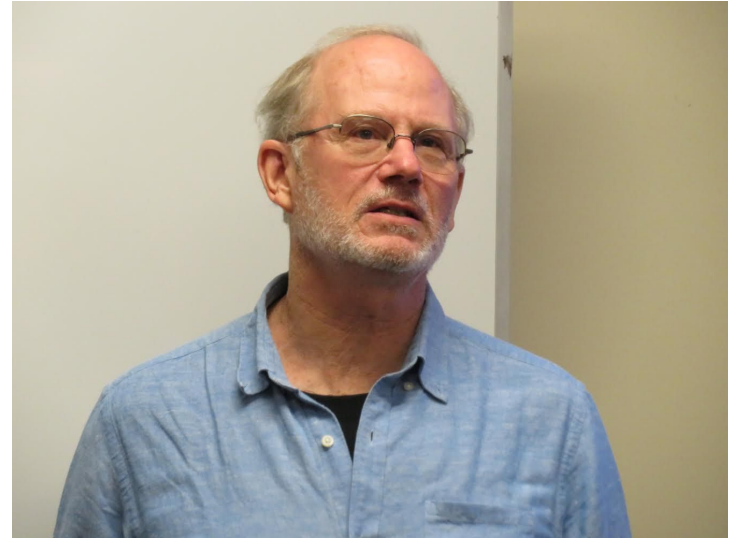


An introduction to implementing LD and temporal methods with NeEstimator

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A moment of appreciation: Robin Waples

- University of Washington
- Salmonid biology
- Consultant on the PIRE project
- Master of N_e
- Co-creator of NeEstimator



NeEstimator overview

- Easy to execute wrapper for several different methods of estimating N_e from genetic data
- Several implementations
 - **Command-line version**
 - Java-based GUI
 - R (package RLDNe)
- Current version: NeEstimator v2 (Do et al., 2014)

NeEstimator GUI

Ne Estimator

File Run Help

INPUT

Directory:

Choose File: ☒ List Files with extensions TXT, GEN, DAT only

Data Format: ☒ GENEPOP ☐ FSTAT

Line 1:

OUTPUT

Directory:

File Name: ☒ Use Default Name (uncheck to edit)

☐ Main and Tabular-Format Output Files to be appended

ADDITIONAL OUTPUT FILES: (Names are preset, not editable) ☐ With Tab delimiter in the format

☐ Output File(s) in tabular format, for the top critical value(s). Name(s):

☐ Output LD Burrows coefficients Name:

Only for populations in range: (max = 50), and only for the top critical value(s)

☐ Output File for Frequency Data for all loci- details up to 100 loci. Name:

Only for populations in range: (max = 50)

☒ Output for missing data if any. Name:

Methods

☒ Linkage Disequilibrium - Model:

☒ Random Mating ☐ Monogamy

☒ Heterozygote Excess

☒ Molecular Coancestry

☒ Temporal, Plan II

Critical Values (ignored by Coancestry)

☐ For LD only: Exclude singleton alleles

☒ Also run without frequency restriction

Options

☐ No Output for Confidence Intervals

☐ Population range to run:

☐ Up to individual per pop:

☐ Restrict Loci by

☐ Ranges:

☒ Omitting Loci:

☐ LD locus pairing across chromosomes

Inputs: genepop format (.geno or .genepop)

Lower Arkansas + Rattlesnake																
NC_048407_1_271437, NC_048407_1_9657082, NC_048407_1_9661459, NC_048407_1_10124451, NC_048407_1_10124452																
POP																
4THST,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0101	0101	0101	0102	0101	0202	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0102	0102	0101	0101	0101	0102	0102
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0000	0102	0102	0101	0101	0202	0202
ARK10,	0000	0101	0101	0101	0101	0101	0101	0202	0101	0000	0202	0101	0101	0101	0202	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0102	0102	0102	0101	0101	0101	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0101	0101	0101	0102	0101	0202	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0000	0101	0101	0102	0101	0202	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0101	0101	0101	0102	0101	0102	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0000	0202	0101	0101	0101	0202	0202
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0000	0102	0101	0102	0101	0102	0102
ARK10,	0102	0101	0101	0101	0101	0101	0101	0202	0101	0000	0102	0101	0102	0101	0202	0202

Useful for microsatellite or SNP data

Format conversion: pgdspider (java) or adegenet (R)

Methods: Linkage Disequilibrium (LD)

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INVITED TECHNICAL REVIEW

MOLECULAR ECOLOGY
RESOURCES WILEY

Practical application of the linkage disequilibrium method for estimating contemporary effective population size: A review

Robin S. Waples 

Measuring linkage disequilibrium

Recall Hardy-Weinberg proportions: for unlinked loci $P_{A,B} = P_A P_B$

A simple measure of **disequilibrium**: $D_{A,B} = P_{A,B} - P_A P_B$

NeEstimator uses an adjusted metric (r^2) that is independent of allele frequencies and is always positive

From linkage disequilibrium to N_e

Expected r^2 can be calculated from recombination fraction (c) and N_e ...

$$E(r^2) = \frac{c^2 + (1-c)^2}{2N_e c(2-c)} = \frac{\gamma}{N_e},$$

...and N_e can be back-calculated from r^2 !

$$\hat{N}_e = \frac{\gamma}{r^2 - 1/n}$$

And if we know (or assume) loci are unlinked ($c=0.5$), this calculation is even simpler

$$\hat{N}_e = \frac{1}{(r^2 - 1/n)} \frac{1}{3} = \frac{1}{3r^2},$$

Confidence intervals

Parametric:

$$\text{CV}(\hat{N}_e) \approx \left(1 + \frac{1}{\gamma} \frac{N_e}{n}\right) \sqrt{2/k}$$

Waples, 2014

Bootstrap: re-sample SNPs (with replacement) x times, calculate N_e using x resampled datasets, and tabulate variation among re-sampled estimates

Assumptions and considerations: sampling/loci

Loci assumed to be unlinked

Sample sizes (number of individuals) may need to be *large*, particularly to estimate reasonable confidence intervals

Increasing the number of loci independent loci can increase precision, but subject to diminishing returns

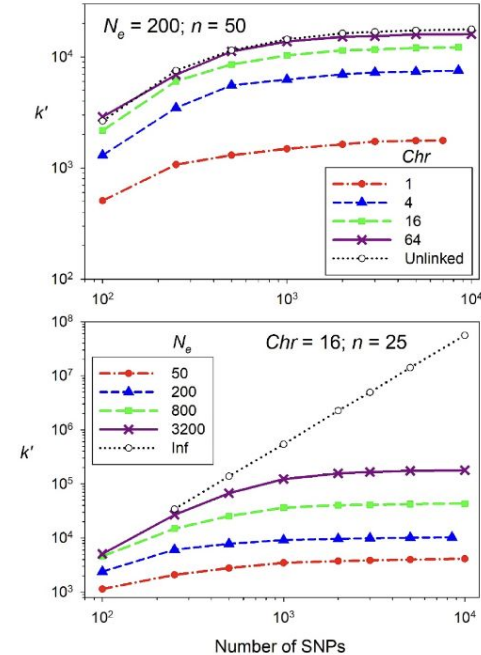


FIGURE 4 Effective degrees of freedom=effective number of locus pairs (k') for r^2 as a function of the number (L) of diallelic (SNP) loci used to calculate r^2 . k' was calculated from simulated data based on the rate of decline in $\text{var}(r^2)$ as more loci were used. Top: Effect of number of chromosomes (Chr), with $N_e = 200$ and $n = 50$. Bottom: effect of N_e with $Chr = 16$ and $n = 25$. Modified from Waples et al. (2022).

Assumptions and considerations: biases

Mutation and selection don't strongly bias the method

Population structure and immigration can introduce strong biases

Systems with overlapping generations, non-random mating systems, and age structure require special consideration

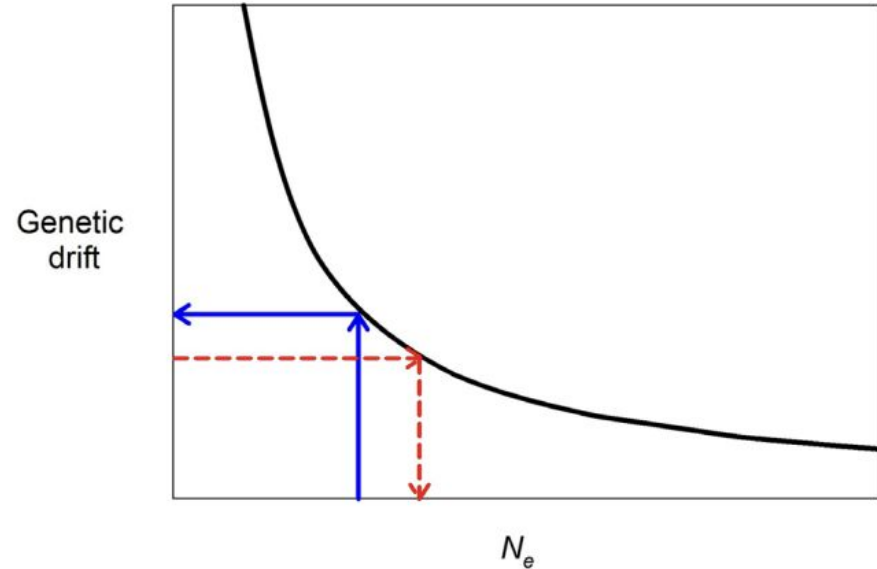
Temporal method: conceptual framework

Variance in allele frequencies (F)
over time is related to N_e

$$N_e \sim t/2F$$

3 different ways of calculating F

- F_e (Nei and Tajima, 1981)
- F_k (Pollak, 1983)
- F_s (Jorde and Ryman, 2007)



Temporal method: sampling

Plan 1: sampling after reproduction,
or sampling with replacement

Plan 2: sampling before
reproduction (without replacement)

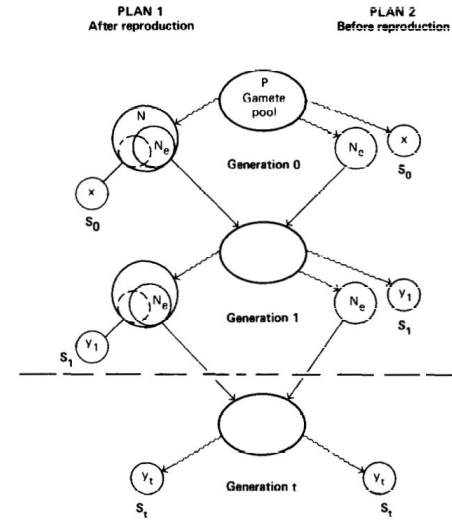


FIGURE 1.—Two sampling plans considered in the analysis. In both plans, P is frequency of an allele in gamete pool preceding generation 0, and x and y_t are allele frequencies in samples (of S_0 and S_t individuals) for genetic analysis taken at generations 0 and t , respectively. N is total population size at time of the initial sample, and N_e is variance effective population size. *Plan I*: sample S_0 is taken after reproduction, so it may contain some of $2N_e$ genes representing effective population size. Sample allele frequencies x and y_t are positively correlated with respect to P because samples S_0 and S_t are derived from same population (size N) at generation 0. *Plan II*: sample is taken before reproduction and not replaced, so the samples S_0 and N_e are mutually exclusive and can be considered to be independent binomial draws from initial gamete pool. Total population size is not a factor, and x and y_t are uncorrelated.

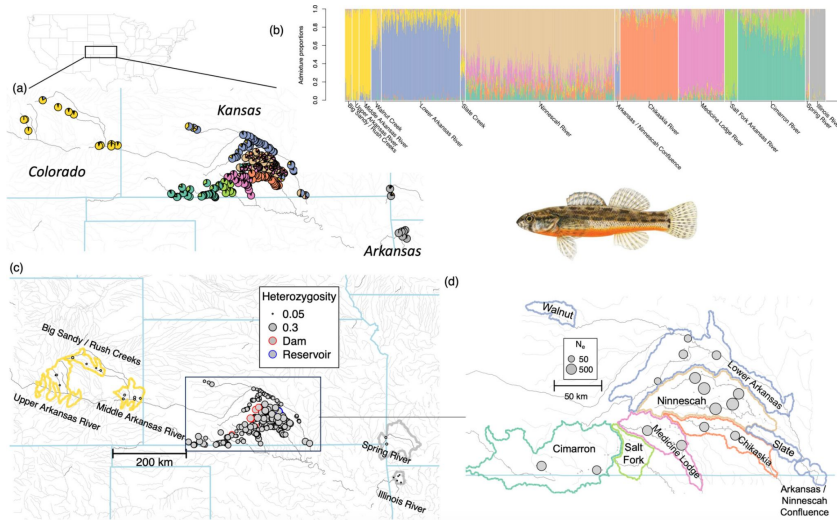
Time considerations

Temporal estimated of N_e don't represent contemporary N_e - instead, they represent a geometric average of N_e per generation over the time period sampled

Since drift accumulates over time, the number of generations between the sampling intervals can affect accuracy. More generations = greater signal of drift (to a point)...

Exercise: datasets

LD method: Arkansas darters



Temporal method: PIRE fish

S. delicatulus



H. miniatus



T. zosterophora



The Philippines PIRE Project

