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# Optics Letters

## Ultrasonic detection based on polarization-dependent optical reflection

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Owing to their extremely wide bandwidths, pure optical ultrasonic detection methods are gaining increasing interest. In this Letter, we proposed a simple ultrasonic detector that is based on the polarization-dependent optical reflection. When the acoustic wave reaches the liquid-glass interface, the acoustic pressure changed the relative refractive index between two media, leading to perturbations in the reflectance of the optical probe beam in glass. Unlike previous studies that detected the modulations in the intensity of the reflected beam, our method, named “polarization-dependent reflection ultrasonic detection (PRUD),” detects the intensity difference between two polarization components of the same probe beam. The PRUD significantly increased the sensitivity. Besides a phantom study, we also successfully detect weak photoacoustic waves in an *in vivo* animal experiment. This novel method can provide a simple way for ultrasonic detection, which will have great potential for ultrasound and photoacoustic imaging and sensing. © 2017 Optical Society of America

**OCIS codes:** (170.0110) Imaging systems; (170.5120) Photoacoustic imaging; (110.7170) Ultrasound; (130.6010) Sensors.

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Ultrasonic detectors with broadbands, small sizes, and high sensitivity are desired for both ultrasound imaging and the emerging photoacoustic tomography (PAT). It is particularly valuable for PAT since the large-scale differences in various optical absorbers could lead to extremely wide bandwidth photoacoustic pressure waves [1–4]. Although various kinds of piezoelectric-material-based ultrasound transducers have been commercially available and widely used, these transducers generally have limited bandwidths (centered at their resonant frequency), and the electrical noise significantly increases as the transducer becomes smaller. Due to their high sensitivity, superior wide bandwidths, small effective active sizes, as well as other good characteristics, several pure optical ultrasonic detection methods have been developed, including the Mach–Zehnder interferometer [5], the surface displacement interferometer [6], full-field speckle interferometry [7], the Fabry–Perot polymer [8], the microring resonator [9,10], the low-coherence interferometer [11], and the surface plasmon resonance detector [12].

Among these pure optical methods, one of the simplest is based on the modulations of light reflectivity at an interface due to the refractive index variations caused by the ultrasonic pressure [13,14]. A typical setup of this method was presented in Fig. 1. The optical power incident on the interface is  $P_0$  and the reflected power is  $R_{\text{opt}}P_0$ , where  $R_{\text{opt}}$  represents the reflectivity.

According to the Fresnel’s reflection law, the optical intensity reflectivity on the interface can be derived as

$$R_s = \left[ \frac{\sin(r_w - r_g)}{\sin(r_w + r_g)} \right]^2, \quad R_p = \left[ \frac{\tan(r_g - r_w)}{\tan(r_g + r_w)} \right]^2, \quad (1)$$

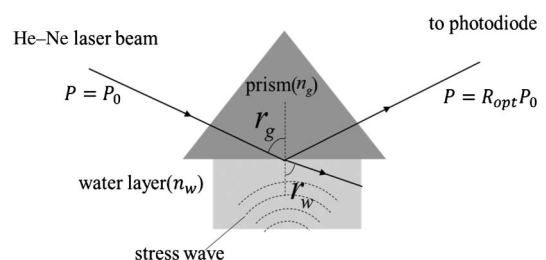
where  $R_s$  and  $R_p$  represent the optical intensity reflectivity of a  $p$ -wave and an  $s$ -wave, respectively, and the reflectivity for unpolarized light is  $R_{\text{opt}} = (R_s + R_p)/2$ ;  $r_g$  and  $r_w$  are angles between the incident beam and the refraction beam with the normal of the interface, respectively. The angles are related according to Snell’s law:

$$n_g \sin r_g = n_w \sin r_w, \quad (2)$$

where  $n_g$  and  $n_w$  represent the refractive indices of the glass prism and water, respectively. Although the acoustic pressure can change both  $n_g$  and  $n_w$ , the contribution to the reflectivity modulation from  $n_g$  is much smaller than that from  $n_w$ , as discussed in [15]. Therefore, only  $n_w$  close to the interface was considered in this Letter. According to the previous literature, the influence of the pressure on  $n_w$  is

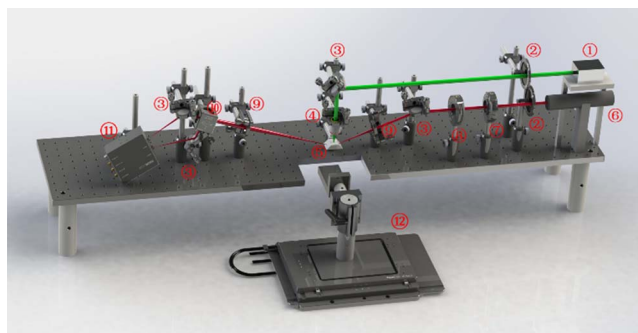
$$dn_w/dn_p = 1.35 \times 10^{-10} \text{ Pa}^{-1}. \quad (3)$$

Because of the laser instability, spontaneously environmental variation (dust, mechanical vibration, etc.), as well as the electrical noise background from the detector, the minimum pressure



**Fig. 1.** Typical ultrasonic detector based on the reflectivity of the light on the interface.

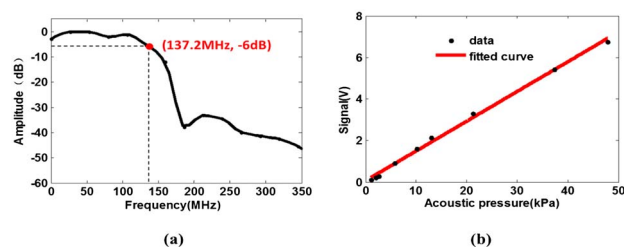




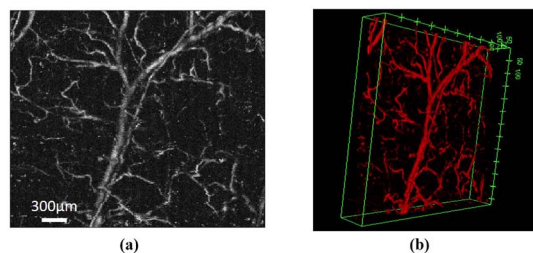
**Fig. 3.** Overall PRUD system design. ① pulsed excitation laser, ② neutral-density filter, ③ mirror, ④ objective lens, ⑤ trapezoid prism, ⑥ continuous-wave laser, ⑦ polarizer, ⑧ quarter-wave plate, ⑨ lens, ⑩ polarization beam splitter, ⑪ balanced detector, and ⑫ electrical displacement platform.

Next, we performed a photoacoustic microscopy *in vivo* imaging on this PRUD setup. Owing to the transparent prism, we can easily shine the excitation light from the top to perform the reflection mode imaging, where the target was put under the prism. This epi-illumination and detection configuration is crucial for *in vivo* study. We used the same laser as the bandwidth test, but slightly focused it through an objective lens (NA = 0.1). The lateral resolution was determined by the optical focusing, which was measured to be about 14  $\mu\text{m}$  by imaging a 10  $\mu\text{m}$  tungsten wire. We used a Kunming mouse for our *in vivo* experiments. The mouse was first anesthetized by an intraperitoneal injection of chloral hydrate solution at 0.1 mg/g; then we fixed it on the electrical translational stage (MS-2000, Applied Scientific Instrumentation) and raster scanned the ear (about 200  $\mu\text{m}$  thick) with 600 nJ/pulse and 6  $\mu\text{m}$  step size without data averaging. The animal's body temperature was maintained with an infrared lamp. All research complied with protocols approved by the Institutional Animal Care and the Use Committee of Peking University. The mouse recovered to its normal behavior condition after the experiments, and there was no observable damage to its ear. Figure 5 shows the maximum amplitude projected image and its 3D reconstruction of the mouse ear. A 3D animation visualization is also available online (see Visualization 1). The image acquisition time was 15 min.

In summary, we developed a broadband and sensitive ultrasonic detector, PRUD, which relied on the polarization-dependent optical reflection. Unlike works in previous literatures, the PRUD detects the intensity difference between the two polarization states by using a balanced detector. The NEP is substantially improved from the scale of 1 bar to 1 kPa. Compared with traditional photoacoustic microscopy that uses a piezo-electric-material-based transducer, the PRUD has a much



**Fig. 4.** Characterization of the PRUD. (a) Frequency response and (b) linearity characterization.



**Fig. 5.** PA image of a mouse ear using PRUD. (a) Maximum amplitude projected image and (b) screenshot of the 3D visualization (see Visualization 1).

broader bandwidth of over 100 MHz with a flat response, and the effective size of the PRUD detector is also much smaller. Moreover, the transparent tapered prism without any coating is very convenient for reflection mode photoacoustic imaging. We demonstrated its superior performance by an *in vivo* PAM imaging of a mouse ear. The NEP and bandwidth of this system are primarily limited by the electrical noise and bandwidth of the balanced detector, which still can be improved in future work. Compared with other current pure-optical ultrasonic detectors, we believe the PRUD is one of the simplest one. Thus, this PRUD would have a great potential to be used in both ultrasound and photoacoustic imaging.

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## REFERENCES

1. S. Zackrisson, S. M. W. Y. van de Ven, and S. S. Gambhir, *Cancer Res.* **74**, 979 (2014).
2. P. Beard, *Interface Focus* **1**, 602 (2011).
3. C. Li and L. V. Wang, *Phys. Med. Biol.* **54**, R97 (2009).
4. L. V. Wang and S. Hu, *Science* **335**, 1458 (2012).
5. G. Paltauf, R. Nuster, M. Haltmeier, and P. Burgholzer, *Appl. Opt.* **46**, 3352 (2007).
6. S. A. Carp, A. Guerra Iii, and S. Q. Duque, Jr., and V. Venugopalan, *Appl. Phys. Lett.* **85**, 5772 (2004).
7. J. Horstmann, H. Spahr, C. Bui, M. Munter, and R. Brinkmann, *Phys. Med. Biol.* **60**, 4045 (2015).
8. E. Zhang, J. Laufer, and P. Beard, *Appl. Opt.* **47**, 561 (2008).
9. Z. Xie, S.-L. Chen, T. Ling, L. J. Guo, P. L. Carson, and X. Wang, *Opt. Express* **19**, 9027 (2011).
10. B. Dong, H. Li, Z. Zhang, K. Zhang, S. Chen, C. Sun, and H. F. Zhang, *Optica* **2**, 169 (2015).
11. Y. Wang, C. Li, and R. K. Wang, *Opt. Lett.* **36**, 3975 (2011).
12. T. X. Wang, R. Cao, B. Ning, A. J. Dixon, J. A. Hossack, A. L. Klibanov, Q. F. Zhou, A. Wang, and S. Hu, *Appl. Phys. Lett.* **107**, 153702 (2015).
13. G. Paltauf, H. Schmidt-Kloiber, and H. Guss, *Appl. Phys. Lett.* **69**, 1526 (1996).
14. J. Niewisch, *Sens. Actuators A* **25**, 213 (1990).
15. G. Paltauf and H. Schmidt-Kloiber, *J. Appl. Phys.* **82**, 1525 (1997).
16. F. Xing, G.-X. Meng, Q. Zhang, L.-T. Pan, P. Wang, Z.-B. Liu, W.-S. Jiang, Y. S. Chen, and J.-G. Tian, *Nano Lett.* **14**, 3563 (2014).