

# Growth and Functional Analyses of Photosynthetic Strategies of *Prochlorococcus* Strains Under Varying Oxygen

Julie Nadeau<sup>1</sup>, Mireille Savoie<sup>1</sup>, Douglas Campbell<sup>1</sup>

<sup>1</sup>Department of Biochemistry, Mount Allison University

## Introduction

### *Prochlorococcus*:

- Photosynthetic unicellular cyanobacterium [1]
- Contributes most primary production in the open oceans. [2]
- Different strains occupy a wide range of habitats, including Oxygen Minimum Zones. [3]
- With climate change, ocean warming will benefit *Prochlorococcus*, but also cause decreased oxygen solubility [2]

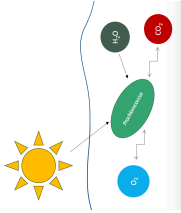


Figure 1: The biological interactions of *Prochlorococcus* with oxygen and light

## Objectives

1. Determine whether *Prochlorococcus* strains are constitutively able to accommodate changes in oxygen, or whether they acclimate over a period of time to different levels of oxygen.
2. Provide insights into the potential ecological niches of *Prochlorococcus* strains.

## Methods

### Biological Analysis of Growth Rates

Using a Multi-Cultivator, two strains of *Prochlorococcus* (MED4, MIT9313) were monitored for OD680 (Chlorophyll and scattering) and OD720 (cell scattering). Under 22°C, 12h photoperiod of blue light (450 ± 45 nm), and combinations of dissolved O<sub>2</sub> (250, 25, 2 µM) and light levels (30, 90, 180 µmol photons m<sup>-2</sup> s<sup>-1</sup>).

### Biological Functional Measurements

Exposed samples under 250, 25, 2 µM O<sub>2</sub> and a series of increasing light levels to track 'light response' curves of Photosystem II electron transport, using Solisense FRR1 instrument. Photosystem I and Photosystem II electron transport in parallel, using Dual-PAM-100 instrument.

## Results & Discussion

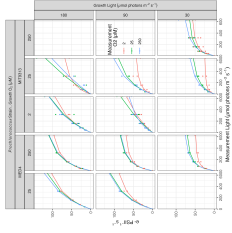


Figure 2: Light response curves of PSI electron transport (PspI1-e2) vs measurement light (µmol photons m<sup>-2</sup> s<sup>-1</sup>) for *Prochlorococcus* strains MED4 and MIT9313, after growth under constant O<sub>2</sub> (250, 25, 2 µM) and 180 µmol photons m<sup>-2</sup> s<sup>-1</sup> for 24h. Lines show that curve fits are used for the data.

- Both strains show significant short term responses of electron transport to decreasing oxygen. Growth under 2 µM O<sub>2</sub> diminishes the short term effects of changing measurement oxygen, indicating growth acclimation to oxygen status.

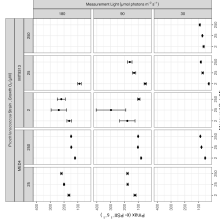


Figure 3: Maximum PSII electron transport rate (PmaxII) vs measurement light (µmol photons m<sup>-2</sup> s<sup>-1</sup>) for *Prochlorococcus* strains MED4 and MIT9313, after growth under constant O<sub>2</sub> (250, 25, 2 µM) and 180 µmol photons m<sup>-2</sup> s<sup>-1</sup> for 24h. Error bars represent standard deviation.

- Strain MED4 shows increasing Pmax values across increasing measurement oxygen concentrations, indicating short term responses to varying oxygen levels. Strain MIT9313 also increases with increasing light but not with increasing O<sub>2</sub>.
- Strain MIT9313 shows interactive effects of measurement oxygen, growth oxygen concentration and growth light on Pmax values, particularly between the lowest (2 µM) and highest (250 µM) oxygen concentrations, indicating acclimating adaptation to varying oxygen levels.

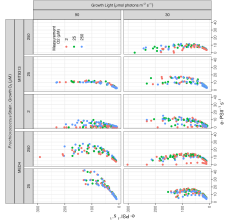


Figure 4: PSI electron transport (PspI1-e2) vs PSI electron transport (PspI1-e2) for *Prochlorococcus* strains MED4 and MIT9313 after growth under different combinations of light level (0 or 180 µmol photons m<sup>-2</sup> s<sup>-1</sup>) and oxygen concentration (2, 25, 250 µM). Error bars represent standard deviation.

- Directly comparing PSI to PSII electron transport shows that in MED4 growth under 25 µM O<sub>2</sub> decreases PSI electron transport.
- In contrast, in MIT9313, PSI electron transport remains more consistent across growth O<sub>2</sub> concentration.

## Conclusion & Next Steps

*Prochlorococcus* shows both long and short term responses to oxygen.

- Cell pellets for future transcriptomic analyses.
- Monitoring electron carrier reduction status through Whole Cell Absorbance Spectra.

## References

1. Becken A.G., Chisholm S.W., Binder B.L. *In: The Molecular Biology of Prochlorococcus and Synechococcus*. Martin, C. (ed.), 1995. Birkbeck Press, New York, 1-15.
2. Fencham P., Gargues J., Gosselin J., Ravelo J., Zhai L., Bao N., et al. *Prochlorococcus and Synechococcus: The Marine Cyanobacteria*. In: *Marine Microbiology and Systematics*. Proceedings of the 10th International Conference on Marine Microbiology, 1995. Springer, 1-15.
3. Paul S., Datta A., Bui S.K., Das S., Datta C. *Marine cyanobacteria: oxygen and redox regulation in the marine cyanobacteria Prochlorococcus*. BMC Genomics. 2010;11:101.