using the sampler

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1 Using the sampler

hlm_gibbs is a generic gibbs sampling framework for spatially-correlated variance components models. The current supported models are:

- hlm.both contains specifications with correlated errors in both levels, with the first statement se/sma describing the lower level and the second statement se/sma describing the upper level. In addition, MVCM, the multilevel variance components model with no spatial correlation, is in the both namespace.
- hlm.lower contains two specifications, se/sma, that can be used for a variance components model with correlated lower-level errors.
- hlm.upper contains two specifications, se/sma that can be used for a variance components model with correlated upper-level errors.

Depending on the structure of the model, you need at least: - X, data at the lower level - Y, system response in the lower level - membership or Delta, the membership vector relating each observation to its group or the "dummy variable" matrix encoding the same information.

Then, if spatial correlation is desired, M is the "upper-level" weights matrix and W the lower-level weights matrix.

Finally, there are many configuration and tuning options that can be passed in at the start, or assigned after the model is initialized.

First, though, let's set up some data for a model on southern counties relating DNL90 to G189, BLK90, and HR90, with statewide average FH90 as an upper-level predictor.

```
In [1]: #seaborn is required for the traceplots
    import pysal as ps
    import hlm
    import numpy as np
    import pandas as pd
    import seaborn as sns
    import matplotlib.pyplot as plt
    %matplotlib inline
```

First, we'll will read in the data and assign to the correct response, upper-, and lower-level data.

```
X = data[['GI89', 'BLK90', 'HR90']].values
N = X.shape[0]
Z = data.groupby('STATE_NAME')['FH90'].mean()
Z = Z.values.reshape(-1,1)
J = Z.shape[0]
Y = data.DNL90.values.reshape(-1,1)
```

Then, we'll construct some weights from the files.

With the data, upper-level weights, and lower-level weights, we can construct a membership vector *or* a dummy data matrix. For now, I'll create the membership vector.

```
In [4]: membership = data.STATE_NAME.apply(lambda x: W2.id_order.index(x)).values
```

But, we could also build the dummy variable matrix using pandas, if we have a suitable categorical variable:

Every call to the sampler is of the following form:

```
sampler(Y, X, W, M, Z, membership, Delta, n_samples,
**configuration)
```

Where W, M are passed if appropriate, Z is passed if used, and only one of membership or Delta is required. In the end, Z is appended to X, so the effects pertaining to the upper level will be at the tail end of the β effects vector. If both Delta and membership are supplied, they're verified against each other to ensure that they agree before they are used in the model.

Every sampler uses, either in whole or in part, hlm.both.generic, which implements the full generic sampler discussed in the working paper. For efficiency, the upper-level samplers modify this runtime to avoid processing the full lower-level covariance matrix.

Thus, with the data and a membership vector, a model can be fit by calling:

The results and state of the sampler are stored within the vcsma object. I'll step through the most important parts of this object.

2 trace

The quickest way to get information out of the model is via the trace object. This is where the results of the tracked parameters are stored each iteration. Any variable in the sampler state can be added to the tracked params. Trace objects are essentially dictionaries with the keys being the name of the tracked parameter and the values being a list of each iteration's sampler output.

```
In [7]: vcsma.trace.varnames
Out[7]: ['Betas', 'Sigma2', 'Tau2', 'Lambda', 'Alphas']
```

In this case, Lambda is the upper-level moving average parameter, Alphas is the vector of correlated group-level random effects, Tau2 is the upper-level variance, Betas are the marginal effects, and Sigma2 is the lower-level error variance.

I'm working to rename these to be consistent with the draft naming convention, but am writing unittests first to make sure that the rename doesn't induce errors.

I've written two helper functions for working with traces. First is to just dump all the output into a pandas dataframe, which makes it super easy to do work on the samples, or write them out to csv and assess convergence in R's coda package.

```
In [8]: trace_dataframe = vcsma.trace.to_df()
In [9]: trace_dataframe.mean()
Out[9]: Lambda
                      0.766717
        Sigma2
                      1.277206
        Tau2
                      0.232024
        Alphas_0
                      0.308541
        Alphas 1
                      0.742316
        Alphas_2
                      0.731663
        Alphas_3
                      0.433992
        Alphas_4
                      0.379480
        Alphas_5
                     -0.866960
        Alphas_6
                      0.148806
        Alphas_7
                      0.098672
        Alphas_8
                     -0.754410
        Alphas_9
                     -0.558129
        Alphas_10
                     -0.133320
        Alphas_11
                      0.003053
        Alphas 12
                     -0.325798
        Alphas_13
                     -0.337959
        Alphas_14
                      0.130551
        Alphas_15
                      0.098818
        Betas 0
                       9.093728
                    -11.844991
        Betas_1
```

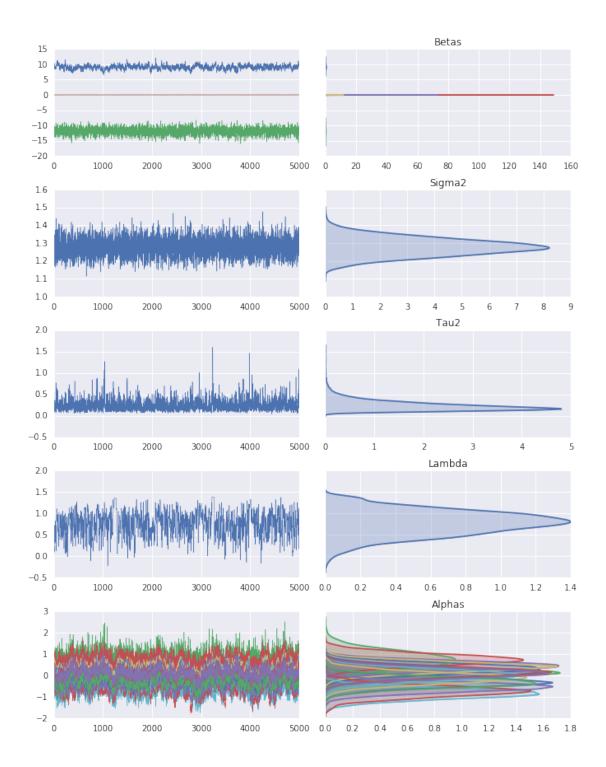
```
Betas_2 0.003461
Betas_3 0.030215
Betas_4 -0.043534
dtype: float64
```

The second is a method to plot the traces:

In [10]: fig, ax = vcsma.trace.plot()

```
plt.tight_layout()
    plt.show()

/home/ljw/anaconda3/envs/py3/lib/python3.5/site-packages/statsmodels/nonparametric,
    y = X[:m/2+1] + np.r_[0,X[m/2+1:],0]*1j
```



The trace object is a subclass of the standard python dictionary that allows for its elements to be accessed by dot notation, .get, or by the typical name-specific slicing:

Out [11]: 0.69088744240431255

In the future, it may make sense to modify the slicing behavior to allow for iteration-specific subsetting, so that trace[100:] would give the trace with the first 100 elements removed.

In addition, data in the trace is stored in lists. The sampler appends new values to this list each iteration.

If you're working with the data substantially, just use trace.to_df, since pandas dataframes are much easier to work with.

To get stuff like posterior quantiles, you can use the attendant pandas dataframe functionality, like describe.

<pre>In [12]: df = vcsma.trace.to_df()</pre>							
In [13]:	n [13]: df.describe()						
Out[13]:	count mean std min 25% 50% 75% max	Lambda 5000.000000 0.766717 0.283057 -0.222647 0.576475 0.782937 0.967878 1.382690	Sigma2 5000.000000 1.277206 0.049026 1.114109 1.243308 1.276352 1.309424 1.476384	Tau2 5000.000000 0.232024 0.127130 0.041333 0.147964 0.201060 0.282579 1.601098	Alphas_0 5000.000000 0.308541 0.251061 -0.593088 0.143531 0.310582 0.477174 1.192098	Alphas_1 5000.000000 0.742316 0.412403 -0.880090 0.464534 0.731781 1.016415 2.503563	\
	count mean std min 25% 50% 75% max	Alphas_2 5000.000000 0.731663 0.273122 -0.537455 0.548285 0.731244 0.916221 1.694635	Alphas_3 5000.000000 0.433992 0.226161 -0.316243 0.285678 0.437807 0.590868 1.153007	Alphas_4 5000.000000 0.379480 0.242647 -0.437794 0.220639 0.387947 0.543126 1.209314	Alphas_5 5000.000000 -0.866960 0.246681 -1.756178 -1.028922 -0.861425 -0.697662 -0.005890	Alphas_6 5000.000000 0.148806 0.259058 -0.752014 -0.018897 0.153212 0.322672 1.096372	
	count mean std min 25% 50% 75% max		Alphas_11 5000.000000 0.003053 0.277827 -0.973372 -0.182646 0.006931 0.188394 0.937809	Alphas_12 5000.000000 -0.325798 0.238633 -1.120906 -0.482853 -0.325915 -0.160284 0.492381	Alphas_13 5000.000000 -0.337959 0.250540 -1.250463 -0.502870 -0.339225 -0.167577 0.561262	Alphas_14 5000.000000 0.130551 0.243599 -0.656771 -0.029217 0.136570 0.297502 0.919789	
	count mean	Alphas_15 5000.000000 0.098818	Betas_0 5000.000000 9.093728	Betas_1 5000.000000 -11.844991	Betas_2 5000.000000 0.003461	Betas_3 5000.000000 0.030215	\

std 0.243712 0.691172 1.124104 0.002612 0.005278

min	-0.842041	6.635997	-16.048654	-0.006154	0.010171
25%	-0.061944	8.640080	-12.589831	0.001684	0.026780
50%	0.099924	9.098844	-11.855798	0.003459	0.030152
75%	0.259934	9.547547	-11.061595	0.005230	0.033846
max	1.059585	12.124211	-8.216864	0.013521	0.049684
	Betas_4				
count	5000.000000				
mean	-0.043534				
std	0.035175				
min	-0.169464				
25%	-0.064759				
50%	-0.042214				
75%	-0.020200				

[8 rows x 24 columns]

0.079243

2.0.1 Formal trace statistics:

max

To compute trace diagnostics, you can use existing python packages. I plan on implementing at least the Gelman-Rubin potential scale reduction factor (PSRF) statistic (an ANOVA-based measure of ergodicity and convergence of multiple chains) and the geweke statistic, and writing or wrapping existing implementations of the PSRF, geweke, and autocorrelation plots and diagnostics.

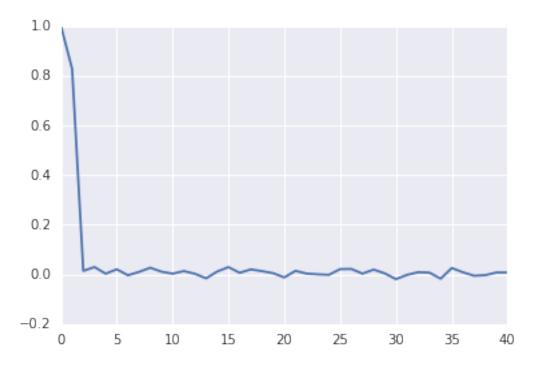
Unfortunately, pymc3's diagnostic functions don't work with arbitrary arrays of values. And, you can't really "initialize" one of their traces and stuff it full of what you want.

Currently, aside from just computing the statistics on the pandas dataframe form of the trace, you can also use stuff like statsmodels to compute the chain autocorrelation:

```
In [14]: from statsmodels.api import tsa
    #if you don't have it, try removing the comment and:
    #! pip install pymc3
```

For example, the partial autocorrelation in the lambda chain over the past 40 lags in the λ sample is:

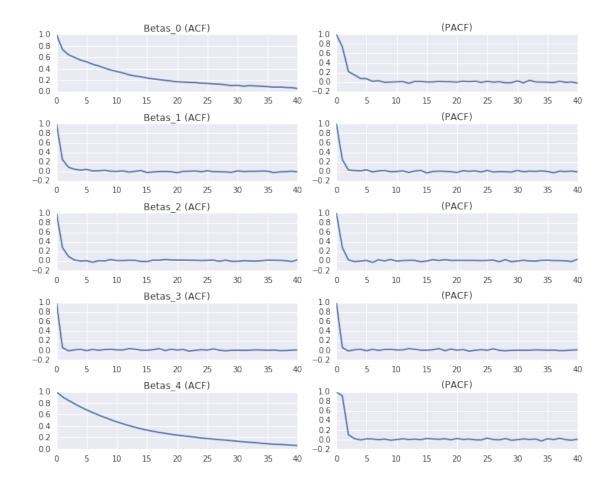
```
In [15]: plt.plot(tsa.pacf(df.Lambda))
Out[15]: [<matplotlib.lines.Line2D at 0x7f8660a72358>]
```



So, the chain is close-to-first order:

We could do this for many parameters, too. An Autocorrelation/Partial Autocorrelation plot can be made of the marginal effects by:

```
In [17]: betas = [c for c in df.columns if c.startswith('Beta')]
    f,ax = plt.subplots(len(betas), 2, figsize=(10,8))
    for i, col in enumerate(betas):
        ax[i,0].plot(tsa.acf(df[col].values))
        ax[i,1].plot(tsa.pacf(df[col].values)) #the pacf plots take a while
        ax[i,0].set_title(col +' (ACF)')
        ax[i,1].set_title('(PACF)')
    f.tight_layout()
    plt.show()
```



This is something again that I'm going to wrap up into a diagnostic plot.

Most of the practitioners I've seen in python output their model results to R to use the coda Bayesian Diagnostics package.

These diagnostics are important checks on validity, and serve to formalize the logic of trace plot inspection.

The most helpful ones (in my opinion) are the Gelman-Rubin diagnostic and the Geweke diagnostic. The Gelman-Rubin statistic, also known as the "potential scale reduction factor," is an ANOVA test between multiple chains that measures how likely it is that the distributions drawn from the trace are the same. So, let's run another chain:

write the dataframe out to a csv and load it in R as an mcmc object:

```
trace <- coda::mcmc(trace, start=500)</pre>
        coda::geweke.diag(trace)
Fraction in 1st window = 0.1
Fraction in 2nd window = 0.5
   Lambda
            Sigma2
                        Tau2 Alphas_0 Alphas_1 Alphas_2 Alphas_3 Alphas_4
-1.903931 -1.952078 -0.552043 -0.415133 -1.108863 -0.363955 -0.196839 -0.425794
Alphas_5 Alphas_6 Alphas_7 Alphas_8 Alphas_9 Alphas_10 Alphas_11 Alphas_12
-0.142884 -0.411852 -0.001707 -0.611931 -0.202954 0.078213 -0.222248 -0.044373
Alphas 13 Alphas 14 Alphas 15
                               Betas 0
                                         Betas 1
                                                   Betas 2
                                                             Betas 3
                                                                       Betas 4
 0.057303 0.121722 -0.227471 -0.109759 -0.312055 -2.204327 0.398186
                                                                      0.316436
```

Use the %Rpush command if you're conducting the analysis from within the notebook:

```
In [21]: trace2 = df
In [22]: %Rpush trace2
In [23]: %%R
         trace2 <- coda::mcmc(trace2, start=500)</pre>
         coda::geweke.diag(trace2)
Fraction in 1st window = 0.1
Fraction in 2nd window = 0.5
   Lambda
             Sigma2
                         Tau2 Alphas_0 Alphas_1 Alphas_2 Alphas_3 Alphas_4
-1.903931 -1.952078 -0.552043 -0.415133 -1.108863 -0.363955 -0.196839 -0.425794
Alphas_5 Alphas_6 Alphas_7 Alphas_8 Alphas_9 Alphas_10 Alphas_11 Alphas_12
-0.142884 \ -0.411852 \ -0.001707 \ -0.611931 \ -0.202954 \ \ 0.078213 \ -0.222248 \ -0.044373
Alphas_13 Alphas_14 Alphas_15
                                Betas_0
                                          Betas_1
                                                    Betas_2
                                                               Betas_3
 0.057303 0.121722 -0.227471 -0.109759 -0.312055 -2.204327
                                                                        0.316436
                                                              0.398186
```

to do the Gelman-Rubin statistic, we need to run another chain. So, we'll run another chain just by re-doing the previous one. There're a few ways to reset a chain, duplicate its state, copy it, or etc.

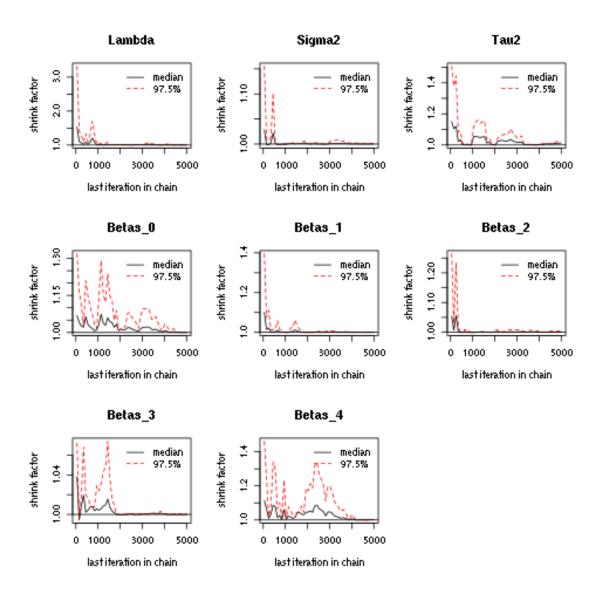
But the simplest is to just rerun the same initial call.

```
/home/ljw/Downloads/projects/hlm_gibbs/hlm/utils.py:91: ComplexWarning: Casting cor
  emax = emax.astype(float)
/home/ljw/Downloads/projects/hlm_gibbs/hlm/utils.py:92: ComplexWarning: Casting cor
  emin = emin.astype(float)
```

I'm going to remove the random effects from this plot so we don't have a huge diagnostic plot:

If the partial scale reduction factor is close to 1, it means the chains are quite similar, and they've likely converged to the same posterior. If it's much larger than 1, it indicates that the between-chain variance is much larger than the within-chain variance, and it's likely that the chains are reflecting different distributions and the MCMC routine hasn't converged to the same steady state twice.

In this case, all of our scale reduction factors are close to 1:



Potential scale reduction factors:

	Point	est.	Upper	C.I.
Lambda		1.00		1.00
Sigma2		1.00		1.00
Tau2		1.01		1.03
Betas_0		1.00		1.00
Betas_1		1.00		1.00
Betas_2		1.00		1.01
Betas_3		1.00		1.00

```
Betas_4 1.00 1.00

Multivariate psrf

1.01
```

3 draw and sample

These two functions are used to call the underlying Gibbs sampler. They take no arguments, and operate on the sampler in place. draw provides a single new sample:

```
In [29]: vcsma.draw()
```

And sample steps forward an arbitrary number of times:

```
In [30]: vcsma.sample(10)
```

At this point, we did 5000 initial samples and 11 extra samples. Thus:

```
In [31]: vcsma.cycles
Out[31]: 5011
```

4 state

This is the collection of current values in the sampler. To be efficient, Gibbs sampling must keep around some of the computations used in the simulation, since sometimes the same terms show up in different conditional posteriors. So, the current values of the sampler are stored in state.

All of the following are tracked in the state:

```
In [32]: print(vcsma.state.varnames)
['Rho_min', 'Tau2_b0', 'Betas_covm', 'Betas_cov0', 'XBetas', 'Betas_mean0', 'Delta?
```

You can construct traces of "extra" parameters in state by passing their names to the extra_traced_params option of the model initialization, or by adding them to the trace & traced_params manually.

```
In [34]: example.traced_params
Out[34]: ['Alphas', 'Betas', 'Sigma2', 'Tau2', 'Lambda', 'DeltaAlphas']
In [35]: example.sample(5)
In [36]: example.trace.DeltaAlphas
Out[36]: [array([[ 0.39952387],
                  [ 0.39952387],
                  [ 0.39952387],
                  . . . ,
                  [ 1.00837255],
                  [ 1.00837255],
                  [ 1.00837255]]), array([[ 0.59357408],
                  [0.59357408],
                  [ 0.59357408],
                  [ 0.44552682],
                  [ 0.44552682],
                  [ 0.44552682]]), array([[ 0.46438779],
                  [0.46438779],
                  [ 0.46438779],
                  [ 0.25678336],
                  [ 0.25678336],
                  [ 0.25678336]]), array([[ 0.27606112],
                  [ 0.27606112],
                  [ 0.27606112],
                  [ 0.39098594],
                  [0.39098594],
                  [ 0.39098594]]), array([[ 0.188163 ],
                  [ 0.188163 ],
                  [ 0.188163 ],
                  . . . ,
                  [0.4715184],
                  [0.4715184],
                  [ 0.4715184]])]
```

5 configs

this is where configuration options for the metropolis steps for the spatial parameter are stored. Each metropolis step has its own entry in configs, which stores a few recognized options. You can access these options directly through the initial call.

```
In [37]: vcsma.configs
Out[37]: {Lambda:{ar:0.4491017964071856, proposal:<scipy.stats._continuous_distns.r</pre>
```

Thus, we can se the previous run's acceptance rate:

```
In [38]: vcsma.configs.Lambda.ar
Out[38]: 0.4491017964071856
```

If you're doing heavy customization, it makes the most sense to first initialize the class without sampling. We did this before when showing how the "extra_traced_params" option worked.

To show, let's initialize a double-level SAR-Error variance components model, but not actually draw anything.

To do this, you pass the option n_samples=0.

This sets up the sampler with a default uninformative configuration. This means the prior precisions are all I \star .001 \star , prior means are all 0, spatial parameters are set to -1/(n-1), and prior scale factors are set arbitrarily.

Options are set by assgning to the relevant property in model.configs.

The model configuration object is another dictionary with a few special methods.

Configuration options are stored for each parameter separately:

```
In [40]: vcsese.configs.varnames
Out[40]: ['Lambda', 'Rho']
In [41]: vcsese.configs
Out[41]: {Lambda:{proposal:<scipy.stats._continuous_distns.norm_gen object at 0x7f8}</pre>
```

So, for example, if we wanted to turn off adaptation in the upper-level parameter, and fix the Metrpolis jump variance to . 25:

Another example might be to fix the prior mean of β to the OLS estimates. One way this could be done would be to pull the Delta matrix out from the state, and estimate:

$$Y = X\beta + \Delta Z + \epsilon$$

using PySAL:

If you wanted to start the sampler at a given starting value, you can do so by assigning that value to the Lambda value in state.

```
In [45]: vcsese.state.Lambda = -.25
```

Changing the spatial parameter priors is also done by changing their prior in state. This prior must be a function that takes a value of the parameter and return the log of the prior probability for that value.

For example, we could assign $P(\lambda) = Beta(2,1)$ and zero if outside (0,1), and asign ρ a truncated $\mathcal{N}(0,.5)$ prior by first defining their functional form:

5.0.1 Performance

The efficiency of the sampler is contingent on the lower-level size. If we were to estimate the draw in a dual-level SAR-Error Variance Components iteration:

```
In [50]: %timeit vcsese.draw()
1 loop, best of 3: 2.7 s per loop
```

It takes about 2 seconds per draw on my Chromebook.

To make it easy to work with the model, you can interrupt and resume sampling at any time:

```
In [51]: %time vcsese.sample(100)
CPU times: user 13min 44s, sys: 22.2 s, total: 14min 7s
Wall time: 5min 21s
In [52]: vcsese.sample(10)
```

5.0.2 Backends

Sometimes, the sample trace is too big to keep in memory at once. So, I wrote tools to keep the trace out of core in a sqlite database.

```
In [53]: from hlm.sqlite import start_sql, trace_from_sql
```

By default, the sampler will not allow you to overwrite the existing model trace on file.

```
In [56]: #!rm model_trace.db
In [57]: vcse = hlm.upper.SE(Y, X, M=W2, membership=membership, n_samples=0, tuning vcse.database = 'model_trace.db'
/home/ljw/Downloads/projects/hlm_gibbs/hlm/utils.py:91: ComplexWarning: Casting cor emax = emax.astype(float)
/home/ljw/Downloads/projects/hlm_gibbs/hlm/utils.py:92: ComplexWarning: Casting cor emin = emin.astype(float)
```

Then, if this option is set, the sample function automatically saves to disk every cycle, to prevent excessive memory pressure. If configuration options can be added to save every k iterations as well.

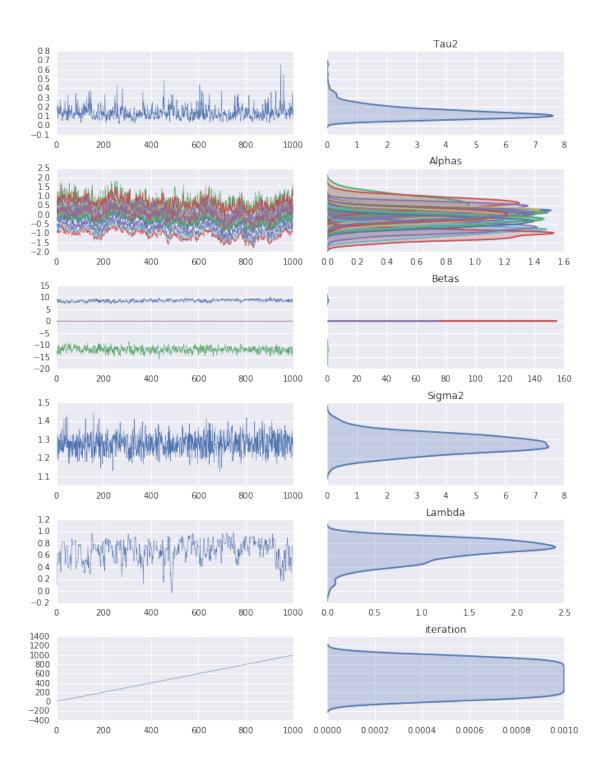
```
In [58]: vcse.sample(1000)
```

Since the trace is being kept on disk and not in the model, the trace in memory consists of a single point:

```
In [59]: vcse.trace
Out[59]: {Betas:[array([[ 7.96862039e+00],
                [-1.07834976e+01],
                [-3.52450551e-03],
                [ 3.48360192e-02]])], Sigma2:[1.2118589894190246], Tau2:[0.0838380
                [ 1.04561945],
                [ 1.12482033],
                [ 0.61758172],
                [ 0.38954034],
                [-0.64827243],
                [ 0.2696091 ],
                [ 0.20314601],
                [-0.89072907],
                [-0.3924088],
                [0.43979513],
                [-0.09320049],
                [-0.2246557],
                [ 0.0248566 ],
                [0.08191254],
```

[0.20912466]])]}

To get the full set of values, you can either read it in explicitly or work with it like it's a database using python's sqlite3 library.



6 Under the Hood

6.0.1 Non-Model tools

Most of the tools in the package are stored in relevant python files in the top level or a dedicated subfolder. Explaining a few:

- abstracts.py the abstract class machinery to iterate over a sampling loop. This is where the "mechanics" of the sampler is defined.
- trace.py the file containing tools and methods for the Trace class definition.
- sqlite.py the file containing model serialization tools and the out-of-core trace storage tools.
- plotting/ a folder containing tools for visualizing model output.
- steps.py a file containing step methods, like metropolis, slice, or inversion, used for the non-analytical posteriors.
- verify.py a file containing the checks on user input that are run for the User classes.

6.0.2 The implementation of a Model

The package is implemented so that every "model type" first sends off to the hlm.both.Base_Generic, which sets up the state, trace, and priors.

Models are added by writing a model.py file and possibly a sample.py file. The model.py file defines a Base/User class pair (like spreg) that sets up the state and trace. It must define hyperparameters, and can precompute objects used in the sampling loop.

The sampling loop itself is defined in sample.py. It must be a function that takes the model and updates it in place with a new draw from the joint posterior. This file can also split out each parameter's sample step into their own functions, which would allow users to configure alternative step methods on the fly.

Any extra functions used in the setup or estimation is stored in their util.py file.

So, if I write a new model, like a varying-intercept model with endogenously-lagged intercepts, I would write: - sample.py, describing a single sampling loop:

```
def sample(VISAR):
    st = VISAR.state
    diff = st.Ay - st.XBetas - st.Delta.dot(st.Alphas)
...
```

• model.py, describing the subclass to setup the model, maybe looking like:

```
class Base_VISAR(hlm.abstracts.Sampler_Mixin):
    def __init__(self, Y, X, M, membership=None, Delta=None):
    ...
```

Somewhere in the model definition, model._sample must be assigned to the loop defined in sample.py. Then, all of the rest of the mechanics of sampling the model, like the trace maintenance and possible use of the sql backend, are taken care of in the parent class.

Right now, I've organized the directory structure with four folders, both_levels, upper_level, lower_level, and hierarchical, which is a catchall for the spatial econometric models with endogenous effects or the Spatially-Varying Coefficient Process model.

The models implemented for the variance components specifications work by using hlm.both_levels.generic and customizing the structure of the covariance in each level. These covariance variables are stored in the state.Psi_1, for the lower-level covariance, and state.Psi_2 for the upper-level covariance.

For example:

```
In [62]: vcse.state.Psi_1 #lower-level covariance
Out[62]: <function hlm.utils.ind_covariance>
In [63]: vcse.state.Psi_2 #upper-level covariance
Out[63]: <function hlm.utils.se_covariance>
In [64]: vcsma.state.Psi_2 #upper-level covariance
Out[64]: <function hlm.utils.sma_covariance>
```

The functions that generate the covariance matrices are stored in hlm.utils. They can be arbitrarily overwritten for alternative covariance specifications.

Thus, if we want to sample a model with a new covariance specification, then a function that returns the covariance (the $\Psi(\lambda)$ function), must be written and assigned to the correct level in the hierarchy.

Models with higher levels can also be added by extending the sample loop in hlm.both_levels.generic to sample from more than two levels.