## **Peroxisome**

A **peroxisome** (IPA: [peg'.mksi\_soum]) [1] is a membrane-bound organelle (formerly known as a microbody), found in the cytoplasm of virtually all eukarvotic cells.[2] Peroxisomes are oxidative organelles. Frequently, molecular oxygen serves as a co-substrate, from which hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) is then formed. Peroxisomes owe their name to hydrogen peroxide generating and scavenging activities. They perform key roles in lipid metabolism and the conversion of reactive oxygen species. Peroxisomes are involved in the catabolism of very long chain fatty acids, branched chain fatty acids, bile acid intermediates (in the liver), D-amino acids, and polyamines, the reduction of reactive oxygen species - specifically hydrogen peroxide[3] - and the plasmalogens, biosynthesis of i.e., phospholipids critical for the normal function of mammalian brains and lungs. [4] They also contain approximately 10% of the total activity of two enzymes (Glucose-6-phosphate dehydrogenase and 6-Phosphogluconate dehydrogenase) in the pentose phosphate pathway, [5] which is important for energy metabolism. [4] It is vigorously debated whether peroxisomes are involved in isoprenoid and cholesterol synthesis in animals. 4 Other known peroxisomal functions include the glyoxylate cycle in germinating seeds ("glyoxysomes"), photorespiration leaves.[6] glycolysis trypanosomes in ("glycosomes"), and methanol and/or amine oxidation and assimilation in some yeasts.

### **Contents**

History

Structural Design

**Metabolic functions** 

Peroxisome assembly

Peroxisome interaction and communication

Associated medical conditions

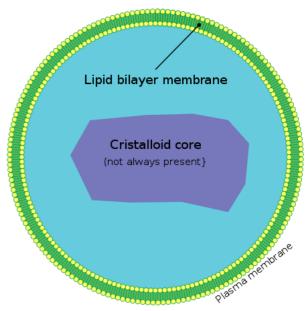
Genes

**Evolutionary origins** 

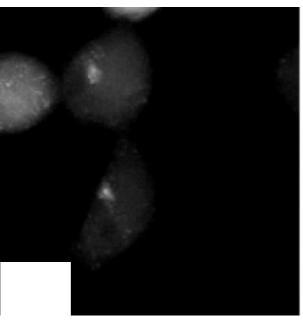
Other related organelles

See also

References



Basic structure of a peroxisome

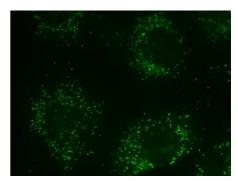


Distribution of peroxisomes (white) in <u>HEK 293</u> cells during mitosis

#### **External links**

## **History**

Peroxisomes (microbodies) were first described by a Swedish doctoral student, J. Rhodin in 1954. They were identified as organelles by the Belgian cytologist Christian de Duve in 1967. De Duve and co-workers discovered that peroxisomes contain several oxidases involved in the production of hydrogen peroxide  $(H_2O_2)$  as well as catalase involved in the decomposition of  $H_2O_2$ 



Peroxisome in rat neonatal cardiomyocyte

to oxygen and water. Due to their role in peroxide metabolism, De Duve named them "peroxisomes", replacing the formerly used morphological term "microbodies". Later, it was described that firefly luciferase is targeted to peroxisomes in mammalian cells, allowing the discovery of the import targeting signal for peroxisomes, and triggering many advances in the peroxisome biogenesis field. [9][10]

## Structural Design

Peroxisomes are small (0.1–1  $\mu$ m diameter) subcellular compartments (organelles) with a fine, granular matrix and surrounded by a single biomembrane which are located in the cytoplasm of a cell. [11][12] Compartmentalization creates an optimized environment to promote various metabolic reactions within peroxisomes required to sustain cellular functions and viability of the organism.

The number, size and protein composition of peroxisomes are variable and depend on cell type and environmental conditions. For example, in baker's yeast ( $\underline{S.\ cerevisiae}$ ), it has been observed that, with good glucose supply, only a few, small peroxisomes are present. In contrast, when the yeasts were supplied with long-chain fatty acids as sole carbon source up to 20 to 25 large peroxisomes can be formed. [13]

### **Metabolic functions**

A major function of the peroxisome is the breakdown of <u>very long chain fatty acids</u> through <u>beta oxidation</u>. In animal cells, the long fatty acids are converted to <u>medium chain fatty acids</u>, which are subsequently shuttled to <u>mitochondria</u> where they eventually are broken down to carbon dioxide and water. In yeast and plant cells, this process is carried out exclusively in peroxisomes. [14][15]

The first reactions in the formation of <u>plasmalogen</u> in animal cells also occur in peroxisomes. Plasmalogen is the most abundant phospholipid in <u>myelin</u>. Deficiency of plasmalogens causes profound abnormalities in the myelination of <u>nerve cells</u>, which is one reason why many <u>peroxisomal disorders</u> affect the nervous system. Peroxisomes also play a role in the production of <u>bile acids important</u> for the absorption of fats and fat-soluble vitamins, such as vitamins A and K. Skin disorders are features of genetic disorders affecting peroxisome function as a result.

The specific metabolic pathways that occur exclusively in mammalian peroxisomes are: [4]

- α-oxidation of phytanic acid
- β-oxidation of very-long-chain and polyunsaturated fatty acids
- biosynthesis of plasmalogens
- conjugation of cholic acid as part of bile acid synthesis

Peroxisomes contain oxidative <u>enzymes</u>, such as <u>D-amino acid oxidase</u> and <u>uric acid oxidase</u>. However the last enzyme is absent in humans, explaining the disease known as <u>gout</u>, caused by the accumulation of uric acid. Certain enzymes within the peroxisome, by using molecular oxygen, remove hydrogen atoms from specific organic substrates (labeled as R), in an oxidative reaction, producing hydrogen peroxide ( $H_2O_2$ , itself toxic):

$$RH_2 + O_2 \rightarrow R + H_2O_2$$

Catalase, another peroxisomal enzyme, uses this  $H_2O_2$  to oxidize other substrates, including <u>phenols</u>, formic acid, formaldehyde, and alcohol, by means of the peroxidation reaction:

 $H_2O_2 + R'H_2 \rightarrow R' + 2H_2O$ , thus eliminating the poisonous hydrogen peroxide in the process.

This reaction is important in liver and kidney cells, where the peroxisomes detoxify various toxic substances that enter the blood. About 25% of the ethanol that humans consume by drinking alcoholic beverages is oxidized to acetaldehyde in this way. [14] In addition, when excess  $H_2O_2$  accumulates in the cell, catalase converts it to  $H_2O$  through this reaction:

$$2\mathrm{H}_2\mathrm{O}_2 
ightarrow 2\mathrm{H}_2\mathrm{O} + \mathrm{O}_2$$

In higher plants, peroxisomes contain also a complex battery of antioxidative enzymes such as superoxide dismutase, the components of the ascorbate-glutathione cycle, and the NADP-dehydrogenases of the pentose-phosphate pathway. It has been demonstrated that peroxisomes generate superoxide  $(O_2^{\bullet-})$  and nitric oxide (NO) radicals. [17][18]

There is evidence now that those reactive oxygen species including peroxisomal  $H_2O_2$  are also important signalling molecules in plants and animals and contribute to healthy ageing and agerelated disorders in humans. [19]

The peroxisome of plant cells is polarised when fighting fungal penetration. Infection causes a <u>glucosinolate</u> molecule to play an antifungal role to be made and delivered to the outside of the cell through the action of the peroxisomal proteins (PEN2 and PEN3). [20]

Peroxisomes in mammals and humans also contribute to anti-viral defense. [21] and the combat of pathogens [22]

# Peroxisome assembly

Peroxisomes can be derived from the smooth endoplasmic reticulum under certain experimental conditions and replicate by membrane growth and division out of pre-existing organelles. Peroxisome matrix proteins are translated in the cytoplasm prior to import. Specific amino acid sequences (PTS or peroxisomal targeting signal) at the *C-terminus* (PTS1) or *N-terminus* (PTS2) of peroxisomal matrix proteins signals them to be imported into the organelle by a targeting factor. There are currently 36 known proteins involved in peroxisome biogenesis and maintenance, called peroxins, which participate in the process of peroxisome assembly in different organisms. In mammalian cells there are 13 characterized peroxins. In contrast to protein import into the endoplasmic reticulum (ER) or mitochondria, proteins do not need to be unfolded to be imported into the peroxisome lumen. The matrix protein import receptors, the peroxins PEX5 and PEX7, accompany their cargoes (containing a PTS1 or a PTS2 amino acid sequence, respectively) all the way to the peroxisome where they release the cargo into the peroxisomal matrix and then return to the cytosol – a step named *recycling*. A special way of peroxisomal protein targeting is called piggy

backing. Proteins that are transported by this unique method do not have a canonical PTS, but rather bind on a PTS protein to be transported as a complex. A model describing the import cycle is referred to as the *extended shuttle mechanism*. There is now evidence that ATP hydrolysis is required for the recycling of receptors to the cytosol. Also, <u>ubiquitination</u> is crucial for the export of PEX5 from the peroxisome to the cytosol. The biogenesis of the peroxisomal membrane and the insertion of peroxisomal membrane proteins (PMPs) requires the peroxins PEX19, PEX3, and PEX16. PEX19 is a PMP receptor and chaperone, which binds the PMPs and routes them to the peroxisomal membrane, where it interacts with PEX3, a peroxisomal integral membrane protein. PMPs are then inserted into the peroxisomal membrane.

The degradation of peroxisomes is called pexophagy. [29]

#### Peroxisome interaction and communication

The diverse functions of peroxisomes require dynamic interactions and cooperation with many organelles involved in cellular lipid metabolism such as the endoplasmic reticulum (ER), mitochondria, lipid droplets, and lysosomes. [30]

Peroxisomes interact with mitochondria in several metabolic pathways, including  $\beta$ -oxidation of fatty acids and the metabolism of reactive oxygen species. Both organelles are in close contact with the endoplasmic reticulum (ER) and share several proteins, including organelle fission factors. Peroxisomes also interact with the endoplasmic reticulum (ER) and cooperate in the synthesis of ether lipids (plasmalogens) which are important for nerve cells (see above). In filamentous fungi, peroxisomes move on microtubules by 'hitchhiking,' a process involving contact with rapidly moving early endosomes. Physical contact between organelles is often mediated by membrane contact sites, where membranes of two organelles are physically tethered to enable rapid transfer of small molecules, enable organelle communication and are crucial for coordination of cellular functions and hence human health. Alterations of membrane contacts have been observed in various diseases.

#### **Associated medical conditions**

 $\frac{\text{Peroxisomal disorders}}{\text{system as well as}} \text{ are a class of medical conditions that typically affect the human nervous system as well as many other organ systems. Two common examples are <math>\frac{\text{X-linked}}{\text{System}}$  adrenoleukodystrophy and peroxisome biogenesis disorders.

#### Genes

*PEX* genes encode the protein machinery ("peroxins") required for proper peroxisome assembly, as described above. Membrane assembly and maintenance requires three of these (peroxins 3, 16, and 19) and may occur without the import of the matrix (lumen) enzymes. Proliferation of the organelle is regulated by Pex11p.

Genes that encode peroxin proteins include:  $\underline{PEX1}$ ,  $\underline{PEX2}$  (PXMP3),  $\underline{PEX3}$ ,  $\underline{PEX5}$ ,  $\underline{PEX6}$ ,  $\underline{PEX7}$ ,  $\underline{PEX9}$ ,  $\underline{^{[36][37]}}$   $\underline{PEX10}$ ,  $\underline{PEX11A}$ ,  $\underline{PEX11B}$ ,  $\underline{PEX11G}$ ,  $\underline{PEX12}$ ,  $\underline{PEX13}$ ,  $\underline{PEX14}$ ,  $\underline{PEX16}$ ,  $\underline{PEX19}$ ,  $\underline{PEX26}$ ,  $\underline{PEX28}$ ,  $\underline{PEX30}$ , and  $\underline{PEX31}$ . Between organisms, PEX numbering and function can differ.

## **Evolutionary origins**

The protein content of peroxisomes varies across species or organism, but the presence of proteins common to many species has been used to suggest an <a href="endosymbiotic">endosymbiotic</a> origin; that is, peroxisomes evolved from bacteria that invaded larger cells as parasites, and very gradually evolved a symbiotic

relationship. [38] However, this view has been challenged by recent discoveries. [39] For example, peroxisome-less mutants can restore peroxisomes upon introduction of the wild-type gene.

Two independent evolutionary analyses of the peroxisomal <u>proteome</u> found homologies between the peroxisomal import machinery and the <u>ERAD</u> pathway in the <u>endoplasmic</u> reticulum, [40][41] along with a number of metabolic enzymes that were likely recruited from the <u>mitochondria</u>. [41] Recently, it has been suggested that the peroxisome may have had an <u>actinobacterial</u> origin; [42] however, this is controversial. [43]

## Other related organelles

Other organelles of the <u>microbody</u> family related to peroxisomes include <u>glyoxysomes</u> of <u>plants</u> and filamentous fungi, glycosomes of kinetoplastids, and Woronin bodies of filamentous fungi.

#### See also

Peroxisome proliferator-activated receptor

### References

- 1. "Definition of PEROXISOME" (https://www.merriam-webster.com/dictionary/peroxisome). www.merriam-webster.com. Retrieved 2019-10-30.
- Islinger M, Voelkl A, Fahimi HD, Schrader M (November 2018). "The peroxisome: an update on mysteries 2.0" (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6182659). Histochemistry and Cell Biology. 150 (5): 443–471. doi:10.1007/s00418-018-1722-5 (https://doi.org/10.1007%2Fs00418-018-1722-5). PMC 6182659 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6182659). PMID 30219925 (https://pubmed.ncbi.nlm.nih.gov/30219925).
- 3. Bonekamp NA, Völkl A, Fahimi HD, Schrader M (2009). "Reactive oxygen species and peroxisomes: struggling for balance". *BioFactors*. **35** (4): 346–55. doi:10.1002/biof.48 (https://doi.org/10.1002%2Fbiof.48). PMID 19459143 (https://pubmed.ncbi.nlm.nih.gov/19459143). S2CID 7502822 (https://api.semanticscholar.org/CorpusID:7502822).
- 4. Wanders RJ, Waterham HR (2006). "Biochemistry of mammalian peroxisomes revisited". *Annual Review of Biochemistry*. **75**: 295–332. doi:10.1146/annurev.biochem.74.082803.133329 (https://doi.org/10.1146%2Fannurev.biochem.74.082803.133329). PMID 16756494 (https://pubmed.ncbi.nlm.nih.gov/16756494).
- Antonenkov, Vasily D. (Jul 1989). "Dehydrogenases of the pentose phosphate pathway in rat liver peroxisomes" (https://doi.org/10.1111%2Fj.1432-1033.1989.tb14898.x). European Journal of Biochemistry. 183 (1): 75–82. doi:10.1111/j.1432-1033.1989.tb14898.x (https://doi.org/10.1111%2 Fj.1432-1033.1989.tb14898.x). ISSN 0014-2956 (https://www.worldcat.org/issn/0014-2956). PMID 2753047 (https://pubmed.ncbi.nlm.nih.gov/2753047).
- 6. Evert RF, Eichhorn SE (2006). Esau's Plant Anatomy: Meristems, Cells, and Tissues of the Plant Body: Their Structure, Function, and Development. John Wiley & Sons. ISBN 9780471738435.
- 7. Rhodin, J (1954). "Correlation of ultrastructural organization and function in normal and experimentally changed proximal tubule cells of the mouse kidney". *Doctorate Thesis. Karolinska Institutet, Stockholm*.
- 8. de Duve C (April 1969). "The peroxisome: a new cytoplasmic organelle". *Proceedings of the Royal Society of London. Series B, Biological Sciences*. **173** (1030): 71–83. Bibcode:1969RSPSB.173...71D (https://ui.adsabs.harvard.edu/abs/1969RSPSB.173...71D). doi:10.1098/rspb.1969.0039 (https://doi.org/10.1098%2Frspb.1969.0039). PMID 4389648 (https://pubmed.ncbi.nlm.nih.gov/4389648). S2CID 86579094 (https://api.semanticscholar.org/CorpusID: 86579094).