

# Group Assignment

February 18, 2018

## 1 Biology

### 1.1 MSC

In an embryo a mesenchymal stem cell is a pluripotent progenitor cell which divides many times and eventually differentiating through a series of separate and unique lineage transitions into a variety of end-stage phenotypes: cartilage, bone, tendon, ligament, marrow stroma, connective tissues. The term, mesenchyme, is derived from the Greek meaning middle (meso) infusion and refers to the ability of mesenchymatous cells to spread and migrate in early embryonic development between the ectodermal and endodermal layers. This characteristic migratory, space-filling ability is the key element of all wound repair in adult organisms involving mesenchymal cells in skin (dermis), bone (periosteum), or muscle (perimysium).

They have the ability to form unique developmental structures, or, in adults to form repair blastemas that are able to achieve regenerative repair, a fact that makes the study of these mesenchymal stem cells crucial as it provides the basis for the evolution of a new therapeutic technology of self-cell repair. Several important advances allow us to consider the possibility of using a patient's own mesenchymal stem cells as starting material for tissue repair protocols. Mesenchymal stem cells must exist to maintain the living organisms, just as hematopoietic stem cells must exist to support both red and white blood cell turnover.

We might be able to isolate such human mesenchymal stem cells and place them in cell culture, where we could mitotically expand their numbers. Eventually, if we had enough of these cells, we could reintroduce them into the original donor in a manner that guaranteed that they would massively differentiate into a specific tissue, such as cartilage or bone, at a transplantation or repair site. Immunorejection would not be a problem because the donor and host would be one and the same.

### 1.2 Morphogen & Pattern Formation

A morphogen is a substance that establishes a graded distribution and elicits distinct cellular responses in a dose-dependent manner that governs the pattern of tissue de-

development in the process of morphogenesis or pattern formation. They function to provide individual cells within a field with positional information, which is interpreted to give rise to spatial patterns. Morphogens can consist of cytoplasmic proteins, such as transcription factors that form a gradient by diffusion within a single cell or syncytium, or secreted signaling molecules that travel from cell to cell. In most cases, morphogens guide the generation of different cell types in a specific spatial order, usually by inducing unique transcriptional responses in a dose dependent manner. Once the gradient has formed, cells must distinguish small differences in morphogen concentration and store this information even after the gradient has dissipated.

Pattern formation is the process by which a specialized cell, the fertilized egg, divides to give rise to seemingly identical cells. Over time and space, these cells take on distinct fates and eventually become organized into functional organ systems. Within a single organ, such as the heart, multiple classes of differentiated cells must synchronize their duties to ensure that the organ as a whole is functional. One of the unifying principles of development is the idea that embryogenesis proceeds via the iterative process of generating naive fields of cells, and then providing cells within each field with unique positional information, which they then interpret to give rise to spatial patterns. Through this process, the embryo is sequentially subdivided, initially along the major body axes, and then into smaller, and more refined units such as organ primordia that are further partitioned and patterned

## 2 Model

### 2.1 Parameter selection

In order to proceed this reaction-diffusion system for VMC pattern formation process, Alan Garfinkel and his partners made the following choices. The first choice is that they treated this model as 2D spatial domain in 100\*100 uniform mesh. Meanwhile the activator and inhibitor are BMP-2 and MGP corresponding to  $U(x,y)$  and  $V(x,y)$  (effective concentrations). The method to calculate the final concentrations for BMP-2 and MGP is the use of second order Runge-Kutta method. Moreover the chemical kinetics they chose were the interactions of BMP-2 with MGP. Additionally, in order to govern the model equations, the initial condition is described as following The initial condition of  $U$  and  $V$  that are uniformly distributed with a 2% random perturbations as well as neumann conditions (no-flux at boundary) for their boundary conditions. The whole equations are the following:

In these above equations,  $D$  is the ratio of diffusion coefficient of BMP-2 and MGP which is  $D_u/D_v$  (BMP-2 and MGP respectively).  $\gamma$  is the scaling parameter which is equal to  $\frac{L^2}{D_v T_C}$ , where  $L$  is the length of the domain which is the actual size in their experiments,  $T_C$  is the time for BMP-2 synthesis.  $k$  is the saturating value.  $c$  and  $e$  are the first order degradation rates for BMP-2 and MGP.  $S$  is source term.

More precisely, in equation 1, BMP-2 will activate its own production autocatalyt-

ically which will saturate. Hence Garfinkel et al chose  $\frac{U^2}{1+kU^2}$ , a sigmoidal form, to govern this autocatalysis. V is treated as the inhibitor in the denominator. There is a degradation for U at rate c. In equation 2,  $U^2$  is used to represent that BMP-2 spurs MGP in non-linear manner. There is a degradation for V at rate e. S represents the exogenous MGP which is added by purpose.

In order to run the equations smoothly, the parameters were chosen carefully. Firstly, Garfinkel et al took the initial value of  $Du$ ,  $1 \times 10^{-8} \text{cm}^2/\text{sec}$ , directly from Entchev et als work which was calculated by using GFP labeling and imaging. Then they considered the diffusion of large amount of molecules in nonlinear slowing manner. Due to acid PH, dimerization of BMP-2 and actual tissue culture, the estimated value reduced to  $0.15 \times 10^{-8} \text{cm}^2/\text{sec}$ . Because there is no directly calculated value for diffusivity of MGP, they estimated an theoretical approximate value,  $30 \times 10^{-8} \text{cm}^2/\text{sec}$ , which was based on empirical formulas. Meanwhile, this approximate value is similar to the diffusion coefficient of amyloid B which is  $50 \times 10^{-8} \text{cm}^2/\text{sec}$ . Finally, the value of D was calculated by  $\frac{D_u}{D_v} = \frac{1}{200}$ .

In the previous experiment which was done by Ghosh-Choudhury et al, it shows that BMP-2 autoregulates in a saturating manner. However, due to unknown level of MGP, it is difficult to establish the precise value of k. In Garfinkel et als model, they chose  $k=0.65$ .

In order to calculate , it is essential that production rates are known. In previous work, they have found the upper limit for the production rate which is in embryonic kidney cells with a cytomegalovirus (CMV) promoter. They also established the rates for BMP-2 production in calcifying vascular cells and in endothelial cells which were both . Furthermore, FLAG-tagged MGP and newly developed ELISA suggested that the production rates of BMP-2 and MGP are similar. These reports can inform that the upper limit for the production rates is and the lower limit is .

As mentioned above, is equal to . In the actual cell culture, the is 4cm, they chose is 1 ng/hr ( ). Hence the

Garfinkel et al estimated the degradation rate of BMP-2 directly and conservatively from the calculation of Entchev et als work. Hence they took  $c = 0.01$ . MGP has more rapidly degradation than BMP-2 based on their unpublished work. The ratio for is approximately . Thus they took  $e = 0.02$ .

The source term S is to be added in order to make tripling pattern formation which means the source term needs to be three times the initial concentration of MGP. In the model, Garfinkel et al chose the initial value of MGP scaled as 2. Then for the 1000 time steps, the S needs to be 0.06 per time steps, totally 6 which will make tripling pattern formation.

## References

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- [2] Jan L. Christian. *Morphogen gradients in Development: from form to function*  
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