Nuclear magnetic resonance

Nuclear magnetic resonance (NMR) is physical phenomenon in which nuclei in a strong constant magnetic field are perturbed by a weak oscillating magnetic field (in the near field[1]) and respond by producing an electromagnetic signal with a frequency characteristic of the magnetic field at the nucleus. This process occurs near resonance, when the oscillation frequency matches the intrinsic frequency of the nuclei, which depends on the strength of the static magnetic field, the chemical environment, and the magnetic properties of the isotope involved; in practical applications with static magnetic fields up to ca. 20 tesla, the frequency is similar to VHF and UHF television broadcasts (60–1000 MHz). NMR results from specific magnetic properties of certain atomic nuclei. Nuclear magnetic resonance spectroscopy is widely used to determine the structure of organic molecules in solution and study molecular physics and crystals as well as non-crystalline materials. NMR is also routinely used in advanced medical imaging techniques, such as in magnetic resonance imaging (MRI).



Bruker 700 MHz nuclear magnetic resonance (NMR) spectrometer.

The most commonly used nuclei are ¹H and ¹³C, although isotopes of many other elements can be studied by high-field NMR spectroscopy as well. In order to interact with the magnetic field in the spectrometer, the nucleus must have an intrinsic <u>nuclear magnetic moment</u> and <u>angular momentum</u>. This occurs when an isotope has a nonzero <u>nuclear spin</u>, meaning an odd number of <u>protons</u> and/or <u>neutrons</u> (see <u>Isotope</u>). <u>Nuclides</u> with even numbers of both have a total spin of zero and are therefore NMR-inactive.

A key feature of NMR is that the <u>resonance</u> frequency of a particular sample substance is usually directly proportional to the strength of the applied magnetic field. It is this feature that is exploited in imaging techniques; if a sample is placed in a non-uniform magnetic field then the resonance frequencies of the sample's nuclei depend on where in the field they are located. Since the resolution of the imaging technique depends on the magnitude of the magnetic field <u>gradient</u>, many efforts are made to develop increased gradient field strength.

The principle of NMR usually involves three sequential steps:

- The alignment (polarization) of the magnetic nuclear spins in an applied, constant <u>magnetic field</u>
 B₀.
- The perturbation of this alignment of the nuclear spins by a weak oscillating magnetic field, usually referred to as a radio-frequency (RF) pulse. The oscillation frequency required for significant perturbation is dependent upon the static magnetic field (**B**₀) and the nuclei of observation.
- The detection of the NMR signal during or after the RF pulse, due to the voltage induced in a detection coil by precession of the nuclear spins around **B**₀. After an RF pulse, precession usually occurs with the nuclei's intrinsic <u>Larmor frequency</u> and, in itself, does not involve transitions between spin states or energy levels. [1]

The two magnetic fields are usually chosen to be perpendicular to each other as this maximizes the NMR signal strength. The frequencies of the time-signal response by the total magnetization (\mathbf{M}) of the nuclear spins are analyzed in NMR spectroscopy and magnetic resonance imaging. Both use applied magnetic fields (\mathbf{B}_0) of great strength, often produced by large currents in superconducting coils, in order to achieve dispersion of response frequencies and of very high homogeneity and stability in order to deliver spectral resolution, the details of which are described by chemical shifts, the Zeeman effect, and Knight shifts (in metals). The information provided by NMR can also be increased using hyperpolarization, and/or using two-dimensional, three-dimensional and higher-dimensional techniques.

NMR phenomena are also utilized in <u>low-field NMR</u>, NMR spectroscopy and MRI in the Earth's magnetic field (referred to as Earth's field NMR), and in several types of magnetometers.

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History

Nuclear magnetic resonance was first described and measured in molecular beams by <u>Isidor Rabi</u> in 1938, by extending the Stern–Gerlach experiment, and in 1944, Rabi was awarded the <u>Nobel Prize in Physics</u> for this work. In 1946, <u>Felix Bloch</u> and <u>Edward Mills Purcell</u> expanded the technique for use on liquids and solids, for which they shared the <u>Nobel Prize</u> in <u>Physics</u> in 1952. [4][5]

Yevgeny Zavoisky likely observed nuclear magnetic resonance in 1941, well before Felix Bloch and Edward Mills Purcell, but dismissed the results as not reproducible.

Russell H. Varian filed the "Method and means for correlating nuclear properties of atoms and magnetic fields", U.S. Patent 2,561,490 (https://patents.google.com/patent/US2561490) on July 24, 1951. Varian Associates developed the first NMR unit called NMR HR-30 in 1952. [6]

Purcell had worked on the development of <u>radar</u> during World War II at the <u>Massachusetts Institute</u> of <u>Technology</u>'s <u>Radiation Laboratory</u>. His work during that project on the production and detection of <u>radio frequency power</u> and on the absorption of such RF power by matter laid the foundation for his discovery of NMR in bulk matter.

Rabi, Bloch, and Purcell observed that magnetic nuclei, like $^1\!\underline{H}$ and $^{31}\!\underline{P}$, could absorb RF energy when placed in a magnetic field and when the RF was of a frequency specific to the identity of the nuclei. When this absorption occurs, the nucleus is described as being *in resonance*. Different atomic nuclei within a molecule resonate at different (radio) frequencies for the same magnetic field strength. The observation of such magnetic resonance frequencies of the nuclei present in a molecule allows any trained user to discover essential chemical and structural information about the molecule.

The development of NMR as a technique in <u>analytical chemistry</u> and <u>biochemistry</u> parallels the development of electromagnetic technology and advanced <u>electronics</u> and their introduction into civilian use.

Theory of nuclear magnetic resonance

Nuclear spin and magnets

All nucleons, that is <u>neutrons</u> and <u>protons</u>, composing any atomic <u>nucleus</u>, have the intrinsic quantum property of <u>spin</u>, an intrinsic angular momentum analogous to the classical angular momentum of a spinning sphere. The overall spin of the nucleus is determined by the <u>spin quantum number S</u>. If the numbers of both the protons and neutrons in a given <u>nuclide</u> are even then S = 0, i.e. there is no overall spin. Then, just as electrons pair up in nondegenerate <u>atomic orbitals</u>, so do even numbers of protons or even numbers of neutrons (both of which are also <u>spin $\frac{1}{2}$ </u> particles and hence fermions), giving zero overall spin.

However, a proton and neutron will have lower energy when their spins are parallel, not antiparallel. This parallel spin alignment of distinguishable particles does not violate the Pauli exclusion principle. The lowering of energy for parallel spins has to do with the quark structure of these two nucleons. As a result, the spin ground state for the deuteron (the nucleus of deuterium, the 2 H isotope of hydrogen), which has only a proton and a neutron, corresponds to a spin value of $\mathbf{1}$, not of zero. On the other hand, because of the Pauli exclusion principle, the tritium isotope of hydrogen must have a pair of anti-parallel spin neutrons (of total spin zero for the neutron-spin pair), plus a proton of spin $\frac{1}{2}$. Therefore, the tritium total nuclear spin value is again $\frac{1}{2}$, just like for the simpler, abundant hydrogen isotope, 1 H nucleus (the *proton*). The NMR absorption frequency for tritium is also similar to that of 1 H. In many other cases of *non-radioactive* nuclei, the overall spin is also non-zero. For example, the 27 Al nucleus has an overall spin value $S = \frac{5}{2}$.

A non-zero spin \vec{S} is always associated with a non-zero magnetic dipole moment, $\vec{\mu}$, via the relation

$$ec{\mu}=\gammaec{S}$$

where γ is the gyromagnetic ratio. Classically, this corresponds to the proportionality between the angular momentum and the magnetic dipole moment of a spinning charged sphere, both of which are vectors parallel to the rotation axis whose length increases proportional to the spinning frequency. It is the magnetic moment and its interaction with magnetic fields that allows the observation of NMR signal associated with transitions between nuclear spin levels during resonant RF irradiation or caused by Larmor precession of the average magnetic moment after resonant irradiation. Nuclides with even numbers of both protons and neutrons have zero <u>nuclear magnetic dipole moment</u> and hence do not exhibit NMR signal. For instance, ${}^{18}O$ is an example of a nuclide that produces no NMR signal, whereas ${}^{13}C$, ${}^{31}P$, ${}^{35}Cl$ and ${}^{37}Cl$ are nuclides that do exhibit NMR spectra. The last two nuclei have spin S > 1/2 and are therefore quadrupolar nuclei.

Electron spin resonance (ESR) is a related technique in which transitions between electronic rather than nuclear spin levels are detected. The basic principles are similar but the instrumentation, data analysis, and detailed theory are significantly different. Moreover, there is a much smaller number of molecules and materials with unpaired electron spins that exhibit ESR (or electron paramagnetic resonance (EPR)) absorption than those that have NMR absorption spectra. On the other hand, ESR has much higher signal per spin than NMR does.

Values of spin angular momentum

Nuclear <u>spin</u> is an intrinsic <u>angular momentum</u> that is quantized. This means that the magnitude of this angular momentum is quantized (i.e. S can only take on a restricted range of values), and also that the x, y, and z-components of the angular momentum are quantized, being restricted to integer or half-integer multiples of \hbar . The integer or half-integer quantum number associated with the spin component along the z-axis or the applied magnetic field is known as the <u>magnetic quantum number</u>, m, and can take values from +S to -S, in integer steps. Hence for any given nucleus, there are a total of 2S + 1 angular momentum states.

The z-component of the angular momentum vector (\vec{S}) is therefore $S_z = m\hbar$, where \hbar is the reduced Planck constant. The z-component of the magnetic moment is simply:

$$\mu_z = \gamma S_z = \gamma m \hbar.$$

Spin energy in a magnetic field

Consider nuclei with a spin of one-half, like $\frac{^{1}}{H}$, $\frac{^{13}}{C}$ or $\frac{^{19}}{F}$. Each nucleus has two linearly independent spin states, with $m = \frac{1}{2}$ or $m = -\frac{1}{2}$ (also referred to as spin-up and spin-down, or sometimes α and β spin states, respectively) for the z-component of spin. In the absence of a magnetic field, these states are degenerate; that is, they have the same energy. Hence the number of nuclei in these two states will be essentially equal at thermal equilibrium.

If a nucleus is placed in a magnetic field, however, the two states no longer have the same energy as a result of the interaction between the nuclear magnetic dipole moment and the external magnetic field. The energy of a magnetic dipole moment $\vec{\mu}$ in a magnetic field \mathbf{B}_0 is given by:

$$E = -\vec{\mu} \cdot \mathbf{B}_0 = -\mu_x B_{0x} - \mu_y B_{0y} - \mu_z B_{0z}.$$

Usually the z-axis is chosen to be along \mathbf{B}_0 , and the above expression reduces to:

$$E=-\mu_{\rm z}B_0$$
,

or alternatively:

$$E=-\gamma m\hbar B_0$$
 .

As a result, the different nuclear spin states have different energies in a non-zero magnetic field. In less formal language, we can talk about the two spin states of a spin $\frac{1}{2}$ as being *aligned* either with or against the magnetic field. If γ is positive (true for most isotopes used in NMR) then $m = \frac{1}{2}$ is the lower energy state.

The energy difference between the two states is:

$$\Delta E = \gamma \hbar B_0$$
 ,

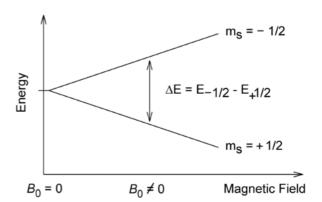
and this results in a small population bias favoring the lower energy state in thermal equilibrium. With more spins pointing up than down, a net spin magnetization along the magnetic field \mathbf{B}_0 results.

Precession of the spin magnetization

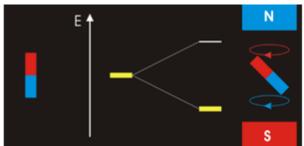
A central concept in NMR is the precession of the spin magnetization around the magnetic field at the nucleus, with the angular frequency

$$\omega = -\gamma B$$

where $\omega = 2\pi\nu$ relates to the oscillation frequency ν and B is the magnitude of the field. This means that the spin magnetization, which is proportional to the sum of the spin vectors of nuclei in magnetically equivalent sites (the expectation value of the spin vector in quantum mechanics), moves on a cone around the **B** field. This is analogous to the precessional motion of the axis of a tilted



Splitting of nuclei spin energies in an external magnetic field



An intuitive model. Nuclei with spin have magnetic moments (spin magnetic moments). By itself, there is no energetic difference for any particular orientation of the nuclear magnet (only one energy state, on the left), but in an external magnetic field there is a high-energy state and a low-energy state depending on the relative orientation of the magnet to the external field, and in thermal equilibrium, the low-energy orientation is preferred. The average orientation of the magnetic moment will precess around the field. The external field can be supplied by a large magnet and also by other nuclei in the vicinity.

spinning top around the gravitational field. In quantum mechanics, ω is the Bohr frequency $\Delta E/\hbar$ of the S_x and S_y expectation values. Precession of non-equilibrium magnetization in the applied magnetic field \mathbf{B}_0 occurs with the Larmor frequency

$$\omega_L = 2\pi\nu_L = -\gamma B_0$$

without change in the populations of the energy levels because energy is constant (time-independent Hamiltonian).^[8]

Magnetic resonance and radio-frequency pulses

A perturbation of nuclear spin orientations from equilibrium will occur only when an oscillating magnetic field is applied whose frequency $v_{\rm rf}$ sufficiently closely matches the Larmor precession frequency $v_{\rm L}$ of the nuclear magnetization. The populations of the spin-up and -down energy levels then undergo Rabi oscillations, which are analyzed most easily in terms of precession of the spin magnetization around the effective magnetic field in a reference frame rotating with the frequency $v_{\rm rf}$. The stronger the oscillating field, the faster the Rabi oscillations or the precession around the effective field in the rotating frame. After a certain time on the order of 2–1000 microseconds, a resonant RF pulse flips the spin magnetization to the transverse plane, i.e. it makes an angle of 90° with the constant magnetic field \mathbf{B}_0 ("90° pulse"), while after a twice longer time, the initial magnetization has been inverted ("180° pulse"). It is the transverse magnetization generated by a resonant oscillating field which is usually detected in NMR, during application of the relatively weak RF field in old-fashioned continuous-wave NMR, or after the relatively strong RF pulse in modern pulsed NMR.

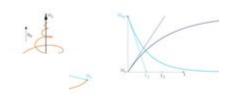
Chemical shielding

It might appear from the above that all nuclei of the same nuclide (and hence the same γ) would resonate at exactly the same frequency. This is not the case. The most important perturbation of the NMR frequency for applications of NMR is the "shielding" effect of the surrounding shells of electrons. [10] Electrons, similar to the nucleus, are also charged and rotate with a spin to produce a magnetic field opposite to the applied magnetic field. In general, this electronic shielding reduces the magnetic field at the nucleus (which is what determines the NMR frequency). As a result, the frequency required to achieve resonance is also reduced. This shift in the NMR frequency due to the electronic molecular orbital coupling to the external magnetic field is called chemical shift, and it explains why NMR is able to probe the chemical structure of molecules, which depends on the electron density distribution in the corresponding molecular orbitals. If a nucleus in a specific chemical group is shielded to a higher degree by a higher electron density of its surrounding molecular orbital, then its NMR frequency will be shifted "upfield" (that is, a lower chemical shift), whereas if it is less shielded by such surrounding electron density, then its NMR frequency will be shifted "downfield" (that is, a higher chemical shift).

Unless the local symmetry of such molecular orbitals is very high (leading to "isotropic" shift), the shielding effect will depend on the orientation of the molecule with respect to the external field (\mathbf{B}_0). In solid-state NMR spectroscopy, magic angle spinning is required to average out this orientation dependence in order to obtain frequency values at the average or isotropic chemical shifts. This is unnecessary in conventional NMR investigations of molecules in solution, since rapid "molecular tumbling" averages out the chemical shift anisotropy (CSA). In this case, the "average" chemical shift (ACS) or isotropic chemical shift is often simply referred to as the chemical shift.

Relaxation

The process of population relaxation refers to nuclear spins that return to thermodynamic equilibrium in the magnet. This process is also called $\underline{T_1}$, "spin-lattice" or "longitudinal magnetic" relaxation, where T_1 refers to the mean time for an individual nucleus to return to its thermal equilibrium state of the spins. After the nuclear spin population has relaxed, it can be probed again, since it is in the initial, equilibrium (mixed) state.

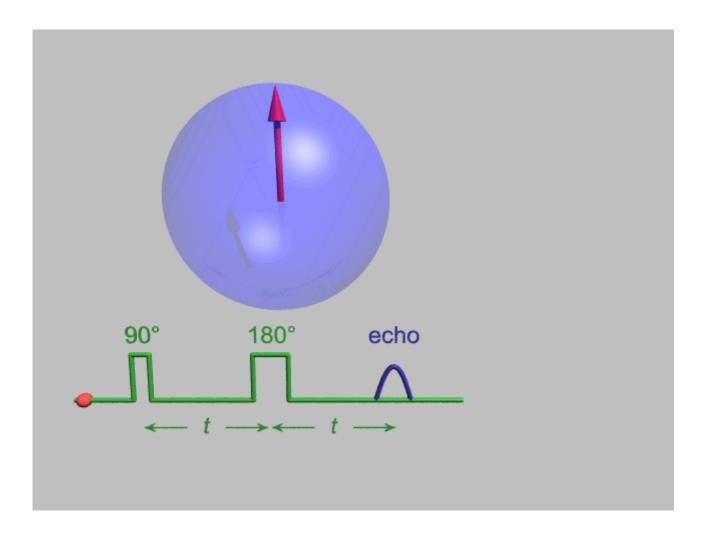


Visualization of the T_1 and T_2 relaxation times.

The precessing nuclei can also fall out of alignment with each other and gradually stop producing a signal. This is called T_2 or

transverse relaxation. Because of the difference in the actual relaxation mechanisms involved (for example, intermolecular versus intramolecular magnetic dipole-dipole interactions), T_1 is usually (except in rare cases) longer than T_2 (that is, slower spin-lattice relaxation, for example because of smaller dipole-dipole interaction effects). In practice, the value of T_2^* which is the actually observed decay time of the observed NMR signal, or free induction decay (to $\frac{1}{e}$ of the initial amplitude immediately after the resonant RF pulse), also depends on the static magnetic field inhomogeneity, which is quite significant. (There is also a smaller but significant contribution to the observed FID shortening from the RF inhomogeneity of the resonant pulse). In the corresponding FT-NMR spectrum—meaning the Fourier transform of the free induction decay—the T_2^* time is inversely related to the width of the NMR signal in frequency units. Thus, a nucleus with a long T_2 relaxation time gives rise to a very sharp NMR peak in the FT-NMR spectrum for a very homogeneous ("well-shimmed") static magnetic field, whereas nuclei with shorter T_2 values give rise to broad FT-NMR peaks even when the magnet is shimmed well. Both T_1 and T_2 depend on the rate of molecular motions as well as the gyromagnetic ratios of both the resonating and their strongly interacting, next-neighbor nuclei that are not at resonance.

A <u>Hahn echo</u> decay experiment can be used to measure the dephasing time, as shown in the animation below. The size of the echo is recorded for different spacings of the two pulses. This reveals the decoherence which is not refocused by the 180° pulse. In simple cases, an <u>exponential decay</u> is measured which is described by the T_2 time.



NMR spectroscopy

NMR spectroscopy is one of the principal techniques used to obtain physical, chemical, electronic and structural information about molecules due to the chemical shift of the resonance frequencies of the nuclear spins in the sample. Peak splittings due to J- or dipolar couplings between nuclei are also useful. NMR spectroscopy can provide detailed and quantitative information on the functional groups, topology, dynamics and three-dimensional structure of molecules in solution and the solid state. Since the area under an NMR peak is usually proportional to the number of spins involved, peak integrals can be used to determine composition quantitatively.

Structure and molecular dynamics can be studied (with or without "magic angle" spinning (MAS)) by NMR of quadrupolar nuclei (that is, with spin $S > \frac{1}{2}$) even in the presence of magnetic "dipole-dipole" interaction broadening (or simply, dipolar broadening) which is always much smaller than the quadrupolar interaction strength because it is a magnetic vs. an electric interaction effect.



900 MHz, 21.2 T NMR Magnet at HWB-NMR, Birmingham, UK

Additional structural and chemical information may be obtained by performing double-quantum NMR experiments for pairs of spins or quadrupolar nuclei such as ${}^{2}H$. Furthermore, nuclear magnetic resonance is one of the techniques that has been used to design quantum automata, and also build elementary quantum computers. [11][12]

Continuous-wave (CW) spectroscopy

In the first few decades of nuclear magnetic resonance, spectrometers used a technique known as continuous-wave (CW) spectroscopy, where the transverse spin magnetization generated by a weak oscillating magnetic field is recorded as a function of the oscillation frequency or static field strength B_0 . When the oscillation frequency matches the nuclear resonance frequency, the transverse magnetization is maximized and a peak is observed in the spectrum. Although NMR spectra could be, and have been, obtained using a fixed constant magnetic field and sweeping the frequency of the oscillating magnetic field, it was more convenient to use a fixed frequency source and vary the current (and hence magnetic field) in an electromagnet to observe the resonant absorption signals. This is the origin of the counterintuitive, but still common, "high field" and "low field" terminology for low frequency and high frequency regions, respectively, of the NMR spectrum.

As of 1996, CW instruments were still used for routine work because the older instruments were cheaper to maintain and operate, often operating at 60 MHz with correspondingly weaker (non-superconducting) electromagnets cooled with water rather than liquid helium. One radio coil operated continuously, sweeping through a range of frequencies, while another <u>orthogonal</u> coil, designed not to receive radiation from the transmitter, received signals from nuclei that reoriented in solution. As of 2014, low-end refurbished 60 MHz and 90 MHz systems were sold as FT-NMR instruments, and in 2010 the "average workhorse" NMR instrument was configured for 300 MHz.

CW spectroscopy is inefficient in comparison with Fourier analysis techniques (see below) since it probes the NMR response at individual frequencies or field strengths in succession. Since the NMR signal is intrinsically weak, the observed spectrum suffers from a poor signal-to-noise ratio. This can be mitigated by signal averaging, i.e. adding the spectra from repeated measurements. While the NMR signal is the same in each scan and so adds linearly, the random noise adds more slowly – proportional to the square root of the number of spectra (see random walk). Hence the overall signal-to-noise ratio increases as the square-root of the number of spectra measured.

Fourier-transform spectroscopy

Most applications of NMR involve full NMR spectra, that is, the intensity of the NMR signal as a function of frequency. Early attempts to acquire the NMR spectrum more efficiently than simple CW methods involved illuminating the target simultaneously with more than one frequency. A revolution in NMR occurred when short radio-frequency pulses began to be used, with a frequency centered at the middle of the NMR spectrum. In simple terms, a short pulse of a given "carrier" frequency "contains" a range of frequencies centered about the carrier frequency, with the range of excitation (bandwidth) being inversely proportional to the pulse duration, i.e. the Fourier transform of a short pulse contains contributions from all the frequencies in the neighborhood of the principal frequency. The restricted range of the NMR frequencies made it relatively easy to use short (1 - 100 microsecond) radio frequency pulses to excite the entire NMR spectrum.

Applying such a pulse to a set of nuclear spins simultaneously excites all the single-quantum NMR transitions. In terms of the net magnetization vector, this corresponds to tilting the magnetization vector away from its equilibrium position (aligned along the external magnetic field). The out-of-equilibrium magnetization vector then precesses about the external magnetic field vector at the NMR frequency of the spins. This oscillating magnetization vector induces a voltage in a nearby pickup coil, creating an electrical signal oscillating at the NMR frequency. This signal is known as the free induction decay (FID), and it contains the sum of the NMR responses from all the excited spins. In order to obtain the frequency-domain NMR spectrum (NMR absorption intensity vs. NMR frequency) this time-domain signal (intensity vs. time) must be Fourier transformed. Fortunately, the

development of Fourier transform (FT) NMR coincided with the development of <u>digital computers</u> and the digital <u>Fast Fourier Transform</u>. Fourier methods can be applied to many types of spectroscopy. (See the full article on Fourier transform spectroscopy.)

Richard R. Ernst was one of the pioneers of pulsed NMR and won a Nobel Prize in chemistry in 1991 for his work on Fourier Transform NMR and his development of multi-dimensional NMR spectroscopy.

Multi-dimensional NMR spectroscopy

The use of pulses of different durations, frequencies, or shapes in specifically designed patterns or *pulse sequences* allows the NMR spectroscopist to extract many different types of information about the molecules in the sample. In multi-dimensional nuclear magnetic resonance spectroscopy, there are at least two pulses and, as the experiment is repeated, the pulse timings are systematically varied and the oscillations of the spin system are probed point by point in the time domain. Multidimensional Fourier transformation of the multidimensional time signal yields the multidimensional spectrum. In two-dimensional nuclear magnetic resonance there will be one systematically varied time period in the sequence of pulses, which will modulate the intensity or phase of the detected signals. In 3D NMR, two time periods will be varied independently, and in 4D NMR, three will be varied. The remaining "dimension" is always provided by the directly detected signal.

There are many such experiments. In some, fixed time intervals allow (among other things) magnetization transfer between nuclei and, therefore, the detection of the kinds of nuclear-nuclear interactions that allowed for the magnetization transfer. Interactions that can be detected are usually classified into two kinds. There are *through-bond* and *through-space* interactions, the latter being a consequence of dipolar couplings in solid-state NMR and of the <u>nuclear Overhauser effect</u> in solution NMR. Experiments of the nuclear Overhauser variety may be employed to establish distances between atoms, as for example by 2D-FT NMR of molecules in solution.

Although the fundamental concept of 2D-FT NMR was proposed by <u>Jean Jeener</u> from the <u>Free University of Brussels</u> at an international conference, this idea was largely developed by <u>Richard Ernst</u> who won the 1991 <u>Nobel prize in Chemistry</u> for his work in FT NMR, including multi-dimensional FT NMR, and especially 2D-FT NMR of small molecules. [17] Multi-dimensional FT NMR experiments were then further developed into powerful methodologies for studying molecules in solution, in particular for the determination of the structure of <u>biopolymers</u> such as <u>proteins</u> or even small nucleic acids. [18]

In 2002 <u>Kurt Wüthrich</u> shared the <u>Nobel Prize</u> in <u>Chemistry</u> (with <u>John Bennett Fenn</u> and <u>Koichi</u> Tanaka) for his work with protein FT NMR in solution.

Solid-state NMR spectroscopy

This technique complements X-ray crystallography in that it is frequently applicable to molecules in an amorphous or <u>liquid-crystalline</u> state, whereas crystallography, as the name implies, is performed on molecules in a crystalline phase. In electronically conductive materials, the <u>Knight shift</u> of the resonance frequency can provide information on the mobile charge carriers. Though nuclear magnetic resonance is used to study the structure of solids, extensive atomic-level structural detail is more challenging to obtain in the solid state. Due to broadening by <u>chemical shift anisotropy</u> (CSA) and dipolar couplings to other nuclear spins, without special techniques such as <u>MAS</u> or dipolar decoupling by RF pulses, the observed spectrum is often only a broad Gaussian band for non-quadrupolar spins in a solid.

Professor Raymond Andrew at the <u>University</u> of Nottingham in the UK pioneered the development of high-resolution <u>solid-state nuclear magnetic resonance</u>. He was the first to report the introduction of the <u>MAS</u> (magic angle sample spinning; MASS) technique that allowed him to achieve spectral resolution in solids sufficient to distinguish between chemical groups with either different chemical shifts or distinct <u>Knight shifts</u>. In MASS, the sample is spun at several kilohertz around an axis that makes the so-called <u>magic angle</u> $\theta_{\rm m}$ (which is ~54.74°, where $3\cos^2\theta_{\rm m}$ -1 = 0) with respect to the direction of the static magnetic field **B**₀; as a result of such magic angle sample spinning, the broad chemical shift anisotropy bands are averaged to their corresponding average (isotropic) chemical shift values. Correct alignment of the sample rotation axis as close as possible to $\theta_{\rm m}$ is essential for cancelling out the chemical-shift anisotropy broadening. There are different angles for the sample spinning relative to the applied field for the averaging of electric quadrupole interactions and paramagnetic interactions, correspondingly ~30.6° and ~70.1°. In amorphous materials, residual line broadening remains since each segment is in a slightly different environment, therefore exhibiting a slightly different NMR frequency.

Dipolar and J-couplings to nearby ¹H nuclei are usually removed by radio-frequency pulses applied at the ¹H frequency during signal detection. The concept of cross polarization developed by Sven Hartmann and Erwin Hahn was utilized in transferring magnetization from protons to less sensitive nuclei by M.G. Gibby, Alex Pines and John S. Waugh. Then, Jake Schaefer and Ed Stejskal demonstrated the powerful use of cross polarization under MAS conditions (CP-MAS) and proton decoupling, which is now routinely employed to measure high resolution spectra of low-abundance and low-sensitivity nuclei, such as carbon-13, silicon-29, or nitrogen-15, in solids. Significant further signal enhancement can be achieved by dynamic nuclear polarization from unpaired electrons to the nuclei, usually at temperatures near 110 K.

Sensitivity

Because the intensity of nuclear magnetic resonance signals and, hence, the sensitivity of the technique depends on the strength of the magnetic field the technique has also advanced over the decades with the development of more powerful magnets. Advances made in audio-visual technology have also improved the signal-generation and processing capabilities of newer instruments.

As noted above, the sensitivity of nuclear magnetic resonance signals is also dependent on the presence of a magnetically susceptible nuclide and, therefore, either on the natural abundance of such nuclides or on the ability of the experimentalist to artificially enrich the molecules, under study, with such nuclides. The most abundant naturally occurring isotopes of hydrogen and phosphorus (for example) are both magnetically susceptible and readily useful for nuclear magnetic resonance spectroscopy. In contrast, carbon and nitrogen have useful isotopes but which occur only in very low natural abundance.

Other limitations on sensitivity arise from the quantum-mechanical nature of the phenomenon. For quantum states separated by energy equivalent to radio frequencies, thermal energy from the environment causes the populations of the states to be close to equal. Since incoming radiation is equally likely to cause stimulated emission (a transition from the upper to the lower state) as absorption, the NMR effect depends on an excess of nuclei in the lower states. Several factors can reduce sensitivity, including:

- Increasing temperature, which evens out the population of states. Conversely, low temperature NMR can sometimes yield better results than room-temperature NMR, providing the sample remains liquid.
- Saturation of the sample with energy applied at the resonant radiofrequency. This manifests in both CW and pulsed NMR; in the first case (CW) this happens by using too much continuous power that keeps the upper spin levels completely populated; in the second case (pulsed), each pulse (that is at least a 90° pulse) leaves the sample saturated, and four to five times the

(longitudinal) relaxation time ($5T_1$) must pass before the next pulse or pulse sequence can be applied. For single pulse experiments, shorter RF pulses that tip the magnetization by less than 90° can be used, which loses some intensity of the signal, but allows for shorter *recycle delays*. The optimum there is called an *Ernst angle*, after the Nobel laureate. Especially in solid state NMR, or in samples containing very few nuclei with spin (diamond with the natural 1% of carbon-13 is especially troublesome here) the longitudinal relaxation times can be on the range of hours, while for proton-NMR they are more in the range of one second.

Non-magnetic effects, such as electric-<u>quadrupole</u> coupling of spin-1 and spin- $\frac{3}{2}$ nuclei with their local environment, which broaden and weaken absorption peaks. $\frac{14}{N}$, an abundant spin-1 nucleus, is difficult to study for this reason. High resolution NMR instead probes molecules using the rarer $\frac{15}{N}$ isotope, which has spin- $\frac{1}{2}$.

Isotopes

Many isotopes of chemical elements can be used for NMR analysis. [19]

Commonly used nuclei:

- 1H, the most commonly used spin-1/2 nucleus in NMR investigation, has been studied using many forms of NMR. Hydrogen is highly abundant, especially in biological systems. It is the nucleus most sensitive to NMR signal (apart from 3H which is not commonly used due to its instability and radioactivity). Proton NMR produces narrow chemical shift with sharp signals. Fast acquisition of quantitative results (peak integrals in stoichiometric ratio) is possible due to short relaxation time. The 1H signal has been the sole diagnostic nucleus used for clinical magnetic resonance imaging (MRI).
- ²H, a spin 1 nucleus commonly utilized as signal-free medium in the form of <u>deuterated solvents</u> during proton NMR, to avoid signal interference from hydrogen-containing solvents in measurement of ¹H solutes. Also used in determining the behavior of lipids in lipid membranes and other solids or liquid crystals as it is a relatively non-perturbing label which can selectively replace ¹H. Alternatively, ²H can be detected in media specially labeled with ²H. Deuterium resonance is commonly used in high-resolution <u>NMR spectroscopy</u> to monitor drifts in the magnetic field strength (lock) and to improve the homogeneity of the external magnetic field.
- 3He, is very sensitive to NMR. There is a very low percentage in natural helium, and subsequently has to be purified from 4He. It is used mainly in studies of endohedral fullerenes, where its chemical inertness is beneficial to ascertaining the structure of the entrapping fullerene.
- $\frac{^{11}B}{^{11}B}$, more sensitive than $\frac{^{10}B}{^{11}B}$, yields sharper signals. The nuclear spin of ^{10}B is 3 and that of ^{11}B is $\frac{3}{2}$. Quartz tubes must be used because <u>borosilicate</u> glass interferes with measurement.
- 13 C spin-1/2, is widely used, despite its relative paucity in naturally occurring carbon (approximately 1.1%). It is stable to nuclear decay. Since there is a low percentage in natural carbon, spectrum acquisition on samples which have not been experimentally enriched in 13 C takes a long time. Frequently used for labeling of compounds in synthetic and metabolic studies. Has low sensitivity and moderately wide chemical shift range, yields sharp signals. Low percentage makes it useful by preventing spin-spin couplings and makes the spectrum appear less crowded. Slow relaxation means that spectra are not integrable unless long acquisition times are used.
- ¹⁴N, spin-1, medium sensitivity nucleus with wide chemical shift. Its large <u>quadrupole</u> moment interferes in acquisition of high resolution spectra, limiting usefulness to smaller molecules and functional groups with a high degree of symmetry such as the headgroups of lipids.

- $\frac{15}{N}$, spin- $\frac{1}{2}$, relatively commonly used. Can be used for labeling compounds. Nucleus very insensitive but yields sharp signals. Low percentage in natural nitrogen together with low sensitivity requires high concentrations or expensive isotope enrichment.
- $\frac{17}{O}$, spin- $\frac{5}{2}$, low sensitivity and very low natural abundance (0.037%), wide chemical shift range (up to 2000 ppm). Quadrupole moment causing line broadening. Used in metabolic and biochemical studies in studies of chemical equilibria.
- $\frac{^{19}\text{F}}{^{2}}$, spin- $\frac{1}{2}$, relatively commonly measured. Sensitive, yields sharp signals, has a wide chemical shift range.
- $\frac{31}{2}$ P, spin- $\frac{1}{2}$, 100% of natural phosphorus. Medium sensitivity, wide chemical shifts range, yields sharp lines. Spectra tend to have a moderate amount of noise. Used in biochemical studies and in coordination chemistry where phosphorus containing ligands are involved.
- ³⁵Cl and ³⁷Cl, broad signal. ³⁵Cl is significantly more sensitive, preferred over ³⁷Cl despite its slightly broader signal. Organic chlorides yield very broad signals. Its use is limited to inorganic and ionic chlorides and very small organic molecules.
- 43 Ca, used in biochemistry to study calcium binding to DNA, proteins, etc. Moderately sensitive, very low natural abundance.
- 195 Pt, used in studies of catalysts and complexes.

Other nuclei (usually used in the studies of their complexes and chemical bonding, or to detect presence of the element):

⁵⁵Mn

⁵⁷F<u>e</u>

⁶¹Ni

Co

'Сu,

Applications

NMR is extensively used in medicine in the form of <u>magnetic resonance imaging</u>. NMR is used industrially mainly for routine analysis of chemicals. The technique is also used, for example, to measure the ratio between water and fat in foods, monitor the flow of corrosive fluids in pipes, or to study molecular structures such as catalysts. [20]

Medicine

The application of nuclear magnetic resonance best known to the general public is magnetic resonance imaging for medical diagnosis and magnetic resonance microscopy in research settings. However, it is also widely used in biochemical studies, notably in NMR spectroscopy such as proton NMR, carbon-13 NMR, deuterium NMR and phosphorus-31 NMR. Biochemical information can also be obtained from living tissue (e.g. human brain tumors) with the technique known as in vivo magnetic resonance spectroscopy or chemical shift NMR microscopy.

These spectroscopic studies are possible because nuclei are surrounded by orbiting electrons, which are charged particles that generate small, local magnetic fields that add to or subtract from the external magnetic field, and so will partially shield the



Medical MRI

nuclei. The amount of shielding depends on the exact local environment. For example, a hydrogen bonded to an <u>oxygen</u> will be shielded differently from a hydrogen bonded to a carbon atom. In addition, two hydrogen nuclei can interact via a process known as <u>spin-spin coupling</u>, if they are on the same molecule, which will split the lines of the spectra in a recognizable way.

As one of the two major spectroscopic techniques used in metabolomics, NMR is used to generate metabolic fingerprints from biological fluids to obtain information about disease states or toxic insults.

Chemistry

By studying the peaks of nuclear magnetic resonance spectra, chemists can determine the structure of many compounds. It can be a very selective technique, distinguishing among many atoms within a molecule or collection of molecules of the same type but which differ only in terms of their local chemical environment. NMR spectroscopy is used to unambiguously identify known and novel compounds, and as such, is usually required by scientific journals for identity confirmation of synthesized new compounds. See the articles on <u>carbon-13 NMR</u> and <u>proton NMR</u> for detailed discussions.

A chemist can determine the identity of a compound by comparing the observed nuclear precession frequencies to known frequencies. Further structural data can be <u>elucidated</u> by observing <u>spin-spin</u> <u>coupling</u>, a process by which the precession frequency of a nucleus can be influenced by the spin orientation of a chemically bonded nucleus. Spin-spin coupling is easily observed in NMR of hydrogen-1 (¹H NMR) since its natural abundance is nearly 100%.

Because the nuclear magnetic resonance timescale is rather slow, compared to other spectroscopic methods, changing the temperature of a T_2^* experiment can also give information about fast reactions, such as the <u>Cope rearrangement</u> or about structural dynamics, such as ring-flipping in <u>cyclohexane</u>. At low enough temperatures, a distinction can be made between the axial and equatorial hydrogens in cyclohexane.

An example of nuclear magnetic resonance being used in the determination of a structure is that of buckminsterfullerene (often called "buckyballs", composition C_{60}). This now famous form of carbon has 60 carbon atoms forming a sphere. The carbon atoms are all in identical environments and so should see the same internal **H** field. Unfortunately, buckminsterfullerene contains no hydrogen and so 13 C nuclear magnetic resonance has to be used. 13 C spectra require longer acquisition times since carbon-13 is not the common isotope of carbon (unlike hydrogen, where 1 H is the common isotope). However, in 1990 the spectrum was obtained by R. Taylor and co-workers at the University of Sussex and was found to contain a single peak, confirming the unusual structure of buckminsterfullerene. [21]

Purity determination (w/w NMR)

While NMR is primarily used for structural determination, it can also be used for purity determination, provided that the structure and <u>molecular weight</u> of the compound is known. This technique requires the use of an <u>internal standard</u> of known purity. Typically this standard will have a high molecular weight to facilitate accurate weighing, but relatively few protons so as to give a clear peak for later integration e.g. <u>1,2,4,5-tetrachloro-3-nitrobenzene</u>. Accurately weighed portions of the standard and sample are combined and analysed by NMR. Suitable peaks from both compounds are selected and the purity of the sample is determined via the following equation.

$$ext{Purity} = rac{w_{ ext{std}} imes n[ext{H}]_{ ext{std}} imes MW_{ ext{spl}}}{w_{ ext{spl}} imes MW_{ ext{std}} imes n[ext{H}]_{ ext{spl}}} imes P$$

Where:

- *w*_{std}: weight of internal standard
- w_{spl}: weight of sample
- *n*[H]_{std}: the integrated area of the peak selected for comparison in the standard, corrected for the number of protons in that functional group
- *n*[H]_{spl}: the integrated area of the peak selected for comparison in the sample, corrected for the number of protons in that functional group
- MW_{std}: molecular weight of standard
- MW_{spl}: molecular weight of sample
- P: purity of internal standard

Non-destructive testing

Nuclear magnetic resonance is extremely useful for analyzing samples non-destructively. Radio-frequency magnetic fields easily penetrate many types of matter and anything that is not highly conductive or inherently <u>ferromagnetic</u>. For example, various expensive biological samples, such as <u>nucleic acids</u>, including <u>RNA</u> and <u>DNA</u>, or <u>proteins</u>, can be studied using nuclear magnetic resonance for weeks or months before using destructive biochemical experiments. This also makes nuclear magnetic resonance a good choice for analyzing dangerous samples.

Segmental and molecular motions

In addition to providing static information on molecules by determining their 3D structures, one of the remarkable advantages of NMR over X-ray crystallography is that it can be used to obtain important dynamic information. This is due to the orientation dependence of the chemical-shift, dipole-coupling, or electric-quadrupole-coupling contributions to the instantaneous NMR frequency in an anisotropic molecular environment. [22] When the molecule or segment containing the NMR-

observed nucleus changes its orientation relative to the external field, the NMR frequency changes, which can result in changes in one- or two-dimensional spectra or in the relaxation times, depending of the correlation time and amplitude of the motion.

Data acquisition in the petroleum industry

Another use for nuclear magnetic resonance is <u>data acquisition</u> in the <u>petroleum industry</u> for <u>petroleum</u> and <u>natural gas</u> exploration and recovery. Initial research in this domain began in the <u>1950s</u>, however, the first commercial instruments were not released until the early <u>1990s</u>. <u>[23]</u> A <u>borehole</u> is drilled into rock and sedimentary strata into which nuclear magnetic resonance logging equipment is lowered. Nuclear magnetic resonance analysis of these boreholes is used to measure rock porosity, estimate permeability from pore size distribution and identify pore fluids (water, oil and gas). These instruments are typically low field NMR spectrometers.

NMR logging, a subcategory of electromagnetic logging, measures the induced magnet moment of hydrogen nuclei (protons) contained within the fluid-filled pore space of porous media (reservoir rocks). Unlike conventional logging measurements (e.g., acoustic, density, neutron, and resistivity), which respond to both the rock matrix and fluid properties and are strongly dependent on mineralogy, NMR-logging measurements respond to the presence of hydrogen. Because hydrogen atoms primarily occur in pore fluids, NMR effectively responds to the volume, composition, viscosity, and distribution of these fluids, for example oil, gas or water. NMR logs provide information about the quantities of fluids present, the properties of these fluids, and the sizes of the pores containing these fluids. From this information, it is possible to infer or estimate:

- The volume (porosity) and distribution (permeability) of the rock pore space
- Rock composition
- Type and quantity of fluid hydrocarbons
- Hydrocarbon producibility

The basic core and log measurement is the T_2 decay, presented as a distribution of T_2 amplitudes versus time at each sample depth, typically from 0.3 ms to 3 s. The T_2 decay is further processed to give the total pore volume (the total porosity) and pore volumes within different ranges of T_2 . The most common volumes are the bound fluid and free fluid. A permeability estimate is made using a transform such as the Timur-Coates or SDR permeability transforms. By running the log with different acquisition parameters, direct hydrocarbon typing and enhanced diffusion are possible.

Flow probes for NMR spectroscopy

Recently, real-time applications of NMR in liquid media have been developed using specifically designed flow probes (flow cell assemblies) which can replace standard tube probes. This has enabled techniques that can incorporate the use of high performance liquid chromatography (HPLC) or other continuous flow sample introduction devices. [24]

Process control

NMR has now entered the arena of real-time process control and process optimization in oil refineries and petrochemical plants. Two different types of NMR analysis are utilized to provide real time analysis of feeds and products in order to control and optimize unit operations. Time-domain NMR (TD-NMR) spectrometers operating at low field (2–20 MHz for ¹H) yield free induction decay data that can be used to determine absolute https

and food manufacturing as well as <u>coal</u> analysis. High resolution FT-NMR spectrometers operating in the 60 MHz range with shielded permanent magnet systems yield high resolution HNMR spectra of refinery and petrochemical streams. The variation observed in these spectra with changing physical and chemical properties is modeled using <u>chemometrics</u> to yield predictions on unknown samples. The prediction results are provided to <u>control systems</u> via analogue or digital outputs from the spectrometer.

Earth's field NMR

In the Earth's magnetic field, NMR frequencies are in the audio frequency range, or the very low frequency and ultra low frequency bands of the radio frequency spectrum. Earth's field NMR (EFNMR) is typically stimulated by applying a relatively strong dc magnetic field pulse to the sample and, after the end of the pulse, analyzing the resulting low frequency alternating magnetic field that occurs in the Earth's magnetic field due to free induction decay (FID). These effects are exploited in some types of magnetometers, EFNMR spectrometers, and MRI imagers. Their inexpensive portable nature makes these instruments valuable for field use and for teaching the principles of NMR and MRI.

An important feature of EFNMR spectrometry compared with high-field NMR is that some aspects of molecular structure can be observed more clearly at low fields and low frequencies, whereas other aspects observable at high fields are not observable at low fields. This is because:

- Electron-mediated heteronuclear <u>J-couplings</u> (<u>spin-spin couplings</u>) are field independent, producing clusters of two or more frequencies separated by several Hz, which are more easily observed in a fundamental resonance of about 2 kHz."Indeed it appears that enhanced resolution is possible due to the long spin relaxation times and high field homogeneity which prevail in EFNMR."

 [25]
- Chemical shifts of several <u>ppm</u> are clearly separated in high field NMR spectra, but have separations of only a few millihertz at proton EFNMR frequencies, so are usually not resolved.

Zero field NMR

In zero field NMR all magnetic fields are shielded such that magnetic fields below 1 nT (nanotesla) are achieved and the nuclear precession frequencies of all nuclei are close to zero and indistinguishable. Under those circumstances the observed spectra are no-longer dictated by chemical shifts but primarily by *J*-coupling interactions which are independent of the external magnetic field. Since inductive detection schemes are not sensitive at very low frequencies, on the order of the *J*-couplings (typically between 0 and 1000 Hz), alternative detection schemes are used. Specifically, sensitive magnetometers turn out to be good detectors for zero field NMR. A zero magnetic field environment does not provide any polarization hence it is the combination of zero field NMR with hyperpolarization schemes that makes zero field NMR attractive.

Quantum computing

NMR <u>quantum computing</u> uses the <u>spin</u> states of nuclei within molecules as <u>qubits</u>. NMR differs from other implementations of quantum computers in that it uses an <u>ensemble</u> of systems; in this case, molecules.

Magnetometers

Various magnetometers use NMR effects to measure magnetic fields, including proton precession magnetometers (PPM) (also known as proton magnetometers), and Overhauser magnetometers. See also Earth's field NMR.

SNMR

Surface magnetic resonance (or magnetic resonance sounding) is based on the principle of Nuclear magnetic resonance (NMR) and measurements can be used to indirectly estimate the water content of saturated and unsaturated zones in the earth's subsurface. [26] SNMR is used to estimate aquifer properties, including quantity of water contained in the aquifer, Porosity, and Hydraulic conductivity.

Makers of NMR equipment

Major NMR instrument makers include <u>Thermo Fisher Scientific</u>, <u>Magritek</u>, <u>Oxford Instruments</u>, <u>Bruker</u>, <u>Spinlock SRL</u>, <u>General Electric</u>, <u>JEOL</u>, <u>Kimble Chase</u>, <u>Philips</u>, <u>Siemens AG</u>, and formerly Agilent Technologies, Inc. (who own Varian, Inc.).

See also

- Benchtop NMR spectrometer
- Carbon-13 NMR
- Chemical shift
- Dynamic nuclear polarisation (DNP)
- Earth's field NMR (EFNMR)
- Free induction decay (FID)
- In vivo magnetic resonance spectroscopy (MRS)
- *J*-coupling
- Larmor equation (Not to be confused with Larmor formula).
- Larmor precession
- Liquid nitrogen
- Low field NMR
- Magic angle spinning
- Magnetometer
- Magnetic resonance imaging (MRI)
- Magnetic resonance (quantum mechanics) for physical & mathematical understanding

- NMR crystallography
- NMR spectra database
- NMR spectroscopy
- NMR Microscopy
- Nuclear magnetic resonance in porous media
- Nuclear quadrupole resonance (NQR)
- Protein dynamics
- Protein NMR
- Proton NMR
- Rabi cycle
- Relaxometry
- Relaxation (NMR)
- Spin echo
- Solid-state NMR
- Structure-based assignment
- Zero field NMR
- SNMR

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