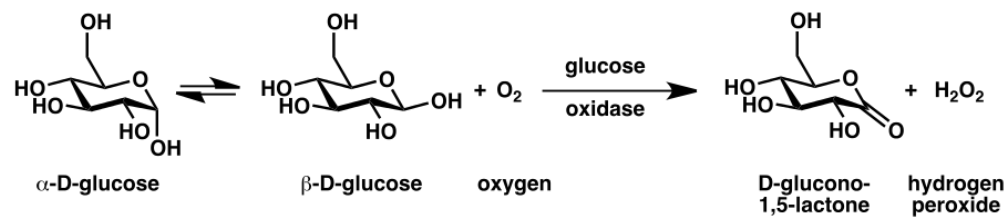


Glucose oxidase

The **glucose oxidase** enzyme (**GOx** or **GOD**) also known as **notatin** (EC number 1.1.3.4) is an oxidoreductase that catalyses the oxidation of glucose to hydrogen peroxide and D-glucono- δ -lactone. This enzyme is produced by certain species of fungi and insects and displays antibacterial activity when oxygen and glucose are present.^[2]



Reaction catalyzed by glucose oxidase

Glucose oxidase is widely used for the determination of free glucose in body fluids (medical testing), in vegetal raw material, and in the food industry. It also has many applications in biotechnologies, typically enzyme assays for biochemistry including biosensors in nanotechnologies.^{[3][4]} It was first isolated by Detlev Müller in 1928 from *Aspergillus niger*.^[5]

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Function

Several species of fungi and insects synthesize glucose oxidase, which produces hydrogen peroxide, which kills bacteria.[2]

Notatin, extracted from antibacterial cultures of *Penicillium notatum*, was originally named **Penicillin A**, but was renamed to avoid confusion with penicillin.^[6] Notatin was shown to be identical to **Penicillin B** and glucose oxidase, enzymes extracted from other molds besides *P. notatum*;^[7] it is now generally known as glucose oxidase.^[8]

Early experiments showed that notatin exhibits *in vitro* antibacterial activity (in the presence of glucose) due to hydrogen peroxide formation.^{[6][9]} *In vivo* tests showed that notatin was not effective in protecting rodents from *Streptococcus haemolyticus*, *Staphylococcus aureus*, or salmonella, and caused severe tissue damage at some doses.^[9]

Glucose oxidase is also produced by the hypopharyngeal glands of honeybee workers and deposited into honey where it acts as a natural preservative. GOx at the surface of the honey reduces atmospheric O₂ to hydrogen peroxide (H₂O₂), which acts as an antimicrobial barrier.^[10]

Structure

GOx is a dimeric protein, the 3D structure of which has been elucidated. The active site where glucose binds is in a deep pocket. The enzyme, like many proteins that act outside of cells, is covered with carbohydrate chains.

Mechanism

At pH 7, glucose exists in solution in cyclic hemiacetal form as 63.6% β -D-glucopyranose and 36.4% α -D-glucopyranose, the proportion of linear and furanose form being negligible. The glucose oxidase binds specifically to β -D-glucopyranose and does not act on α -D-glucose. It oxidises all of the glucose in solution because the equilibrium between the α and β anomers is driven towards the β side as it is consumed in the reaction.^[3]

Glucose oxidase catalyzes the oxidation of β -D-glucose into D-glucono-1,5-lactone, which then hydrolyzes into gluconic acid.

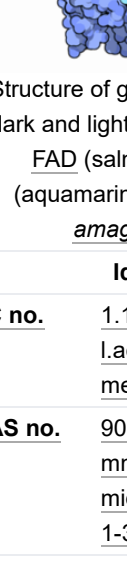
In order to work as a catalyst, GOx requires a coenzyme, flavin adenine dinucleotide (FAD). FAD is a common component in biological oxidation-reduction (redox) reactions. Redox reactions involve a gain or loss of electrons from a molecule. In the GOx-catalyzed redox reaction, FAD works as the initial electron acceptor and is reduced to FADH⁻.^[11] Then FADH⁻ is oxidized by the final electron acceptor, molecular oxygen (O₂), which can do so because it has a higher reduction potential. O₂ is then reduced to hydrogen peroxide (H₂O₂).

Applications

Glucose monitoring

Glucose oxidase is widely used coupled to peroxidase reaction that visualizes colorimetrically the formed H_2O_2 , for the determination of free glucose in sera or blood plasma for diagnostics, using spectrometric assays manually or with automated procedures, and even point-of-use rapid assays.^{[3][8]}

Similar assays allow the monitoring of glucose levels in fermentation, bioreactors, and to control glucose in vegetal raw material and food products. In the glucose oxidase assay, the glucose is first oxidized, catalyzed by glucose oxidase, to produce gluconate and hydrogen peroxide. The hydrogen peroxide is then oxidatively coupled with a chromogen to produce a colored compound which may be measured spectroscopically. For example, hydrogen peroxide together with 4-amino-antipyrine (4-AAP) and phenol in the presence of peroxidase yields a red quinoneimine dye that can be measured at 505 nm. The absorbance at 505 nm is proportional to the concentration of glucose in the sample.

<h1>Glucose oxidase</h1> 	
<p>Structure of glucose oxidase dimer (dark and light blue) complexed with <u>FAD</u> (salmon) and glycans (aquamarine) from <u><i>Penicillium amagasakiense</i></u>.^[1]</p>	
<h2>Identifiers</h2>	
EC no.	1.1.3.4 (https://www.qmul.ac.uk/sbcs/iubmb/enzyme/EC1/1/3/4.html)
CAS no.	9001-37-0 (http://www.commonchemistry.org/ChemicalDetail.aspx?ref=9001-37-0&title=)
<h2>Databases</h2>	
IntEnz	IntEnz view (https://www.ebi.ac.uk/intenz/query?cmd=SearchEC&ec=1.1.3.4)
BRENDA	BRENDA entry (http://www.brenda-enzymes.org/enzyme.php?ecno=1.1.3.4)
ExPASy	NiceZyme view (https://enzyme.expasy.org/EC/1.1.3.4)
KEGG	KEGG entry (https://www.genome.jp/dbget-bin/www_bget?enzyme+1.1.3.4)
MetaCyc	metabolic pathway (https://biocyc.org/META/substring-search?type=NIL&object=1.1.3.4)
PRIAM	profile (http://priam.prabi.fr/cgi-bin/PRIAM_profiles/CurrentRelease.pl?EC=1.1.3.4)
PDB structures	<u>RCSB PDB</u> (https://www.rcsb.org/search?q=rcsb_polymer_entity.rcsb_ec_lineage.id:1.1.3.4) <u>PDBe</u> (https://www.ebi.ac.uk/pdbe/entry/search/index?ec_number:1.1.3.4) <u>PDBsum</u> (https://www.ebi.ac.uk/thornton-srv/databases/cgi-bin/enzymes/GetPage.pl?ec_number=1.1.3.4)
Gene Ontology	<u>AmiGO</u> (http://amigo.geneontology.org/amigo/term/GO:0046562) / <u>QuickGO</u> (https://www.ebi.ac.uk/QuickGO/term/GO:0046562)
<h2>Search</h2>	
PMC	articles (https://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&term=1.1.3.4%5BEC/RN%20Number%5D%20AND%20pubmed%20pmc%20local%5Bsb%5D)
PubMed	articles (https://www.ncbi.nlm.nih.gov/entrez/query.fcgi?db=pubmed&term=1.1.3.4%5BEC/RN%20Number%5D)
NCBI	proteins (https://www.ncbi.nlm.nih.gov/protein?term=1.1.3.4%5BEC/RN%20Number%5D)

