

Photosynthetic Adaptations of Polar Phytoplankton to Extreme Low Light

Natasha Madeleine Ryan Supervised By Dr. Douglas A. Campbell Spring Phytoplankton Blooms

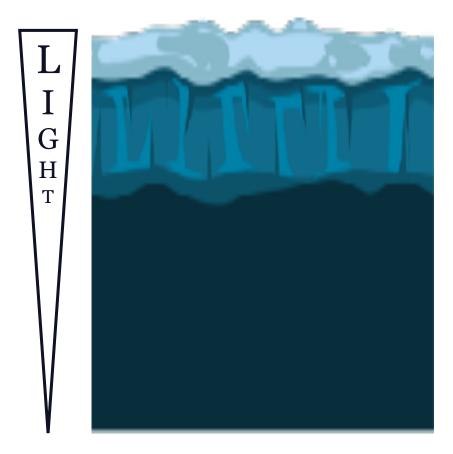


Ardyna & Arrigo, 2020. Nature Climate Change

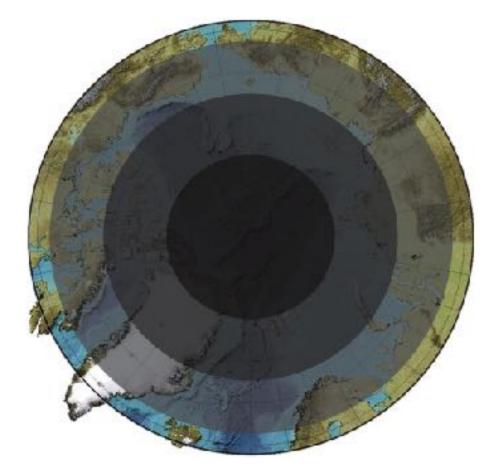
Phytoplankton Bloom in the Barents Sea, Joshua Stevens, NASA

Photic Constraints

Light Attenuation



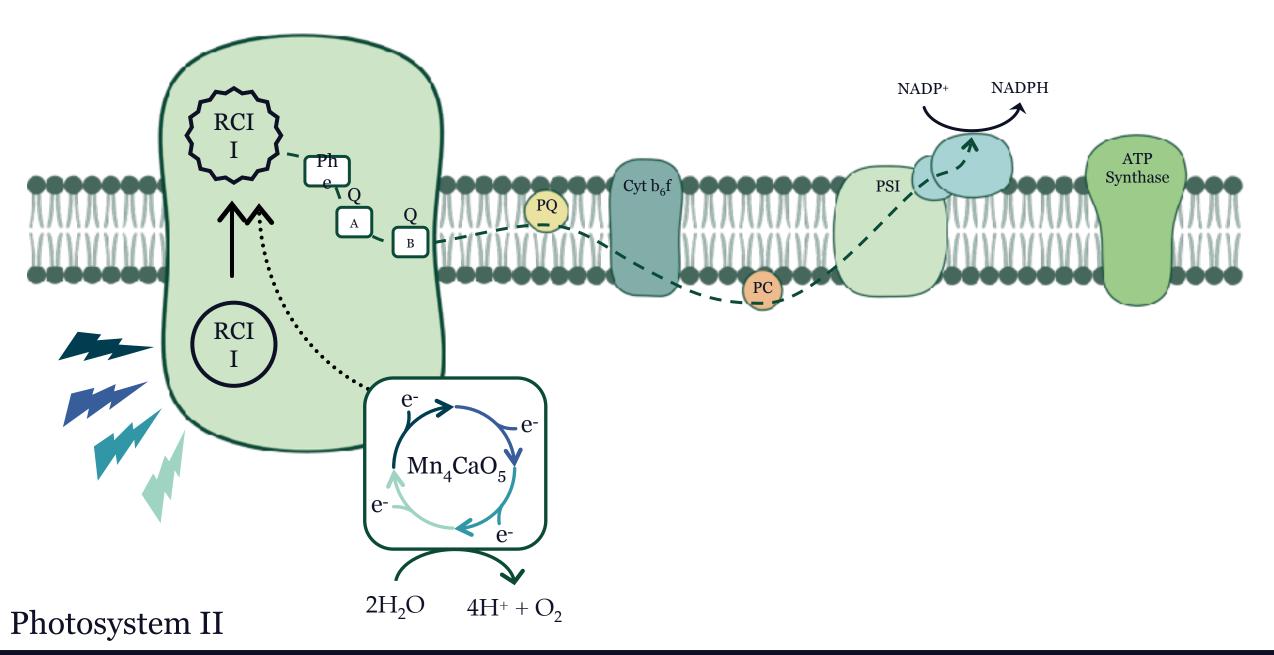
Polar Night



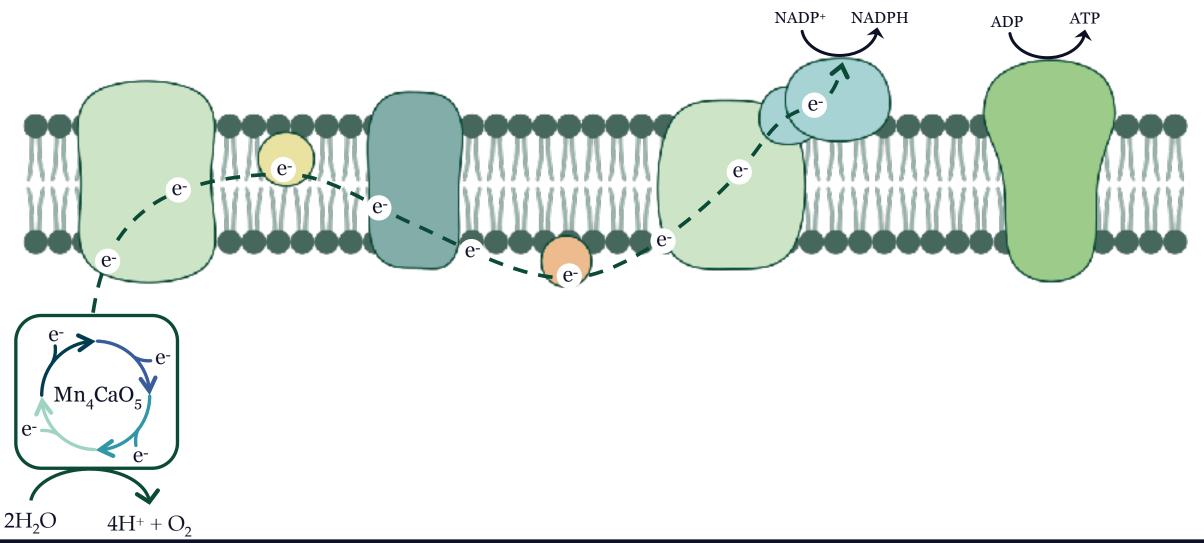
Have polar phytoplankton adapted to extremely low light by increasing their efficiency of photosynthetic energy conversion?

Have polar phytoplankton adapted to extremely low light by increasing their efficiency of photosynthetic energy conversion?

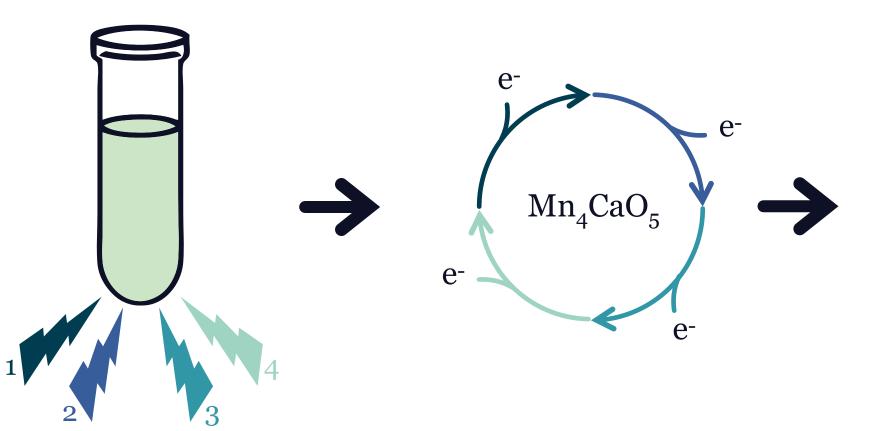
Hypothesized Mechanism: suppressing energetically wasteful charge recombinations in Photosystem II

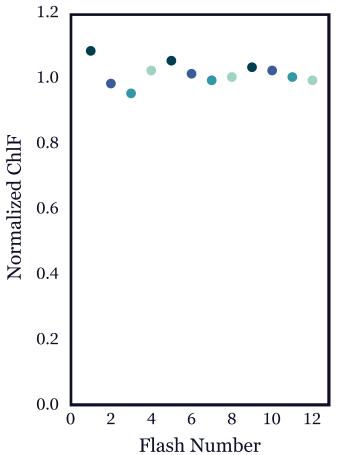


Stable Cycling

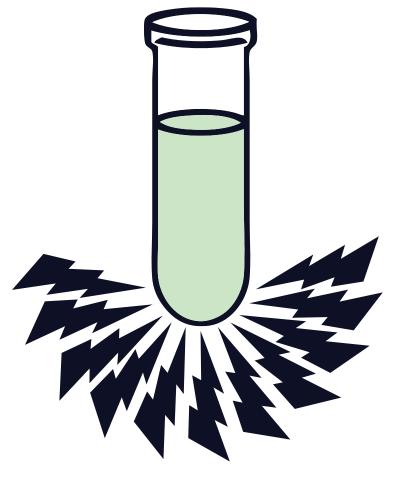


St-ChlF Approach



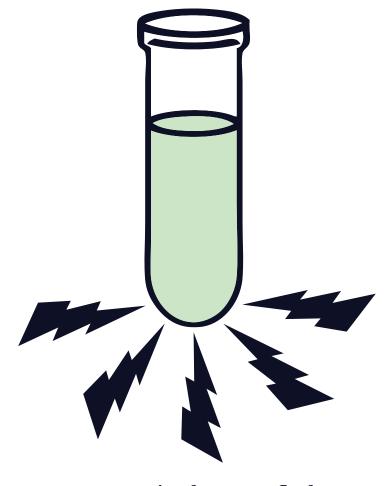


High Light



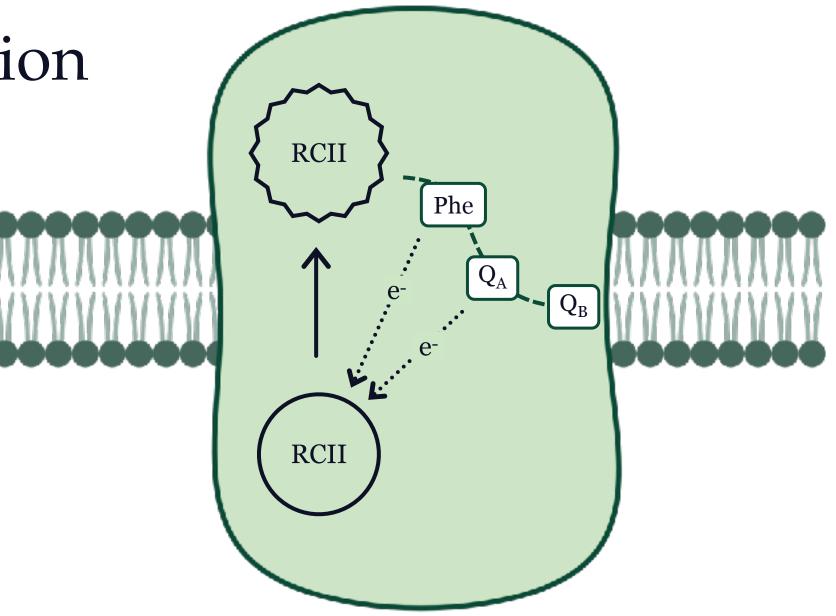
Shorter spacing between flashes

Low Light

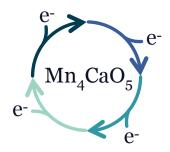


Longer spacing between flashes

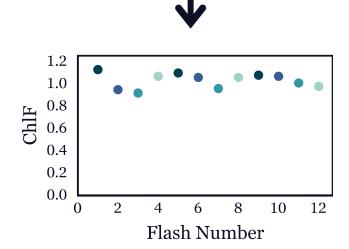
Recombination



Idealized PSII Population

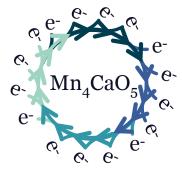


Synchronized Cycling



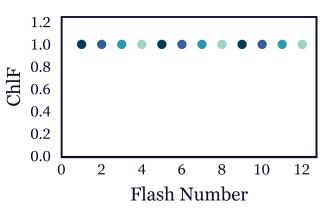
Periodic ChlF Oscillations

PSII Population with Recombination



Desynchronized Cycling





Damped ChlF Oscillations

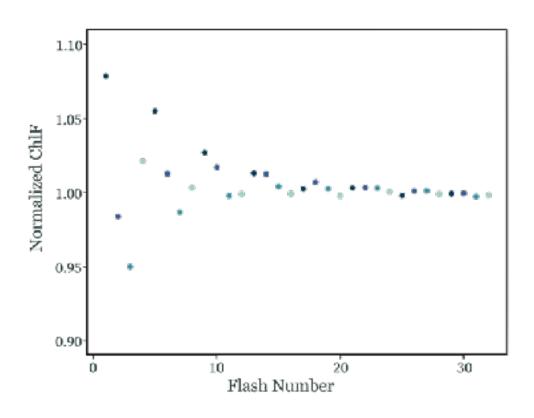
Wavelet Transformation

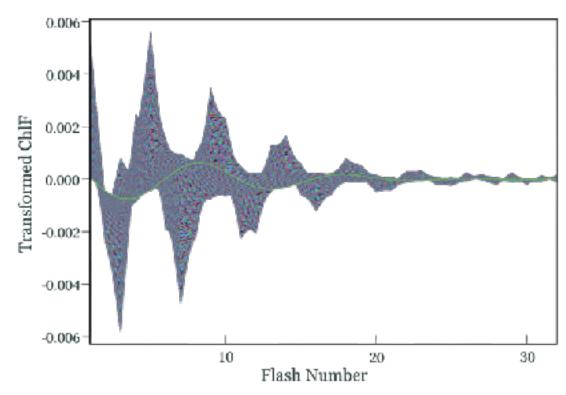
Sample Plots: Fragilariopsis cylindrus at 0°C and 0.6 µmol photons m-2s-1

Raw Fluorescence Data



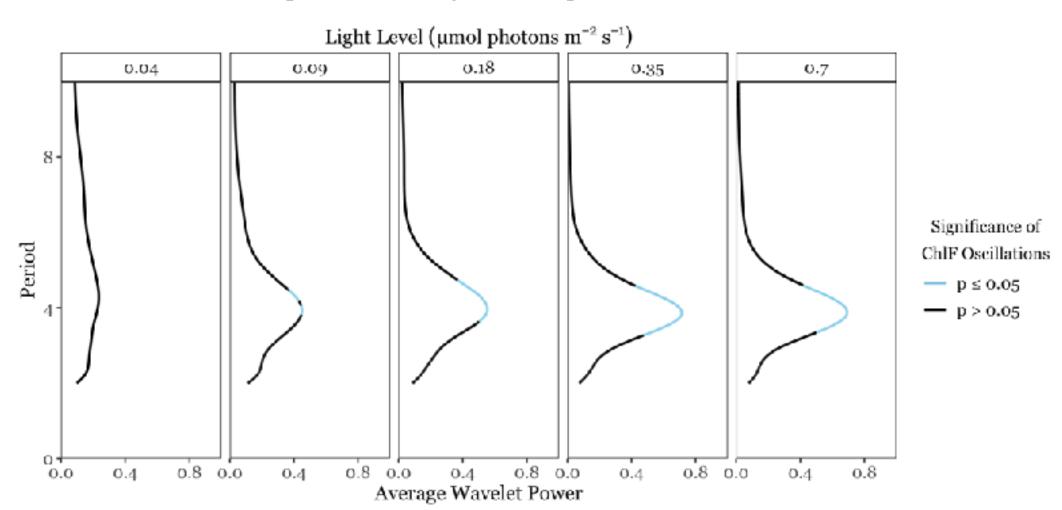
Fourier Wavelets





ChlF Periodicity

Sample Plot: Chlamydomonas priscuii at 8°C

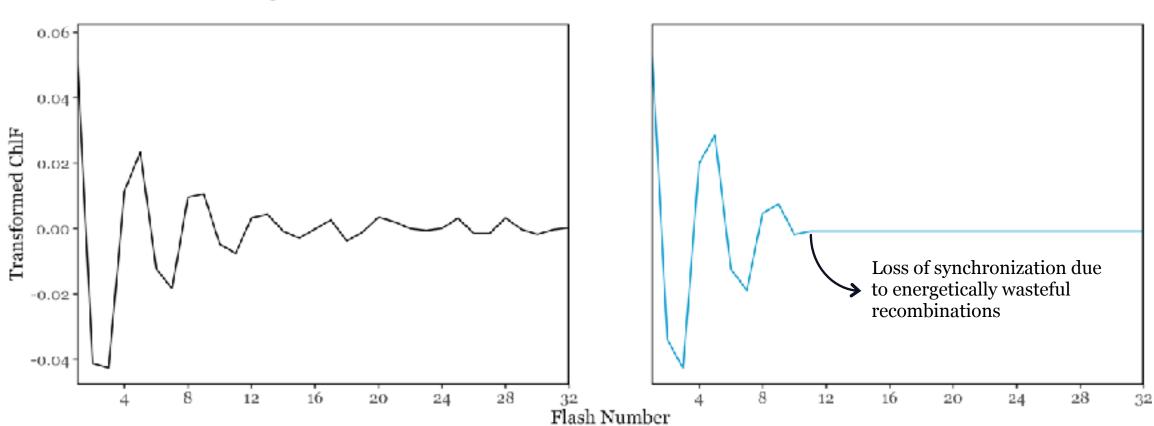


Duration of Cycling

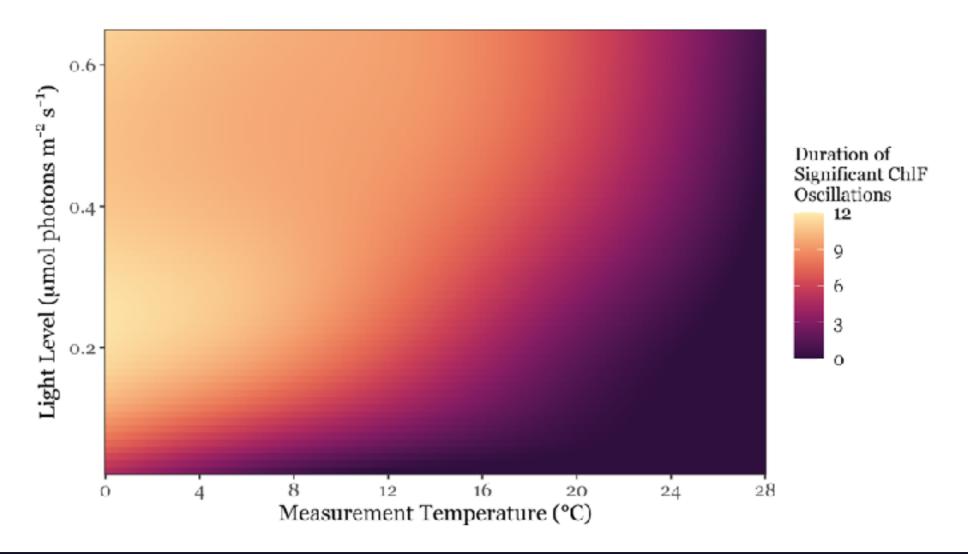
Sample Plot: *Chlamydomonas priscuii* at 8°C & 0.7 µmol photons m⁻²s⁻¹

Original Wavelet

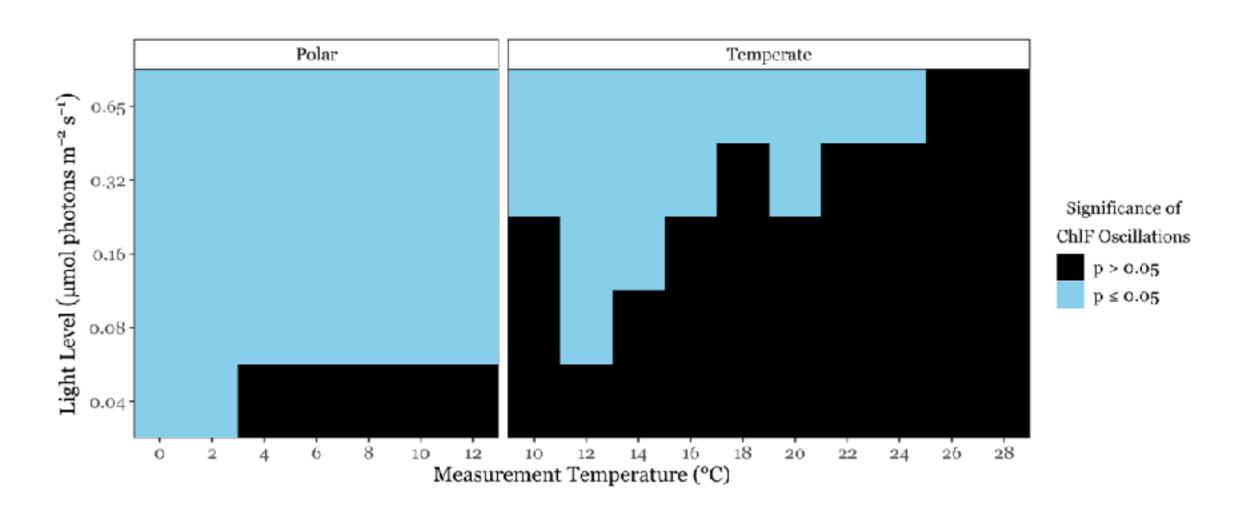
Reconstruction at $\alpha = 0.05$



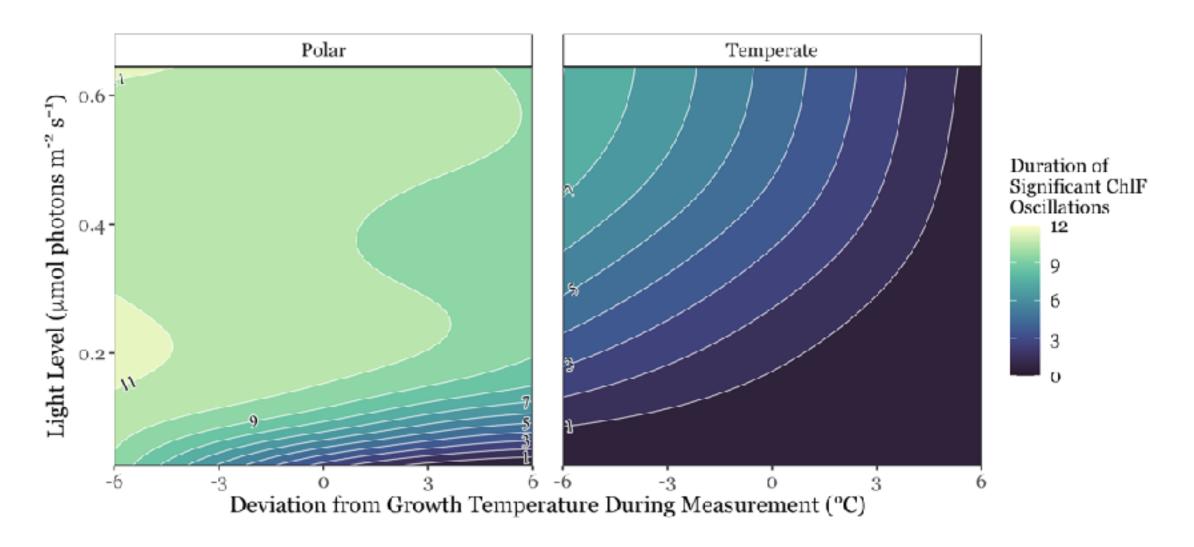
Individual strains sustain ChlF oscillations longer under measurement at **higher** light levels & **lower** temperatures



Polar strains exhibited synchronous cycling under a **broader** range of conditions



Polar strains sustained **longer** cycling under low light & comparable temperatures



- Within strains, longer cycling was observed at higher light levels and colder temperatures
- Polar strains exhibit significant cycling across a broader range of conditions than temperate strains
- 3 Under low light & comparable temperatures, polar strains sustain longer synchronous cycling than temperate strains

Polar phytoplankton exhibit the capacity to suppress energetically wasteful charge recombinations & sustain energetically efficient photosynthesis under extremely low light

Acknowledgements

I thank my supervisor:
Dr. Douglas A. Campbell

as well as Campbell Lab members:

Dr. Maximilian Berthold, Dr. Sylwia Sliwinska-Wilczewska, Naaman Omar,

Mireille Savoie, & Miranda Corkum

and our partners at

The International Research Laboratory Takuvik at Université de Laval

& Cvetkovska Lab at the University of Ottawa











References

Ardyna, M., & Arrigo, K. R. (2020). Phytoplankton dynamics in a changing Arctic Ocean. Nature Climate Change, 10(10), Article 10.

Berge, J., Renaud, P. E., Darnis, G., Cottier, F., Last, K., Gabrielsen, T. M., Johnsen, G., Seuthe, L., Weslawski, J. M., Leu, E., Moline, M., Nahrgang, J., Søreide, J. E., Varpe, Ø., J., Daase, M., & Falk-Petersen, S. (2015). In the dark: A review of ecosystem processes during the Arctic polar night. *Progress in Oceanography*, 139, 258–271.

Berman-Frank, I., Campbell, D., Ciotti, A., Erickson, Z., Fujiki, T., Halsey, K., Hickman, A., Gorbunov, M., Hughes, D., Kolber, Z., Moore, M., Oxborough, K., Prasil, O., Robinson, C., Ryan-Keogh, T., Silsbe, G., Simis, S., Thomalla, S., & Varkey, D. (2023). *Application of Single Turnover Active Chlorophyll Fluorescence for Phytoplankton***Productivity Measurements. Version 2.0 [Report]. Scientific Committee on Oceanic Research (SCOR) Working Group 156.

Gates, C., Ananyev, G., & Dismukes, G. C. (2020). Realtime kinetics of the light driven steps of photosynthetic water oxidation in living organisms by "stroboscopic" fluorometry. *Biochimica et Biophysica Acta (BBA) - Bioenergetics*, 1861(8), 148212.

Hancke, K., Lund-Hansen, L. C., Lamare, M. L., Højlund Pedersen, S., King, M. D., Andersen, P., & Sorrell, B. K. (2018). Extreme Low Light Requirement for Algae Growth Underneath Sea Ice: A Case Study From Station Nord, NE Greenland. *Journal of Geophysical Research: Oceans*, 123(2), 985–1000.

Kawakami, K., & Shen, J.-R. (2018). Purification of fully active and crystallizable photosystem II from thermophilic cyanobacteria. In Methods in Enzymology (Vol. 613, pp. 1–16). Elsevier.

Keren, N., Berg, A., van Kan, P. J. M., Levanon, H., & Ohad, I. (1997). Mechanism of photosystem II photoinactivation and D1 protein degradation at low light: The role of back electron flow. *Proceedings of the National Academy of Sciences*, *94*(4), 1579–1584.

Kirk, J. T. O. (2011). Light and Photosynthesis in Aquatic Ecosystems (3rd ed.). Cambridge University Press.

Mukhopadhyay, S., Mandal, S. K., Bhaduri, S., & Armstrong, W. H. (2004). Manganese Clusters with Relevance to Photosystem II. Chemical Reviews, 104(9), 3981–4026.

Randelhoff, A., Lacour, L., Marec, C., Leymarie, E., Lagunas, J., Xing, X., Darnis, G., Penkerc'h, C., Sampei, M., Fortier, L., D'Ortenzio, F., Claustre, H., & Babin, M. (2020). Arctic mid-winter phytoplankton growth revealed by autonomous profilers. *Science Advances*, *6*(39), eabc2678.

Rappaport, F., Cuni, A., Xiong, L., Sayre, R., & Lavergne, J. (2005). Charge Recombination and Thermoluminescence in Photosystem II. Biophysical Journal, 88(3), 1948–1958.

Raven, J. A., Kübler, J. E., & Beardall, J. (2000). Put out the light, and then put out the light. Journal of the Marine Biological Association of the United Kingdom, 80(1), 1–25.

Roesch, A., & Schmidbauer, H. (2018). WaveletComp: Computational wavelet analysis [Manual].

Schuback, N., Tortell, P. D., Berman-Frank, I., Campbell, D. A., Ciotti, A., Courtecuisse, E., Erickson, Z. K., Fujiki, T., Halsey, K., Hickman, A. E., Huot, Y., Gorbunov, M. Y., Hughes, D. J., Kolber, Z. S., Moore, C. M., Oxborough, K., Prášil, O., Robinson, C. M., Ryan-Keogh, T. J., ... Varkey, D. R. (2021). Single-Turnover Variable Chlorophyll Fluorescence as a Tool for Assessing Phytoplankton Photosynthesis and Primary Productivity: Opportunities, Caveats and Recommendations. Frontiers in Marine Science, 8.

Theis, F. J., & Meyer-Bäse, A. (2010). Spectral Transformations. In Biomedical Signal Analysis: Contemporary Methods and Applications (1st ed., p. 42). MIT Press.

Vass, I. (2011). Role of charge recombination processes in photodamage and photoprotection of the photosystem II complex. *Physiologia Plantarum*, 142(1), 6–16.

Zaharieva, I., & Dau, H. (2019). Energetics and Kinetics of S-State Transitions Monitored by Delayed Chlorophyll Fluorescence. Frontiers in Plant Science, 10.