Changing diel growth symmetries and light-capture in PhycoCyanin and PhycoErythrin-rich picocyanobacteria, across photic regimes and growth phases

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# Abstract

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Picocyanobacteria are the most abundant phytoplankters in aquatic ecosystems, and arecrucial to the optical properties of ocean water, influencing its color and transparency. We grew two PhycoCyanin(PC)-rich and two PhycoErythrin(PE)-rich strains of *Synechococcus*, under a matrix of 4 photoperiods and 6 light levels. Using high frequency measurements, we found the strains were generally opportunistic in exploiting higher light diel light doses to achieve faster growth rates, although some strains suffered photoinhibition of growth under 900 µmol photons m−2s−1 and 24 h photoperiod. Using whole-cell absorbance spectra we showed that the PE-rich strains always had a higher Photosynthetically Usable Radiation (PUR)/Photosynthetically Active Radiation (PAR) ratio than did the PC-rich strains. In general, the PUR/PAR ratio decreased with increasing light. We observed an increase in cell-specific pigment content during initial growth, followed by a sharp decrease as cells transitioned to pre-stationary phase. Our results show the PE-rich strains are stronger light harvesting competitors, but the PC-rich strains may have lower N-quotients for their light capture system. These differences may help explain differential seasonal prevalence of PE-rich and PC-rich picocyanobacteria, in terms of costs of exploitation of different photic regimes.

# Introduction

The photic regime, comprised of light level (PAR), duration (photoperiod), and spectral quality, is a pivotal influence on the growth and productivity of phytoplankton within aquatic ecosystems. In polar regions, characterized by prolonged periods of wintertime darkness and continuous daylight during summer, phytoplankton encounter unique challenges. Light is the primary limiting factor for biomass production in winter, suppressing phytoplankton growth and metabolic activity, whereas the extended daylight in summer boosts photosynthetic activity [1]. In temperate regions, seasonal variation in light-limitation is less pronounced, but phytoplankton are still influenced by daily and seasonal fluctuations. There is a clear contrast between more favorable conditions for phytoplankton growth in spring and summer, compared to fall and winter [2,3]. In the tropics, daylight remains nearly constant throughout the year [4], and phytoplankton productivity is rather controlled by nutrients resupply into the euphotic zone [5,6] and zooplankton grazing [7].

Cyanobacteria growth undergoes distinct phases, including lag phase, exponential growth phase, stationary phase, and death phase [8]. During the lag phase, cyanobacteria acclimate to the environment and prepare for active growth by synthesizing essential cellular components. The exponential growth phase is marked by rapid cell division and biomass accumulation, fueled by optimal environmental conditions and nutrient availability. As nutrient levels become limited, algae enter the stationary phase, characterized by a balance between cell division and death, leading to a plateau in population growth. The death phase occurs when resources are depleted, and the algae experience cell death and decomposition, contributing to nutrient recycling in aquatic ecosystems [8]. Cell death may also be associated with the release of toxins into the environment. Understanding the temporal progression of growth phases is essential for predicting cyanobacterial activity and their impact on ecosystem dynamics over time.

*Synechococcus*, a diverse genus of picocyanobacteria, exhibits a nearly ubiquitous distribution spanning diverse geographical regions [9], while demonstrating a remarkable range of adaptations to environmental conditions. *Synechococcus*’ capacity to thrive across diverse marine and freshwater habitats positions it as a pivotal agent in energy and nutrient transfer within food webs, and serves as a link connecting the microbial loop with higher trophic levels, offering direct sustenance to grazers, including zooplankton and small fish [10]. *Synechococcus*, as one of the two dominant picocyanobacterial genera in oceanic waters, also significantly affects light attenuation and availability for other photosynthetic organisms, and influences the ocean colour, allowing for satellite detection of *Synechococcus* rich communities [11,12]. General relations among optical absorption spectra and pigment compositions have been used to determine diagnostic pigment indices of major phytoplankton functional types [13–15]. Modeling suggests that *Synechococcus* abundance will rise due to climate warming [9]. The projected changes may vary geographically and may include shifts in the spatial distribution of the main picocyanobacteria, as well as changes in the proportions among the *Synechococcus* sp. lineages [16]. However, knowledge about the impact of these environmental changes on the occurrence and ecophysiology of various picocyanobacterial phenotypes is not sufficiently known.

*Synechococcus* exhibits significant phenotypic diversity across many lineages, encompassing strains rich in PhycoErythrin (PE-rich) or PhycoCyanin (PC-rich) [17,18]. These phycobilin pigment-proteins are pivotal for light absorption during photosynthesis and confer distinctive colours to the picocyanobacteria. The disparate light preferences between PC-rich and PE-rich *Synechococcus* sp. strains influence their ecological niches. PC-rich strains thrive in environments with elevated light levels, such as surface waters and coastal regions, where blue light predominates. PE-rich strains exhibit adaptation to low-light conditions, primarily inhabiting the deeper layers of the water column where green light prevails. These differences result in PC-rich and PE-rich *Synechococcus* sp. strains predominantly occupying complementary habitats [16,17,19].

Photic regimes and growth phases of PC-rich and PE-rich *Synechococcus* sp. may drive spatial and temporal variability of *Synechococcus* biomass and community lineage composition within aquatic environments, relating to varying metabolic costs between physiological strategies. The aim of this research was to determine whether photic regimes and growth phases affect diel growth symmetries and light-capture in PC-rich and PE-rich *Synechococcus* sp.

# Material and Methods

## Culture condition and experimental setup

Two non-axenic PhycoCyanin(PC)-rich (CCBA\_056 or CCBA\_077) and two PhycoErythrin(PE)-rich (CCBA\_048 or CCBA\_127) strains of *Synechococcus* were obtained from Culture Collection of Baltic Algae (CCBA; <https://ccba.ug.edu.pl/pages/en/home.php>). Pre-cultures of picocyanobacteria strains were kept in Tissue Culture Flasks (VWR International, Cat. No. 10062-872, PA, USA) and were transferred to fresh f/2 media [20] at salinity of 8 PSU every two weeks, under a photoperiod of 12 h and Photosynthetically Active Radiation (PAR) of 10 µmol photons m−2s−1 supplied from cool white fluorescent tubes, at 22℃.

Cultures of each strain were grown in 8 x 80 mL round bottom glass tubes in a Multi-Cultivator MC 1000-OD (Photon Systems Instruments, Drásov, Czech Republic). Each culture tube contained 75 mL of f/2 medium inoculated with and 5 mL of growing pre-culture, to achieve exponential growth from the beginning of the experiment, with little to no lag phase upon inoculation.

Cultures grew at 22℃, with photoperiods of 8, 12, 16, or 24 h, with peak Photosynthetically Active Radiation (PAR) of 30, 90, 180, 300, 600, or 900 µmol photons m−2s−1 supplied from white LED lamps independently to each culture tube. To reflect the natural movement of the sun, the photoperiods of 8-16 h were applied in the shape of a sine wave, while the 24-hour photoperiod was applied in a square shape. Since the area under a sine curve is 1/2 the area under a square of equal width, the 24 h square photoperiod cultures received 4 times the diel photon dose of the 12 h sine photoperiod cultures.

The cultures of picocyanobacteria were acclimatized for one day to the new conditions corresponding to the incubation conditions of the proper culture. Tubes contained Glass Aeration Tubes and were closed with a silicone inert stopper perforated by an aeration input tube extending to the bottom of the culture tube, and a pressure outlet tube. Aeration with a total air flow rate of around 1,100 mL min−1 distributed across 8 tubes for ~ 140 mL min−1 tube−1 ensures mixing and provides sufficient air/CO2 supply to cultures across the entire culture volume. Cultivation and monitoring functions (light, temperature, optical density, and aeration gas) of the Multi-Cultivator system was controlled via the Photobioreactor Control Software (Photon Systems Instruments, Drásov, Czech Republic).

## The growth curve and logistic growth rate analysis

Picocyanobacterial growth was monitored every 5 minutes by automatically recording OD680, OD720, and ΔOD (ΔOD = OD680-OD720) for 14 days, independently for each culture tube. The exceptions were experiments conducted with a photoperiod of 24 h and light of 900 µmol photons m−2s−1, which lasted 7 days.

Based on the obtained measurements of growth we determined exponential chlorophyll specific exponential growth rates (µ) by fitting logistic growth curves to plots of the chlorophyll *a* proxy of ΔOD vs. time for each combination of strain, photoperiod, and peak PAR. We used a modified Levenberg-Marquardt fitting algorithm [**minpack.lm?**].

## The AccLen and growth symmetry analysis

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## Determining the number of cells

The number of picocyanobacterial cells was calculated using linear regression models based on cell concentration (N mL−1) and OD at 680 nm. The OD of cultures was measured using a CLARIOstar Plus Plate Reader (BMG, Labtech, Ortenberg, Germany) and calculation of the cell number was conducted using the PAMAS S40 GO Particle counter (PAMAS Partikelmess- und Analysesysteme GmbH, Rutesheim, Germany). The linear correlations between N and OD680 for individual strains were used to estimate the number of cells based on OD measurements obtained from Multi-Cultivator system (Tab. S1 in the Supplementary materials).

## Whole-cell absorbance spectra measurements

Absorbance measurements on intact cells in suspension were conducted in OLIS CLARiTY 17 UV/Vis/NIR with integrating cavity upgrade spectrophotometer (On-Line Instrument Systems, Inc., Bogart, GA, USA) according to the method described by [21] with modifications. In an experiment, identical 8 mL solutions that contained f/2 medium, were added to both the sample and reference observation cavities of the spectrophotometer. After recording a baseline from 375 to 710 nm, 1 mL was withdrawn from the sample cavity and replaced with 1 mL of the cell suspension of tested picocyanobacteria. The pathlength corrected absorbance per cm was performed by determining the Javorfi coefficients [22] as described in the equipment manual.

## Estimating Photosynthetically Usable Radiation (PUR)

Using whole-cell absorbance spectra of *Synechococcus* sp. cultures as described above (Fig. ??) we estimated Photosynthetically Usable Radiation (PUR) according to the method proposed by [23]. Initially, we normalized the obtained whole-cell absorbances (AbsNorm440) and emission spectra of the white LED lamps (EmNorm440) to a reference wavelength of 440 nm. The PUR value, which is the ratio of the normalized sum of absorbance and emission spectra to the sum of normalized emission spectra multiplied by the intensity of the tested light (PAR) was calculated (Eq. (1)).

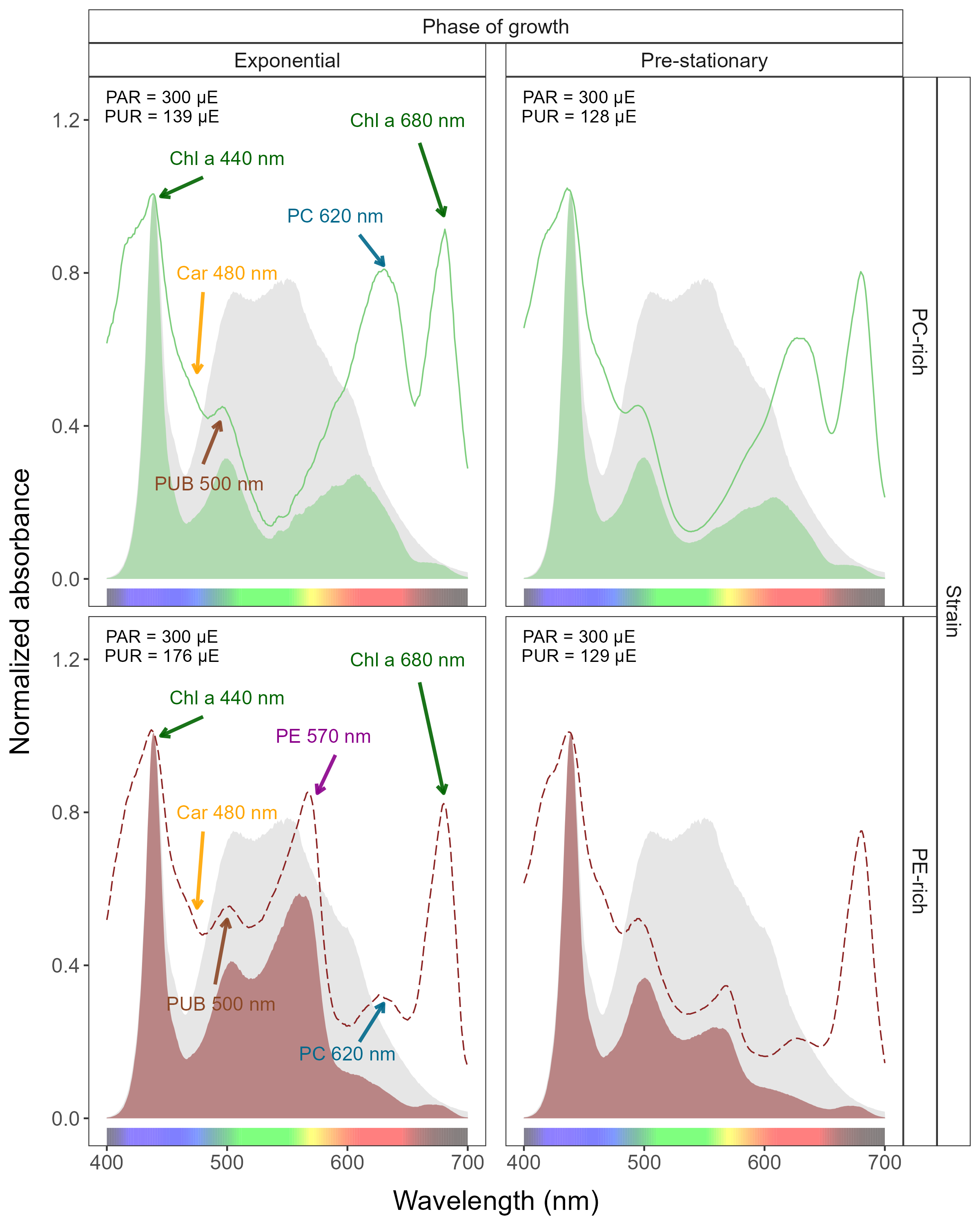


Figure 1: Example of representative whole-cell absorbance spectra of PC-rich (solid green lines) or PE-rich (dashed red lines)cultures of *Synechococcus* sp., normalized to absorbance at 440 nm, measured from the the exponential or pre-stationary phases of growth, together with emission spectra of the white LED lamps (Photosynthetically Active Radiation (PAR), normalized to emission at 440 nm (light gray area), of 300 µmol photons m−2s−1) used for culture growth. Estimated Photosynthetically Usable Radiation (PUR) is shown as a green area for the PC-rich strain and a red area for the PE-rich strain. Peaks characteristic of known pigments are labelled; Chl *a*, chlorophyll *a*; PC, phycocyanin; PEB-rich PE, phycoerythin-rich phycoerythrin; PUB-rich PE, phycourobilin-rich phycoerythrin, Car, carotenoids.

## Pigment content analysis

The pigment content: chlorophyll *a* (Chl *a*), carotenoids (Car), phycoerythrin (PE), phycocyanin (PC), and allophycocyanin (APC) in *Synechococcus* sp. cultures over time was estimated with previously determined linear correlations between pigment content obtained by extraction technique and absorbance values of individual pigment peaks (nm) obtained from the whole-cell absorbance spectra (Tab. S2 in the Supplementary materials). Total amount of phycobilin pigments (Phyco) for individual strains was obtained by adding the content of PE, PC, and APC.

Pigments extraction were performed using formula from [24] for Ch *a* and Car concentrations. PE, PC, and APC were calculated based on [25]. The extracts contained photosynthetic pigments were measured using a CLARIOstar Plus Plate Reader (BMG, Labtech, Ortenberg, Germany), at wavelengths of 480, 665, and 750 nm for Chl *a* and Car calculation and at 565, 620, 650, and 750 nm for PE, PC, and APC. The values of individual pigment peaks (nm) from the whole-cell absorbance spectra were obtained by Olis-modernized Cary 14 UV/Vis/NIR with Integrating Sphere upgrade spectrophotometer (On-Line Instrument Systems, Inc., Bogart, GA, USA). For the linear model, the following wavelengths were analyzed: 480 (Car), 565 (PE), 620 (PC), 650 (APC), and 665 (Chl *a*) nm.

## Estimating cumulative diel PAR

Based on the length and shape of the photoperiod (sine wave for photoperiod of 8-16 h; square for photoperiod of 24 h) and the given light level, we estimated the value of the cumulative diel photon dose. For a photoperiod arranged in the shape of a sine wave we using Eq. (2). For a continuous 24 h photoperiod we used Eq. (3).

## Changes effective absorption cross section of PSII and PSII flux per unit volume

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Kolber, Z., Klimov, D., Ananyev, G., Rascher, U., Berry, J., & Osmond, B. (2005). Measuring photosynthetic parameters at a distance: laser induced fluorescence transient (LIFT) method for remote measurements of photosynthesis in terrestrial vegetation. Photosynthesis research, 84, 121-129.

Oxborough, K., Moore, C. M., Suggett, D. J., Lawson, T., Chan, H. G., & Geider, R. J. (2012). Direct estimation of functional PSII reaction center concentration and PSII electron flux on a volume basis: a new approach to the analysis of Fast Repetition Rate fluorometry (FRRf) data. Limnology and Oceanography: Methods, 10(3), 142-154.

Keller, B., Vass, I., Matsubara, S., Paul, K., Jedmowski, C., Pieruschka, R., … & Muller, O. (2019). Maximum fluorescence and electron transport kinetics determined by light-induced fluorescence transients (LIFT) for photosynthesis phenotyping. Photosynthesis Research, 140, 221-233.

Calculation of the absorption cross section of PSII photochemistry (3C3PSII’; nm2 quanta-1) under ambient light through an iterative curve fit to the saturation phase of an Fast Repetition Rate fluorometry (FRRf) single turnover (ST) measurement of tested picocyanobacteria were obtained using SoliSense laser-induced fluorescence transient (LIFT) fluorometer equipped with a prototype temperature control unit (LIFT-REM, Soliense Inc., New York, USA). PSII flux per unit volume (JVPSII; xxxx add proper unit) was calculated according to the method proposed by [Campbell?; Oxborough et al., 2012].

PC-rich and PE-rich picocyanobacteria were measured under diel peak PAR growth light under a blue LED (Ex 445 nm) and orange (Ex 590 nm) excitation. Excitation protocols were used to manipulate the level of photosynthetic activity and chlorophyll fluorescence (ChlF). Flash Power for blue excitation was 60000 and for orange excitation was 14000 µmol photons m−2s−1. The intensity of the blue and orange LIFT LED in DC mode and excitation power were calibrated using a quantum sensor (LI-250, LI-COR, Inc.). Data were collected during an rapid light curve (RLC) sequence during which light intensity was increased from 0 to 320 µmol photons m−2s−1 and then decreased from 320 to 0 µmol photons m−2s−1 with a 1s pause in darkness between measurements. Acquisitions were made at 10-s intervals. [Campbell and Kolber?; Kolber et al., 2005; Oxborough et al., 2012].

## Statistical analysis

The statistical differences for the growth rate, pigments content, pigments ratio or PUR/PAR ratio obtained at different light levels and photoperiods were tested using the two-way ANOVA without replication [26,27]. In the technique of ANOVA without replication, the sample observation size is one, meaning that there was only a single observation for each combination of nominal variables. In this case, the analysis was performed using the means of both the variables as well as the total mean of considering every observation as a single cluster. If the ANOVA revealed significant differences among groups, the post-hoc Tukey HDS test was used for pairwise analysis. Levels of significant difference were: \* *p* < 0.05; \*\* *p* < 0.01; \*\*\* *p* < 0.001.

The absolute growth rate (AGR), the total daily growth (TDG), and related metrics were computed after applying a spline smoothing filter to the initial OD680 curve. The trends of change in the growth symmetry (GS) were tested using the Mann-Kendall test [28] on the ±5 photoperiods to avoid noise during the first few days of the experiment. The sigmoid curve was also fitted to compare with the maximum absolute growth increment (maxAGI) position, and define the transition point between exponential growth and pre-stationary phase.

All analysis of obtained results was conducted using R version 4.3.0 [29] running under RStudio [30]. To determine significant differences in studied experiments the “stats” v. 3.6.2 R standard packages were used. This package provides basic statistical functions, including the *aov()* function for ANOVA, *TukeyHSD()* function for Tukey’s test, and the *SSlogis()* function used to fit the sigmoid curve. Manuscript was prepared as Rmarkdown document [31]. Figures were plotted using “ggplot” [32] R package.

# Results

## Changes in logistic growth rates

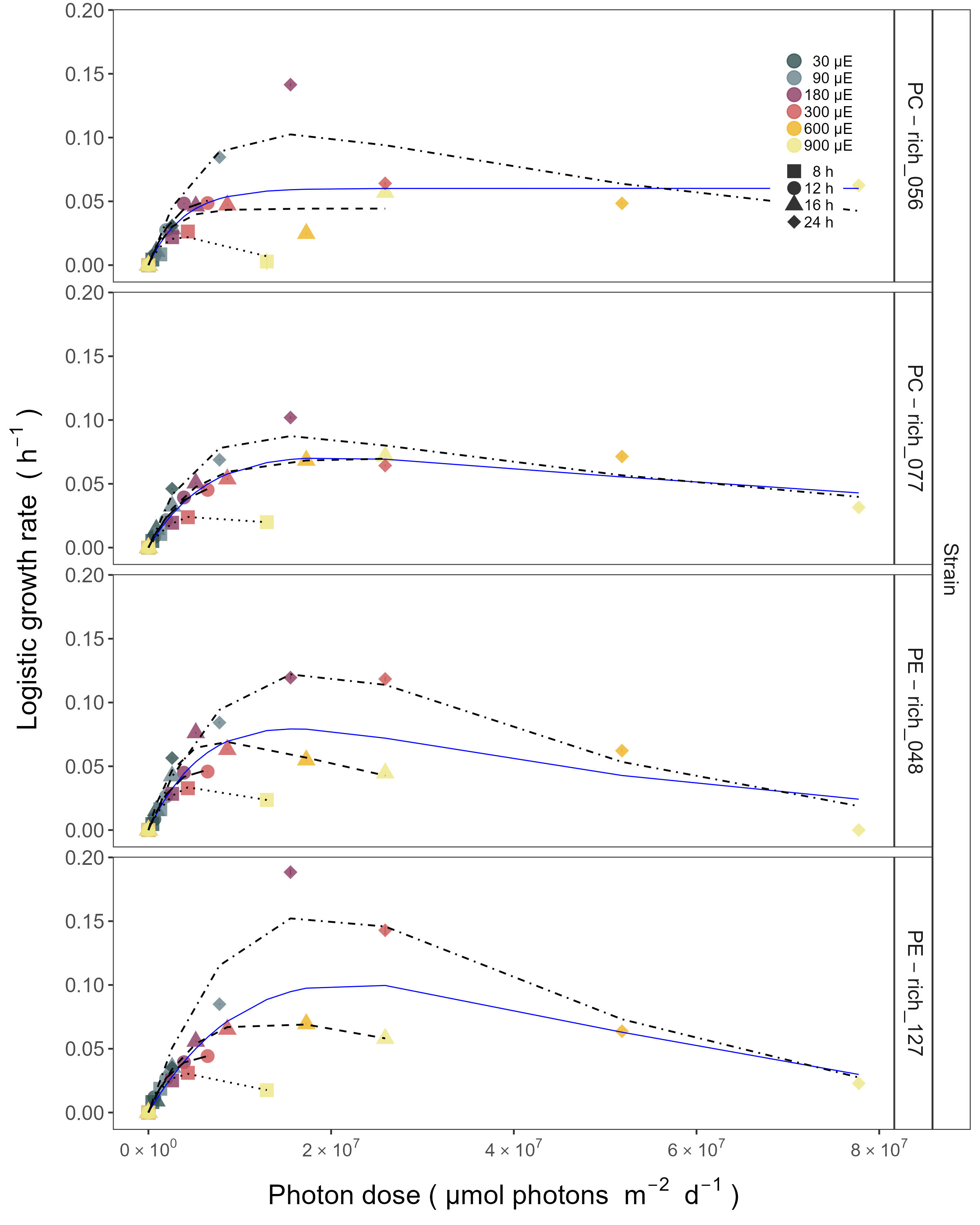


Figure 2: Chlorophyll specific exponential growth rates, estimated from logistic fits of chlorophyll proxy OD680-OD720 vs. elapsed time, for two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. grown at 30, 90, 180, 300, 600, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, 16, or 24 h. Growth rates (+/- SE from logistic model; SE falls within symbol sizes) are plotted vs. cumulative diel µmol photons m−2d−1, and pool fit with curve (Harrison and Platt, 1986) was shown as solid blue line. Separate lines fit for growth under 8 (dotted line), 12 (longdash line), 16 (dashed line), or 24 (twodash line) h of photoperiod when they were significantly different (ANOVA, *p* < 0.05) from the pool fit.

## Changes of diel growth symmetry

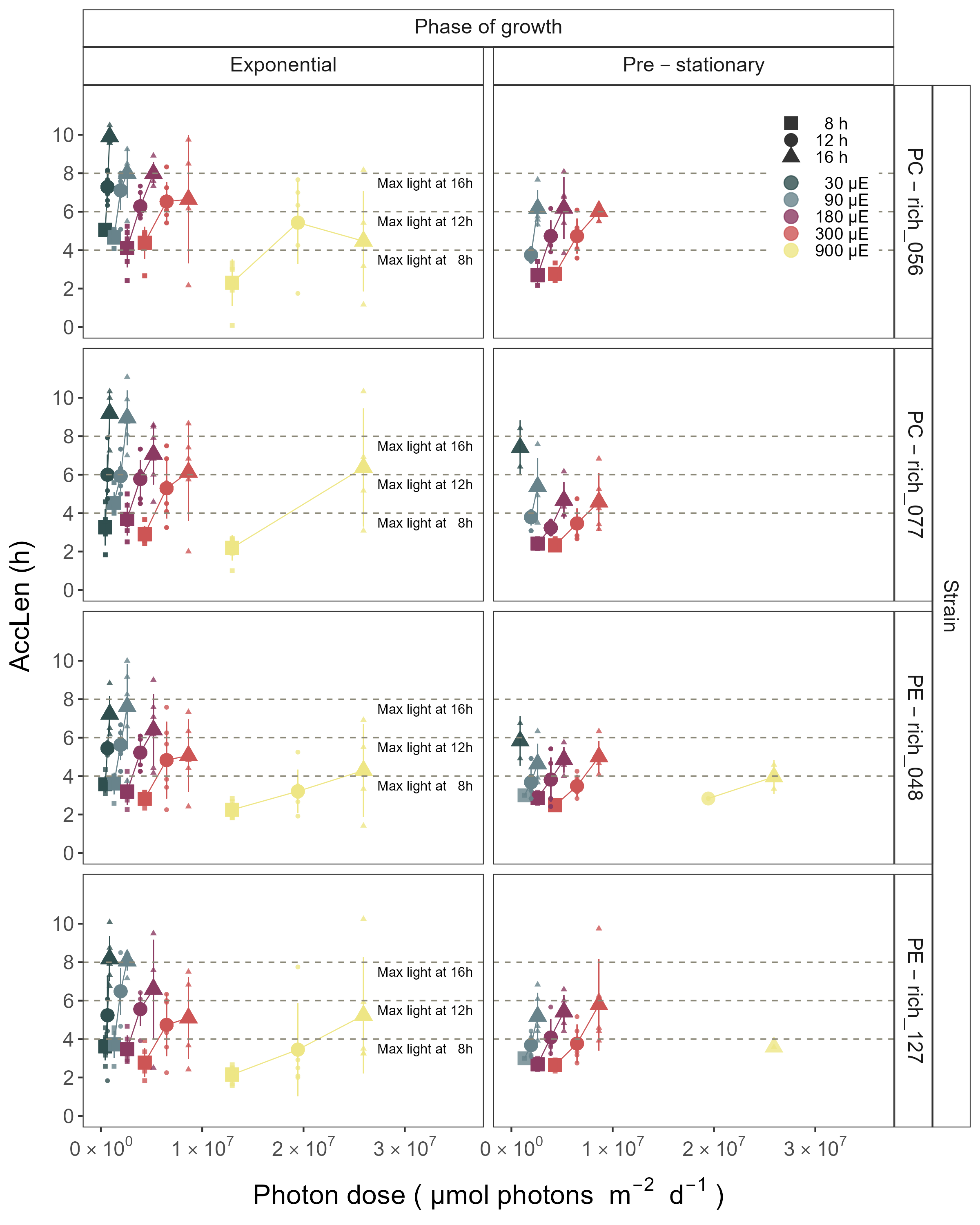


Figure 3: Hours of photoperiod to reach maximum hourly growth increment (AccLen), for two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. grown at 30, 90, 180, 300, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, or 16 h. The horizontal lines indicate the time (h) to reach the maximum light for a given photoperiod; 4 h for the 8 h photoperiod; 6 h for the 12 h photoperiod; or 8 h for the 16 h photoperiod. Figure represents all data (small symbols) and means (big symbols) for n = 0-5 days from exponential phase, prior to reaching maximum absolute hourly growth; or from pre-stationary growth phase, after reaching maximum absolute hourly growth.

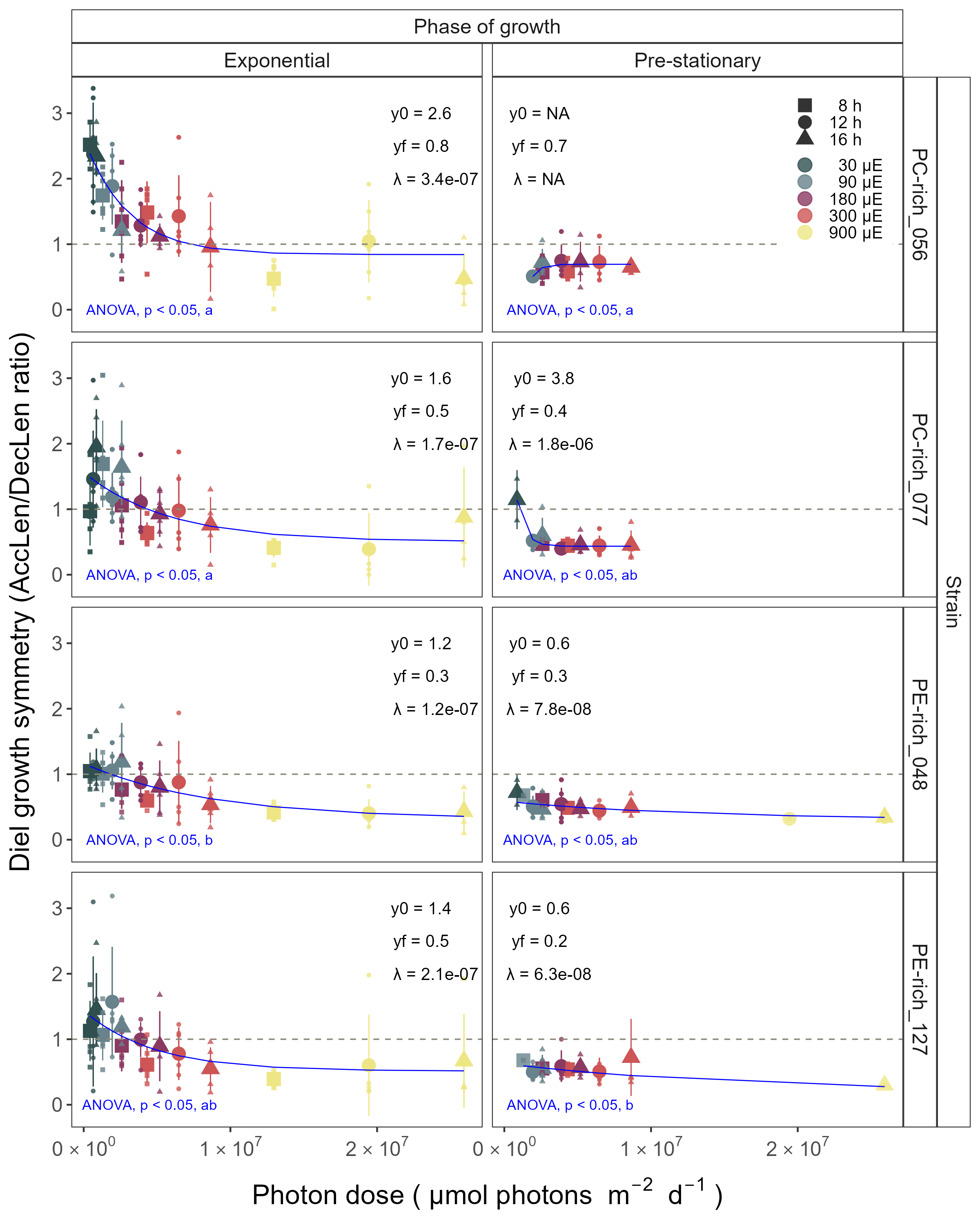


Figure 4: Index of diel growth symmetry (AccLen/DecLen), for two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. grown at 30, 90, 180, 300, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, or 16 h. Figure represents all data (small symbols) and means (big symbols) for n = 0-5 days from exponential phase, prior to reaching maximum absolute hourly growth; or from pre-stationary growth phase, after reaching maximum absolute hourly growth. Blue solid line shows single phase exponential decay fit. Single phase exponential decay parameters were also presented. Different lowercase letters indicate significant differences between the fit models for strains across diel growth symmetry and cumulative diel µmol photons m−2d−1 for a given phase of growth (ANOVA; *p* < 0.05).

## Changes of TDG

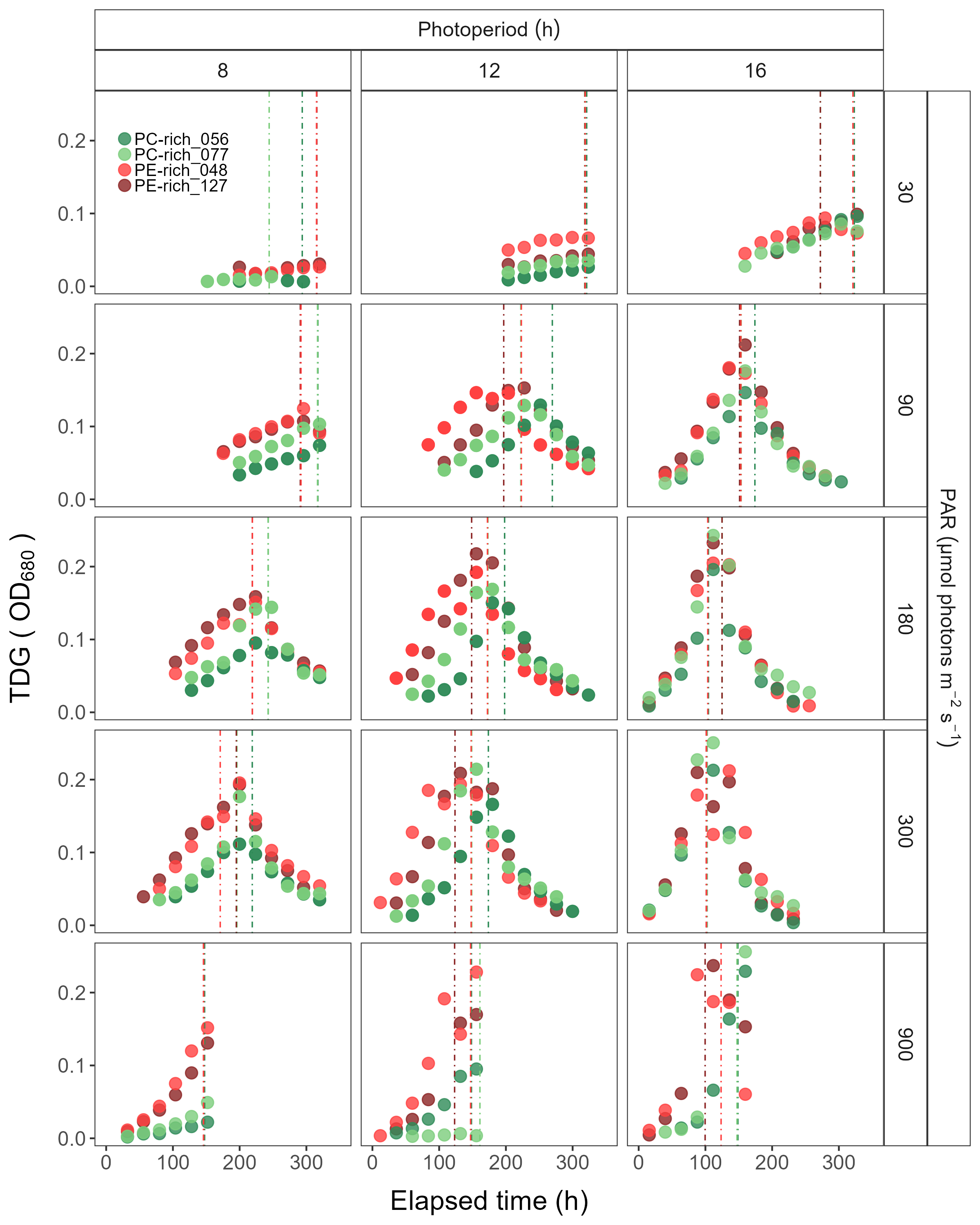


Figure 5: Changes of TDG (tracked as daily change in OD680 increment) of two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. grown at 30, 90, 180, 300, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, or 16 h. The vertical lines represent the time when the strains reached their maximum absolute hourly growth (tMaxAG).

## Changes in total Phyco/Chl a ratio and PUR/PAR ratio

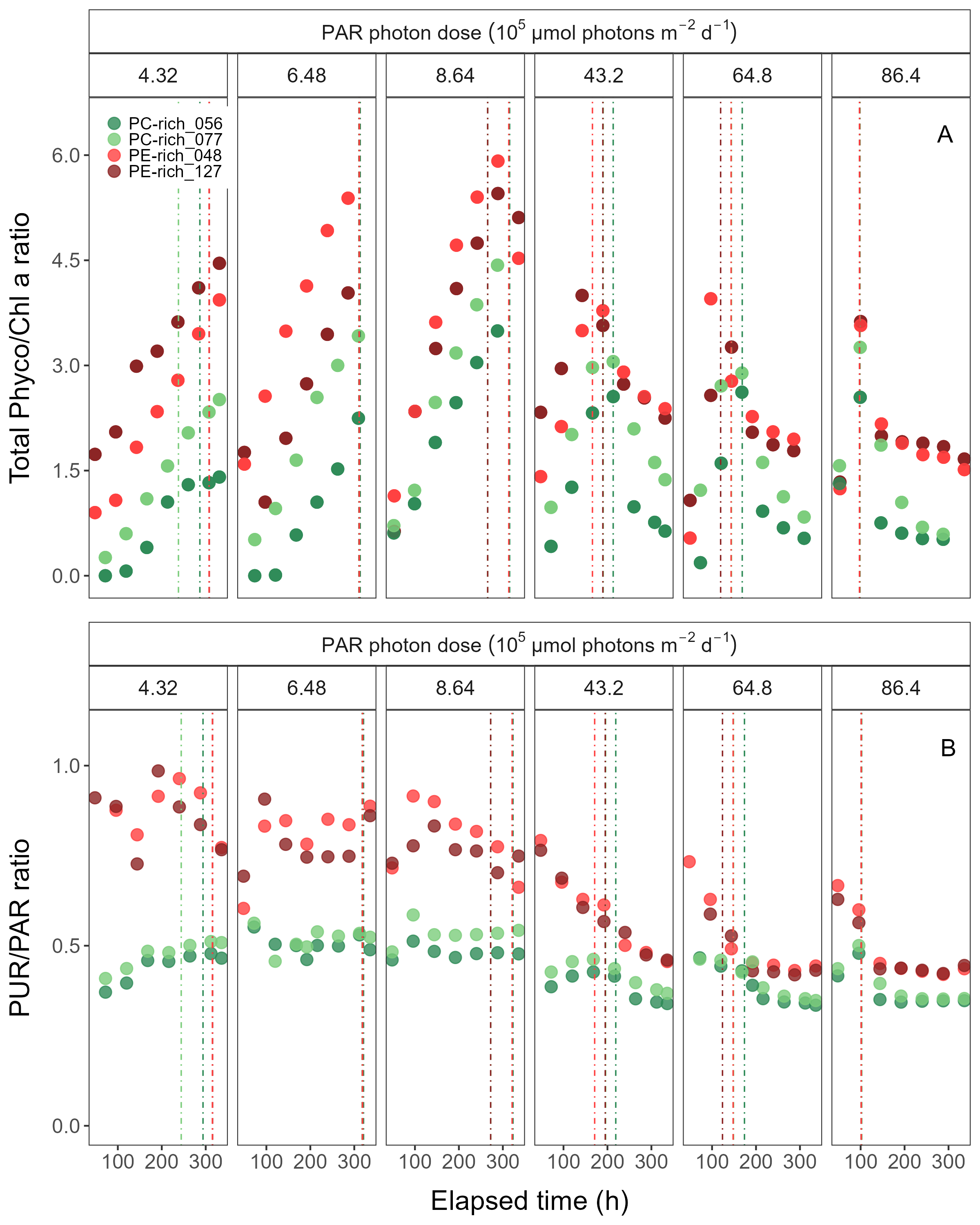


Figure 6: Changes of total Phyco/Chl *a* ratio and PUR/PAR ratio of two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. at selected cumulative diel µmol photons m−2d−1 over time (h). The vertical lines represent the time when the strains reached their maximum absolute hourly growth (tMaxAG).

## Changes effective absorption cross section of PSII

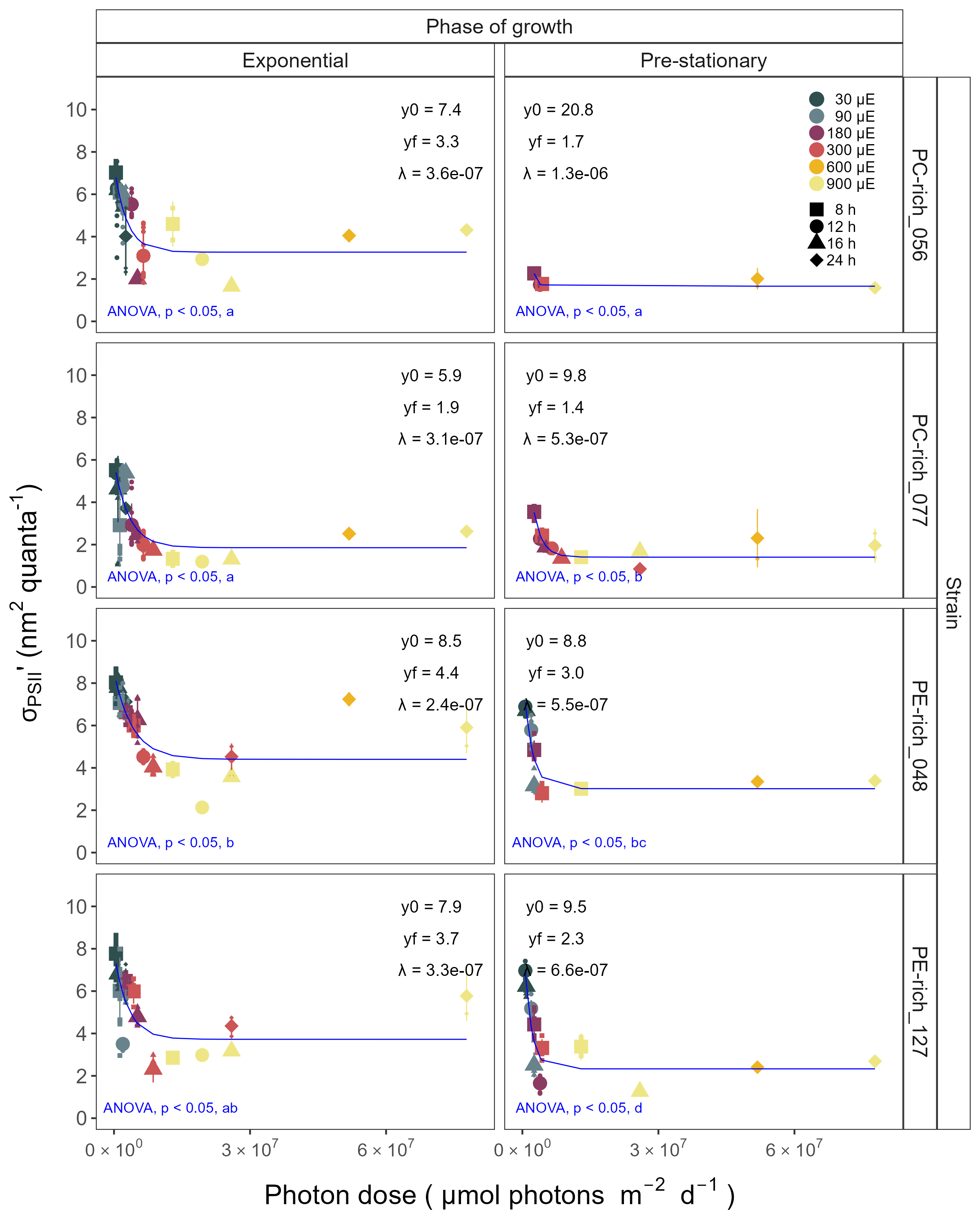


Figure 7: Effective absorption cross section of PSII (σPSII‘; nm2 quanta-1) measured under diel peak PAR growth light under Ex590 nm (orange) excitation in two PhycoCyanin(PC)-rich cultures (056, 077) and two PhycoErythrin(PE)-rich cultures (048, 127) (Culture Collection of Baltic Algae) of *Synechococcus* sp. grown at 30, 90, 180, 300, 600, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, 16, or 24 h. Blue solid line shows single phase exponential decay fit. Single phase exponential decay parameters were also presented. Different lowercase letters indicate significant differences between the fit models for strains across σPSII’ and cumulative diel µmol photons m−2d−1 for a given phase of growth (ANOVA; *p* < 0.05).

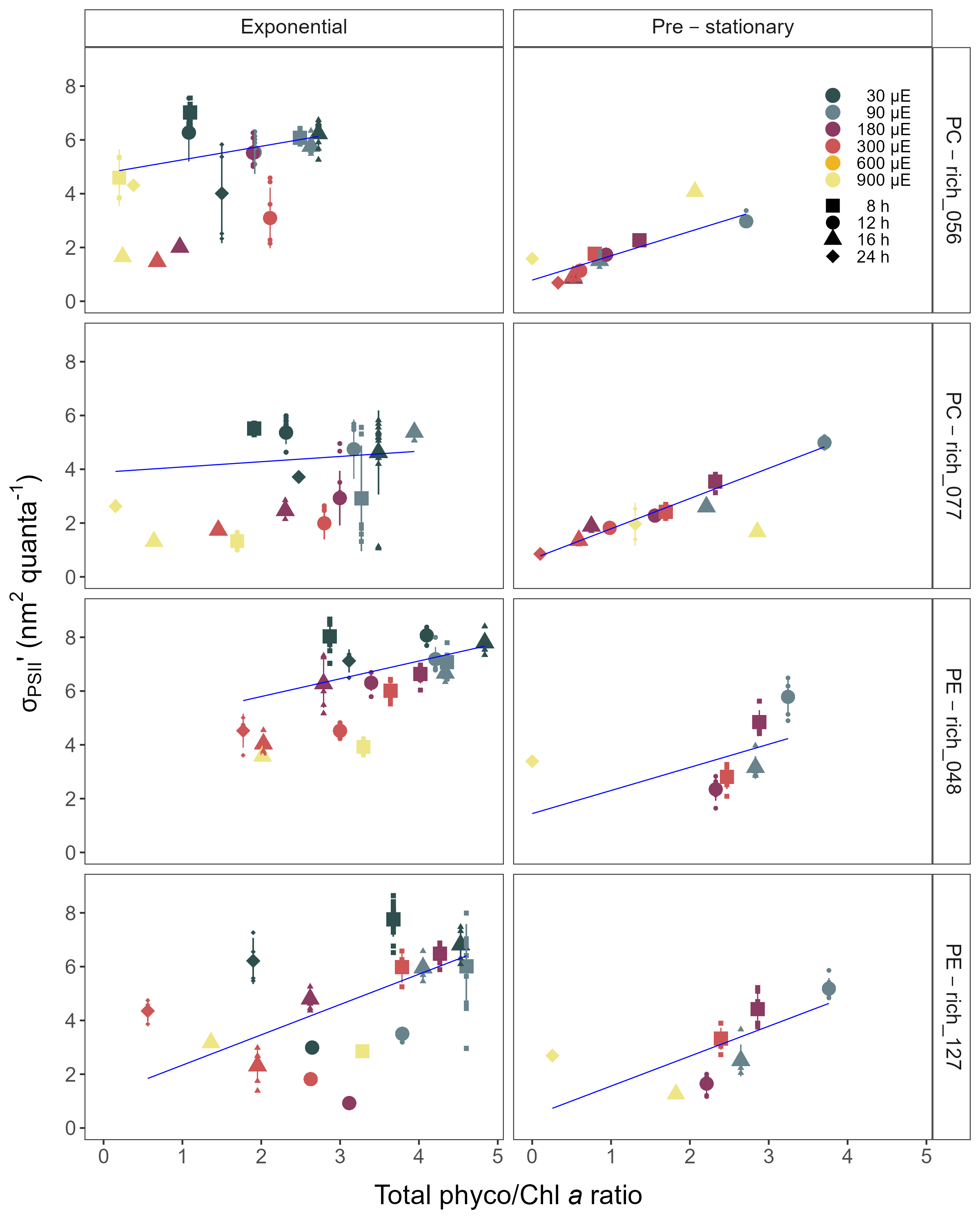


Figure 8: Changes of effective absorption cross section of PSII (σPSII’; nm2 quanta-1) measured under diel peak PAR growth light under Ex590 nm (orange) excitation in relation to the total Phyco/Chl *a* ratio of two PhycoCyanin(PC)-rich cultures (Culture Collection of Baltic Algae; 056, 077) and two PhycoErythrin(PE)-rich cultures (Culture Collection of Baltic Algae; 048, 127) of *Synechococcus* sp. grown at 30, 90, 180, 300, 600, or 900 peak diel PAR µmol photons m−2s−1; and photoperiods of 8, 12, 16, or 24 h. Blue solid line shows linnear model fit.

# Discussion

## The role of photoperiod for picocyanobacteria growth in aquatic ecosystems

Photoperiod, the duration of daily light exposure, plays an important role in influencing the growth and productivity of picocyanobacteria within aquatic ecosystems. In this work, we have shown that not only the daily dose of light, but also the length of exposure affected the picocyanobacteria growth rate. The PE-rich and PC-rich strains of *Synechococcus* sp. showed faster logistic growth rates with increasing photoperiod, including constant light conditions. Most of the strains were able to survive even under dose of light of 77,760,000 µmol photons m−2 per day. In addition, one of PC-rich strains showed the fastest growth rate in these extreme conditions.

Phytoplankton are highly sensitive to changes in photoperiod, which serves as a key environmental cue for their metabolic activities and life cycle events [2,33,34]. The duration of light exposure within a day regulates various physiological processes, including photosynthesis, growth, reproduction, and nutrient assimilation in phytoplankton. Changes in photoperiod trigger adaptive responses, shaping the temporal dynamics and community structure of phytoplankton. Understanding the impact of photoperiod on picocyanobacteria dynamics is essential for predicting their responses to environmental changes, including climate-induced alterations in day length and seasonality.

Based on our research, we confirmed that *Synechococcus* sp. can exist and even become the dominant faction of phytoplankton in all geographic zones on Earth as long as they have access to light. However, we predict that that some of PC-strains of *Synechococcus* sp. may be less numerous than PE-strains in surface waters (where the light intensity could be extremely high) when the photoperiod is quite low (autumn and winter in temperate zones and tropical water throughout the year). On the other hand, in regions with a longer photoperiod (summer in the temperate zone and summer at the poles), PC-strains may become dominant species in the surface waters. Our research has also highlighted the possibility of occurrence of both PE-rich and PC-rich *Synechococcus* sp. in conditions of continuous irradiation. Thus, it can be predicted that *Synechococcus* may become the dominant fraction of phytoplankton during the Arctic summer near the poles regions regardless of their genetic lineages and pigments composition.

## The importance of Photosynthetically Active Radiation (PAR) for picocyanobacteria growth

Light intensity is critical factor that significantly influence the growth and productivity of picocyanobacteria within aquatic ecosystems. Photosynthetically Active Radiation (PAR) refers to the spectral range of solar radiation (approximately 400-700 nm) that is capable of driving photosynthesis [23]. In our work, the PE-rich and PC-rich *Synechococcus* sp. strains showed faster logistic growth rates with increasing light, although some strains suffered photoinhibition. The *Synechococcus* sp. strains reach their plateau in the light intensity range of 180-300 µmol photons m−2s−1. Growth at 900 µmol photons m−2s−1 was also noted but not as efficient as under moderate light. The exception was one PC-rich strain, which under this condition reached the maximum growth rate.

Light intensity, a measure of the amount of PAR reaching a specific area, directly affects the physiology of picocyanobacteria [18,35,36]. Optimal light intensity levels provide the necessary energy for efficient photosynthesis, promoting phytoplankton growth, reproduction, and biomass production. The availability and distribution of PAR and light intensity in aquatic ecosystems are influenced by cloud cover, water depth, and light attenuation due to water turbidity and suspended particles [37–39]. *Synechococcus* sp., a widely studied picocyanobacterial genus, exhibits remarkable adaptability to different light intensities, particularly under white light conditions. White light encompasses the entire visible spectrum, and *Synechococcus* sp. has developed various strategies to optimize its photosynthetic efficiency across a range of light intensities. Under high-light conditions, *Synechococcus* employs photoprotective mechanisms to prevent the harmful effects of excess light energy. These include the dissipation of excess energy as heat via non-photochemical quenching (NPQ) and the regulation of antenna pigments, such as phycobilisomes, to balance light absorption and energy transfer. In contrast, under low-light conditions, *Synechococcus* sp. increases the expression of light-harvesting complexes to enhance light absorption and capture [40–42].

Numerous studies have highlighted the significance of PAR and light intensity as a key driver of phytoplankton productivity and its influence on ecosystem dynamics, biogeochemical cycling, and food web interactions [e.g., 37,38,39,43]. Our research shows that an increase in light intensity can result in the dominance of both PE-rich and PC-rich picocyanobacteria in aquatic ecosystems and confirmed the possibility of occurrence of *Synechococcus* sp. in extremely high irradiance conditions.

## The importance of Photosynthetically Usable Radiation (PUR) for picocyanobacteria growth

Photosynthetically Usable Radiation (PUR) is the fraction of radiant energy (Photosynthetically Active Radiation; PAR) of such wavelength that it can be absorbed by the cyanobacteria and algae. Thus, PUR is always smaller than PAR (PUR < PAR). PUR depends on the spectral composition of the submarine radiant energy as well as on the pigment composition [23]. In this work, we showed that the PE-rich strains always had a higher PUR/PAR ratio than the PC-rich strains. The PUR/PAR ratio decreased with increasing light in the PE-rich strains, while it initially increased under low light and short photoperiod in the PC-rich strains.

PUR plays a fundamental role in the growth and productivity of phytoplankton within aquatic ecosystems [4,44,45]. Phytoplankton, as primary producers, heavily mainly on PUR for their energy acquisition through photosynthesis. The availability of PUR directly influences the photosynthetic rates and overall metabolic activity of phytoplankton. High levels of PUR promote optimal photosynthetic efficiency, leading to enhanced growth, reproduction, and biomass accumulation. Conversely, insufficient or suboptimal PUR availability can limit the metabolic processes and growth of phytoplankton. The spatial and temporal distribution of PUR within aquatic ecosystems is influenced by various factors, including solar zenith angle, water depth, water clarity, and the presence of light-absorbing substances such as dissolved organic matter [23,45]. Understanding the dynamics and availability of PUR is crucial for comprehending the variability of picocyanobacteria communities in different aquatic environments. As we face ongoing environmental changes, including alterations in light regimes due to climate change and human activities, assessing the impact of changing PUR on picocyanobacteria communities becomes increasingly important for predicting and managing the response of aquatic ecosystems. Our results indicate that PE-rich strains of *Synechococcus* sp., due to their high content of phycoerythrin, can better use the available radiation. Therefore, their long-term dominance in the environment can be postulated, especially in places where access to light is limited.

## The changes in pigment content of picocyanobacteria

Temporal variations in cell-specific pigment content of *Synechococcus* sp. were observed during the growth phase, characterized by an initial increase followed by a sharp decrease. These trends exhibited dependency on growth, light intensity, and photoperiod, manifesting subsequent to the attainment of daily maximum absolute growth. Maximum pigment content was documented under conditions of low irradiance and extended photoperiod. Moreover, PC-rich strains had more pigments in the cell compared to PE-rich strains of *Synechococcus* sp.

Pigment dynamics are profoundly influenced by the prevailing light regimes. Primary photosynthetic pigments in *Synechococcus* sp. comprise chlorophyll *a*, responsible for light energy capture. Under low-light conditions, picocyanobacteria tend to increase their chlorophyll *a* content to enhance light absorption and maximize energy capture for photosynthesis. Conversely, high-light conditions often lead to a decrease in chlorophyll *a* content, serving as a photoprotective mechanism against excessive irradiation. In addition to chlorophyll *a*, picocyanobacteria utilize phycobilins, including phycocyanin and phycoerythrin, as accessory pigments to enhance light harvesting efficiency. Adapting to low-light environments, picocyanobacteria enhance phycobilin production to compensate for limited irradiance, thereby optimizing their photosynthetic capabilities. The chlorophyll/phycobilin ratio serves as a valuable indicator of the prevailing light conditions and the balance between chlorophyll-based and phycobilin-based light harvesting strategies. Elevated light intensities result in a decreased chlorophyll/phycobilin ratio as picocyanobacteria allocate resources towards efficient phycobilin-mediated light capture. These intricate changes in pigment composition and ratios represent vital adaptations that enable picocyanobacteria to optimize photosynthetic efficiency and thrive in dynamic light environments [46–48].

# Conclusion

Understanding the influence of light intensity and photoperiod on the dynamics of picocyanobacteria is imperative for predicting their spatial distribution across various geographic regions and their response to observed environmental changes. Our findings have substantiated that *Synechococcus* sp., irrespective of its genetic lineages and pigment composition, can thrive and even dominate the phytoplankton community worldwide when exposed to sufficient light. Furthermore, our investigations have demonstrated the survival capacity of both PE-rich and PC-rich *Synechococcus* sp. strains under conditions of exceptionally high and continuous irradiation. Consequently, it can be predicted that *Synechococcus* sp. has the potential to emerge as the prevailing phytoplankton component during the Arctic summer near polar regions. Nevertheless, our results showed the PE-rich strains are stronger light-harvesting competitors as they tend to live deeper in the water column, but the PC-rich strains may have lower N-quotients for their light capture system. Additionally, we anticipate that PC-rich strains of *Synechococcus* sp. could be less abundant than PE-rich strains in surface waters, where light intensity tends to be extremely high, especially during periods of reduced photoperiod, such as autumn and winter in temperate zones and throughout the year in tropical waters. Conversely, in regions characterized by an extended photoperiod i.e., summer in the temperate zone and summer at the poles, PC-rich strains may assume dominance in surface waters. These differences may help explain differential seasonal prevalences of *Synechococcus* sp., in terms of the costs of exploitation of different photic regimes.

# References

1. Arrigo KR. Sea ice ecosystems. Annual Review of Marine Science. 2014;6: 439–467. doi:[10.1146/annurev-marine-010213-135103](https://doi.org/10.1146/annurev-marine-010213-135103)

2. Huisman J, Arrayás M, Ebert U, Sommeijer B. How Do Sinking Phytoplankton Species Manage to Persist? The American Naturalist. 2002;159: 245–254. doi:[10.1086/338511](https://doi.org/10.1086/338511)

3. Holtrop T, Huisman J, Stomp M, Biersteker L, Aerts J, Grébert T, et al. Vibrational modes of water predict spectral niches for photosynthesis in lakes and oceans. Nature Ecology & Evolution. 2021;5: 55–66. doi:[10.1038/s41559-020-01330-x](https://doi.org/10.1038/s41559-020-01330-x)

4. Behrenfeld MJ, O’Malley RT, Siegel DA, McClain CR, Sarmiento JL, Feldman GC, et al. Climate-driven trends in contemporary ocean productivity. Nature. 2006;444: 752–755. doi:[10.1038/nature05317](https://doi.org/10.1038/nature05317)

5. Hutchins DA, Boyd PW. Marine phytoplankton and the changing ocean iron cycle. Nature Climate Change. 2016;6: 1072–1079. doi:[10.1038/nclimate3147](https://doi.org/10.1038/nclimate3147)

6. Li Q, Legendre L, Jiao N. Phytoplankton responses to nitrogen and iron limitation in the tropical and subtropical Pacific Ocean. Journal of Plankton Research. 2015;37: 306–319. doi:[10.1093/plankt/fbv008](https://doi.org/10.1093/plankt/fbv008)

7. Christaki U, Jacquet S, Dolan JR, Vaulot D, Rassoulzadegan F. Growth and grazing on Prochlorococcus and Synechococcus by two marine ciliates. Limnology and Oceanography. 1999;44: 52–61. doi:[10.4319/lo.1999.44.1.0052](https://doi.org/10.4319/lo.1999.44.1.0052)

8. Reynolds CS. The Ecology of Phytoplankton. Cambridge University Press; 2006.

9. Flombaum P, Gallegos JL, Gordillo RA, Rincón J, Zabala LL, Jiao N, et al. Present and future global distributions of the marine Cyanobacteria Prochlorococcus and Synechococcus. Proceedings of the National Academy of Sciences. 2013;110: 9824–9829. doi:[10.1073/pnas.1307701110](https://doi.org/10.1073/pnas.1307701110)

10. Li WKW. Composition of ultraphytoplankton in the central North Atlantic. Marine Ecology Progress Series. 1995;122: 1–8. Available: <https://www.jstor.org/stable/24852252>

11. Bracher A, Bouman HA, Brewin RJW, Bricaud A, Brotas V, Ciotti AM, et al. Obtaining Phytoplankton Diversity from Ocean Color: A Scientific Roadmap for Future Development. Frontiers in Marine Science. 2017;4.

12. Xi H, Losa SN, Mangin A, Soppa MA, Garnesson P, Demaria J, et al. Global retrieval of phytoplankton functional types based on empirical orthogonal functions using CMEMS GlobColour merged products and further extension to OLCI data. Remote Sensing of Environment. 2020;240: 111704. doi:[10.1016/j.rse.2020.111704](https://doi.org/10.1016/j.rse.2020.111704)

13. Vidussi F, Claustre H, Manca BB, Luchetta A, Marty J-C. Phytoplankton pigment distribution in relation to upper thermocline circulation in the eastern Mediterranean Sea during winter. Journal of Geophysical Research: Oceans. 2001;106: 19939–19956. doi:[10.1029/1999JC000308](https://doi.org/10.1029/1999JC000308)

14. Fishwick JR, Aiken J, Barlow R, Sessions H, Bernard S, Ras J. Functional relationships and bio-optical properties derived from phytoplankton pigments, optical and photosynthetic parameters; a case study of the Benguela ecosystem. Journal of the Marine Biological Association of the United Kingdom. 2006;86: 1267–1280. doi:[10.1017/S0025315406014287](https://doi.org/10.1017/S0025315406014287)

15. Hirata T, Hardman-Mountford NJ, Brewin RJW, Aiken J, Barlow R, Suzuki K, et al. Synoptic relationships between surface Chlorophyll-*a* and diagnostic pigments specific to phytoplankton functional types. Biogeosciences. 2011;8: 311–327. doi:[10.5194/bg-8-311-2011](https://doi.org/10.5194/bg-8-311-2011)

16. Six C, Ratin M, Marie D, Corre E. Marine Synechococcus picocyanobacteria: Light utilization across latitudes. Proceedings of the National Academy of Sciences. 2021;118: e2111300118. doi:[10.1073/pnas.2111300118](https://doi.org/10.1073/pnas.2111300118)

17. Haverkamp THA, Schouten D, Doeleman M, Wollenzien U, Huisman J, Stal LJ. Colorful microdiversity of Synechococcus strains (picocyanobacteria) isolated from the Baltic Sea. The ISME Journal. 2009;3: 397–408. doi:[10.1038/ismej.2008.118](https://doi.org/10.1038/ismej.2008.118)

18. Aguilera A, Alegria Zufia J, Bas Conn L, Gurlit L, Śliwińska-Wilczewska S, Budzałek G, et al. Ecophysiological analysis reveals distinct environmental preferences in closely related Baltic Sea picocyanobacteria. Environmental Microbiology. 2023;n/a: 1–12. doi:[10.1111/1462-2920.16384](https://doi.org/10.1111/1462-2920.16384)

19. Six C, Finkel ZV, Irwin AJ, Campbell DA. Light Variability Illuminates Niche-Partitioning among Marine Picocyanobacteria. PLOS ONE. 2007;2: e1341. doi:[10.1371/journal.pone.0001341](https://doi.org/10.1371/journal.pone.0001341)

20. Guillard RRL. Culture of Phytoplankton for Feeding Marine Invertebrates. In: Smith WL, Chanley MH, editors. Culture of Marine Invertebrate Animals: Proceedings 1st Conference on Culture of Marine Invertebrate Animals Greenport. Boston, MA: Springer US; 1975. pp. 29–60. doi:[10.1007/978-1-4615-8714-9\_3](https://doi.org/10.1007/978-1-4615-8714-9_3)

21. Blake R, Griff M. In situ spectroscopy on intact Leptospirillum ferrooxidans reveals that reduced cytochrome 579 is an obligatory intermediate in the aerobic iron respiratory chain. Frontiers in Microbiology. 2012;3.

22. Jávorfi T, Erostyák J, Gál J, Buzády A, Menczel L, Garab G, et al. Quantitative spectrophotometry using integrating cavities. Journal of Photochemistry and Photobiology B: Biology. 2006;82: 127–131. doi:[10.1016/j.jphotobiol.2005.10.002](https://doi.org/10.1016/j.jphotobiol.2005.10.002)

23. Morel A. Available, usable, and stored radiant energy in relation to marine photosynthesis. Deep Sea Research. 1978;25: 673–688. doi:[10.1016/0146-6291(78)90623-9](https://doi.org/10.1016/0146-6291(78)90623-9)

24. Strickland JD, Parsons TR. Practical Hand Book of Seawater Analysis. Fisheries Research Board of Canada. 1972;167 (2nd edition): 1–311. doi:[DOI: http://dx.doi.org/10.25607/OBP-1791](DOI:%20http://dx.doi.org/10.25607/OBP-1791)

25. Bennett A, Bogorad L. COMPLEMENTARY CHROMATIC ADAPTATION IN A FILAMENTOUS BLUE-GREEN ALGA. Journal of Cell Biology. 1973;58: 419–435. doi:[10.1083/jcb.58.2.419](https://doi.org/10.1083/jcb.58.2.419)

26. Alin A, Kurt S. Testing non-additivity (interaction) in two-way ANOVA tables with no replication. Statistical methods in medical research. 2006;15: 63–85. doi:[10.1191/0962280206sm426oa](https://doi.org/10.1191/0962280206sm426oa)

27. Medley DM, Mitzel HE, Doi AN. Analysis-of-Variance Models and their use in a Three-Way Design without Replication. The Journal of Experimental Education. 1956;24: 221–229. doi:[10.1080/00220973.1956.11010542](https://doi.org/10.1080/00220973.1956.11010542)

28. Toutenburg H. Hollander, M., D. A. Wolfe: Nonparametric statistical methods. John Wiley & Sons, New York-Sydney-Tokyo-Mexico City 1973. Biometrische Zeitschrift. 1975;17: 526–526. doi:[10.1002/bimj.19750170808](https://doi.org/10.1002/bimj.19750170808)

29. R Core Team R. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing. Vienna, Austria. 2019.

30. Team RStudio. RStudio: Integrated Development Environment for R. Boston, MA: RStudio, Inc. https://support.posit.co/hc/en-us/articles/206212048-Citing-RStudio; 2015.

31. Handel A. Andreas Handel - Custom Word formatting using R Markdown. https://www.andreashandel.com/posts/2020-10-07-custom-word-format/; 2020.

32. Wickham H. Data Analysis. In: Wickham H, editor. Ggplot2: Elegant Graphics for Data Analysis. Cham: Springer International Publishing; 2016. pp. 189–201. doi:[10.1007/978-3-319-24277-4\_9](https://doi.org/10.1007/978-3-319-24277-4_9)

33. Alberte RS, Tel-Or E, Packer L, Thornber JP. Functional organisation of the photo-synthetic apparatus in heterocysts of nitrogen-fixing cyanobacteria. Nature. 1980;284: 481–483. doi:[10.1038/284481a0](https://doi.org/10.1038/284481a0)

34. LaRoche J, Robicheau BM. The Pelagic Light-Dependent Microbiome. In: Stal LJ, Cretoiu MS, editors. The Marine Microbiome. Cham: Springer International Publishing; 2022. pp. 395–423. doi:[10.1007/978-3-030-90383-1\_9](https://doi.org/10.1007/978-3-030-90383-1_9)

35. Śliwińska-Wilczewska S, Konarzewska Z, Wiśniewska K, Konik M. Photosynthetic Pigments Changes of Three Phenotypes of Picocyanobacteria Synechococcus sp. Under Different Light and Temperature Conditions. Cells. 2020;9: 2030. doi:[10.3390/cells9092030](https://doi.org/10.3390/cells9092030)

36. Śliwińska-Wilczewska S, Cieszyńska A, Maculewicz J, Latała A. Ecophysiological characteristics of red, green, and brown strains of the Baltic picocyanobacterium *Synechococcus* sp. a laboratory study. Biogeosciences. 2018;15: 6257–6276. doi:[10.5194/bg-15-6257-2018](https://doi.org/10.5194/bg-15-6257-2018)

37. Kirk JTO. Light and Photosynthesis in Aquatic Ecosystems. 1983.

38. Field CB, Behrenfeld MJ, Randerson JT, Falkowski P. Primary Production of the Biosphere: Integrating Terrestrial and Oceanic Components. Science. 1998;281: 237–240. doi:[10.1126/science.281.5374.237](https://doi.org/10.1126/science.281.5374.237)

39. Torremorell A, Llames ME, Pérez GL, Escaray R, Bustingorry J, Zagarese H. Annual patterns of phytoplankton density and primary production in a large, shallow lake: The central role of light. Freshwater Biology. 2009;54: 437–449. doi:[10.1111/j.1365-2427.2008.02119.x](https://doi.org/10.1111/j.1365-2427.2008.02119.x)

40. Chen J, Li Y, Jing H, Zhang X, Xu Z, Xu J, et al. Genomic and transcriptomic evidence for the diverse adaptations of Synechococcus subclusters 5.2 and 5.3 to mesoscale eddies. New Phytologist. 2022;233: 1828–1842. doi:[10.1111/nph.17903](https://doi.org/10.1111/nph.17903)

41. Dufresne A, Ostrowski M, Scanlan DJ, Garczarek L, Mazard S, Palenik BP, et al. Unraveling the genomic mosaic of a ubiquitous genus of marine cyanobacteria. Genome Biology. 2008;9: R90. doi:[10.1186/gb-2008-9-5-r90](https://doi.org/10.1186/gb-2008-9-5-r90)

42. Mella-Flores D, Six C, Ratin M, Partensky F, Boutte C, Le Corguillé G, et al. Prochlorococcus and Synechococcus have Evolved Different Adaptive Mechanisms to Cope with Light and UV Stress. Frontiers in Microbiology. 2012;3.

43. Churilova TYa, Suslin VV, Moiseeva NA, Efimova TV. Phytoplankton Bloom and Photosynthetically Active Radiation in Coastal Waters. Journal of Applied Spectroscopy. 2020;86: 1084–1091. doi:[10.1007/s10812-020-00944-0](https://doi.org/10.1007/s10812-020-00944-0)

44. Falkowski P, Scholes RJ, Boyle E, Canadell J, Canfield D, Elser J, et al. The Global Carbon Cycle: A Test of Our Knowledge of Earth as a System. Science. 2000;290: 291–296. doi:[10.1126/science.290.5490.291](https://doi.org/10.1126/science.290.5490.291)

45. Morel A. Optical modeling of the upper ocean in relation to its biogenous matter content (case I waters). Journal of Geophysical Research: Oceans. 1988;93: 10749–10768. doi:[10.1029/JC093iC09p10749](https://doi.org/10.1029/JC093iC09p10749)

46. Chakdar H, Pabbi S. Cyanobacterial Phycobilins: Production, Purification, and Regulation. In: Shukla P, editor. Frontier Discoveries and Innovations in Interdisciplinary Microbiology. New Delhi: Springer India; 2016. pp. 45–69. doi:[10.1007/978-81-322-2610-9\_4](https://doi.org/10.1007/978-81-322-2610-9_4)

47. Stadnichuk IN, Krasilnikov PM, Zlenko DV. Cyanobacterial phycobilisomes and phycobiliproteins. Microbiology. 2015;84: 101–111. doi:[10.1134/S0026261715020150](https://doi.org/10.1134/S0026261715020150)

48. Beale SI. Biosynthesis of Cyanobacterial Tetrapyrrole Pigments: Hemes, Chlorophylls, and Phycobilins. In: Bryant DA, editor. The Molecular Biology of Cyanobacteria. Dordrecht: Springer Netherlands; 1994. pp. 519–558. doi:[10.1007/978-94-011-0227-8\_17](https://doi.org/10.1007/978-94-011-0227-8_17)