Lab Journal 01.04.2023

I. Exploring the dataset

Our dataset is downloaded from here (GitHub repo of the original paper).

```
data = pd.read_csv("ACE2_train_data.csv")
```

(for now we work from the same directory we download our data in).

What is the dataframe we have downloaded?

In the <u>paper</u>, two characteristics of mutated <u>RBD</u>s of <u>COVID-19</u> were predicted: affinity to the human <u>ACE2</u> receptor (binary: binding/non-binding, see <u>here</u> and later <u>here</u>) and antibody escape (for 4 different antibodies). <u>ACE2_train_data.csv</u> should contain the training binding information.

First we inspect the data in order to understand its structure and, insbesondere, technical details like the data delimiter. For that last task specifically we use the <u>csv</u> module of the <u>standard library</u>.

```
import csv

# read in the first ten lines of the file
with open('ACE2_train_data.csv', 'r') as f:
    first_lines = [f.readline() for _ in range(10)]

# use csv.Sniffer to determine the delimiter
dialect = csv.Sniffer().sniff('\n'.join(first_lines))

print(dialect.delimiter)
```

from which we get

```
·
>>,
```

Hence, when constructing our <u>Dataset</u> class, we need to remember to use ',' as a delimiter when reading the dataframe. Next, let's explore the dataset with the help of the standard <u>Pandas</u> instruments: we begin with <u>data.info</u>

```
In [41]: data.info
Out[41]: <bound method DataFrame.info of</pre>
                                                    Unnamed: 0
                                                                               junction_aa consensus_count Label Distance
                      287261 KNAGFNCYNPLETYGFWRTGGVDW
467439 KNEQFNCYGPINAYGFQRTGGEDW
                                                                                           10
                      414422 KNQKFNCYVPLFHYGFWPTVGVGF
          3
                      103144 KNQGFNCYNPLVNYGFYRTNGRSF
                      478954 KNRGFNCYKPLPGYGFQRTDGINW
         4
                                                                                 0
                                                                                            9
          406881
                       16530 KNKGFNCYIPIEDYGFQRTSGRSY
          406882
                       48280
                              KNEGFNCYNPITEYGFWTTSGLDW
                                                                                           10
          406883
                      420449 KNGKENCYHPTVRYGEHPTVGRGY
                      173734
                               KNGQFNCYIPIAGYGFLPTLGVSY
          406885
                      554432
                              KNRGFNCYTPIFKYGFFTTWGRNY
         [406886 rows x 5 columns]>
```

Hence, the name of the columns are [no name], junction_aa, consensus_count, Label, Distance.

Out[51]:

	Unnamed: 0	consensus_count	Label	Distance
count	406886.000000	406886.000000	406886.000000	406886.000000
mean	323195.466440	1.443908	0.500027	9.017398
std	189813.180032	1.423285	0.500001	1.167593
min	0.000000	1.000000	0.000000	1.000000
25%	154232.250000	1.000000	0.000000	8.000000
50%	325632.500000	1.000000	1.000000	9.000000
75%	483459.500000	2.000000	1.000000	10.000000
max	663081.000000	310.000000	1.000000	12.000000

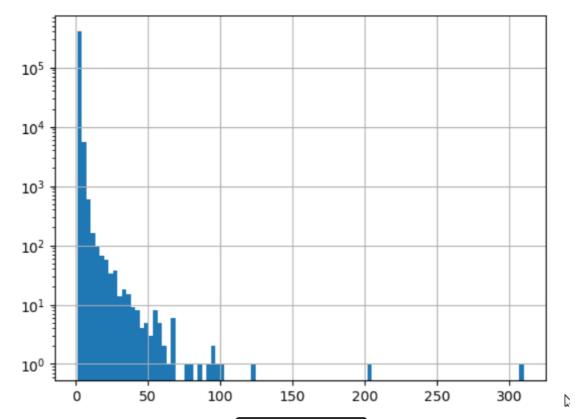
What are their "physical" meanings? The unnamed column is, apparently, just an ID. Distance appears to be the <u>Hamming distance</u> from the reference RBD. The only binary characteristic is Label)

so it should be the binding/non-binding.

junction_aa is the <u>amino acid sequence</u>. consensus_count poses the most questions.

As the following histogram indicates (note the logarithmic scale),

```
In [56]: data.consensus_count.hist(bins=100, log=True)
Out[56]: <AxesSubplot: >
```



the vast majority of data has the value of consensus count equal to 1-10, less then 1% has more, but the maximal value is 300. What exactly is consensus_count, is, unfortunately, not immediately clear from the text of the article. We ignore this parameter for now.

Let's try to build a DataLoader.

This does not work, because we need a vector representation for the junction_aa parameter. Our simplest option is to use one-hot encoding. However, a better way is to use a <u>word embedding</u> method, or, even better, the <u>Doc2Vec</u>. For that, we use the <u>gensim</u> library.

We use the AA-sequences from the train dataset to train a word2vec embedding model:

```
from gensim.models import Word2Vec
vec_size = 10
win_size = 2
epochs = 100

sentences = data.iloc[:, 1].apply(list)

model = Word2Vec(sentences, vector_size=vec_size, window=win_size,
min_count=1)
model.train(sentences, total_examples=len(sentences), epochs=epochs)
```

and save it:

```
model.save('Word2Vec_vs10_ws2_e100.model')
```

The drawback of this method is that we only obtain embeddings for individual letters. To obtain embeddings for whole sentences, we have no better method than to just sum all these vectors and normalize, thus forgetting all structural information. A better option seems to be Doc2Vec, that provides vector embeddings for whole sentences (but ultimately, I suppose, it will end up in a <u>CNN</u> embedding).

```
from gensim.models import Doc2Vec
from gensim.models.doc2vec import TaggedDocument
min_count = 1
docs = data.iloc[:, 1]

documents = [TaggedDocument(words=list(data.iloc[i, 1]), tags=[str(i)])
for i in range(len(data))]

model = Doc2Vec(documents, vector_size=10, window=2, min_count=1, workers=4)
model.save('doc2vec_vs10_ws2_mc1.model')
```

The next steps:

- 1. Create a one-hot embedding for reference
- 2. Create a CNN encoding
- 3. Train a CNN/RNN based on this embedding. Compare the results for different encodings and choose the best. Document the time of work and quality metrics

on ACE2 dataset.

4. Use these embeddings and dataloaders to train a CNN/RNN on one of the four antibody escape problems.

TBR: Word2Vec Tutorial:

https://www.kaggle.com/code/pierremegret/gensim-word2vec-tutorial