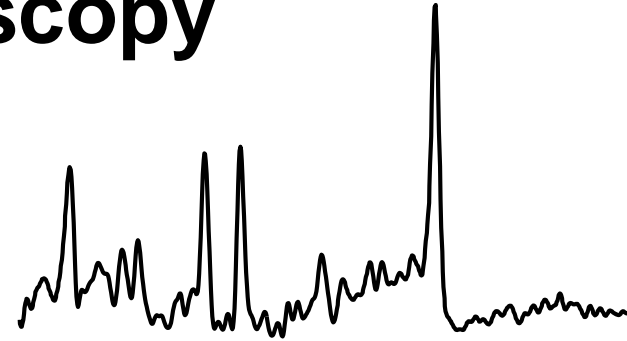


# MR Spectroscopy



T. Lange

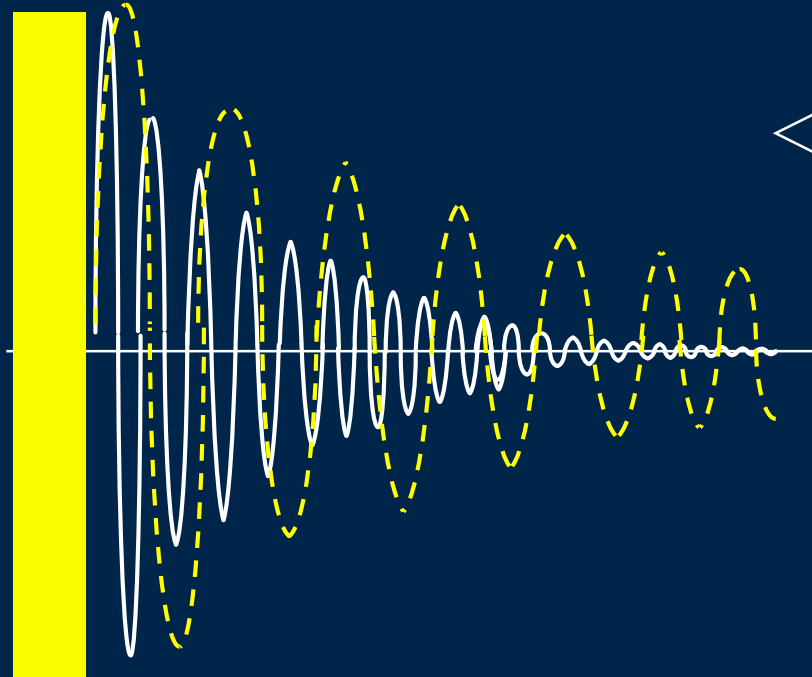
Dept.of Radiology, Medical Physics



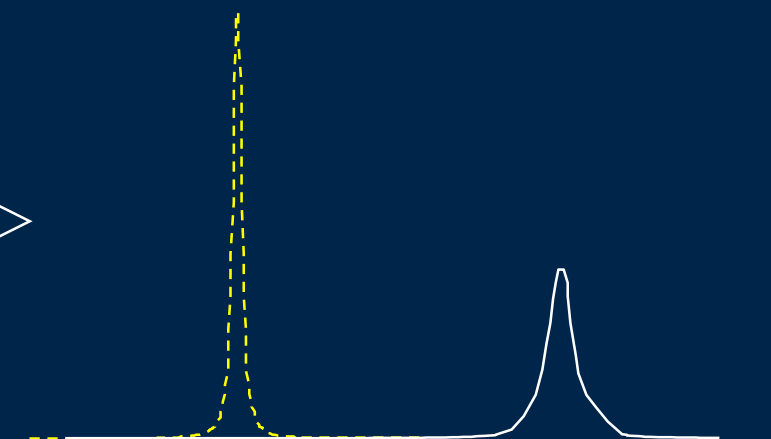
**UNIVERSITÄTS  
KLINIKUM FREIBURG**

# FID and Spectrum

Excitation  
pulse



FT



Spectrum

Free induction decay

# Magnetic Resonance Larmor Frequency

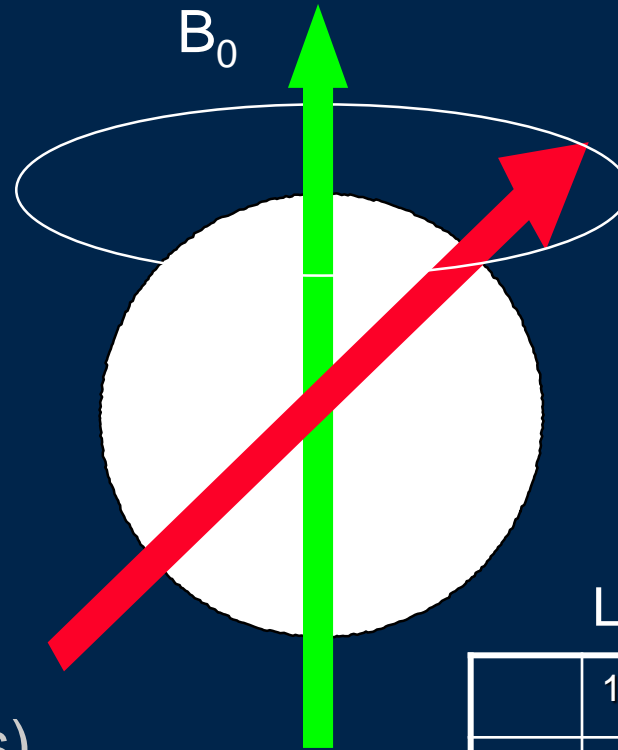
$$\nu_0 = \gamma^* \times B_0$$

$$(\gamma^* = \frac{\gamma}{2\pi})$$

$\gamma$ : gyromagnetic ratio  
(property of nucleus)

$$\gamma^*_{\text{H}} = 42.577 \text{ MHz/T}$$

$$\gamma^*_{\text{P}} = 17.235 \text{ MHz/T}$$



Lamor frequency:

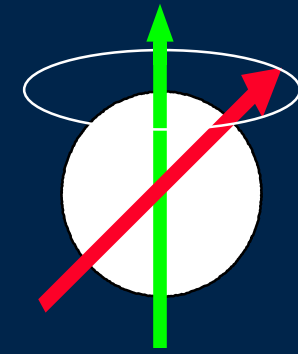
	1.5 T	3 T
$^1\text{H}$	63.86 MHz	127.73 MHz
$^{31}\text{P}$	25.85 MHz	51.7 MHz

# Biologically Important NMR-Visible Nuclei

Relative Sensitivity  $\sim |\gamma|^3 \cdot \text{NA}$  (NA = natural abundance)

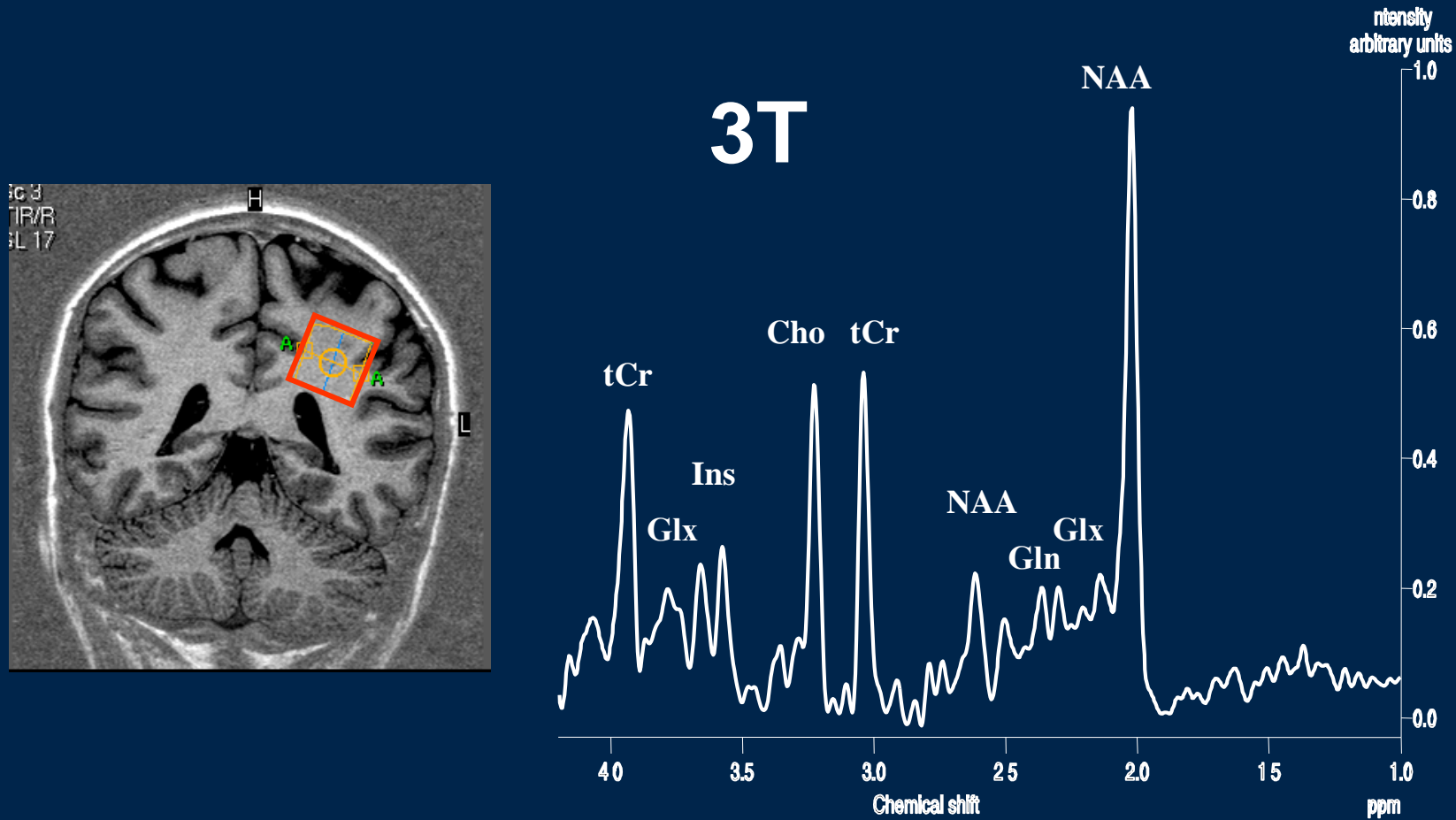
	Spin- quantum number	Gyro- magnetc ratio $\gamma^* = \gamma/2\pi$	Natural abundance [%]	Relative sensitivity for equal number of spins and constant magnetic field strenght	Relative sensitivity corrected for natural abundance
<b><math>^1\text{H}</math></b>	1/2	42.58	99.98	1.00	1.00
<b><math>^{13}\text{C}</math></b>	1/2	10.71	1.11	$1.59 \cdot 10^{-2}$	$1.8 \cdot 10^{-4}$
<b><math>^{14}\text{N}</math></b>	1	3.08	99.64	$1.01 \cdot 10^{-3}$	$1.0 \cdot 10^{-3}$
<b><math>^{17}\text{O}</math></b>	5/2	5.77	0.04	$2.91 \cdot 10^{-2}$	$1.1 \cdot 10^{-5}$
<b><math>^{19}\text{F}</math></b>	1/2	40.06	100.00	$8.30 \cdot 10^{-1}$	$8.3 \cdot 10^{-1}$
<b><math>^{23}\text{Na}</math></b>	3/2	11.26	100.00	$9.27 \cdot 10^{-2}$	$9.3 \cdot 10^{-2}$
<b><math>^{31}\text{P}</math></b>	1/2	17.24	100.00	$6.64 \cdot 10^{-2}$	$6.6 \cdot 10^{-2}$
<b><math>^{39}\text{K}</math></b>	3/2	1.99	93.08	$5.08 \cdot 10^{-4}$	$4.7 \cdot 10^{-4}$
<b><math>^{43}\text{Ca}</math></b>	7/2	2.87	0.14	$6.40 \cdot 10^{-3}$	$9.3 \cdot 10^{-6}$

# Nuclei of Biological Interest



- $^1\text{H}$  metabolites: 1-100 mmol/l in the body, **high sensitivity**  
*Problems:* water suppression (110 mol/l !!),  
fat suppression  
overlapping of metabolite peaks
- $^{31}\text{P}$  ~ 10 mmol/l, important for studying **energy metabolism**  
*Problems:* low sensitivity (100\* lower than  $^1\text{H}$  MRS)  
fast T2 relaxation
- $^{13}\text{C}$  basic atom in organic molecules  
*Problems:* only 1% natural abundance of  $^{13}\text{C}$   
very low sensitivity  
→ Sensitivity can be enhanced with hyperpolarization!

# $^1\text{H}$ Spectrum of Brain at High Field

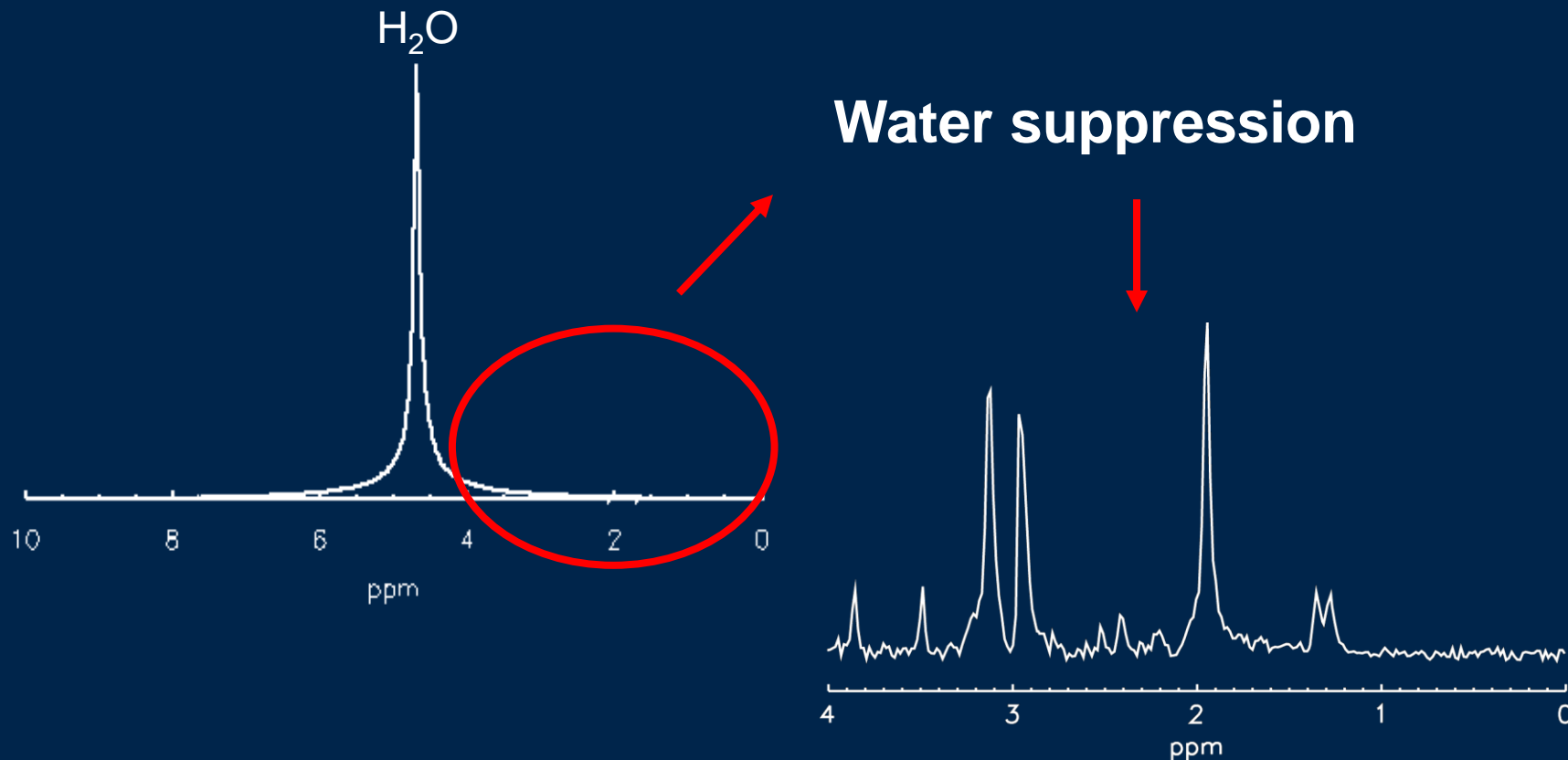


Courtesy: Dept. of Radiology, University of Bonn, Germany

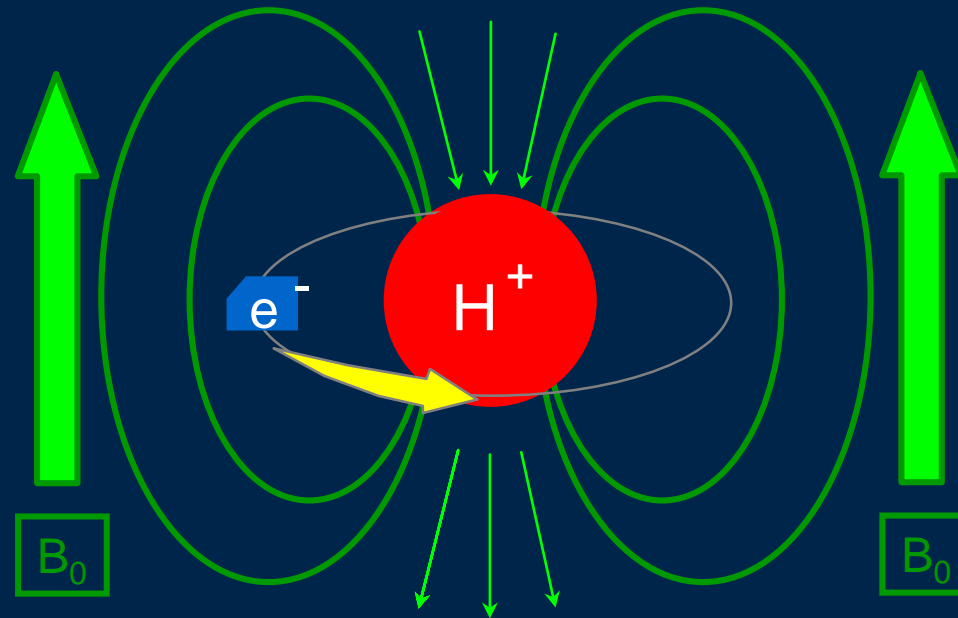
# *In vivo* MRS

Water concentration  $\sim 10^4$  times higher than metabolite concentrations

=> We are interested in the *small* peaks



# Chemical Shift - Shielding effect of the electron shell



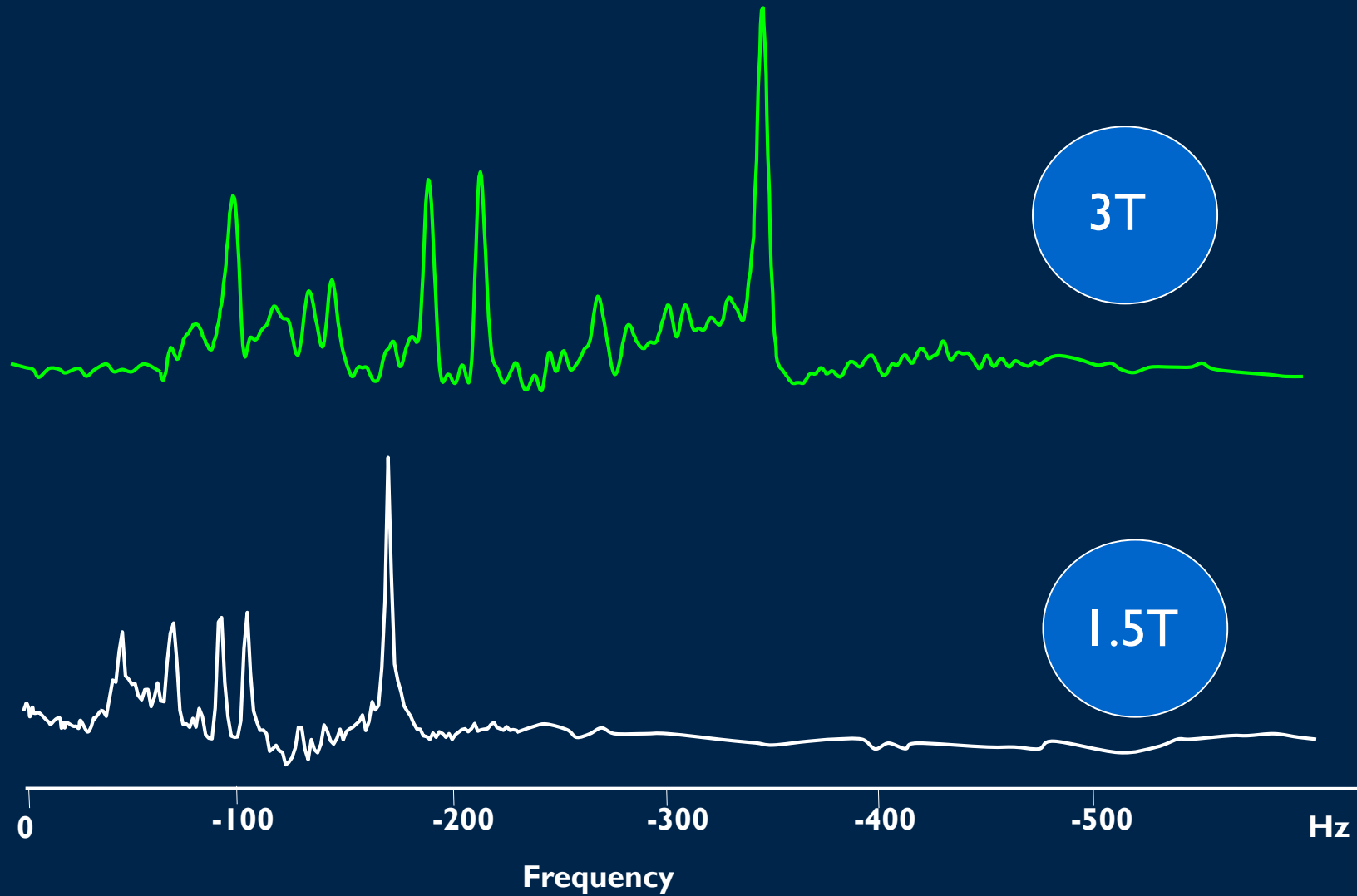
Shielding/deshielding of the nucleus by the electron shell:

- => different chemical environment for nuclei of same species
- => slightly different local magnetic field
- => slightly different Larmor frequency

The chemical shift scales with  $B_0$  and therefore also with the Larmor frequency!



# Hz Scaling

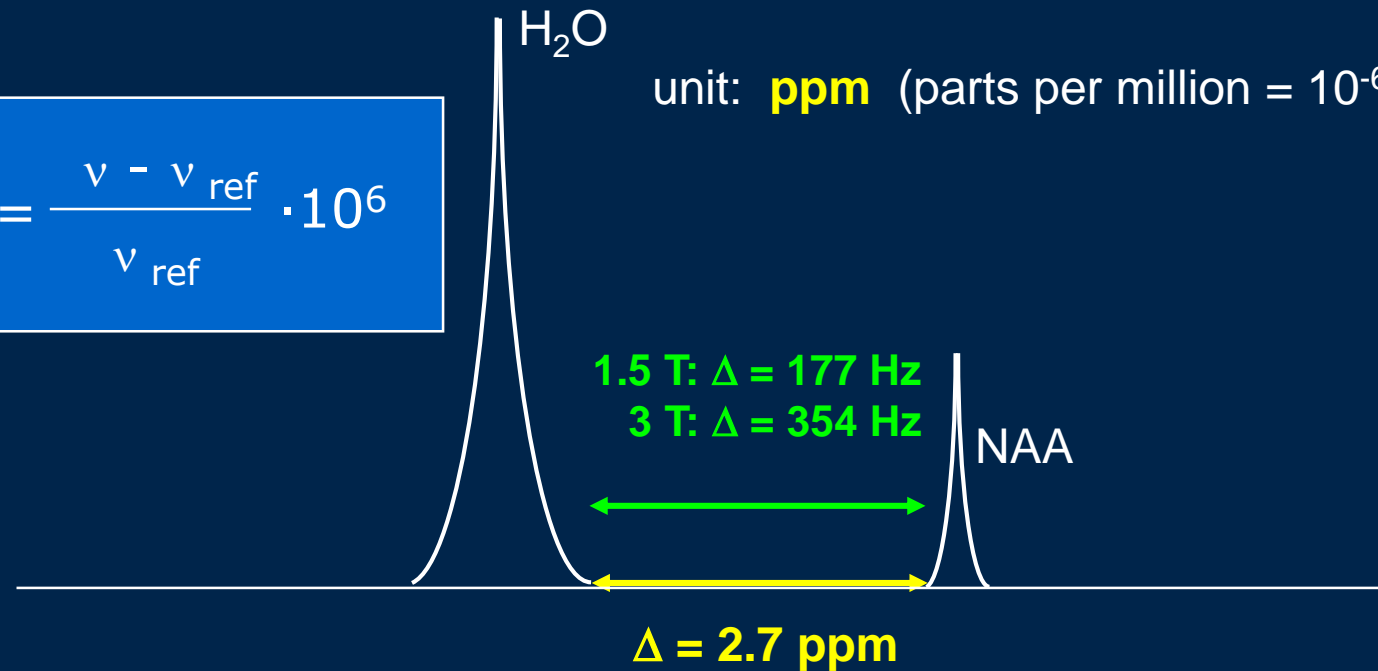


# Chemical Shift

$$\delta [\text{Hz}] = \nu - \nu_{\text{ref}}$$

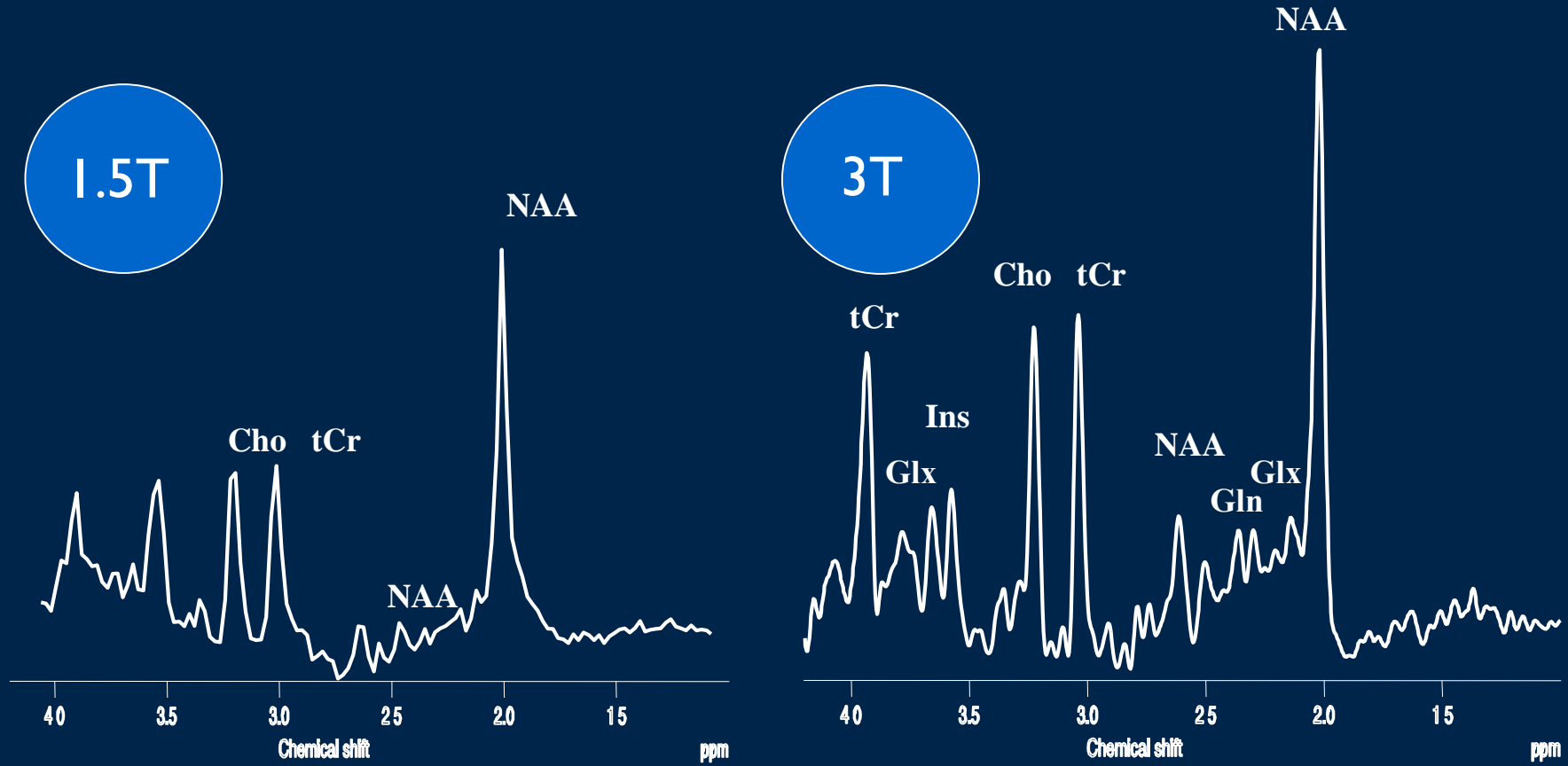
$$\delta [\text{ppm}] = \frac{\nu - \nu_{\text{ref}}}{\nu_{\text{ref}}} \cdot 10^6$$

unit: **ppm** (parts per million =  $10^{-6}$ )

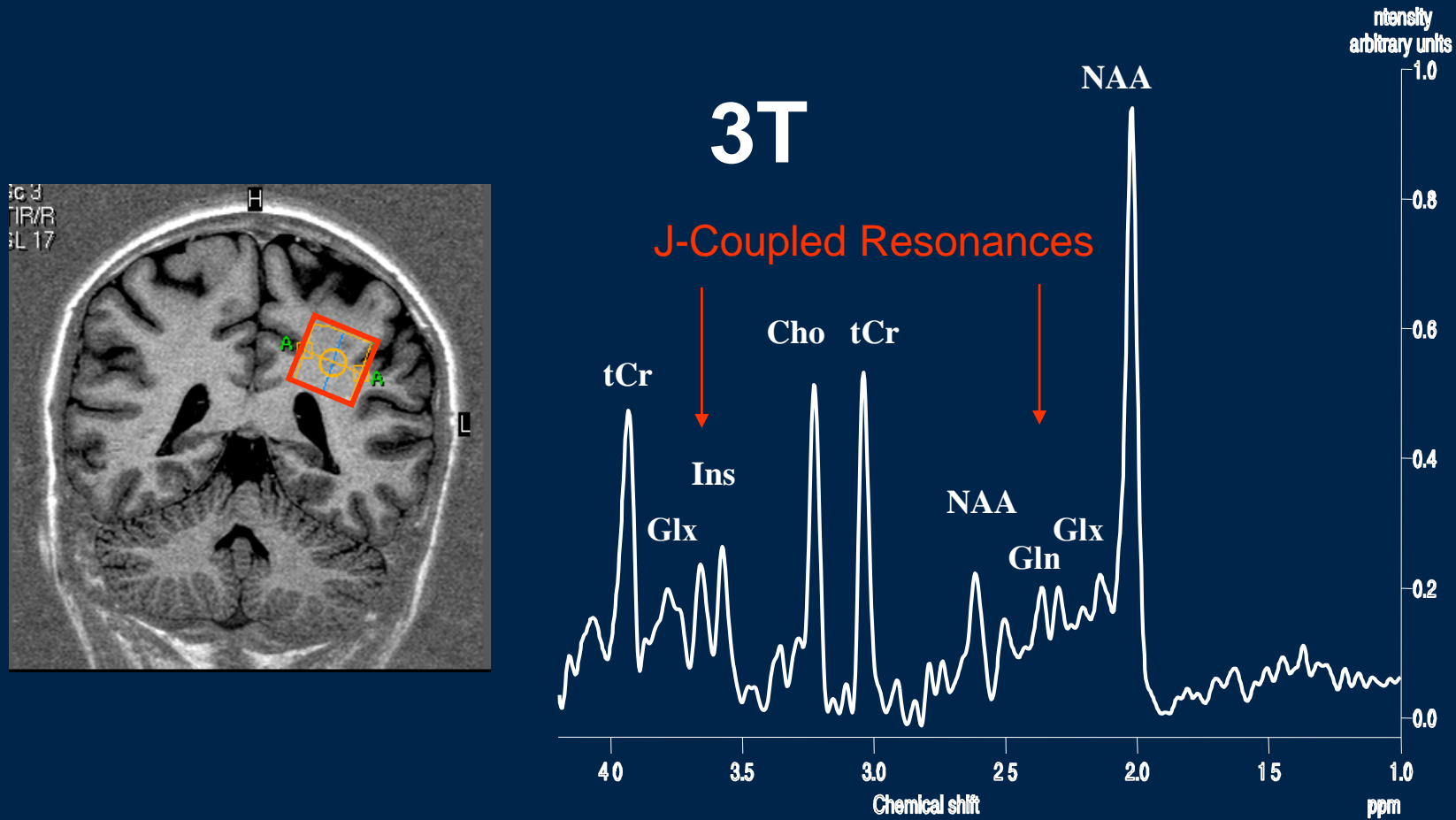


**$^1\text{H}$ :**  $\nu_{\text{ref}} = \nu_{\text{TMS}}$  (TMS: tetramethylsilane  $\text{Si}(\text{CH}_3)_4$ )

# ppm Scaling

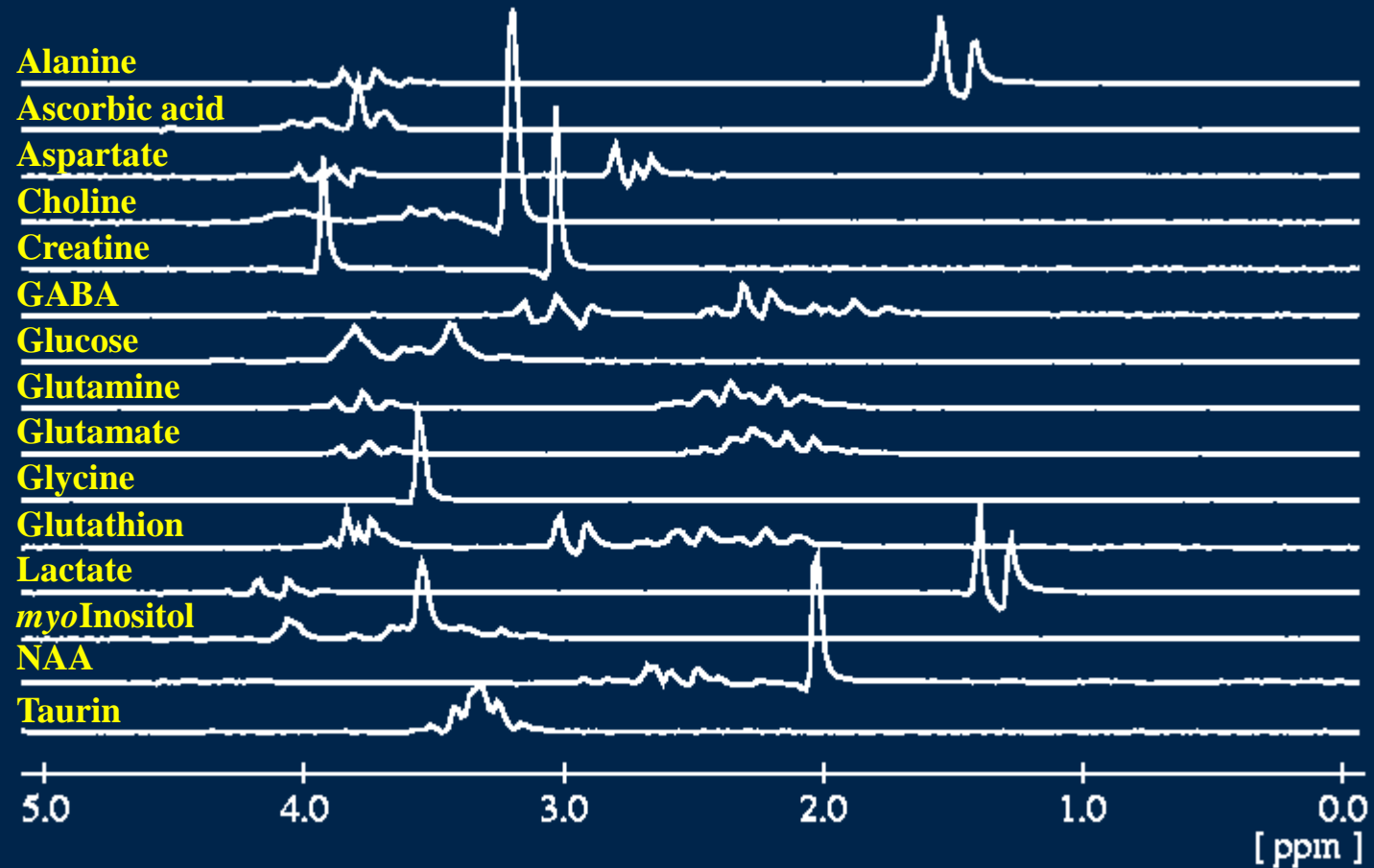


# Hydrogen Spectrum of Brain at High Field

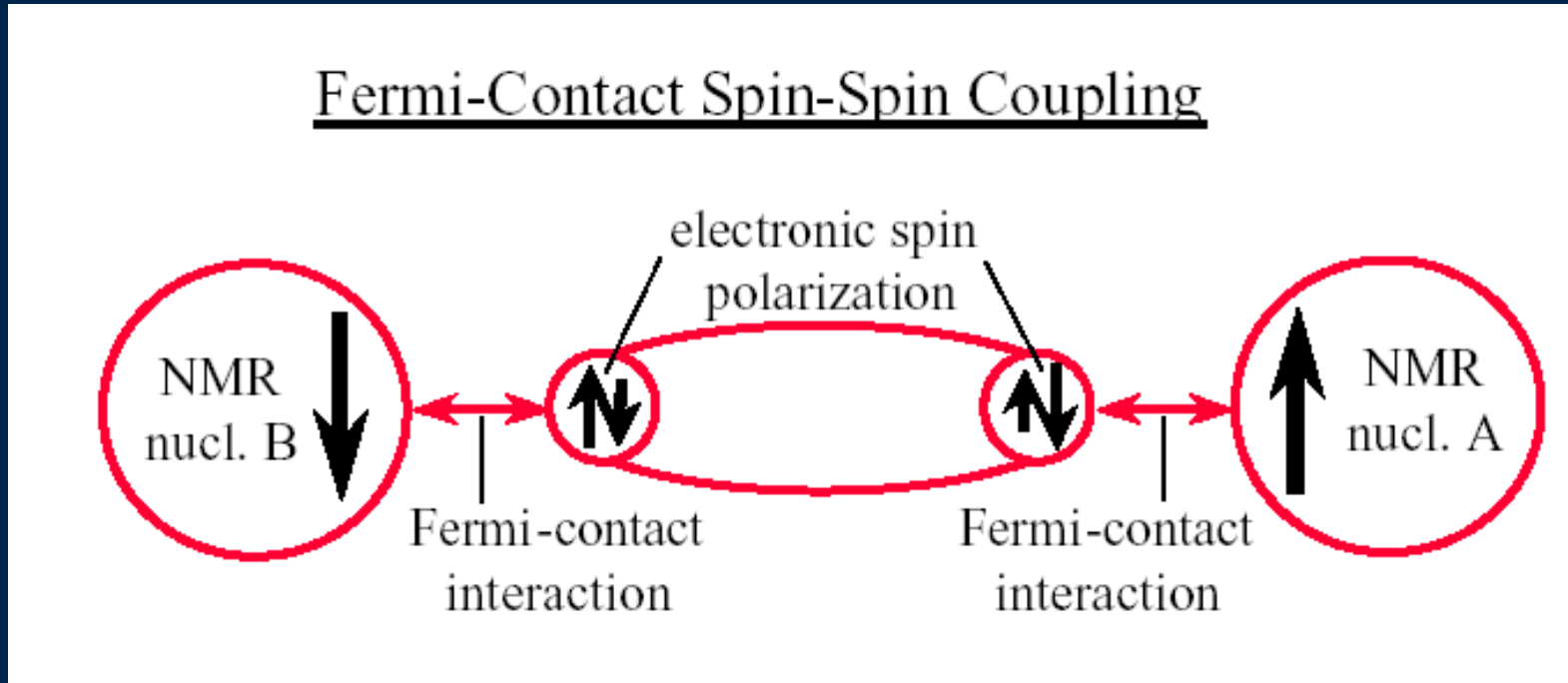


Courtesy: Dept. of Radiology, University of Bonn, Germany

# Metabolites seen in $^1\text{H}$ MR Spectroscopy



# J-coupling

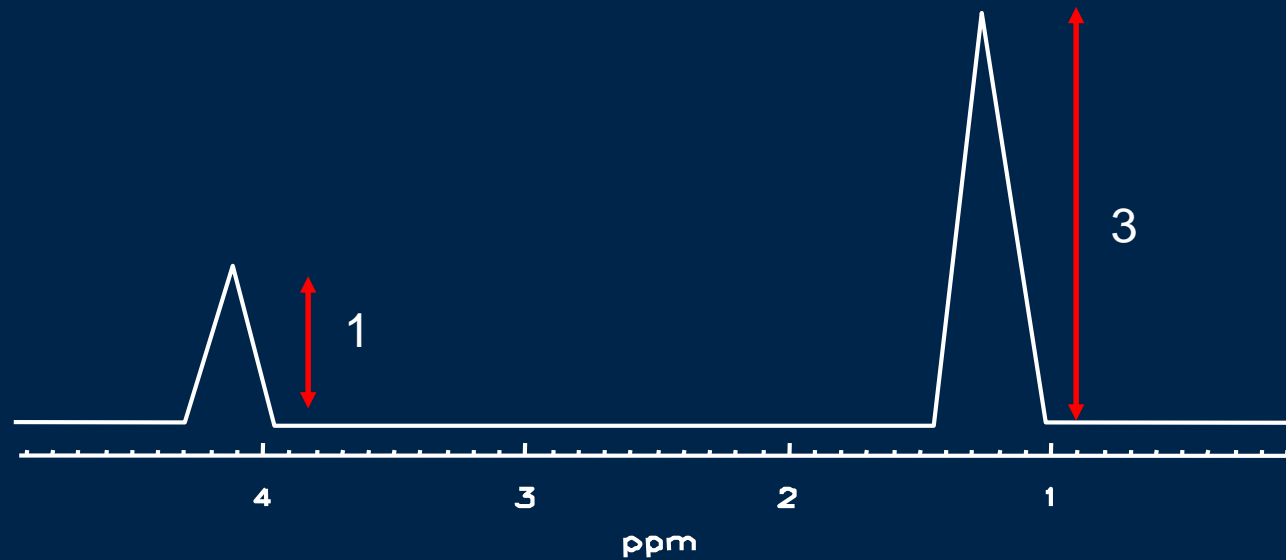
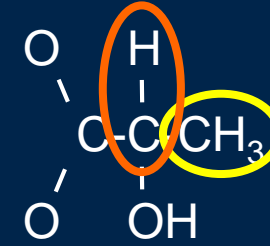


→ Intramolecular interaction

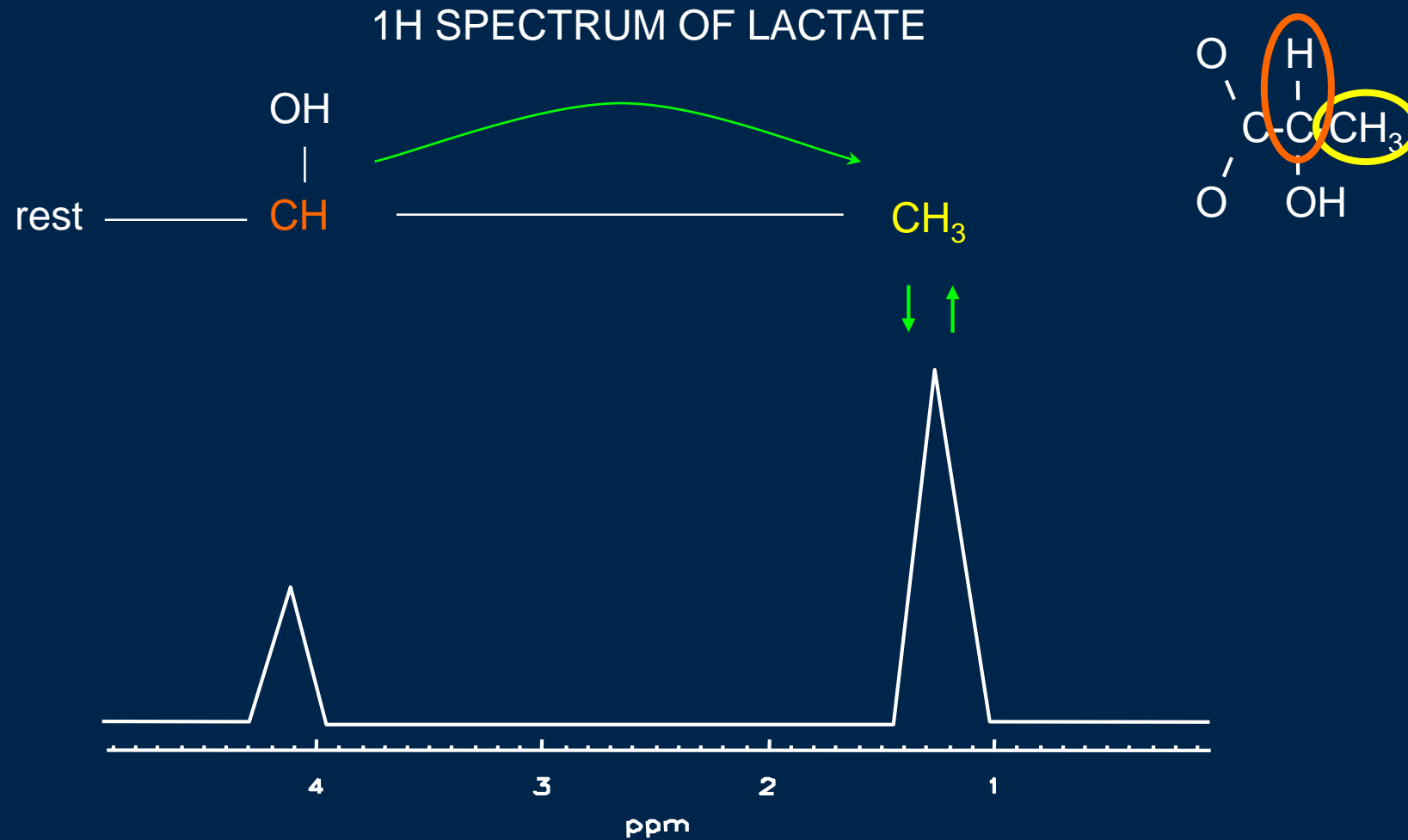
**J-coupling is equal at all field strengths !**

# J-coupling - Fine structure

<sup>1</sup>H SPECTRUM OF LACTATE

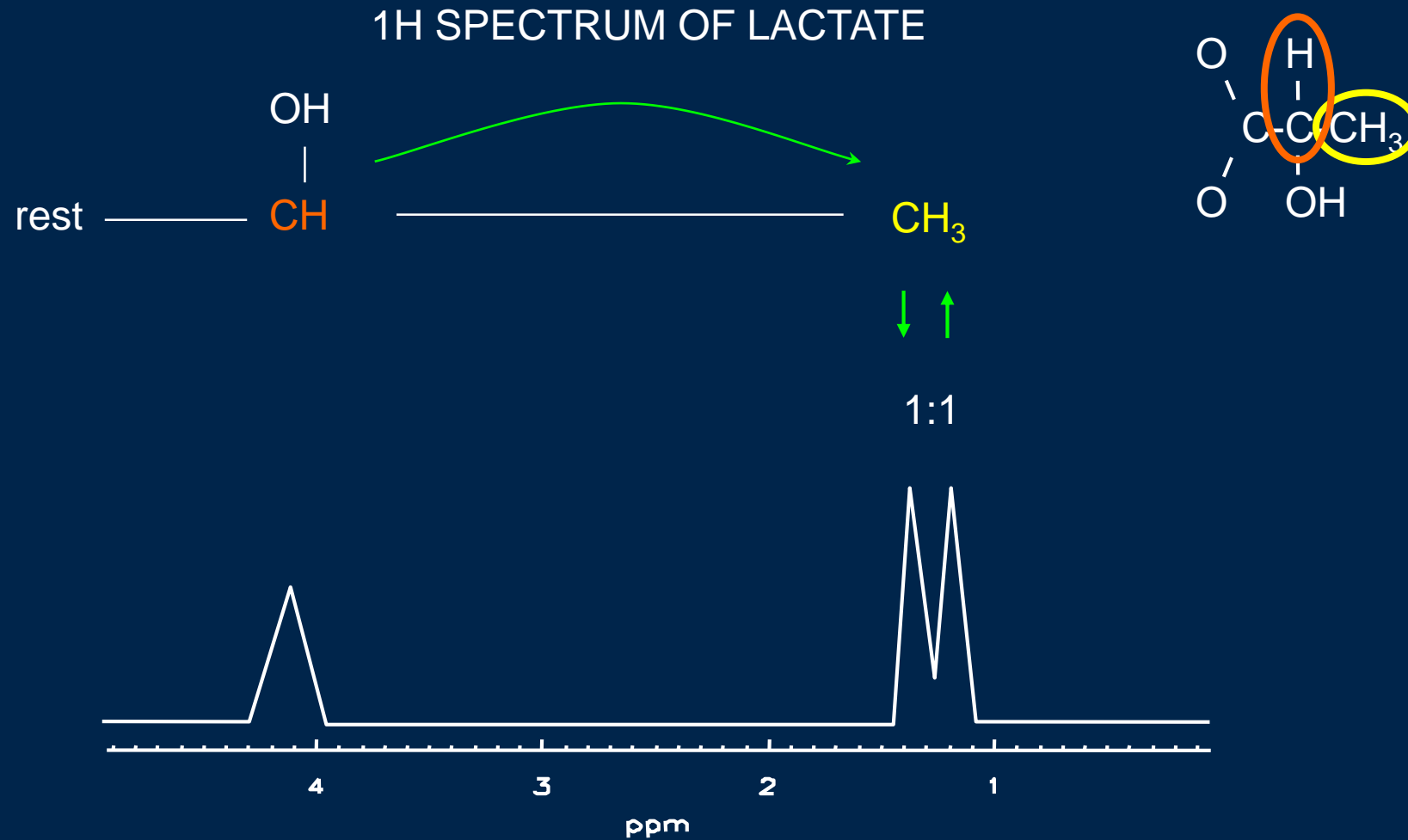


# J-coupling - Fine structure

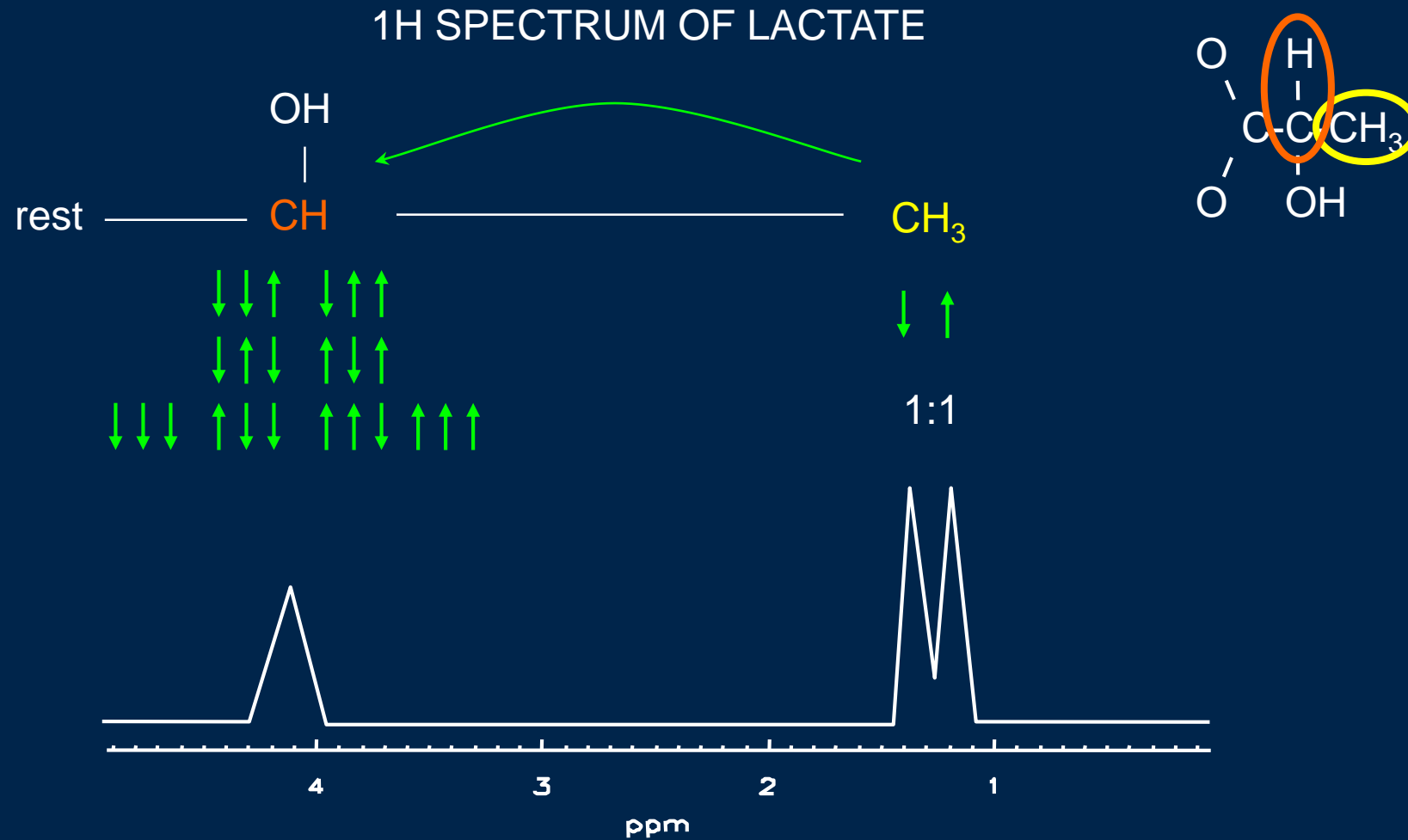




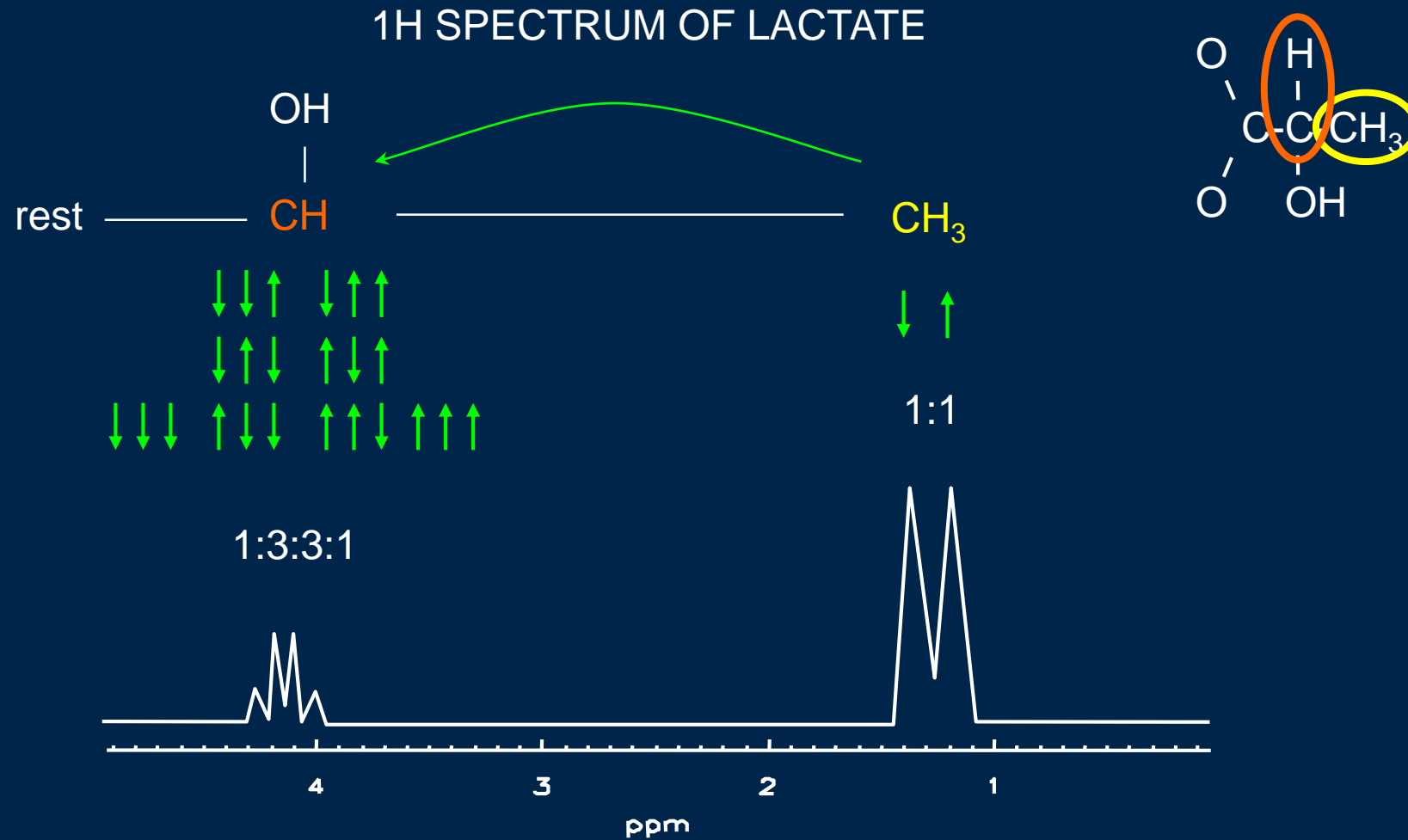
# J-coupling - Fine structure



# J-coupling - Fine structure



# J-coupling - Fine structure

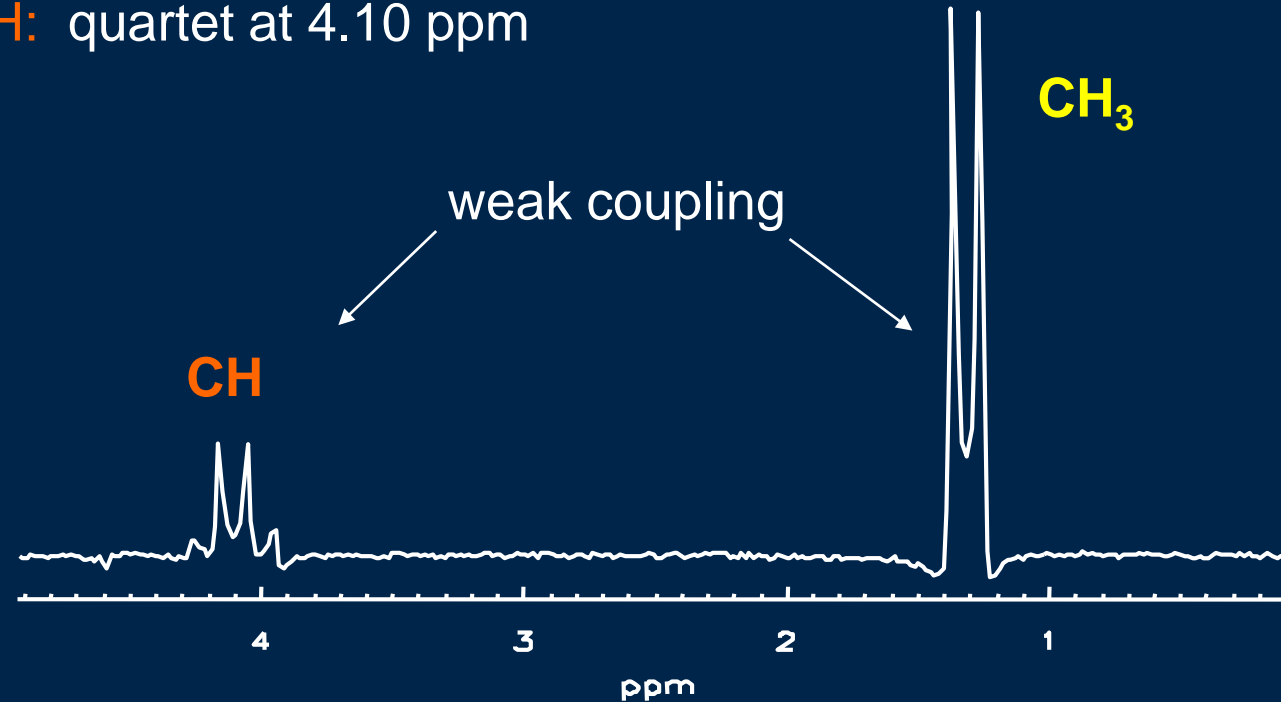
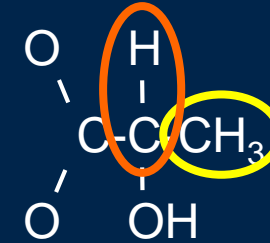


# J-coupling - Fine structure

## <sup>1</sup>H SPECTRUM OF LACTATE

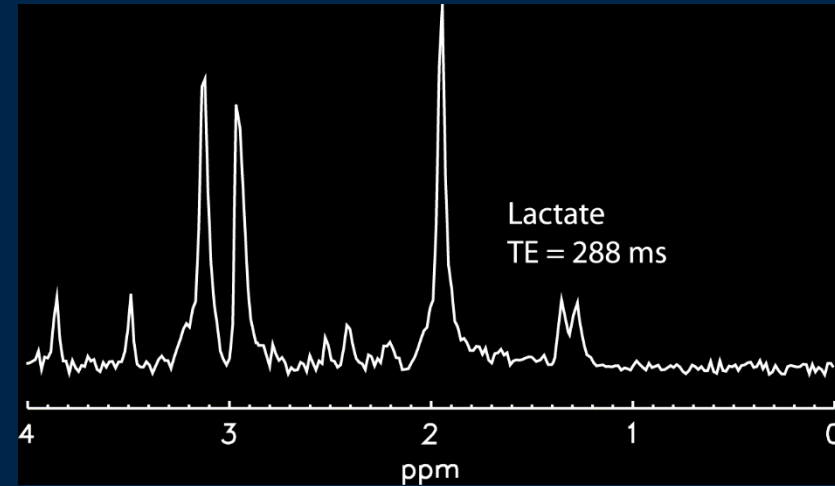
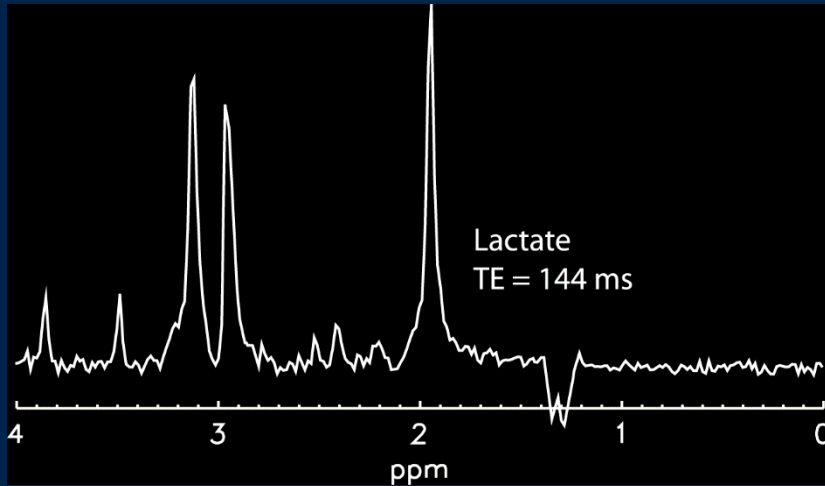
**CH<sub>3</sub>:** doublet at 1.31 ppm

**CH:** quartet at 4.10 ppm



# J-coupling - Evolution

Resonance shape is echo time dependent:



=> multiplet looks

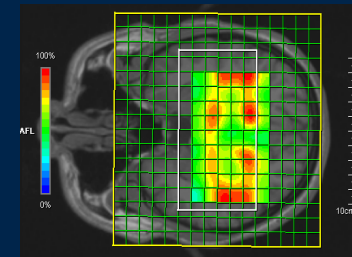
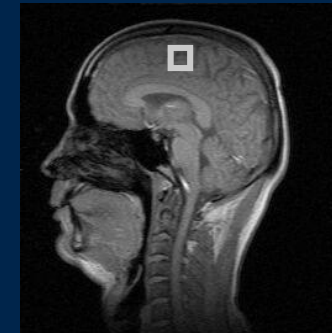
up at  $\text{TE} = n/J$  with even  $n$

down at  $\text{TE} = n/J$  with odd  $n$

# Localisation Techniques

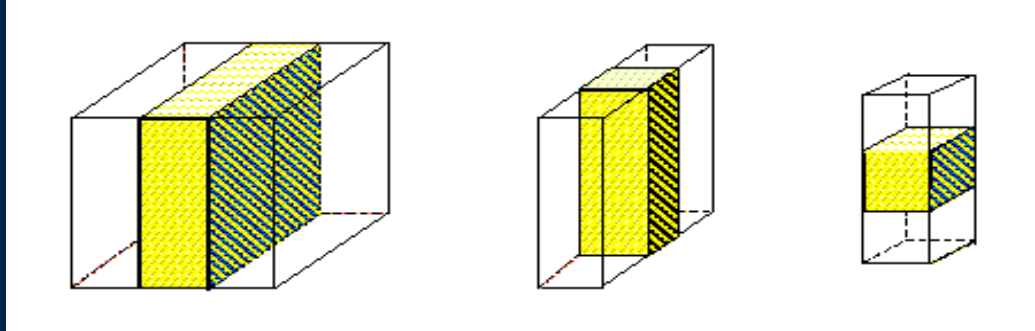
Purpose: Data collection from a well-defined volume of interest (VOI)

- Gradient localization with conventional pulses:
  - **PRESS**: Point RESolved Spectroscopy  
(Bottomley PA, *Ann N Y Acad Sci.* 1987;508:333-48)
  - **STEAM**: STimulated Echo Acquisition Method  
(Haase et al., *Radiology* 1986;160:787-790)
- Gradient localization with adiabatic pulses:
  - **LASER**: Localization by Adiabatic SElective Refocusing  
(Garwood M et al., *J Magn Reson* 2001 Dec;153(2):155-77)
- Localization via phase encoding: Chemical Shift Imaging (CSI)

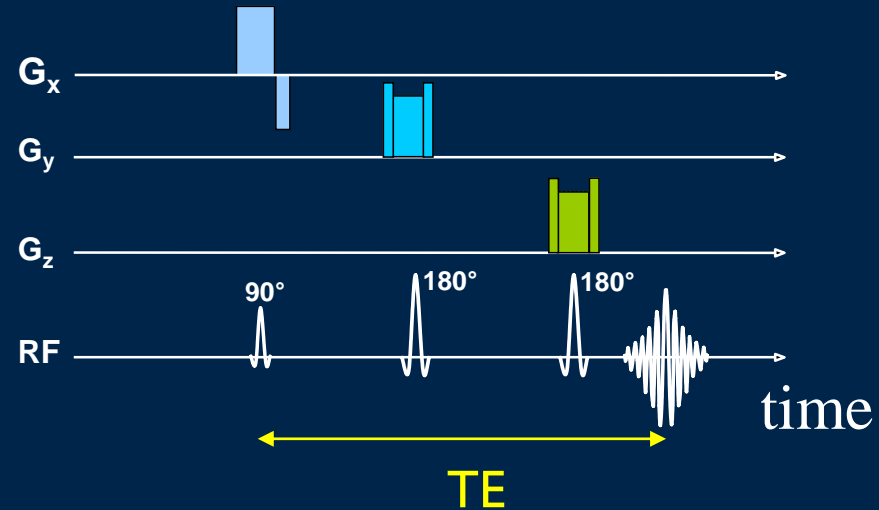


# Localisation by Gradients

In each direction a pulse in combination with a gradient field defines one slice  
=> the intersection of these slices is the selected volume of interest



# PRESS

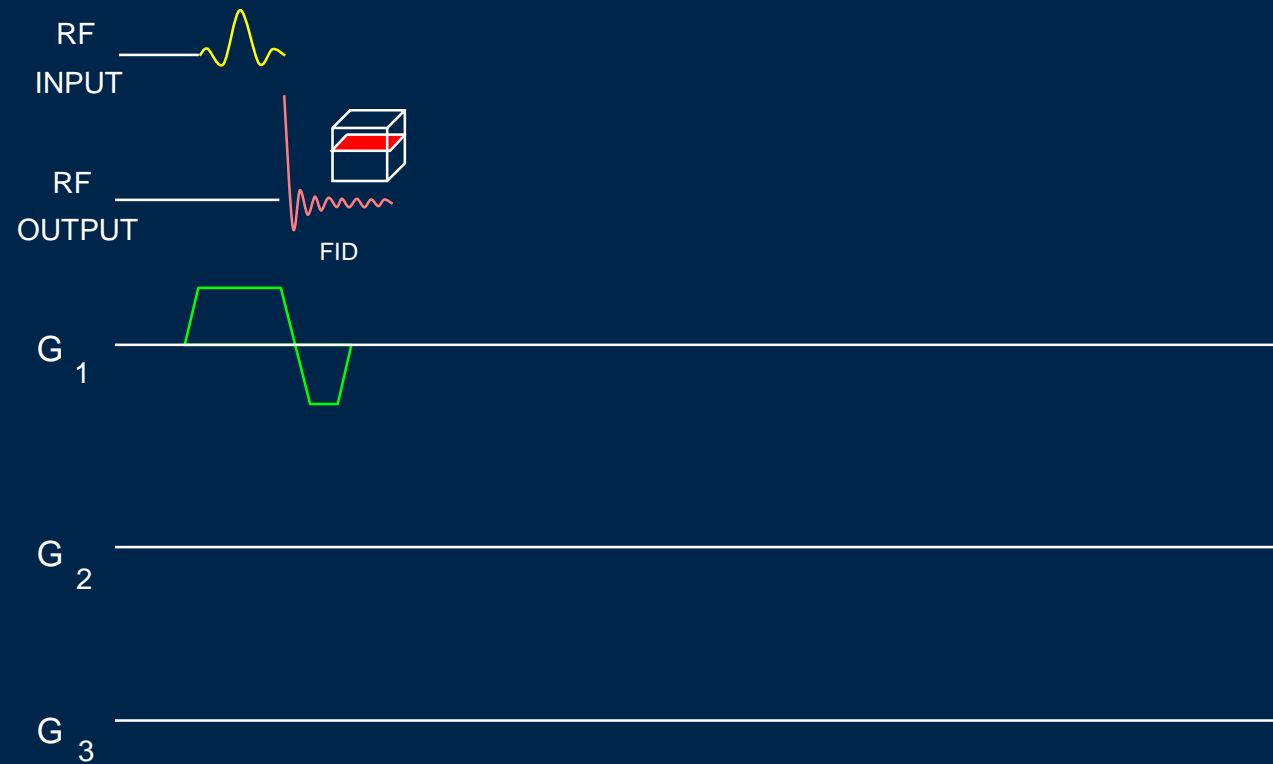


- (Point **RES**olved **S**pectroscopy)
- *selective excitation:*  
double spin echo sequence:  $90^\circ - 180^\circ - 180^\circ -$  echo acquisition
- spoiler/crusher gradients: eliminate unwanted coherences from outside the selected volume



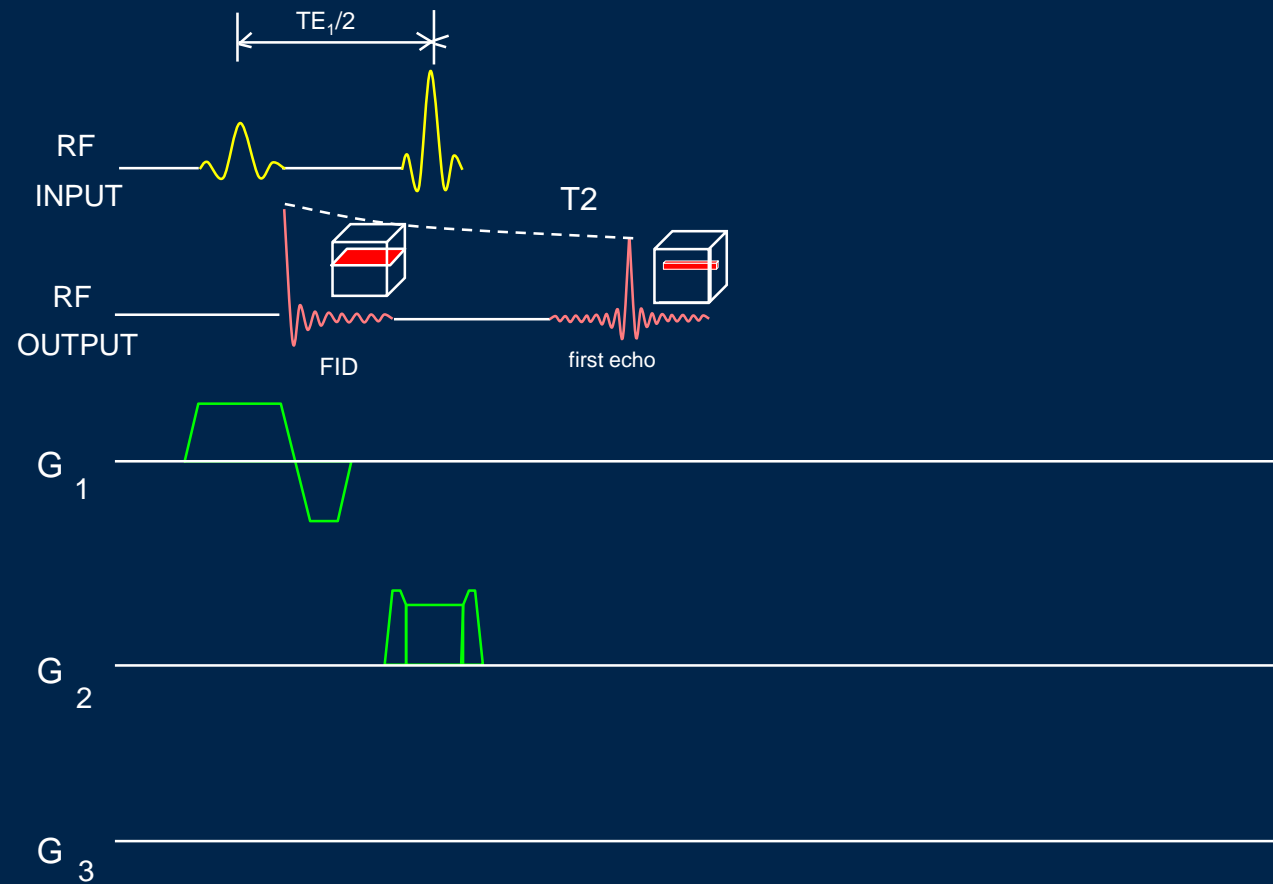
# PRESS

Method of choice for  $^1\text{H}$  spectroscopy



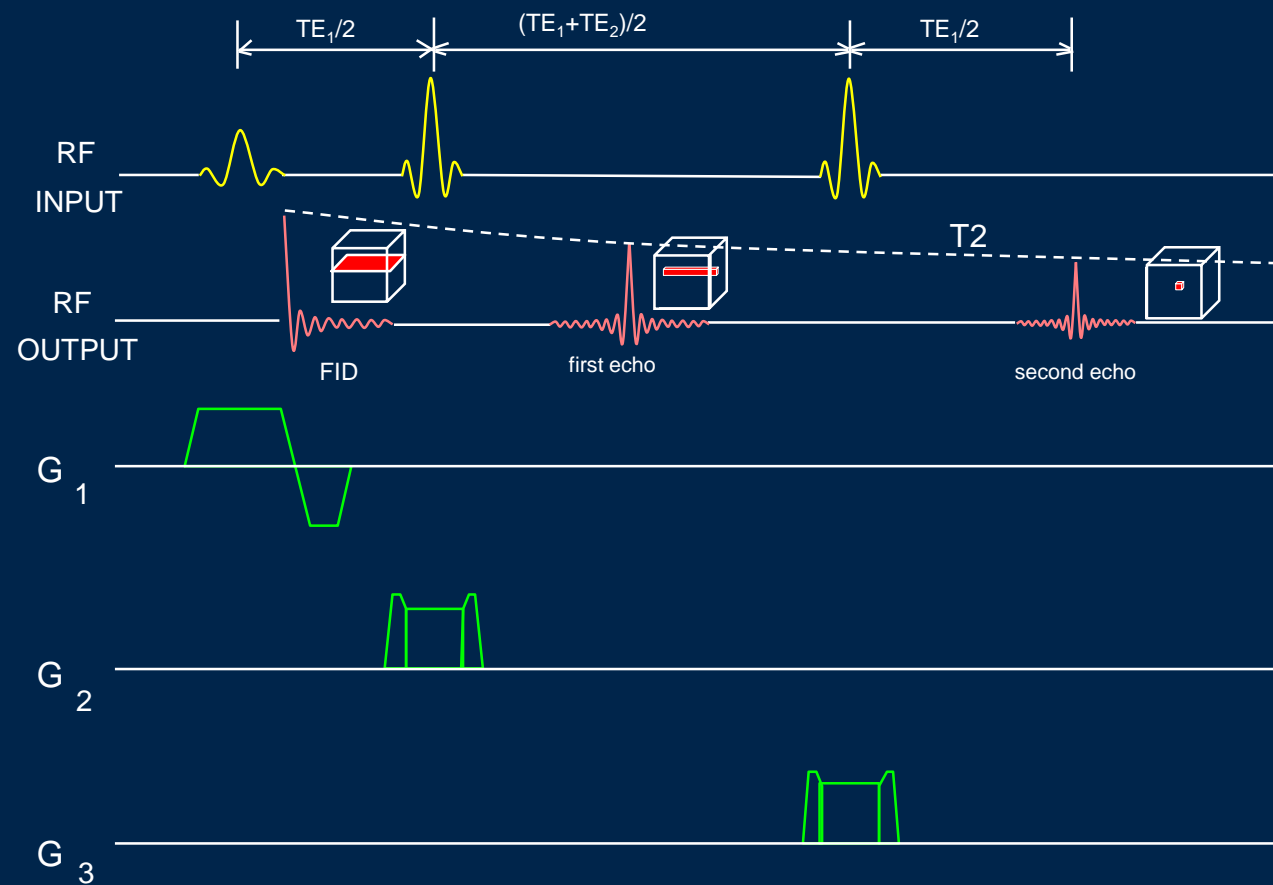
# PRESS

Method of choice for  $^1\text{H}$  spectroscopy

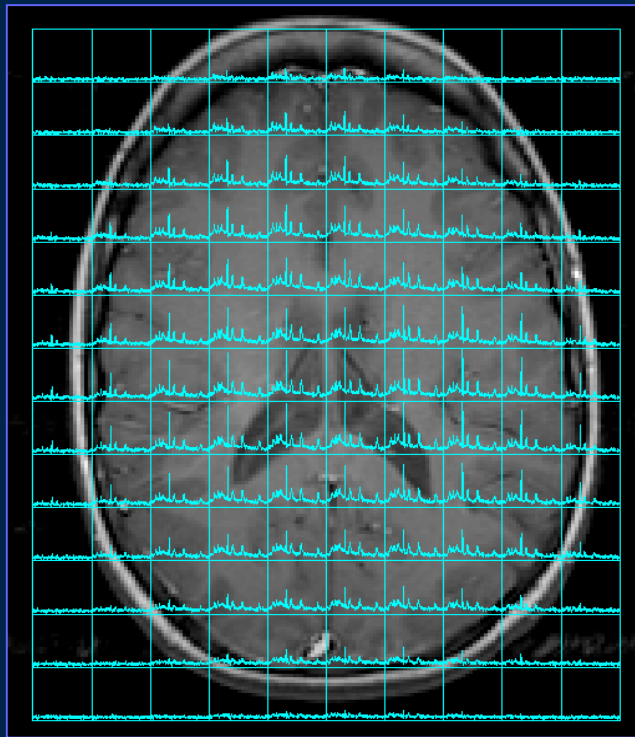


# Echo Volume Selection

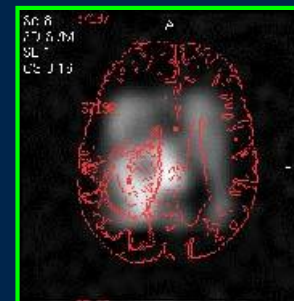
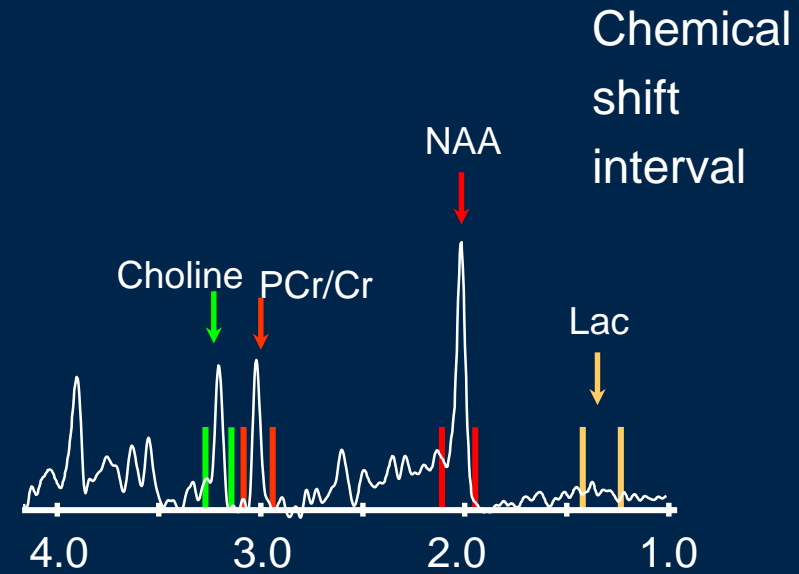
Method of choice for  $^1\text{H}$  spectroscopy



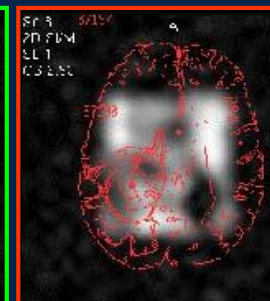
# Spectroscopic Imaging



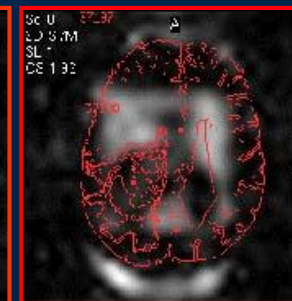
every voxel  
→  
a spectrum



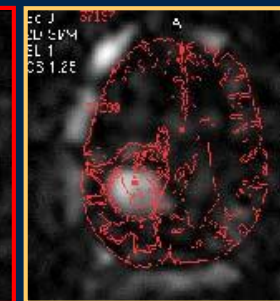
choline



creatine

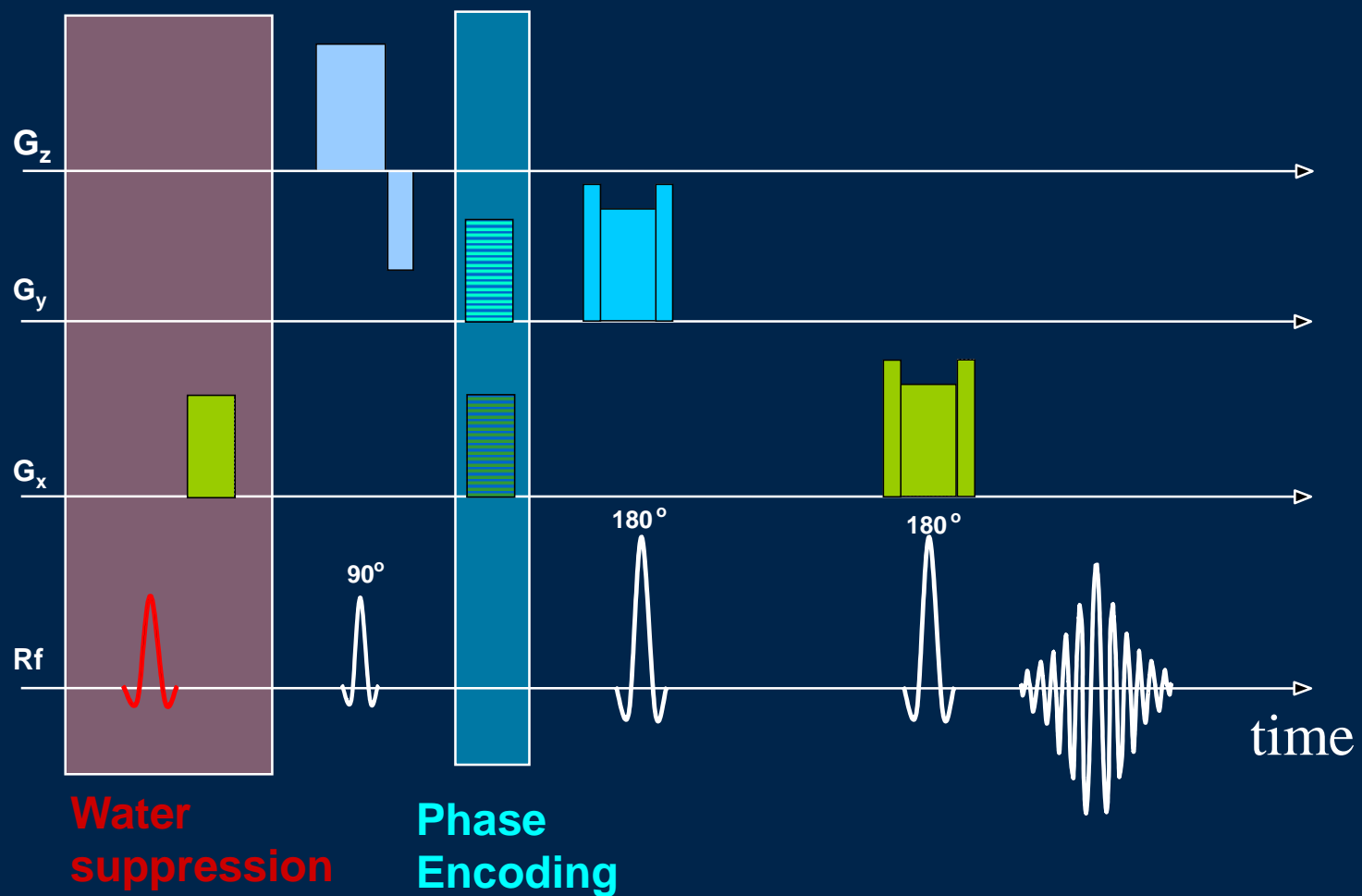


NAA

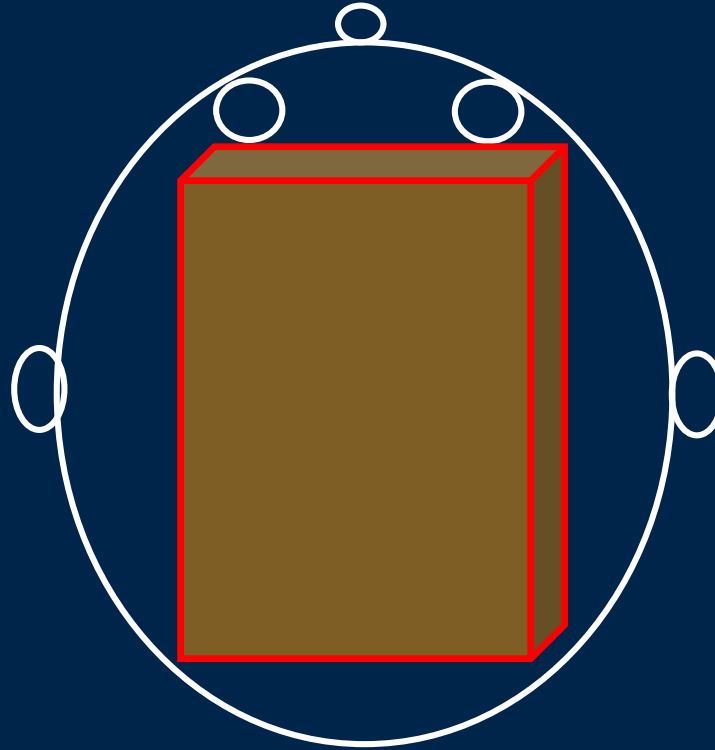


lactate

# 2D-SI Sequence with PRESS Localisation



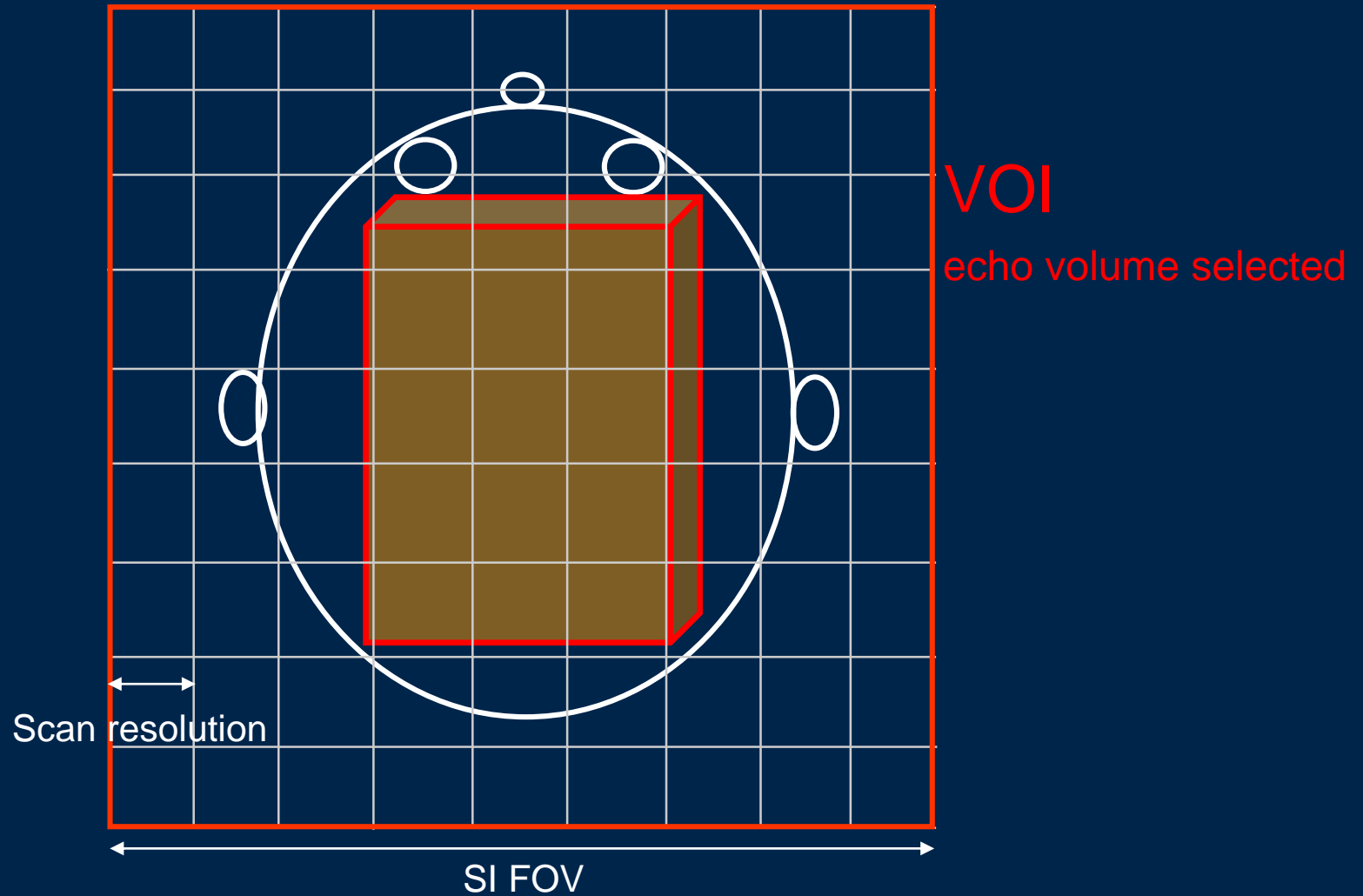
# 2D-Spectroscopic Imaging



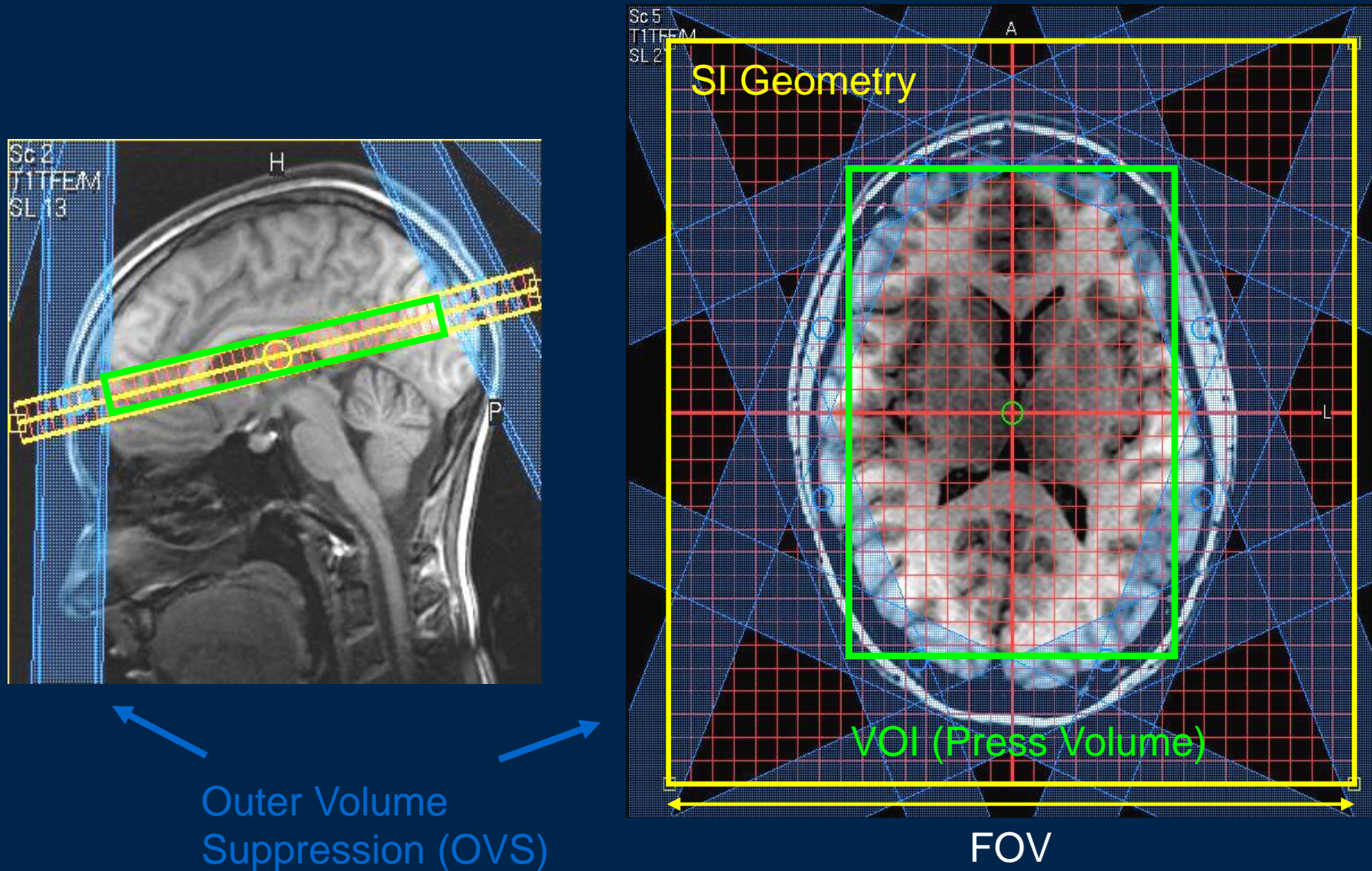
VOI

echo volume selected

# 2D-Spectroscopic Imaging



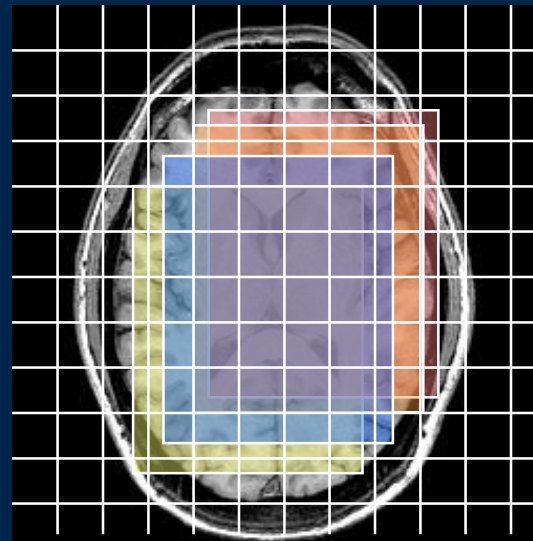
# 2D-SI: Outer Volume Suppression





# 2D-SI Sequence with PRESS Localisation

- **PRESS:** localisation slightly **different** for different metabolites / chemical shifts!

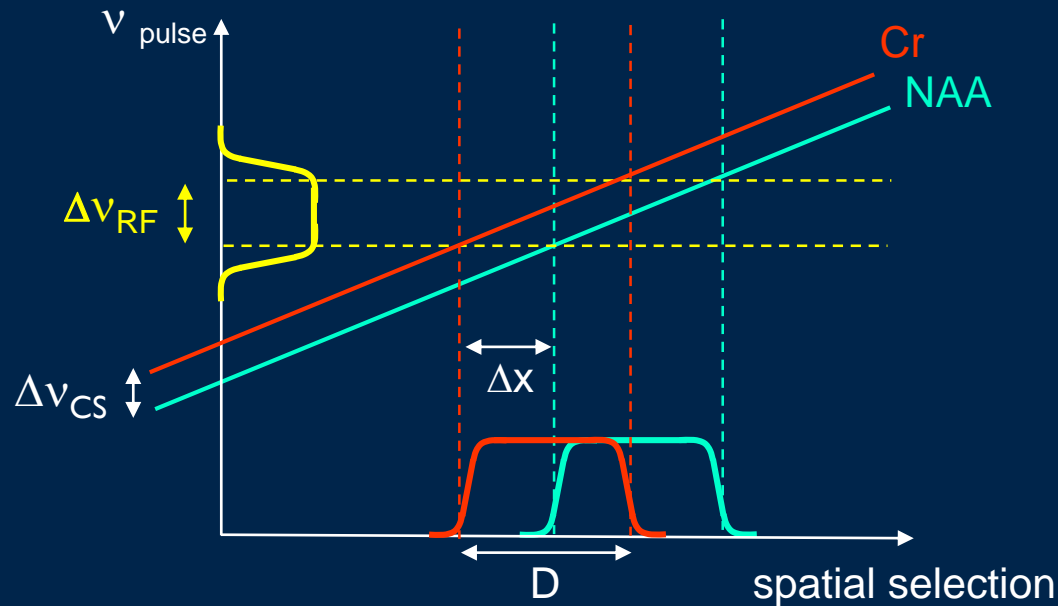


NAA  
Cr  
Cho  
Lac/Lip

- **Phase encoding:** localisation **equal** for all metabolites / chemical shifts !

# Chemical Shift Displacement

Chemical shift displacement artifact:



Gradient strength for given pulse bandwidth  $\Delta \nu_{\text{RF}}$  and slice thickness  $D$  :

$$G_x = \frac{\Delta \nu_{\text{RF}}}{\gamma^* \cdot D}$$

Spatial displacement:

$$\Delta x = \frac{\Delta \nu_{\text{CS}}}{\gamma^* \cdot G_x}$$

=> relative displacement:

$$\frac{\Delta x}{D} = \frac{\Delta \nu_{\text{CS}}}{\Delta \nu_{\text{RF}}}$$

# Spectroscopic Imaging vs. SVS

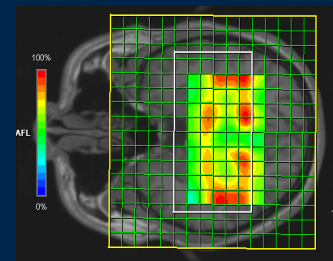
## *Single voxel spectroscopy:*

- 😊 good signal-to-noise ratio
- 😊 Rapid: ~2 to 6 min for 8 cc voxel
- 😞 selected volume is block-shaped  $\neq$  anatomical shape
- 😞 only information on one location

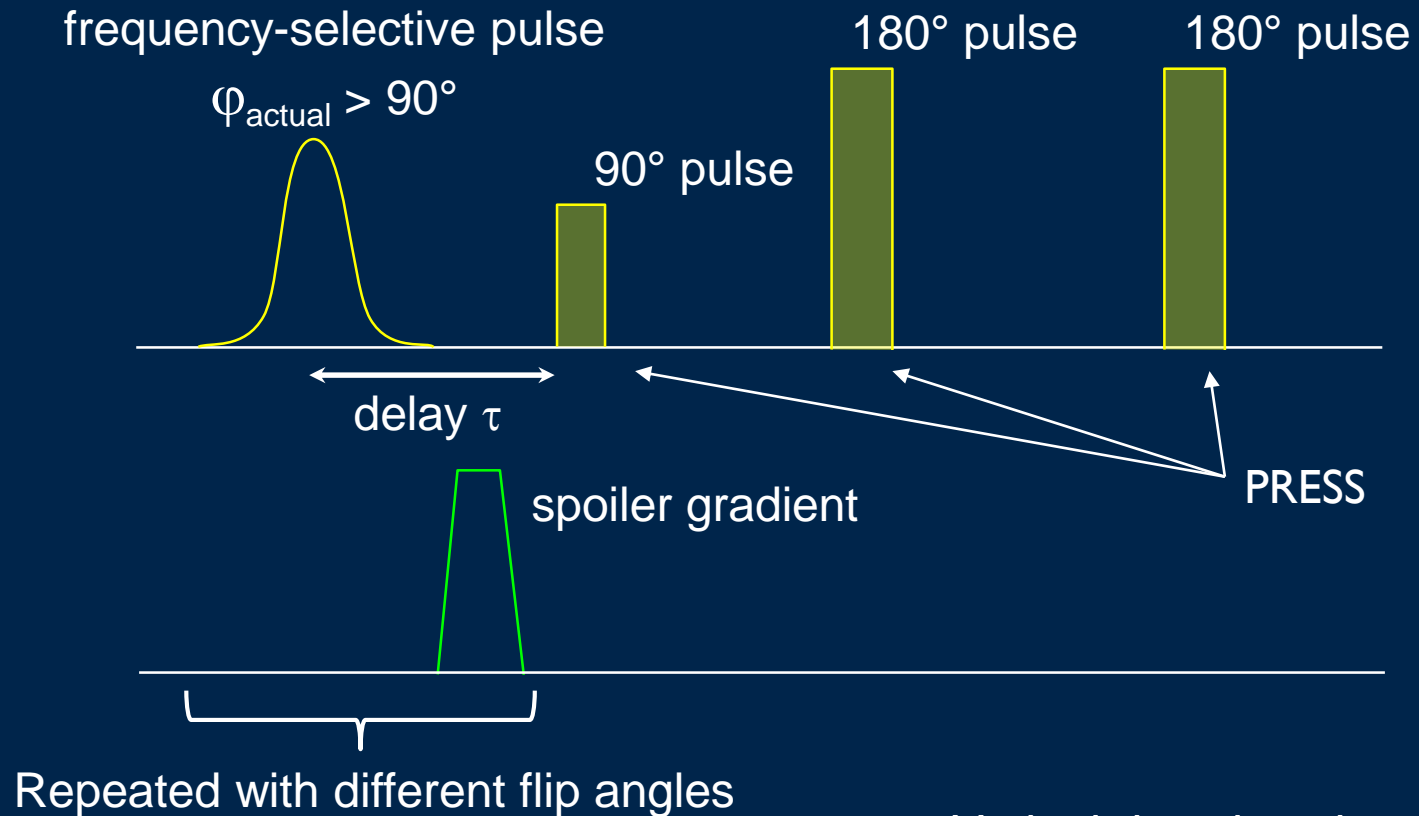


## *Spectroscopic imaging:*

- 😊 overview of spatial distribution of metabolites
- 😊 Usually higher resolution (~1 ml)
- 😞 mostly: longer acquisition times
- 😞 reduced signal-to-noise ratio (shim never as good as for SVS)
- 😞 signal leakage into neighbouring voxels (PSF)
- 😞 Slice should not go through air, bone, major vessels, fat



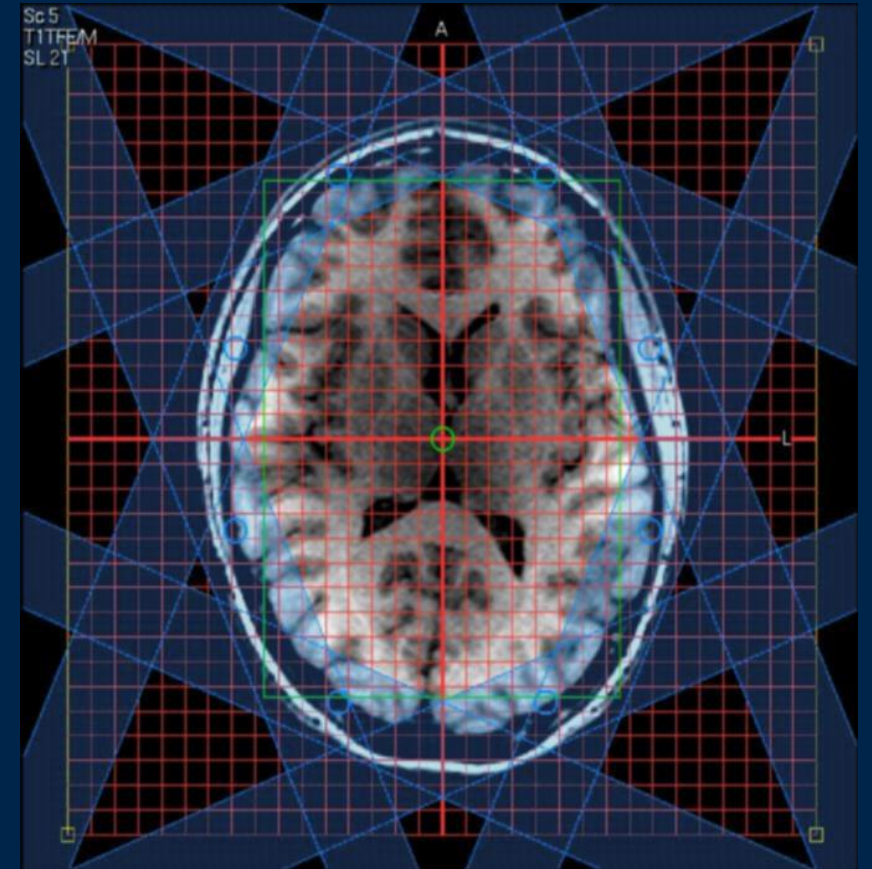
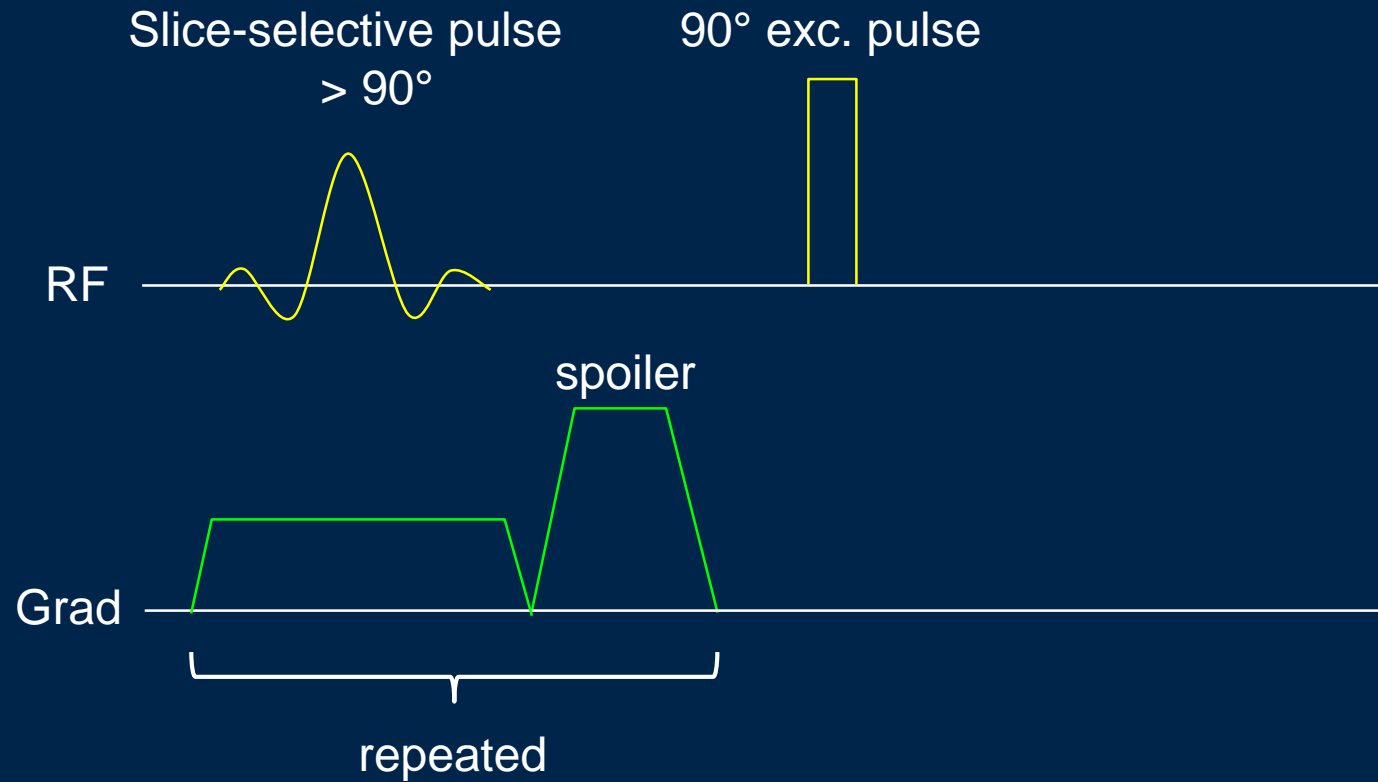
# Water Suppression



Methods based on this approach:

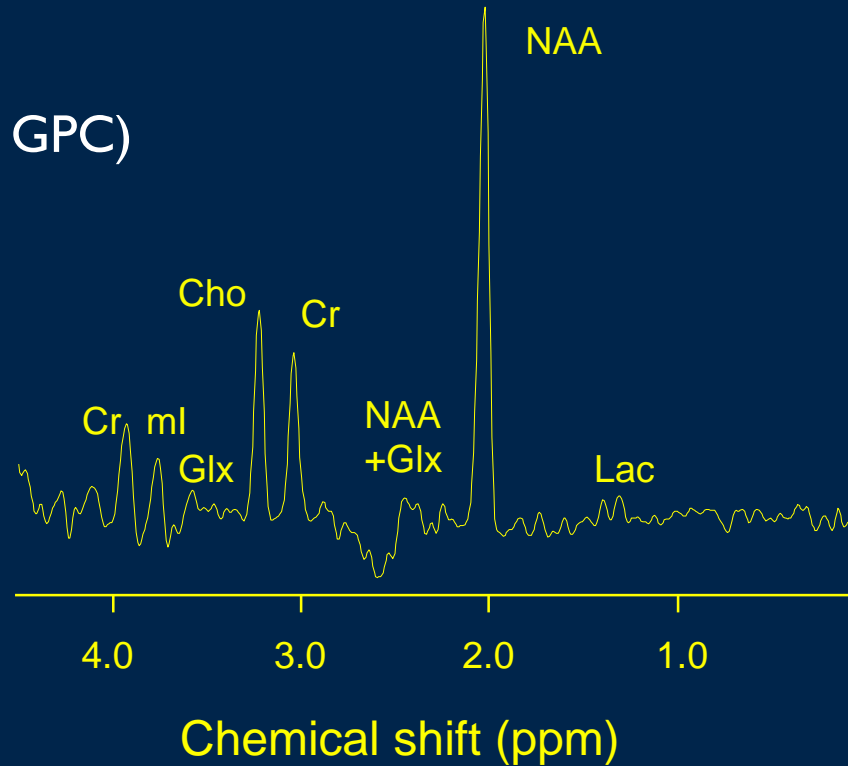
- CHESS (Chemical Shift Selective excitation)
- WET (Water Suppression enhanced through  $T_1$  effects)

# Outer Volume Suppression (OVS)



# Brain Metabolites in $^1\text{H}$ MR Spectroscopy

NAA: N-acetylaspartate  
Cr: creatine + phosphocreatine  
Cho: choline-containing compounds (PCh, GPC)  
Glx: glutamate + glutamine  
ml: myo-inositol  
Lac: lactate



# Literature and Ressources

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- Robin A. de Graaf:  
„In Vivo NMR Spectroscopy: Principles and Techniques“
- James Keeler:  
„Understanding NMR Spectroscopy“
- Malcolm H. Levitt:  
„Spin Dynamics: Basics of Nuclear Magnetic Resonance“
- Jeffrey C. Hoch and Alan S. Stern:  
„NMR Data Processing“

*Toolbox for MRS processing: FID-A*

<https://www.opensourceimaging.org/project/fid-a-advanced-processing-and-simulation-of-mr-spectroscopy/>