
                        10 <list [7]> lrn_4
                        10 <list [7]> lrn_5
                                            <S3: PreprocWr~ <list [7~
## 5
             0.2
## 6
             0.25
                        10 <list [7]> lrn_6
                                           <S3: PreprocWr~ <list [7~
## 7
             0.3
                        10 <list [7] > lrn_7
                                           <S3: PreprocWr~ <list [7~
## 8
             0.35
                        10 <list [7]> lrn_8
                                           <S3: PreprocWr~ <list [7~
## 9
                        10 <list [7]> lrn_9
             0.4
                                           <S3: PreprocWr~ <list [7~
## 10
              0.45
                        10 10 17]> lrn_10 <S3: PreprocWr~ <li>17~
## # ... with 166 more rows
```

C. Extract summary results: (single tib, pre-calculater fit + lambda_opt_list)

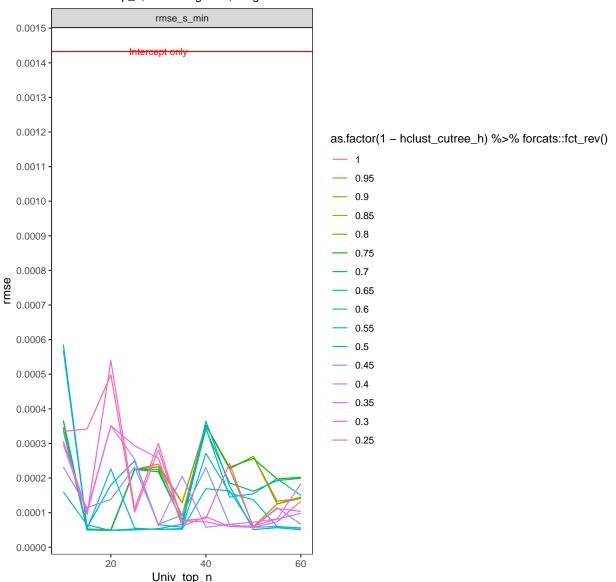
```
TargetName<-task_j %>% getTaskTargetNames
bmr_tib_naive_res<-bmr_tib_naive %>%
  mutate(fit_baked = whole_run %>% map('fit_baked')) %>%
  mutate(s =
                     whole_run %>% map('alt_lambda_list')) %>%
  mutate(Baked_x = whole_run %>% map('Baked_x')) %>%
  mutate(Baked_y = whole_run %>% map('Baked_y')) %>%
  mutate(UnivOnlyCoef = whole_run %>% map('UnivOnlyCoef')) %>%
  mutate(s_lambda.min = s %>% map_dbl('lambda.min')) %>%
                     = map2(fit_baked, s_lambda.min, ~coef(.x, s = .y) %>% tidy)) %>%
  mutate(coef_s_min
  mutate(predict_s_min = pmap(list(fit_baked, Baked_x, s_lambda.min), function(x,y,z)
      \# x = bmr\_tib\_naive\_res\$fit\_baked[[1]]; y = bmr\_tib\_naive\_res\$Baked\_x; z = bmr\_tib\_naive\_res\$s\_lambda.
     predict(object = x, newx = y, s = z) %>% data.frame %>% pull(X1) )) %>%
                     = map2_dbl(predict_s_min, Baked_y, ~sqrt(mean((.x - .y) ^ 2)))) %>%
  mutate(rmse_s_min
  mutate(n_coef_s_min = coef_s_min %>% map_int(~nrow(.x)))
# bmr_tib_naive_res %>% filter(hclust_cutree_h == 0.3, Univ_top_n == 30) %>% t
```

```
# bmr_tib_naive_res$n_coef_s_min
# bmr_tib_naive_res$n_coef_s_1se
# bmr_tib_naive_res$n_coef_s_pushed
```

plot #1: Benchmarking RMSE lines, by {h, top_n}

```
DF_rmse<-bmr_tib_naive_res %>%
  select(hclust_cutree_h, Univ_top_n, rmse_s_min) %>%
# select(hclust_cutree_h, Univ_top_n, rmse_s_min, rmse_s_1se, rmse_s_pushed) %>% gather('s_opt','rmse',
  gather('s_opt','rmse', 3)
## featureless / intercept only
y<-task_j %>% getTaskTargets
y_mean<-y %>% mean
rmse_null<-sqrt(mean((y - y_mean) ^ 2))</pre>
DF_rmse %>% filter(hclust_cutree_h == param.corrplot.n.clusters.h, Univ_top_n == param.Univ.filt.top.n.
## # A tibble: 1 x 4
   hclust_cutree_h Univ_top_n s_opt
                                               rmse
##
               <dbl>
                          <dbl> <chr>
                                              <dbl>
## 1
                 0.3
                             30 rmse_s_min 0.000225
# filter(s_opt %in% c('rmse_s_1se', 'rmse_s_pushed')) %>%
  # filter(hclust_cutree_h %in% c(0.75)) %>%
DF_rmse %>%
  # filter(hclust_cutree_h == param.corrplot.n.clusters.h) %>%
  ggplot() +
  geom_line(aes(x = Univ_top_n, y = rmse, color = as.factor(1-hclust_cutree_h) %% forcats::fct_rev() )
facet_grid(.~s_opt) +
    annotate("text", label = "Intercept only", x = 30, y = rmse_null, size = 3, colour = "red")+
scale_y_continuous(breaks = pretty(c(rmse_null, DF_rmse$rmse), n = 10)) +
  theme_bw() + theme(panel.grid.major = element_blank(), panel.grid.minor = element_blank()) +
  labs(title = 'Sensitivity analysis: RMSE for different pre-processing feature selection criteria',
       subtitle = 'Univariate: top_n, Clustering: 1-h, Single iteration of 100% train/test')
```

Sensitivity analysis: RMSE for different pre–processing feature selection criteria Univariate: top_n, Clustering: 1–h, Single iteration of 100% train/test



plot #2: s_lambda

```
ggplot(DF_rmse_s_lambda) +
   geom_point(aes(x = lambda, y = rmse, col = hclust_cutree_h %>% as.factor, shape = Univ_top_n %>% as.f
   geom_hline(yintercept = rmse_null, colour = "red") +
   facet_wrap(s_opt ~.) +
   labs(title = 'Sensitivity analysis: RMSE and LASSO lambda for different pre-processing feature select

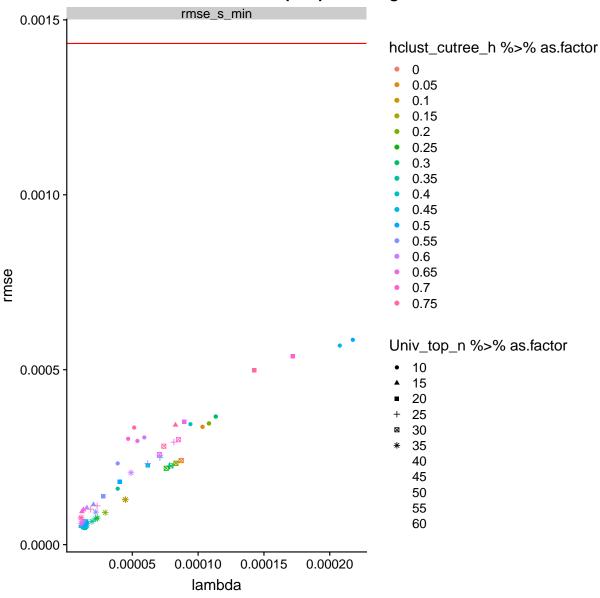
## Warning: The shape palette can deal with a maximum of 6 discrete values

## because more than 6 becomes difficult to discriminate; you have

## 11. Consider specifying shapes manually if you must have them.

## Warning: Removed 80 rows containing missing values (geom_point).
```

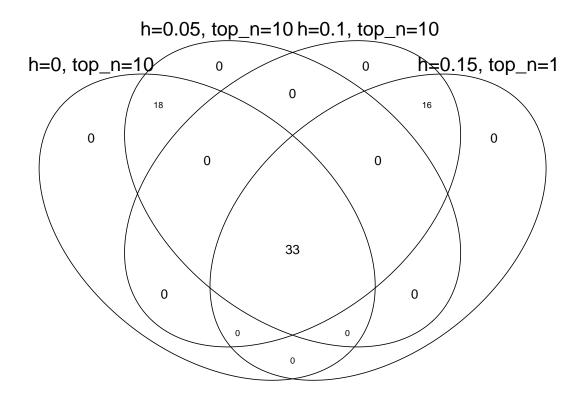
s: RMSE and LASSO lambda for different pre-processing feature selection criteria



```
# library(ggpubr)
# DF_rmse_s_lambda$hclust_cutree_h %<>% as.factor
# ggline(DF_rmse_s_lambda, x = "lambda", y = "rmse", add = c("mean_sd", "jitter"), color = "hclust_cutr")
```

Features overlap accross first 4 runs

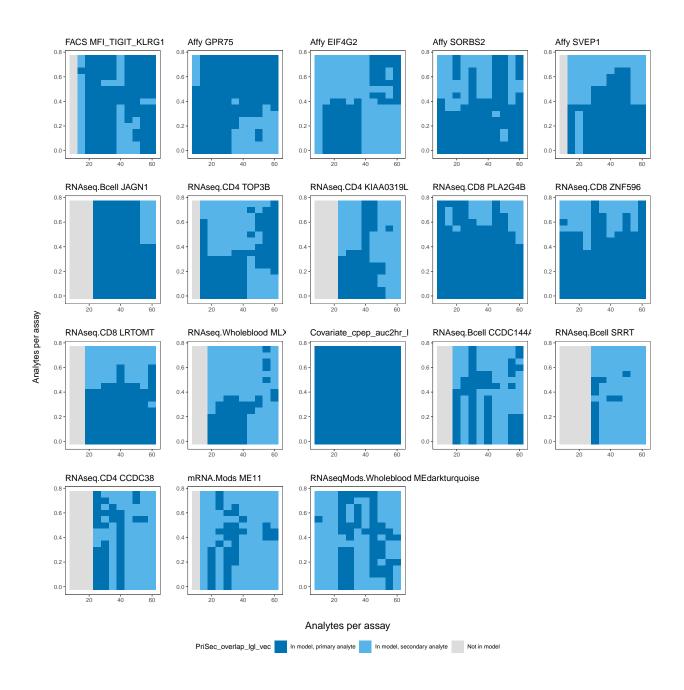
```
bmr_tib_naive_res$coef_s_min %>% setNames(str_c('h=', bmr_tib_naive_res$hclust_cutree_h, ', top_n=', bm
.[1:4] %>%
gplots::venn()
```



Plot 3: Benchmark analytes comparison: overlap between each $\{top_n, h\}i$ vs Gold-Standard $\{top_n = 30, h = 0.3\}$

```
bmr_all_comb<-bmr_tib_naive_res %>%
    select(hclust_cutree_h, Univ_top_n, coef_s_min, UnivOnlyCoef) %>%
    rename(secondary_analytes = UnivOnlyCoef) %>%
    mutate(primary_analytes = coef_s_min %>% map(~.x$row[-1])) %>%
                                                      = map2_lgl(primary_analytes, secondary_analytes, ~all(.x %in% .y))) ## Note
    mutate(check_overlap
# bmr_all_comb$primary_analytes[[1]] %in% bmr_all_comb$secondary_analytes[[1]]
# bmr_all_comb_dim<-bmr_all_comb %>% select(hclust_cutree_h, Univ_top_n) %>% table %>% dim
GS_comb<-bmr_all_comb %>%
    filter(hclust_cutree_h == param.corrplot.n.clusters.h, Univ_top_n == param.Univ.filt.top.n.features)
# REF: bmr_all_comb
GS_comb_primary_tib<-data.frame(GS_Primary_analyte = GS_comb$primary_analytes, stringsAsFactors = FALSE
GS_comb_primary_tib %<>%
    mutate(bmr_all_comb_overlap_matrix_YN = GS_Primary_analyte %>% map(~{
        \# .x = GS\_comb\_primary\_tib GS\_Primary\_analyte[[2]]
        # .x %in% bmr_all_comb$primary_analytes[[40]]
       bmr_all_comb %<>%
        ## Primary
          # .x %in% bmr_all_comb$secondary_analytes[[3]] %>% head
          # sec_list<-bmr_all_comb %>% pull(secondary_analytes)
          # sec_list %>% map(function(sec) .x %in% sec) %>% unlist
           mutate(primary_overlap_lgl_vec = primary_analytes %>%
                             map_lgl(function(primary_comb_i) .x %in% primary_comb_i) %>%
                             setNames(bmr_all_comb %>% unite('h_topN', c('hclust_cutree_h', 'Univ_top_n')) %>%
                                                   pull(h_topN) )) %>%
        ## Secondary
            mutate(secondary_overlap_lgl_vec = secondary_analytes %>%
                             map_lgl(function(secondary_comb_i) .x %in% secondary_comb_i) %>%
                              setNames(bmr_all_comb %>% unite('h_topN', c('hclust_cutree_h', 'Univ_top_n')) %>%
                                                   pull(h_topN) )) %>%
        ## Primary + Secondary (primary over secondary via any() )
            mutate(PriSec_overlap_lgl_vec = map2_chr(primary_overlap_lgl_vec, secondary_overlap_lgl_vec,
                    ~ifelse(.y, ifelse(.x, 'In model, primary analyte', 'In model, secondary analyte'),
                                    'Not in model' )))
        # end internal DF matrix for all {h,top_n} combinations
 }))
# bmr_all_comb$primary_overlap_lgl_vec %>% table
## Nested tibble with TRUE/FALSE values for (vector) overlap
\# GS\_comb\_primary\_tib\$bmr\_all\_comb\_overlap\_matrix\_YN[[1]] \ \%>\% \ select(primary\_overlap\_lgl\_vec, \ secondary\_tib\$bmr\_all\_comb\_overlap\_matrix\_YN[[1]] \ \%>\% \ select(primary\_overlap\_lgl\_vec, \ secondary\_tib\_vec, \ secondary\_tib\_
# GS_comb_primary_tib %<>% mutate(overlap_matrix_YN = bmr_all_comb_overlap_matrix_YN %>% map(~.x$primar
```

```
## coariates were forced to be at all models be definition. they are not part of the pre-processing, th
GS_comb_primary_tib[GS_comb_primary_tib$GS_Primary_analyte=='Covariate_cpep_auc2hr_log_baseline','bmr_a
## Rename analytes
# !!! FOR VISUALIZATION ONLY. NOT RELEVANT IF OTHER ANALYTES ARE SELECTED, OR IN DIFFERENT ORDER !!!
GS_comb_primary_tib$GS_Primary_analyte %<>% str_replace('.ZZZ.',' ')
GS_comb_primary_tib$GS_Primary_analyte[2:5] <-paste('Affy', c('GPR75', 'EIF4G2', 'SORBS2', 'SVEP1'), sep='
# library(RColorBrewer)
myColors <- c("#DDDDDD", "#56B4E9", "#0072B2")</pre>
names(myColors) <- c("Not in model", "In model, secondary analyte", "In model, primary analyte")</pre>
#names(myColors) <- c("TRUE", "FALSE", "NA")</pre>
GS_comb_primary_tib %<>%
  mutate(overlap_matrix_ggplot = map2(bmr_all_comb_overlap_matrix_YN, GS_Primary_analyte, ~
    \# .x = GS_comb_primary_tib\$bmr_all_comb_overlap_matrix_YN[[15]]; .y = GS_comb_primary_tib\$GS_Primar
    # DF<-.x %>% select(hclust_cutree_h, Univ_top_n, PriSec_overlap_lgl_vec)
      #DF$hclust_cutree_h %>% table; DF$Univ_top_n %>% table
      #select(hclust_cutree_h, Univ_top_n, PriSec_overlap_lgl_vec) %>%
      ggplot() + geom_tile(aes(x = Univ_top_n, y = hclust_cutree_h, fill = PriSec_overlap_lgl_vec)) +
        scale_fill_manual(values = myColors) +
        # labs(x = "Analytes per assay", y = "Min correlation in cluster", title = .y) +
        labs(x = "", y = "", title = .y) +
# scale y reverse() +
        theme(legend.position = "none", text = element_text(size=12),
              plot.title = element_text(size = 12)) +
        theme_bw(base_size = 10) + theme(panel.grid.major = element_blank(),
                                         panel.grid.minor = element_blank())
 ))
# GS_comb_primary_tib$overlap_matrix_qqplot[[15]] # MLXIP
## remove some analytes from plot
Exclude_analytes<-c(6, 8, 11, 16, 17)
GS_comb_primary_tib_ordered<-GS_comb_primary_tib[c(c(1:18)[-Exclude_analytes], Exclude_analytes),]
common_legend<-get_legend(GS_comb_primary_tib_ordered$overlap_matrix_ggplot[[1]] + theme(legend.position)
# plot_grid(NULL, common_legend, ncol=1)
Grid<-plot_grid(plotlist = GS_comb_primary_tib_ordered$overlap_matrix_ggplot %>% map(~.x + theme(legend
Grid_xlab<-ggdraw(add_sub(Grid, "Analytes per assay"))</pre>
#Grid_yx_shared<-ggdraw(add_sub(Grid_y_shared, "Min correlation in cluster"))
# Grid_no_axes_shared_legend < -plot_grid(Grid, common_legend, ncol = 1, rel_heights = c(1,.2))
ggpubr::annotate_figure(Grid_xlab, left = 'Analytes per assay', bottom = common_legend)
```



?. Session information

sessionInfo()

```
## R version 3.5.0 (2018-04-23)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 14393)
##
## Matrix products: default
##
## locale:
## [1] LC_COLLATE=English_United States.1252
```

```
## [2] LC_CTYPE=English_United States.1252
## [3] LC_MONETARY=English_United States.1252
## [4] LC NUMERIC=C
## [5] LC_TIME=English_United States.1252
## attached base packages:
## [1] stats
                 graphics grDevices utils
                                               datasets methods
                                                                    base
##
## other attached packages:
## [1] bindrcpp_0.2.2
                          knitr_1.21
                                            JDRFCAV_0.1.0
## [4] impute_1.54.0
                          limma_3.36.1
                                            biobroom_1.12.0
## [7] broom_0.5.0
                          glmnet_2.0-16
                                            foreach_1.4.4
## [10] Matrix_1.2-14
                          mlrCPO_0.3.4
                                            mlr_2.13.9000
## [13] ParamHelpers_1.12 cowplot_0.9.3
                                            ggplot2_3.1.0
## [16] tibble_2.0.1
                          tidyr_0.8.2
                                            stringr_1.3.1
## [19] purrr_0.2.5
                          magrittr_1.5
                                            dplyr_0.7.8
##
## loaded via a namespace (and not attached):
## [1] Rcpp_1.0.0
                            lattice_0.20-35
                                                gtools_3.8.1
   [4] assertthat 0.2.0
                            digest_0.6.18
                                                utf8 1.1.4
## [7] R6_2.3.0
                            plyr_1.8.4
                                                backports_1.1.3
## [10] evaluate_0.12
                            pillar_1.3.1
                                                gplots_3.0.1
## [13] rlang_0.3.1
                            lazyeval_0.2.1
                                                data.table_1.12.0
## [16] gdata 2.18.0
                            checkmate 1.9.1
                                                rmarkdown 1.11
## [19] labeling_0.3
                            splines 3.5.0
                                                munsell_0.5.0
## [22] compiler_3.5.0
                            xfun_0.4
                                                pkgconfig_2.0.2
## [25] BiocGenerics_0.26.0 BBmisc_1.11
                                                htmltools_0.3.6
                            gridExtra_2.3
## [28] tidyselect_0.2.5
                                                codetools_0.2-15
## [31] XML_3.98-1.16
                            fansi_0.4.0
                                                ggpubr_0.1.7
## [34] crayon_1.3.4
                            withr_2.1.2
                                                bitops_1.0-6
## [37] grid_3.5.0
                            nlme_3.1-137
                                                gtable_0.2.0
## [40] scales_1.0.0
                            KernSmooth_2.23-15
                                                cli_1.0.1
## [43] stringi_1.2.4
                            reshape2_1.4.3
                                                parallelMap_1.4
## [46] fastmatch_1.1-0
                            iterators_1.0.10
                                                tools_3.5.0
## [49] forcats 0.3.0
                            Biobase_2.40.0
                                                glue_1.3.0
## [52] parallel_3.5.0
                            survival_2.41-3
                                                yaml_2.2.0
## [55] colorspace_1.4-0
                            caTools_1.17.1.1
                                                bindr_0.1.1
bmr_tib_naive_res<-bmr_tib_naive %>%
  # s lambda.1se
  mutate(s_lambda.1se = s %>% map_dbl('lambda.1se')) %>%
  mutate(coef_s_1se
                    = map2(fit_baked, s_lambda.1se, ~coef(.x, s = .y) %>% tidy)) %>%
  mutate(predict_s_1se = pmap(list(fit_baked, Baked_x, s_lambda.1se), function(x,y,z)
     predict(object = x, newx = y, s = z) %% data.frame %% pull(X1) )) %%
                       = map2_dbl(predict_s_1se, Baked_y, ~sqrt(mean((.x - .y) ^ 2)))) %>%
  mutate(rmse s 1se
  mutate(n_coef_s_1se = coef_s_1se %>% map_int(~nrow(.x))) %>%
# s_lambda.pushed
  mutate(s_lambda.pushed = s %>% map_dbl('pushed_lambda')) %>%
  mutate(coef_s_pushed
                        = map2(fit_baked, s_lambda.pushed, ~coef(.x, s = .y) %>% tidy)) %>%
  mutate(predict_s_pushed = pmap(list(fit_baked, Baked_x, s_lambda.pushed), function(x,y,z)
     predict(object = x, newx = y, s = z) %>% data.frame %>% pull(X1) )) %>%
  mutate(rmse_s_pushed
                        = map2_dbl(predict_s_pushed, Baked_y, ~sqrt(mean((.x - .y) ^ 2)))) %>%
```

```
mutate(n_coef_s_pushed = coef_s_pushed %>% map_int(~nrow(.x)))
```

B.2: automatic via mlr::benchmark(). However, no manual control for s.

```
# benchmark's default require some additional nested resampling. here will force test to be the same 10
## test = 100% train
Task_j_N<-task_j %>% getTaskSize
Holdout_single_pair_AA<-makeFixedHoldoutInstance(train.inds = 1:Task_j_N, test.inds = 1:Task_j_N, size
Holdout_single_pair_AA$desc$predict = 'both'
## prediction by default is done here with the learner's original s
bmr<-benchmark(bmr_tib$lrn_i, task_j, resamplings = Holdout_single_pair_AA, measures = list(rmse))</pre>
library(tidyverse)
## -- Attaching packages ----- tidyverse 1.2.1 -
## v readr
            1.1.1
                     v forcats 0.3.0
                          _____
## -- Conflicts -----
                                                             ## x foreach::accumulate() masks purrr::accumulate()
## x Matrix::expand()
                         masks tidyr::expand()
## x tidyr::extract()
                         masks magrittr::extract()
## x dplyr::filter()
                         masks stats::filter()
## x cowplot::ggsave()
                         masks ggplot2::ggsave()
## x dplyr::lag()
                         masks stats::lag()
## x purrr::set_names()
                         masks magrittr::set_names()
## x foreach::when()
                          masks purrr::when()
library(cowplot)
df <- data.frame(</pre>
 x = rep(c(2, 5, 7, 9, 12), 2),
 y = rep(c(1, 2), each = 5),
 z = factor(rep(1:5, each = 2)),
 W = rep(diff(c(0, 4, 6, 8, 10, 14)), 2)
myColors <- c("#DDDDDD", "#56B4E9", "#0072B2", "#56E4E9", "#0172B2")
gg_master<-df %>% ggplot() + geom_tile(aes(x = x, y = y, fill = z)) +
 scale_fill_manual(values = myColors) +
 labs(x = "Analytes per assay", y = "Min correlation in cluster") +
 # scale_y_reverse() +
 theme(legend.position = "none", text = element_text(size=12), plot.title = element_text(size = 12)) +
 theme_bw(base_size = 10) + theme(panel.grid.major = element_blank(), panel.grid.minor = element_blank
gg_1<-gg_master + labs(title = 'g1')</pre>
gg_2<-gg_master + labs(title = 'g2')</pre>
gg_3<-gg_master + labs(title = 'g3')</pre>
gg_4<-gg_master + labs(title = 'g4')</pre>
gg_5<-gg_master + labs(title = 'g5')
```

```
common_legend<-get_legend(GS_comb_primary_tib_ordered$overlap_matrix_ggplot[[1]] + theme(legend.position)</pre>
pg<-plot_grid(plotlist = GS_comb_primary_tib_ordered$overlap_matrix_ggplot %>% map(~.x + theme(legend.p
plot_grid(pg, common_legend, ncol = 1, rel_heights = c(1,.1))
                                                                             Affy SVEP1
      FACS MFI
                        Affy GPR75
                                         Affy EIF4G:
                                                           Affy SORBS
           40
                         20 40
                                           20
                                              40
                                                             20 40 60
                                                                               20 40
      RNAseq.Bc
                       RNAseq.CI
                                         RNAseq.C[
                                                           RNAseq.CI
                                                                             RNAseq.CI
        20 40
                                              40
                                                             20 40 60
                                                                               20 40
              60
                         20
                            40
                                           20
      RNAseq.C[
                        RNAseq.WI
                                         Covariate_c
                                                           RNAseq.Bc
                                                                             RNAseq.Bc
                                           20 40 60
      RNAseq.CI
                                         RNAseqMods.Wholeblood MEdarkturquoise
                       mRNA.Mod
        20
           40
                         20
    PriSec_overlap_lgl_vec
                             In model, primary analyte
                                                   In model, secondary analyte
                                                                           Not in model
```

13

pg

