# Data Descriptor Template

**Scope Guidelines**

**Data Descriptors** submitted to *Scientific Data* should provide detailed descriptions of valuable research datasets, including the methods used to collect the data and technical analyses supporting the quality of the measurements. Data Descriptors focus on helping others reuse data, rather than testing hypotheses, or presenting new interpretations, methods or in-depth analyses. Relevant datasets must be deposited in an appropriate public repository prior to Data Descriptor submission, and their completeness will be considered during editorial evaluation and peer review. The data must be made publicly available without restriction in the event that the Data Descriptor is accepted for publication (excepting reasonable controls related to human privacy issues or public safety).

### Title

Mass spectrometry peptidomics data from porcine wounds infected with *Staphylococcus aureus* and *Pseudomonas aeruginosa*

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### Abstract

Recently, mass spectrometry based peptidomics studies have proven useful in the identification of biomarkers and bioactive peptide-based therapeutics. Here, we present a dataset consistent of temporal wound fluid peptidomics data from highly defined porcine models. Wound fluids from porcine wounds infected with *Staphylococcus aureus* and *Pseudomonas aeruginosa* were sampled at different timepoints of the infection. Peptides were extracted from the samples, followed by liquid chromatography tandem mass spectrometry analysis in data dependent acquisition mode. The resulting spectra are deposited, allowing for database searching and subsequent peptidomic analysis of the infected wound fluid peptidome.

### Background & Summary

Peptides are short sequences of amino acids which are naturally produced in organisms, both through the translation of mRNA but importantly also through the degradation of proteins. These peptides play important roles in different biological systems, such as being messengers in signalling pathways1 and by providing antimicrobial properties during infection2,3,4. Peptidomics is the study of large sets of peptides from biological samples and has proven to be pivotal in the characterization of peptides and protein degradation under different physiological conditions as well as in the search for therapeutic peptides. Identification and quantification of peptides is necessary to conduct peptidomic analyses. Commonly, liquid chromatographic (LC) separation followed by mass spectrometry (MS) analysis is used for this purpose5.

During a wound infection, a combination of host and pathogen derived proteases create an environment with increased proteolytic activity. These proteases are utilized by the host to remodel tissue and fight the invasion of bacteria. It has also been speculated that similar mechanisms are utilized by the bacteria, to promote colonization6,7,8. By investigating the resulting peptidome during different wound infection conditions, further understanding of this environment and mechanism could be gained.

Chronic wounds pose a large economic burden on society and life quality burden for patients. The prevalence of these wounds is likely to increase with an aging population and more life-style related diseases9. Further, pathogens such as *Staphylococcus aureus* and *Pseudomonas aeruginosa* are two of the most prevalent bacteria in infected wounds10. They are considered of high and critical priority respectively by the World Health Organization due to their developed resistance against current antibiotics11. Therefore, identifying novel means of diagnosing and treating such wounds is of extreme importance.

This study was carried out to identify differences in the peptidomic landscape of wound fluids depending on the presence and type of pathogen, while also employing a newly developed analysis algorithm with the potential to remove large amounts of redundancy in peptidomic datasets12. This was done by generating LC-MS/MS data from wound fluids derived from highly defined infected porcine wounds (Fig. 1). Here, we present descriptions and access to the datasets. We believe that the vastness of the peptidomic data yields many opportunities to analyze it using different methods to gain novel insights about the mechanisms underlying protein degradation in infected wounds.

### Methods

**Study design**

The data generated in this study was utilized to analyze the peptidomic landscape of infected wounds in Hartman et al. (2024). Briefly, wounds were generated on Göttingen minipigs, whereafter they were infected with either *S. aureus* or *P. aeruginosa*, or not infected and used as control. Wound dressings were placed on the wounds, which absorbed the wound fluid generated in the wound healing process (Fig. 1a). The dressings were collected every 24 hours for 2-3 days depending on the sample type. Four of the wounds infected by *S. aureus* on day 1 were infected with *P. aeruginosa* on day 2, creating a double infection. Additionally, some of the wounds infected by *S. aureus* were contaminated by *P. aeruginosa* on the infection day, creating a set of samples which undergo an accidental double infection (Fig. 1b).

The proteomic content of the samples was discarded by filtration (Fig. 1c), and the subsequent peptidomic sample was analyzed with liquid chromatography tandem mass spectrometry (LC-MS/MS) in data dependent acquisition mode on an Evosep One LC (Evosep, Denmark) coupled to a timsTOF Pro MS (Bruker, USA). The data was searched with PEAKS X and deposited to ProteomeXchange (Fig. 1d). A stratified blinded random subset of samples from day 1, containing 4 samples from each single infection and control condition, were re-analyzed. The re-analysis was conducted 8 months after the original samples using the same conditions except for that the MS had been changed to a timsTOF HT (Bruker, USA) (Fig 1b).

**Pig acute wound fluid collection**

Partial thickness wounds were induced to Göttingen minipigs, which were then either infected with *S. aureus* or *P.aeruginosa*, or kept uninfected as control samples. Polyurethane dressings were used to dress the wounds and were changed after 24 and 48 hours. The old dressings were soaked in 10 mM Tris, and the fluids were extracted using a syringe. A subset of the *S. aureus* infected wounds was accidentally cross contaminated with *P. aeruginosa*. Another set of the *S. aureus* infected wounds were infected with *P. aeruginosa* after 24 hours, and these dressings were also collected and extracted 72 hours after the initial *S. aureus* infection. Extracted fluids were then supplemented with Halt Protease Inhibitor Cocktail (Thermo Fisher Scientific, USA) and kept at -80°C until further use.

**Wound fluid peptide extraction**

500 μg of protein (determined with Pierce BCA Protein Assay Kit (Thermo Fisher Scientific, USA) as per provided instructions) was diluted to 100 μl with 10 mM Tris at pH 7.4. 300 μl of 8 M urea diluted in 10 mM Tris at pH 7.4, supplemented with 0.067% RapiGest SF (Waters, USA) was then added followed by an incubation at room temperature for 30 minutes. 100 μl 6 M urea in 10 mM Tris at pH 7.a was added to Microcon - 30 centrifugal filter units and centrifuged at 10000 RCF for 15 minutes at room temperature (RT). After this the wound fluid samples were added to the centrifugal filters, and centrifuged at 10000 RCF for 30 minutes at RT. Finally, another 100 μl of 6 M urea in 10 mM Tris at pH 7.4 was centrifuged through the filter units at 10000 RCF for 5 minutes at RT and the filtrate was stored at -20°C.

**Acidification and solid phase extraction**

1 μl 100% formic acid (FA) was added to 60 μl of each extracted peptide sample. 100 μl 100% acetonitrile (ACN) + 0.1% FA was added to UltraMicro Spin Columns (The Nest Group, USA) which were centrifuged at 800 RCF for 1 minute at room temperature. All further centrifugation steps in this section were performed this way. 100 μl 2% ACN + 0.1% trifluoroacetic acid (TFA) was centrifuged through the columns, twice, before adding the samples and performing an additional centrifugal step. Lastly, 100 μl 70% ACN + 0.1% TFA was centrifuged through the columns to elute the sample which was then dried in an Eppendorf Concentrator plus (Eppendorf, Germany).

**LC-MS/MS**

The dried peptide samples were then dissolved in 30 μl 2% ACN + 0.1% FA, before being loaded onto Evotip Pure columns (Evosep, Denmark) according to the provided instructions, apart from that the samples were not dissolved in 20 µl 0.1% FA before loading. The samples were analyzed by LC/MS-MS on an Evosep One LC (Evosep, Denmark) coupled with a timsTOF Pro mass spectrometer (Bruker, USA). The LC used a EV1137 Performance Column - 15 cm x 150 µm, with 1.5 µm ReproSil-Pur C18 beads (Evosep, Denmark). The accompanying 30 samples per day program was used for separation. The MS used the DDA PASEF mode, with 10 PASEF scans every acquisition cycle. Accumulation and ramp times were set to 100 ms, precursors with a +1 charge were ignored, and target intensity was set to 20000, with dynamic exclusion active, at 0.4 min. Isolation width was set to 2 at 700 Th and 3 at 800 Th.

**Database search**

The data from the LC-MS/MS runs were searched with PEAKS X. UniProtKB reviewed (Swiss-Prot) protein list of pig (organism\_id:9823) proteins was used as a database, but with fibrinogen alpha chain (FIBA\_PIG) and fibrinogen beta chain (FIBB\_PIG) being changed to the UniProt KB unreviewed (TrEMBL) versions F1RX36\_PIG and F1RX37\_PIG. The list was downloaded May 11th, 2023. Data refinement was set to merge scans and correct precursor based on mass and charge states with charges between 1 and 4. It was also set to associate features between 2 and 8. Precursor tolerance was set to 20.0 ppm using monoisotopic mass and fragment tolerance was set to 0.03 Da. 1 modification per peptide was allowed with oxidation (M, +15.99) being the only possible modification. Results were filtered at 1% FDR with ≥1 unique peptide for each protein. FDR was set to be estimated with decoy-function.

### Data Records

Both the raw mass spectrometry data (.d folders generated by Bruker Compass) as well as the database search of the data (.mgf and .mzid.gz generated by PEAKS X) have been uploaded to ProteomeXchange as a part of the public dataset PXD048892 along with a design file for sample identification (.csv) (<https://dx.doi.org/10.6019/PXD048892>). Additionally, similar files from the blinded re-run of samples have been uploaded to ProteomeXchange under the identifier PXD055074 (<https://dx.doi.org/10.6019/PXD055074>).

### Technical Validation

To get an understanding of the data, general characteristics of the different groups were summarized (Fig. 2). Unique peptide overlap was summarized, with more unique peptides appearing in the *S. aureus* (2115)and *P. aeruginosa* (4762) groups compared to the control group (370), while the number of peptides shared by all samples was 1774 (Fig 2.a). The number of identified peptides decreases over time (Fig 2.b). The mean length distribution weighted by the log2 intensities were similar for all sample types (Fig 2.c). The log2 intensities were scaled to a mean of 0 and a unit variance. Thereafter, the dimensionality of the data was reduced using Uniform Manifold Approximation Projection (UMAP). Default settings were used as per the umap-learn python package (Fig 2.d). The data cluster based on infection type and sampling day, suggesting that there are differences in the peptidomes depending on infection type and time point.

To validate that our peptide extraction and mass spectrometry methods are reproducible, 4 wound fluid samples from day 1 and each of the groups *S. aureus*, *P. aeruginosa*, and control were randomly selected and had their peptides extracted again (Fig. 1b). Once extracted, the samples were blinded and analyzed by LC-MS/MS as the previous samples, but using a timsTOF HT (Bruker, USA) MS. It was seen that the unique peptides identified in each sample group corresponded well with peptides previously seen in the group, with fewer observed unique peptides in the blind group due to missing values (Fig 3.a, b, c), and that unique peptide overlap is similar for blinded samples compared to their respective group (Fig 3.d). It can also be seen that the peptide length distribution is retained (Fig 3.e), and that the samples cluster together with the other samples from the same group and day when reducing dimensionality using UMAP (Fig 3.f), showing that the method is robust and replicable.

### Usage Notes

The data was supplied as both raw output files and result files searched with PEAKS X. The raw files can be used for searching the data with different softwares or parameters than the one presented in this study. There are many ways to analyze the data in the resulting result files. MS intensities follow a lognormal distribution, so for comparative analyses the log of the intensities is computed prior to assuming normal distributions. The data contains missing values, and one can deal with these in different ways, e.g. through imputation. Further, to remove technical bias effects, normalization of the peptide intensities is commonly applied.

### Code Availability

No custom code was used to generate the data.

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### Author contributions

FF and EH wrote the manuscript. FF, SK and JP performed the sample preparation. FF and SK performed the mass spectrometry analysis. AS and JM supervised the project.

### Competing interests

### The authors declare no competing interests.

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### Figures

### Figure Legends

Figure 1. Flowchart of experiments. **a** Wounds were generated onto pigs and dressed with polyurethane dressings, which absorb fluids from the wound. **b** Dressings were collected at different points in time and the contained wound fluids were extracted and supplemented with protease inhibitor. **c** Wound fluids were filtered to separate the peptides from the larger molecules found in the samples. **d** Filtered peptides were purified using solid phase extraction, before being analyzed by liquid chromatography-mass spectrometry. The results were then databased searched and uploaded to an online repository.

Figure 2. Characteristics of samples and groups. S.a – *Staphylococcus aureus*, P.a – *Pseudomonas aeruginosa*, Ctrl – Control. **a** Venn diagram showing the distribution of unique peptides between the three sample groups S.a, P.a and Ctrl. **b** The number of unique peptides observed in each sample. **c** The measured intensities of each peptide was log2-transformed and summarized based on number of amino acids, sample group and day. This sum was then divided by the sum of all transformed intensities within that group to generate the relative intensity of peptides for each peptide length for each sample group. **d** Uniform Manifold Approximation and Projection dimensionality reduction based on the log2-transformed intensity for every peptide in each sample.

Figure 3. Characterization and comparison with blinded samples. S.a – *Staphylococcus aureus*, P.a – *Pseudomonas aeruginosa*, Ctrl – Control. **a**

### Tables

### Additional Formatting Information