

# Looking at Genomic Data in R



# We are all genetic mutants!




A baby will contain between 50 and 100 mutations that are not found in either parent  
(number of mutations increases with the age of the father)

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Research Article  
Genetics and Genomics

**Large, three-generation human families reveal post-zygotic mosaicism and variability in germline mutation accumulation**

Thomas A Sasani , Brent S Pedersen, Ziyue Gao, Lisa Baird, Molly Przeworski, Lynn B Jorde , Aaron R Quinlan 

University of Utah, United States; Stanford University, United States; Columbia University, United States

Sep 24, 2019 · <https://doi.org/10.7554/eLife.46922>  

# Words you need to know

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Reference allele: allele in reference genome  
(most common; also called ancestral)

Alternate allele: mutation

Somatic mutation: mutation in DNA of body cells

Germline mutation: mutation in DNA of gametes

# Types of Genomic Variation

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## Single Nucleotide Polymorphism (SNP)

<b>Reference</b>	A	T	A	C	C	G	T	T	C	T	A	G	G
<b>Sample</b>	A	T	A	C	C	A	T	T	C	T	A	G	G

SNP: alternate allele has MAF  $>1\%$  in population

SNV: alternate allele has MAF  $<1\%$  in population

# Types of Genomic Variation

## Insertion/Deletion (Indel)

Reference	A	T	A	C	C	G	T	T	-	-	C	T	A	G	G
Sample	A	T	A	C	C	G	T	T	G	G	C	T	A	G	G

Reference	T	T	G	C	A	A	A	G	G	A	T	C	C
Sample	T	T	G	-	A	A	A	G	G	A	T	C	C

# Types of Genomic Variation

## Copy Number Variant (CNV)

Reference	T	T	G	T	-	-	-	-	-	-	A	A	A	G	G
Sample 1	T	T	G	T	G	T	-	-	-	-	A	A	A	G	G
Sample 2	T	T	G	T	G	T	G	T	G	T	A	A	A	G	G

# Exercise: Genomic Data

# Common types of genomic data files

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FASTQ

FASTA

SAM/BAM

BED

VCF

Variant Call Format





# Variant call format (VCF) file - header

```
##fileformat=VCFv4.0
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=1000GenomesPilot-NCBI36
##phasing=partial
##INFO=<ID=NS,Number=1,Type=Integer,Description="Number of Samples With Data">
##INFO=<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO=<ID=AF,Number=.,Type=Float,Description="Allele Frequency">
##INFO=<ID=AA,Number=1,Type=String,Description="Ancestral Allele">
##INFO=<ID=DB,Number=0,Type=Flag,Description="dbSNP membership, build 129">
##INFO=<ID=H2,Number=0,Type=Flag,Description="HapMap2 membership">
##FILTER=<ID=q10,Description="Quality below 10">
##FILTER=<ID=s50,Description="Less than 50% of samples have data">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT=<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
```

# Variant call format (VCF) file – body of the file

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	
20	14370	rs6054257	G	A	29	PASS	NS=3;DP=14;AF=0.5;DB;H2	
20	17330	.	T	A	3	q10	NS=3;DP=11;AF=0.017	
20	1110696	rs6040355	A	G,T	67	PASS	NS=2;DP=10;AF=0.333,0.667;AA=T;DB	
20	1230237	.	T	.	47	PASS	NS=3;DP=13;AA=T	
20	1234567	microsat1	GTCT		G,GTACT	50	PASS	NS=3;DP=9;AA=G

#FORMAT	NA00001	NA00002	NA00003
GT:GQ:DP:HQ	0 0:48:1:51,51	1 0:48:8:51,51	1/1:43:5:.,.
GT:GQ:DP:HQ	0 0:49:3:58,50	0 1:3:5:65,3	0/0:41:3
GT:GQ:DP:HQ	1 2:21:6:23,27	2 1:2:0:18,2	2/2:35:4
GT:GQ:DP:HQ	0 0:54:7:56,60	0 0:48:4:51,51	0/0:61:2
GT:GQ:DP	0/1:35:4	0/2:17:2	1/1:40:3

# Variant call format (VCF) file – body of the file

#CHROM	POS	ID	REF	ALT	QUAL	INFO
20	14370	rs6054257	G	A	29	NS=3;DP=14;AF=0.5;DB;H2
20	17330	.	T	A	3	q10 NS=3;DP=11;AF=0.017
20	1110696	rs6040355	A	G,T	67	PASS NS=2;DP=10;AF=0.333,0.667;AA=T;DB
20	1230237	.	T	.	47	PASS NS=3;DP=13;AA=T
20	1234567	microsat1	GTCT	G,GTACT	5	NS=3;DP=9;AA=G

#FORMAT	NA00001	NA00002	NA00003
GT:GQ:DP:HQ	0 0:48:1:51,51	1 0:48:8:51,51	1/1:43:5:.,.
GT:GQ:DP:HQ	0 0:49:3:58,50	0 1:3:5:65,3	0/0:41:3
GT:GQ:DP:HQ	1 2:21:6:23,27	2 1:2:0:18,2	2/2:35:4
GT:GQ:DP:HQ	0 0:54:7:56,60	0 0:48:4:51,51	0/0:61:2
GT:GQ:DP	0/1:35:4	0/2:17:2	1/1:40:3

AA - ancestral allele

AC - allele count in genotypes for each ALT allele  
(same order as listed)

AF - allele frequency for each ALT allele  
(same order as listed; used for primary data)

DB - dbSNP membership

H2 - membership in hapmap2

# Variant call format (VCF) file – body of the file

```
#CHROM    POS      ID           REF  ALT     QUAL     FILTER     INFO
20      14370      rs6054257    G    A       29      PASS      NS=3;DP=14;AF=0.5;DB;H2
20      17330      .            T    A       3       q10     NS=3;DP=11;AF=0.017
20      1110696   rs6040355    A    G,T     67      PASS      NS=2;DP=10;AF=0.333,0.667;AA=T;DB
20      1230237   .            T    .       47      PASS      NS=3;DP=13;AA=T
20      1234567   microsat1    GTCT  G,GTACT 50      PASS      NS=3;DP=9;AA=G
```

```
#FORMAT    NA000001    NA000002    NA000003
GT:GQ:DP:HQ    0|0:48:1:51,51  1|0:48:8:51,51  1/1:43:5:.,.
GT:GQ:DP:HQ    0|0:49:3:58,50  0|1:3:5:65,3    0/0:41:3
GT:GQ:DP:HQ    1|2:21:6:23,27  2|1:2:0:18,2    2/2:35:4
GT:GQ:DP:HQ    0|0:54:7:56,60  0|0:48:4:51,51  0/0:61:2
GT:GQ:DP      0/1:35:4        0/2:17:2        1/1:40:3
```

GT - genotype

GQ - genotype quality

HQ - haplotype quality

DP - combined depth across samples, e.g. DP=154

NS - Number of samples with data

# Variant call format (VCF) file – body of the file

```
#CHROM POS ID REF ALT QUAL FILTER INFO
20 14370 rs6054257 G A 29 PASS NS=3;DP=14;AF=0.5;DB;H2
20 17330 . T A 3 q10 NS=3;DP=11;AF=0.017
20 1110696 rs6040355 A G,T 67 PASS NS=2;DP=10;AF=0.333,0.667;AA=T;DB
20 1230237 . T . 47 PASS NS=3;DP=13;AA=T
20 1234567 microsat1 GTCT G,GTACT 50 PASS NS=3;DP=9;AA=G
```

```
#FORMAT NA000001 NA000002 NA000003
GT:GQ:DP:HQ 0|0:48:1:51,51 1|0:48:8:51,51 1/1:43:5:.,.
GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:3:5:65,3 0/0:41:3
GT:GQ:DP:HQ 1|2:21:6:23,27 2|1:2:0:18,2 2/2:35:4
GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51 0/0:51:2
GT:GQ:DP 0/1:35:4 0/2:17:2 1/1:40:3
```

0|0: two copies of reference allele

1|0: one reference allele, one alternate allele

1|2: two different alternate alleles

1/0: one reference allele, one alternate allele, but genome *unphased*

# Genomes make no sense without a guidebook

Annotation:

the process of describing the structure and function of the components of a genome

This helps us figure out what variants might be important for our research question.



# Human Genome Variant Databases

1000 Genomes: <https://www.internationalgenome.org/>

dbSNP: <https://www.ncbi.nlm.nih.gov/snp/>

dbVar: <https://www.ncbi.nlm.nih.gov/dbvar/>

ClinVar: <https://www.ncbi.nlm.nih.gov/clinvar/>

Exome Aggregation Consortium (ExAC):

<https://gnomad.broadinstitute.org/downloads#exac-variants>

Genome Aggregation Database (gnomAD):

<https://gnomad.broadinstitute.org/>

Genome Data Commons: <https://gdc.cancer.gov/>

# Annotation Tools:

ANNOVAR: <https://annovar.openbioinformatics.org/en/latest/>

SnEff: <https://pcingola.github.io/SnpEff/>

SIFT: <https://sift.bii.a-star.edu.sg/>

GATK VariantAnnotator

VariantAnnotation R Package:

<https://bioconductor.org/packages/release/bioc/html/VariantAnnotation.html>

Variant Annotation Integrator (UCSC):

<https://genome.ucsc.edu/cgi-bin/hgVai>

biomaRT:

<https://bioconductor.org/packages/release/bioc/html/biomaRt.html>



# Exercise: Variant Calls